



Identification of Distinct Unmutated Chronic Lymphocytic Leukemia Subsets in Mice Based on Their T Cell Dependency

Simar Pal Singh^{1,2,3}, Marjolein J. W. de Bruijn¹, Mariana P. de Almeida¹, Ruud W. J. Meijers², Lars Nitschke⁴, Anton W. Langerak², Saravanan Y. Pillai^{1†}, Ralph Stadhouders^{1,5} and Rudi W. Hendriks^{1*}

OPEN ACCESS

Edited by:

Martin Turner,
Babraham Institute (BBSRC),
United Kingdom

Reviewed by:

Claudia Ribeiro De Almeida,
University of Oxford, United Kingdom
Ingo Ringshausen,
University of Cambridge,
United Kingdom

*Correspondence:

Rudi W. Hendriks
r.hendriks@erasmusmc.nl

†Present Address:

Saravanan Y. Pillai,
EpiExpressions, Rotterdam,
Netherlands

Specialty section:

This article was submitted to
B Cell Biology,
a section of the journal
Frontiers in Immunology

Received: 25 May 2018

Accepted: 14 August 2018

Published: 13 September 2018

Citation:

Pal Singh S, de Bruijn MJW,
de Almeida MP, Meijers RWJ,
Nitschke L, Langerak AW, Pillai SY,
Stadhouders R and Hendriks RW
(2018) Identification of Distinct
Unmutated Chronic Lymphocytic
Leukemia Subsets in Mice Based on
Their T Cell Dependency.
Front. Immunol. 9:1996.
doi: 10.3389/fimmu.2018.01996

¹ Department of Pulmonary Medicine, Erasmus MC, Rotterdam, Netherlands, ² Department of Immunology, Erasmus MC, Rotterdam, Netherlands, ³ Post-graduate School Molecular Medicine, Erasmus MC, Rotterdam, Netherlands, ⁴ Department of Genetics, University of Erlangen, Erlangen, Germany, ⁵ Department of Cell Biology, Erasmus MC, Rotterdam, Netherlands

Chronic lymphocytic leukemia (CLL) can be divided into prognostically distinct subsets with stereotyped or non-stereotyped, mutated or unmutated B cell receptors (BCRs). Individual subsets vary in antigen specificity and origin, but the impact of antigenic pressure on the CLL BCR repertoire remains unknown. Here, we employed *IgH.TE μ* mice that spontaneously develop CLL, expressing mostly unmutated BCRs of which ~35% harbor V_H11-2/V_k14-126 and recognize phosphatidylcholine. Proportions of V_H11/V_k14-expressing CLL were increased in the absence of functional germinal centers in *IgH.TE μ* mice deficient for CD40L or activation-induced cytidine deaminase. Conversely, *in vivo* T cell-dependent immunization decreased the proportions of V_H11/V_k14-expressing CLL. Furthermore, CLL onset was accelerated by enhanced BCR signaling in *Siglec-G^{-/-}* mice or in mice expressing constitutively active Bruton's tyrosine kinase. Transcriptional profiling revealed that V_H11 and non-V_H11 CLL differed in the upregulation of specific pathways implicated in cell signaling and metabolism. Interestingly, principal component analyses using the 148 differentially expressed genes revealed that V_H11 and non-V_H11 CLL clustered with BCR-stimulated and anti-CD40-stimulated B cells, respectively. We identified an expression signature consisting of 13 genes that were differentially expressed in a larger panel of T cell-dependent non-V_H11 CLL compared with T cell-independent V_H11/V_k14 or mutated *IgH.TE μ* CLL. Parallel differences in the expression of these 13 signature genes were observed between heterogeneous and stereotypic human unmutated CLL. Our findings provide evidence for two distinct unmutated CLL subsets with a specific transcriptional signature: one is T cell-independent and B-1 cell-derived while the other arises upon antigen stimulation in the context of T-cell help.

Keywords: B cell receptors, T cell help, BCR signaling, Bruton's tyrosine kinase, chronic lymphocytic leukemia

INTRODUCTION

Chronic lymphocytic leukemia (CLL) is the most common adult leukemia characterized by an accumulation of monoclonal CD5⁺ mature B cells with low surface immunoglobulin (Ig) expression in peripheral blood (1).

CLL is a clinically and molecularly heterogeneous disease whereby progression is influenced by many factors. One-third of patients can be classified as stereotypic CLL, in which BCRs are highly similar between patients (2). The remaining two-third of CLL either lack or have limited similarity with stereotyped CLL BCRs. This classification provides strong molecular evidence for antigen selection in CLL pathogenesis (2). CLL can also be grouped based on IGHV mutational status (3, 4). Significant (>2%) somatic hypermutation (SHM) is observed in patients with mutated CLL (M-CLL), who often develop indolent disease. SHM is absent in unmutated CLL (U-CLL) which evolves rapidly and has a less favorable prognosis (4). The SHM status provides a robust and stable prognostic marker, independently of clinical stage and other markers (5). Furthermore, it reinforces the role of selection by self-antigens or exogenous antigens in CLL pathogenesis. CLL cells show constitutive activation of several BCR downstream kinases, increasing leukemic cell survival *in vitro* (6). In support, small molecule inhibitors of BCR-associated kinases including Bruton's tyrosine kinase (Btk) have shown impressive clinical anti-tumor activity (7, 8).

Few external antigens that potentially drive CLL *in vivo* have been identified; CLL cells were shown to display antigen-independent, cell-autonomous signaling mediated by auto-recognition (9). Several reports have shown that U-CLL express polyreactive BCRs that bind with low affinity to various auto-antigens generated during apoptosis or oxidation (10, 11). In this respect, they resemble natural antibodies secreted by B-1 cells in mice. B-1 cells are a self-renewing CD5⁺ B cell population with remarkably restricted IGHV gene usage and low or no SHM (12). B-1 cells are thought to be generated based on positive selection, by virtue of their receptor specificities to self-antigens, independent of T-cell help (12). Adding to this complexity, the antigen specificity of U-CLL includes both T cell-independent (TI) and T cell-dependent (TD) antigens (11, 13, 14). On the other hand, M-CLL express BCRs that are believed to bind with high-affinity to auto-antigens and show activation of pathways associated with anergic B cells (15, 16).

Differences regarding BCR reactivity have fueled several theories concerning the cellular origins of CLL. SHM status and transcription profiling indicated that U-CLL and M-CLL are derived from CD5⁺CD27⁻ pre- and CD5⁺CD27⁺ post-germinal center (GC) B cells, respectively (17, 18). Extrafollicular or marginal zone (MZ) B cell responses, involving the activation of low-affinity B cells to TI antigens with low SHM, could also be relevant for CLL (19). Direct *in vivo* evidence for the TD or TI origin of CLL subgroups is still missing, mainly due to a lack of mouse models that spontaneously develop both stereotypic and non-stereotypic, mutated and unmutated CLL (20). In the widely studied *Eμ-TCL1* model, CLL predominantly express unmutated stereotyped *IghV11* or *IghV12* BCRs (21). The *IgH.TEμ* CLL mouse model that we previously generated is

based on sporadic expression of the SV40 large T oncogene in mature B cells (22). This was achieved by SV40 large T insertion in opposite transcriptional orientation into the *IgH* locus D_H-J_H region. In contrast to the *Eμ-TCL1* model, *IgH.TEμ* mice mainly develop unmutated CLL with a diverse *IghV* repertoire, and at low frequencies mutated CLL (20, 22). Because of their mixed sv129xC57BL/6 background, we used IgMa/IgMb allotype expression to define CLL incidence by the accumulation of >70% IgMb⁺ B-cells (22, 23). Aging *IgH.TEμ* mice show accumulation of monoclonal CLL-like CD5⁺CD43⁺IgM⁺IgD^{low}CD19⁺ B cells around nine months of age. Although constitutive Btk signaling was not apparent in primary *IgH.TEμ* CLL cells, CLL development was dependent on Btk. Btk-mediated signaling enhanced leukemogenesis and Btk-deficiency led to a complete rescue from the disease (23). Moreover, primary CLL cells from *IgH.TEμ* mice or stable cell lines generated from these mice had detectable expression of p-Akt and substantial levels of p-S6, both of which function downstream of the BCR (23, 24).

To address the impact of antigenic pressure on BCR selection in CLL, we analyzed the effects of defective T cell help and GC formation, as well as robust antigenic stimulation on CLL development in *IgH.TEμ* mice. We show that there are two distinct unmutated CLL subsets present in the *IgH.TEμ* mouse model. The V_H11-2/V_κ14-126-expressing CLL developed independently of T-cell help. Conversely, non-V_H11 CLL was TD and displayed a specific transcriptional signature associated with non-stereotypic U-CLL in human. These findings provide evidence for differential dependence on T cell help in unmutated CLL in mice and suggest that development of human U-CLL can also be T cell-dependent.

MATERIALS AND METHODS

Mice

Mice (C57BL/6) deficient for *Cd40l* (25), *Aicda* (26) or *Siglec-G* (27), and *Cd19-E-Btk-2* (28) transgenic mice were crossed to *IgH.TEμ* mice (F1 sv129xC57BL/6). CLL development was monitored every 3–6 weeks by screening peripheral blood for a monoclonal B cell expansion using flow cytometry. CLL formation was defined by accumulation of >70% IgMb⁺ B-cells in the peripheral blood of the mice. Mice were sacrificed after detection of CLL. Mice were bred and kept in the Erasmus MC experimental animal facility and experiments were approved by the Erasmus MC committee of animal experiments.

Patients and Healthy Controls

Primary patient material was obtained from peripheral blood from CLL patients, while peripheral blood from healthy controls (>50 years of age) was obtained via Erasmus MC and via Sanquin blood bank (Rotterdam). Diagnostic and control samples were collected upon informed consent and anonymized for further use, following the guidelines of the Institutional Review Board, and in accordance with the declaration of Helsinki. The BCR characteristics of all CLL patients are included in **Supplementary Table 5**. Peripheral blood mononuclear cells (PBMCs) were isolated using Ficoll Hypaque (GE Healthcare, Little Chalfont, UK) according to the manufacturer's instructions.

Naïve mature B cells were isolated from healthy control PBMCs using FACS-purification for CD19+CD27-IgD+ cells. The purity of naïve mature healthy B cell samples was >95% as determined by flow cytometry.

In vivo Immunizations

TD immune responses were induced by i.p. immunization. Primary immunizations were induced in 10-12-week-old mice with 100 µg TNP-KLH on alum. After 5 weeks this was followed by a secondary immunization with 100 µg TNP-KLH in PBS (28).

BCR Sequencing

Primer sequences and PCR condition were previously described (22, 23). PCR products were directly sequenced using the BigDye terminator cycle sequencing kit with AmpliTaq DNA polymerase on an ABI 3130xl automated sequencer (Applied Biosystems). Sequences were analyzed using IMG/Quest (<http://www.imgt.org>), using Ig gene nomenclature as provided by IMG/Quest. All sequences were confirmed in at least one duplicate analysis.

Flow Cytometry Procedure

Preparation of single-cell suspensions of lymphoid organs and lysis of red blood cells were performed according to standard procedures. Cells were (in)directly stained in flow cytometry buffer (PBS, supplemented with 0.25% BSA, 0.5 mM EDTA and 0.05% sodium azide) using the following fluorochrome or biotin-conjugated monoclonal antibodies or reagents: anti-B220 (RA3-6B2), anti-CD19 (ID3), anti-CD5 (53-7.3), anti-CD43 (R2/60), anti-CD23 (B3B4) all from eBioscience and anti-CD138 (281-2), anti-CD95 (Jo2), anti-IgD (11-26), anti-IgM (AF6-78), anti-IgMa (DS-1), anti-Igλ (R26-46), anti-Igκ (187.1), anti-CD21 (7G6), all from BD Biosciences, using conjugated streptavidin (eBioscience) as a second step for biotin-conjugated antibodies.

Leukemic cells (CD19⁺CD5⁺) were stained with fluorescein-labeled phosphatidylcholine (PtC) liposomes (DOPC/CHOL 55:45, Formumax Scientific Inc.) in flow cytometry buffer. Cells were co-stained with anti-CD19, anti-CD43, or anti-CD5 (BD Biosciences).

MACS Cell Sorting

Splenic single-cell suspensions were prepared in magnetic-activated cell sorting (MACS) buffer (PBS/2mM EDTA/0.5%BSA) and naïve splenic B cells from 8–12 week-old WT C57BL/6 mice were purified by MACS, as previously described (24, 29). Non-B cells, B-1 cells, GC B cells, and plasma cells were first labeled with biotinylated antibodies (BD Biosciences) to CD5 (53–7.3), CD11b (M1-70), CD43 (S7), CD95 (Jo2), CD138 (281-2), Gr-1 (RB6-8C5), and TER-119 (PK136) and subsequently with streptavidin-conjugated magnetic beads (Miltenyi Biotec). Purity of MACS-sorted naïve B cells was confirmed by flow cytometry (typically > 99% CD19+ cells). To obtain activated B cells, purified naïve WT B cells were cultured in culture medium [RPMI 1640 (life technologies)/10% FCS (gibco)/50 µg/mL gentamycin(life technologies)/0,05 mM β-mercaptoethanol (Sigma)] in the presence of 10 µg/ml F(ab')₂ anti-IgM (Jackson Immunoresearch) for 12 h.

RNA-Sequencing

RNA was extracted from naïve or activated splenic B cells, as well as from purified (using MACS-purification for CD19+ cells) primary tumors from IgH.TEµ mice with the RNeasy Micro kit (Qiagen) according to manufacturer's instructions. The TruSeq RNA Library Prep kit (Illumina) was used to construct mRNA sequencing libraries that were sequenced on an Illumina HiSeq 2500 (single read mode, 36 bp read length). Raw reads were aligned using Bowtie to murine transcripts (RefSeq database) from the University of California at Santa Cruz (UCSC) mouse genome annotation (NCBI37/mm9) (30). Differential gene expression analysis was performed using DESeq2 (31) with an adjusted *P*-value (false discovery rate; FDR) of *P* < 0.05. Log₂-fold changes and FDR values as calculated by DESeq2 were used to generate a volcano plot using R (R studio version 1.1.383). Normalized gene expression levels quantified as reads per kilobase of a transcript per million mapped reads (RPKM) were used for various clustering approaches (unsupervised hierarchical clustering, supervised clustering, and PCA) that were performed using R and PAST software (<https://folk.uio.no/ohammer/past/>). Visualization of clustering analysis output was performed using R, PAST, and Java TreeView (32). Molecular pathway enrichments were obtained from the online MSigDB database. Gene expression data for anti-CD40 plus IL-4 stimulated follicular B-cells was obtained from previously reported data and downloaded from the Gene Expression Omnibus (GEO; accession number GSE77744) (33). RNA-Seq data generated in this study have been deposited in the GEO database (accession number GSE117713).

Quantitative Real Time PCR Analysis

Samples tested in qRT-PCR were from IgH.TEµ (7 V_H11 and 15 non-V_H11), from IgH.TEµ.Aicda^{-/-} (4 V_H11 and 4 non-V_H11), and from IgH.TEµ.TD (4 non-V_H11) mouse groups. For quantitative RT-PCR analysis, TaqMan probes were employed. Probe Finder software (Roche Applied Science), the Universal Probe Library (Roche Applied Science) and Ensembl genome browser (<http://www.ensembl.org/>) were used for primer and probe design. Taqman Universal Master Mix II, was purchased from Thermo Fisher Scientific. Quantitative RT-PCR was performed by using the 7300 Real Time PCR system (Applied Biosciences) according to manufacturer's instructions. Gene expression was analyzed with an ABI Prism 7300 Sequence Detector and ABI Prism Sequence Detection Software version 1.4 (Applied Biosystems). Cycle-threshold levels were calculated for each gene and the housekeeping gene glyceraldehyde-3-phosphate dehydrogenase (*Gapdh*) was used for normalization of the values. All primer sequences and probe numbers are listed in **Supplementary Table 7**.

Statistical Analysis

Statistical analysis was performed using GraphPad Prism software (San Diego, California, USA) or R. The log rank test was used for calculating the level of significance for survival differences between mouse groups. The Chi-square test was used to determine the significance for BCR usage differences between different mouse groups. To evaluate differences in expression

levels of different genes by qRT-PCR we used a Mann-Whitney *U*-test between two groups or a Kruskal-Wallis test corrected with Dunn's multiple comparison test for more than two groups.

RESULTS

Two Subsets of Unmutated CLL Arise in *IgH.TE μ* Mice

To analyze the BCR repertoire, we aged a panel of *IgH.TE μ* mice and collected blood every 3–6 weeks to monitor CLL incidence. Hereby, CLL incidence was defined by the accumulation of >70% IgMb⁺ B-cells, which displayed a CLL-like CD5⁺CD43⁺IgM⁺IgD^{low}CD19⁺ phenotype (22, 23). We performed sequencing analyses of Ig heavy (*Igh*) and light (*Igl*) chain transcripts and found that a substantial proportion (~36%) of CLL in *IgH.TE μ* mice expressed stereotyped BCRs consisting of the V_H11-2 *Igh* chain, with similar *Igh* CDR3 length and amino acid sequences, and the V_K14-126 *Igl* chain (22, 23) (Supplementary Table 1, Figures 1A,B). The V_H11/V_K14 CLL mice exhibited an earlier disease onset compared with *IgH.TE μ* mice with non-stereotypic (non-V_H11) BCR (mean incidence age 184 days and 219 days, respectively, $p = 0.0175$) (Figure 1C). In wild-type mice the V_H11-2/V_K14-126 BCR is preferentially expressed by B-1 lymphocytes and shows specificity to phosphatidylcholine (PtC) (12). We could confirm PtC-binding specificity of V_H11-2 BCRs on CLL cells (Figure 1D). V_H11 CLL showed decreased surface IgM expression and increased surface IgD expression compared to non-V_H11 CLL (Figures 1E,F). A major proportion (~65%) of the remaining non-V_H11 CLL expressed a J558 V_H1-family BCR with heterogeneous CDR3 length, amino acid sequence and *Igl* chain usage (Supplementary Table 1). V_H1 CLL showed delayed disease onset (mean incidence age 231 days), compared with V_H11 CLL (Supplementary Figure 1).

In conclusion, based on *Ig* gene usage we could distinguish different subsets of unmutated *IgH.TE μ* CLL displaying differential disease onset.

Germinal Center Defects Lead to Increased V_H11/V_K14 Usage in Unmutated CLL

Because V_H11/V_K14-expressing CLL likely originate from B-1 cells, we hypothesized that they should still develop in the absence of functional GCs. Therefore, we investigated their dependence on functional GCs and T cell help by crossing *IgH.TE μ* mice with *Cd40l*^{-/-} or *Aicda*^{-/-} mice. *Cd40l*^{-/-} or *Aicda*^{-/-} mice display a complete lack or aberrant enlargement of GCs, respectively, paralleling the human hyper-IgM syndrome phenotype (26, 34). We monitored CLL incidence, as described above, in cohorts of *Cd40l*-deficient *IgH.TE μ* mice (*IgH.TE μ .Cd40l*^{-/-}, $n = 13$), *Aicda*-deficient *IgH.TE μ* mice (*IgH.TE μ .Aicda*^{-/-}, $n = 26$) and *IgH.TE μ* control littermates, $n = 69$ or $n = 56$, respectively for ~400 days (Figure 2). CLL frequency and onset was not altered in *IgH.TE μ .Cd40l*^{-/-} mice (~59%, compared with ~62% in *IgH.TE μ* control littermates;

$p = 0.99$) or in *IgH.TE μ .Aicda*^{-/-} mice (~62%, compared with ~64% in *IgH.TE μ* control littermates; $p = 0.78$) (Figures 2A,B).

To explore the impact of CD40L or AID-deficiency on BCR usage in CLL, we performed *IghV* and *IglV* sequence analyses in selected CLL samples with high tumor load (>95% IgMb⁺CD5⁺CD43⁺CD19⁺ CLL-like cells) (Supplementary Table 1). Interestingly, usage of the stereotypic V_H11/V_K14 BCR was significantly increased in CLL from *IgH.TE μ .Cd40l*^{-/-} mice ($n = 7/7$, 100%), compared with control *IgH.TE μ* mice ($n = 9/25$, ~36%, Chi-square $p < 0.001$). Also CLL from *IgH.TE μ .Aicda*^{-/-} mice showed increased V_H11/V_K14 usage ($n = 9/13$, ~69%) compared with control littermates ($n = 7/20$, ~35%, Chi-square $p < 0.01$) (Figures 2C,D). These V_H11 CLL also expressed similar *Igh* CDR3 sequences (Supplementary Table 1).

Taken together, these findings indicate that V_H11/V_K14-expressing CLL arise independently of T cell help or GC formation, whereas non-V_H11 CLL is T cell-dependent and reduced in the absence of functional GCs in *IgH.TE μ .Cd40l*^{-/-} and *IgH.TE μ .Aicda*^{-/-} mice.

T-cell Dependent Antigenic Stimulation of B Cells *in vivo* Reduces V_H11/V_K14 Usage in Unmutated *IgH.TE μ* CLL

To directly investigate whether antigenic stimulation in the context of T cell help affects CLL onset and the CLL BCR repertoire, we immunized *IgH.TE μ* mice with TNP-KLH coupled to alum (*IgH.TE μ .TD*, $n = 20$) to induce a TD B cell response. CLL onset did not differ between immunized and non-immunized littermates ($n = 56$) (Figure 3A). At the age of ~400 days, CLL incidence in *IgH.TE μ .TD* mice was ~65% similar to non-immunized control *IgH.TE μ* mice (~62%) (Figure 3A).

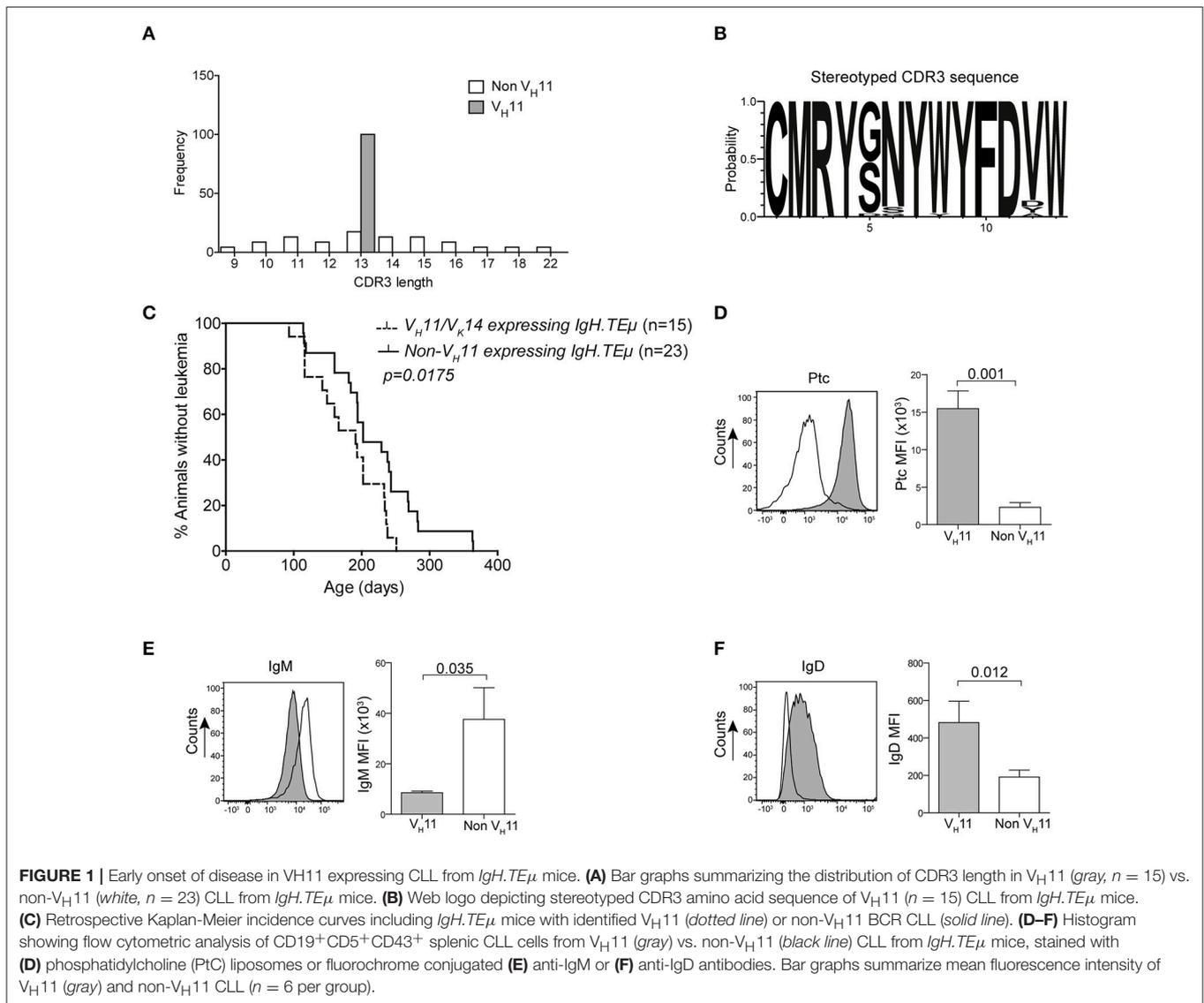
Next, we analyzed *IghV* and *IglV* sequences in CLL samples with high tumor load (>95% IgMb⁺CD5⁺CD43⁺CD19⁺ CLL-like cells). In contrast to control *IgH.TE μ* mice, which showed ~35% ($n = 7/20$) V_H11 usage, only 10% ($n = 1/10$) of *IgH.TE μ .TD* CLL expressed a V_H11/V_K14 BCR (Chi-square $p = 0.09$) (Figure 3B). The majority ($n = 6/9$, 67%) of CLL in *IgH.TE μ .TD* mice expressed a J558/V_H1-family *IghV* gene and we did not observe mutated CLL (Supplementary Table 1).

In summary, we found that robust TD immunization favors development of non-V_H11 CLL.

Enhanced BCR Signaling Accelerates Disease Onset in *IgH.TE μ* Mice

Our findings provide evidence that T cell-derived activation or selection signals, in particular CD40L, shape the BCR repertoire of CLL in *IgH.TE μ* mice, but do not significantly affect disease onset or progression. It is therefore conceivable that in the *IgH.TE μ* mouse model, BCR-derived signals may be more decisive for disease progression.

To monitor the impact of BCR signaling strength on CLL development and *IghV* gene selection, we first crossed *IgH.TE μ* mice with *E-Btk-2* transgenic mice. These mice express the constitutive active E41K-BTK mutant selectively in the B-cell



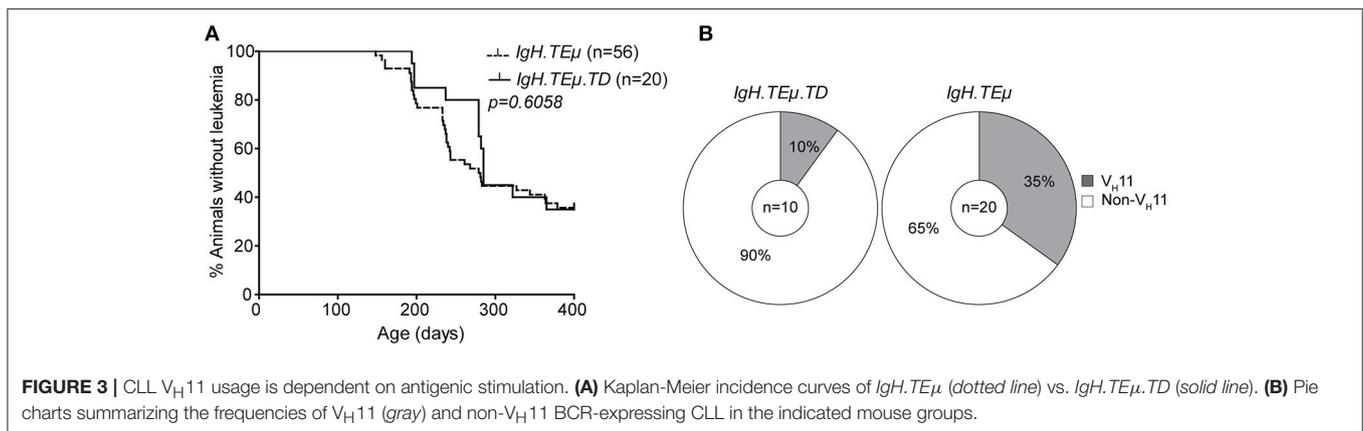
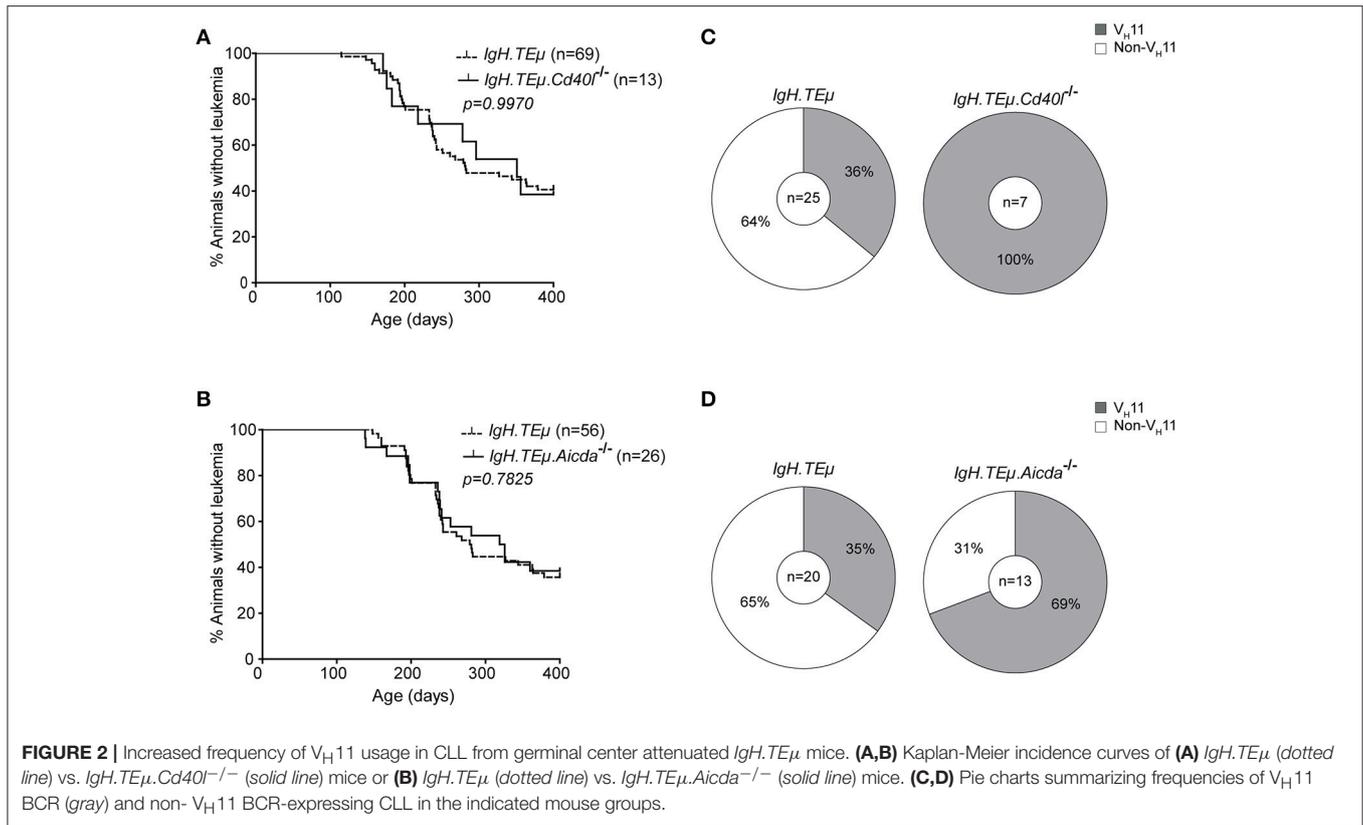
lineage driven by the CD19 promoter (28). The E41K mutation enhances Btk membrane localization and thereby its activation by Syk or Src-family tyrosine kinases (35). *E-Btk-2* mice show defective follicular B cell survival and a relative expansion of splenic B-1 cells (28). Flow cytometry analysis of *E-Btk-2* B-1 cells did not reveal detectable PtC binding, indicating that V_H11 BCR expression was limited (data not shown).

We found that *IgH.TEμ.E-Btk-2* mice ($n = 21$) developed CLL significantly earlier (mean age of onset of ~ 155 days), compared with control *IgH.TEμ* mice (~ 279 days; $p < 0.0001$) (Figure 4A). In addition, *IgH.TEμ.E-Btk-2* mice appeared to have an increased disease frequency ($\sim 90\%$ at ~ 400 days, compared with $\sim 71\%$ for control *IgH.TEμ* mice). Sequence analysis of *Igh* revealed that 1 out of 8 ($\sim 12\%$) tumors from *IgH.TEμ.E-Btk-2* mice expressed a V_H11 BCR, compared with 35% ($n = 9/26$) in the control *IgH.TEμ* group (Figure 4C, Supplementary Table 1). This difference was not statistically significant, but the finding of

a PtC-reactive V_H11 CLL was surprising, since PtC-binding B-1 cells were not detectable in *E-Btk-2* mice. The majority ($\sim 71\%$; $n = 5/7$) of the non- V_H11 BCRs expressed J558/ V_H1 -family *IghV* genes.

To confirm that enhanced BCR signaling affects disease onset, we crossed *IgH.TEμ* mice on a *Siglec-G* deficient background (*IgH.TEμ.Siglec-G^{-/-}*). *Siglec-G* is a negative regulator of BCR-mediated signaling that is expressed in all B cells (27). It is a potent inhibitor of BCR-induced Ca^{2+} signaling and a key regulator of survival and selection of B-1 cells (36). In addition, *Siglec-G*-deficiency abrogates V_H11 usage in B-1 cells (36).

Similar to *IgH.TEμ.E-Btk-2* mice, also *IgH.TEμ.Siglec-G^{-/-}* mice displayed an increased disease frequency ($\sim 93\%$ at ~ 400 days, compared with $\sim 70\%$ for *IgH.TEμ* mice), with significantly accelerated CLL onset (~ 121 days compared with ~ 268 days for *IgH.TEμ* mice; $p < 0.0001$) (Figure 4B). *IghV* analyses showed a $V_H11/V_{\kappa}14$ usage of $\sim 18\%$ ($n = 2/11$) in *IgH.TEμ.Siglec-G^{-/-}*

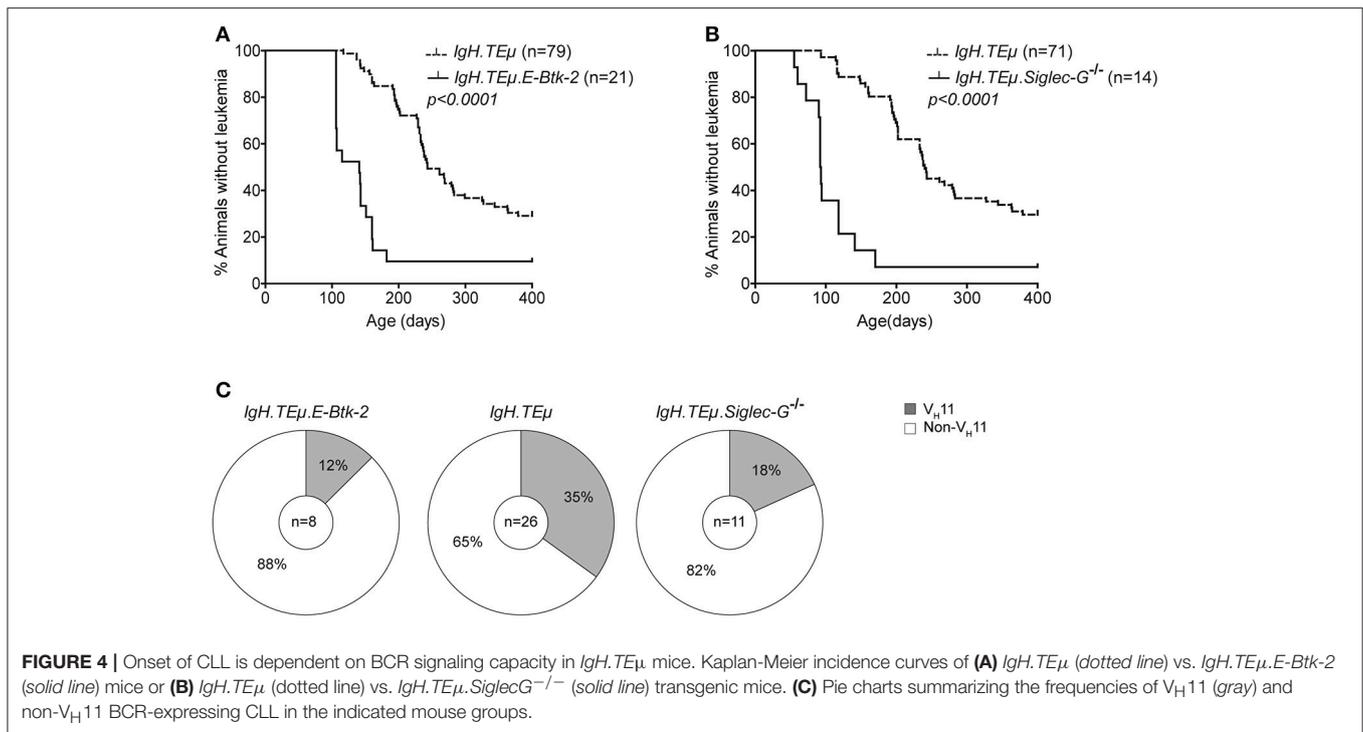


CLL vs. ~35% (9/26) in the control group (Chi-square $p < 0.242$) (Figure 4C, Supplementary Table 1). Only 2/11 $IgH.TE\mu.Siglec-G^{-/-}$ CLL expressed J558/ V_H11 -family $IghV$ genes.

Thus, BCR signaling strength plays an important role in CLL development in $IgH.TE\mu$ mice, whereby enhanced signaling accelerates disease onset. Because $E-Btk-2$ or $Siglec-G^{-/-}$ B-1 cells do not show detectable PtC expression, our findings suggest that those few V_H11 B cells present are efficiently transformed to CLL in $IgH.TE\mu.E-Btk-2$ or $IgH.TE\mu.Siglec-G^{-/-}$ mice. Thus, BCR signaling strength may also affect the BCR repertoire in CLL.

Transcriptome Profiling Identifies Unique Genes and Pathway Aberrations for $V_H11/V\kappa14$ and non- V_H11 CLL $IgH.TE\mu$ Mice

To further explore the biological phenotype of the V_H11 and non- V_H11 CLL subsets, we performed genome-wide gene expression profiling on primary $IgH.TE\mu$ CLL (tumor load >95%) expressing either a V_H11 ($n = 3$) or a non- V_H11 ($n = 3$) BCR. As a reference we included resting unstimulated (un-B, $n = 4$) and anti-IgM stimulated (α IgM-B, $n = 4$) naïve splenic B cells from wild-type mice. Normalized gene



expression values (see Methods for details) were used for principle component analysis (PCA). The first two principal components, which represented $\sim 70\%$ of the total variation among the different samples analyzed, identified three separate clusters, corresponding to un-B, α IgM-B and primary *IgH.TE μ* CLL samples, indicating a strong correlation between biological replicates (**Supplementary Figure 2**).

When we performed differential gene expression analysis (focusing only on genes passing a stringent statistical filter of Benjamini-Hochberg false discovery rate corrected $P < 0.05$), we found 148 differentially expressed genes (**Figure 5A; Supplementary Table 2**). Of these genes, 59 genes were upregulated in V_H11 CLL and 89 genes were upregulated in non-V_H11 CLL. To identify biological processes that underlie the transcriptional differences between V_H11 and non-V_H11 CLL, we performed pathway enrichment analysis using the Molecular Signatures Database (MSigDB) (37). Genes upregulated in V_H11 CLL were functionally enriched for an interferon-mediated response, active Wnt signaling and constitutively active RAF1 signaling (**Figure 5B, Supplementary Table 3A**). On the other hand, genes downregulated in V_H11 CLL were involved in quite diverse pathways, including interleukin-, epidermal growth factor receptor (EGFR)-, vascular endothelial growth factor (VEGF)-mediated signaling, metabolic processes, hypoxia and the UV radiation-induced stress response (**Figure 5B, Supplementary Table 3A**).

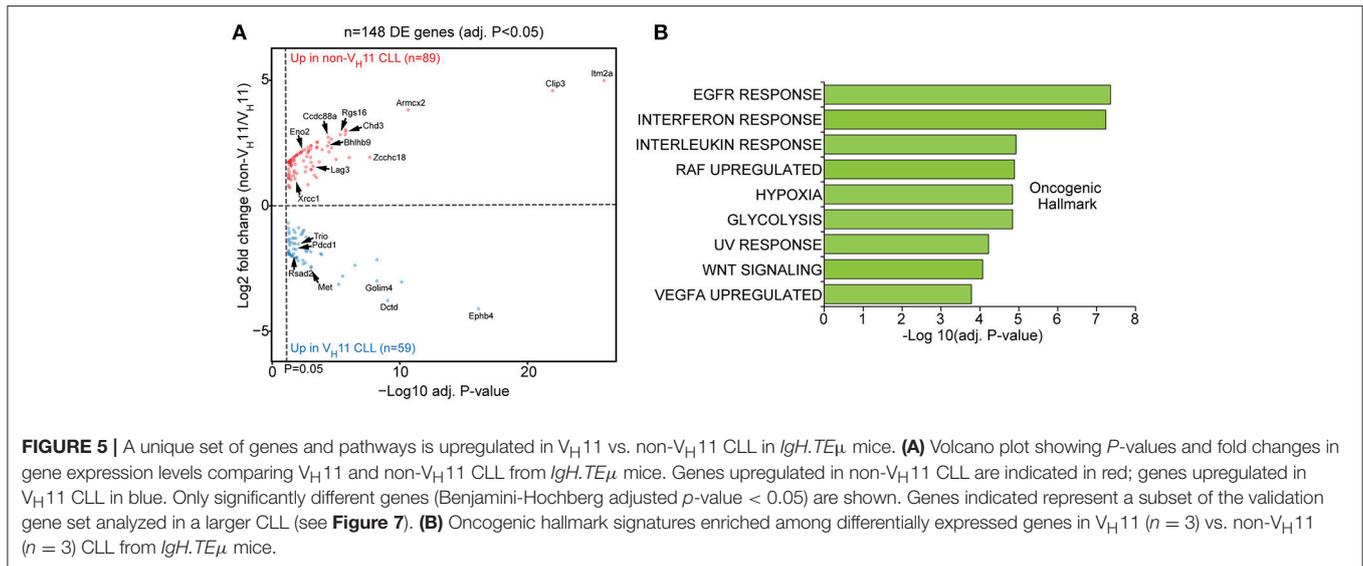
Taken together, these data suggest that in addition to a different origin, V_H11 and non-V_H11 CLL subsets display distinct transcriptional signatures, signifying differential activity of key signaling pathways.

Strong BCR Dependence of V_H11/V_K14 CLL in *IgH.TE μ* Mice

Next, we performed a PCA of the 148 differentially expressed genes between V_H11 and non-V_H11 CLL. To investigate the impact of T-cell-independent BCR stimulation and T-cell-dependent CD40 stimulation on differential gene expression, we included RNA-Seq gene expression values of the 148 genes from the unstimulated and α IgM-stimulated B cells described above, as well as previously reported gene expression values from anti-CD40/IL-4 stimulated follicular B-cells (α -CD40/IL4-B) (33).

The first principal component (PC1) separated both CLL groups and the two stimulated B cell subsets from unstimulated B cells, suggesting *IgH.TE μ* CLL cells share a transcriptional signature related to activated B-cell phenotypes. Interestingly, PC2 revealed a strong similarity between α IgM-stimulated B cells and V_H11 CLL on one hand and between α -CD40/IL4-stimulated B cells and non-V_H11 CLL on the other hand (**Figure 6A**). These findings indicate more prominent BCR stimulation in V_H11 than in non-V_H11 CLL B cells *in vivo* and are consistent with a dependence on T-cell help for non-V_H11 CLL.

To identify the gene signature underlying the clustering of α IgM-stimulated B cells and V_H11 CLL, as well as α -CD40/IL-4-stimulated B cells and non-V_H11 CLL, we performed hierarchical clustering analyses to separate the 148 genes into 4 clusters (**Figure 6B, Supplementary Table 3B**). Cluster 1 consists of 17 genes that were highly correlated between α IgM-stimulated B cells and V_H11 CLL and between α -CD40/IL-4-stimulated B cells and non-V_H11 CLL. Pathway enrichment analysis (**Supplementary Table 3C**) on this cluster revealed overrepresentation of genes involved in interferon response



and KRAS signaling. Clusters 2 (35 genes) and Cluster 3 (56 genes) consist of genes that were highly correlated only between α -CD40/IL-4-stimulated B cells and non- V_H11 CLL or only between α IgM-stimulated B cells and V_H11 CLL, respectively. These clusters were enriched for interferon response/PI3K-AKT signaling genes (cluster 2) or UV response, epithelial-mesenchymal transition, glycolysis, hypoxia, unfolded protein response genes (cluster 3) (**Supplementary Table 3C**). Finally, cluster 4 (enriched for genes involved in the reactive oxygen species pathway) represents genes with low or anti-correlated expression values between the stimulated B cells and CLL. Thus, genes from clusters 1 and 3 signify the clustering of α IgM-stimulated B cells and V_H11 CLL, while genes from clusters 1 and 2 drive the clustering of α -CD40/IL-4-stimulated B cells and non- V_H11 CLL (**Figure 6B**). This analysis was further validated by computing the average correlation strength for each of the four gene clusters with PC2 from our PCA (**Figure 6B**). Indeed, clusters 1 to 3 underlying the α IgM-B cells and V_H11 CLL and the α -CD40/IL-4-B cells and non- V_H11 CLL segregation—and particularly cluster 1 genes—showed significantly stronger correlation values with PC2 than cluster 4 (**Figure 6C**).

Validation of V_H11 /non- V_H11 CLL Gene Expression Differences in Mouse and Human CLL

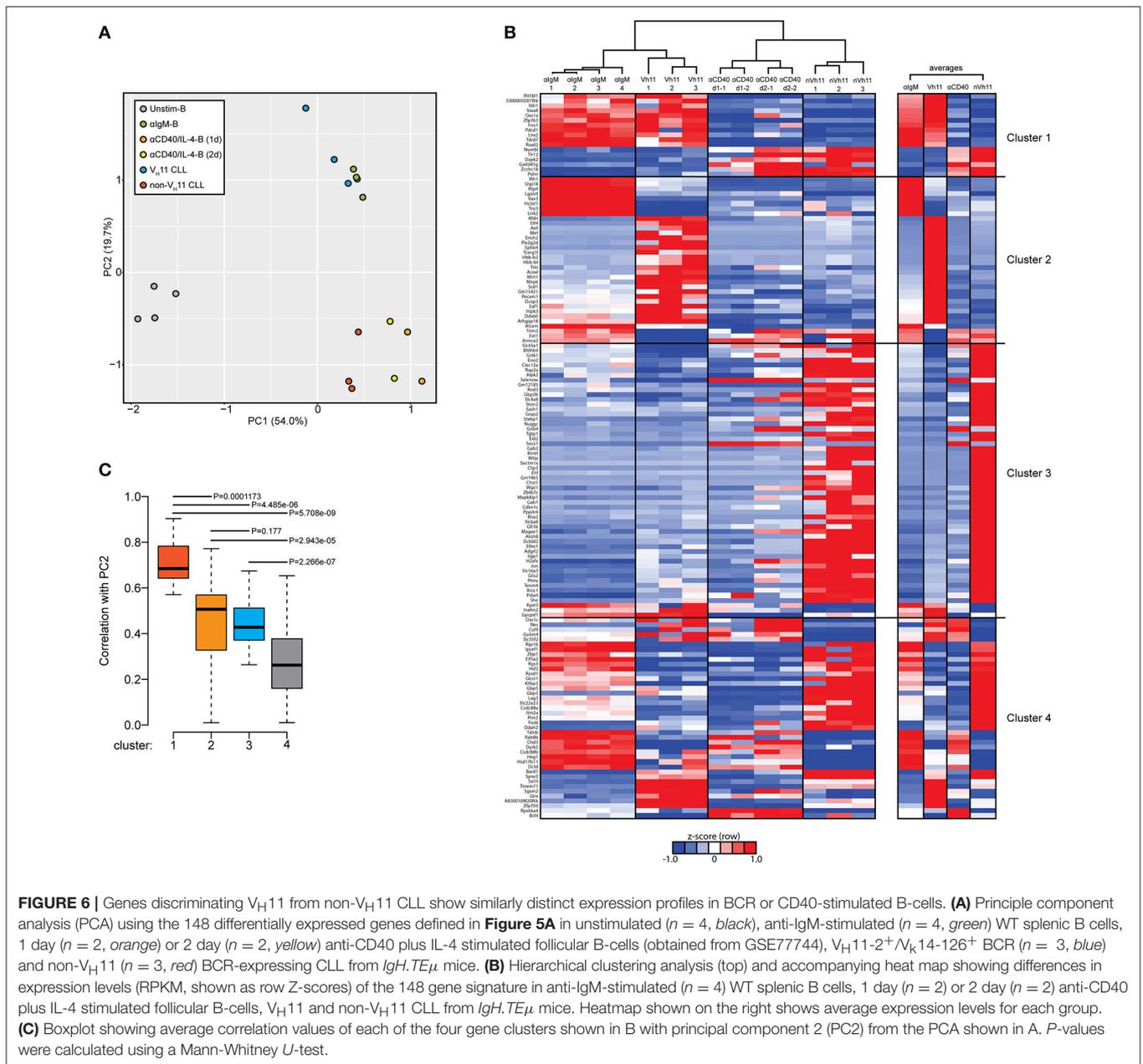
To further strengthen the existence of a unique transcriptional signature that differentiates V_H11 and non- V_H11 CLL B cells, we selected 24 robustly differentially expressed genes for validation. Some of these genes have already been shown to play a role in hematologic malignancies, including CLL (*Pim-2*, *Met*, *Rgs16*, *Ccdc88a*, *Zcchc18*, *Clip3*) (38–42), diffuse large B cell lymphoma, follicular lymphoma (*Vav3*) (43), acute lymphoblastic leukemia (ALL) (*Itm2a*, *Chst1*) (44, 45) or acute myeloid leukemia (AML) (*Chd3*) (46).

Expression levels were validated by quantitative real-time PCR (qRT-PCR) in an extended cohort of 15 V_H11 and 23

non- V_H11 primary CLL from *IgH.TE μ* mice. Naïve Splenic B cells from wild type mice ($n = 4$) were included as controls. Comparison of RNA-Seq (RPKM) and qRT-PCR expression fold changes between the two CLL groups revealed highly correlated trends for these 24 genes (spearman correlation $r = 0.72$; $p < 0.0001$), validating our RNA-Seq analysis when extrapolated to a larger *IgH.TE μ* CLL cohort (**Figure 7A**). qRT-PCR validation showed that 11/24 (~46%) of the selected genes were significantly different ($p < 0.05$) between V_H11 and non- V_H11 CLL (**Supplementary Table 4**, **Figure 7B**). Additionally, 7/24 (~29%) genes were significantly different ($p < 0.05$) between non- V_H11 CLL from *IgH.TE μ* and *IgH.TE μ .Siglec-G $^{-/-}$* mice, which might be related to the altered V_H usage in *Siglec-G $^{-/-}$* mice or the early disease onset in *IgH.TE μ .Siglec-G $^{-/-}$* mice.

Expression of five of these 13 genes that were significantly upregulated in non- V_H11 CLL vs. V_H11 CLL (*Ccdc88a*, *Clip3*, *Zcchc18*, *Chd3*, *Itm2a*) was also evaluated in five mutated *IgH.TE μ* CLL, defined by <97% IghV germline identity [**Supplementary Table 1** and ter Brugge et al. (22)]. Interestingly, qRT-PCR analysis showed that four out of five tested genes (except *Itm2a*) were expressed at low levels in mutated CLL, similar to V_H11 CLL (**Figure 7B**). Thus, non- V_H11 unmutated CLL in *IgH.TE μ* mice represent a unique subset that can be distinguished from V_H11 unmutated and from mutated CLL by a specific transcriptional signature. Furthermore, correlation analyses indicated that within the non-stereotypic subgroup in particular V_H11 CLL represents the most heterogeneous CLL subgroup in *IgH.TE μ* mice ($n = 16$; average spearman r , $\rho = 0.280$; **Supplementary Figure 3**). In these analyses we also found that expression of these five genes is positively correlated in V_H11 CLL ($n = 15$; average spearman r , $\rho = 0.537$) and in the small non- V_H11 /non- V_H11 CLL subgroups ($n = 6$; average spearman r , $\rho = 0.703$) (**Supplementary Figure 3**).

Next, we evaluated the expression of the 13 signature genes in a panel of 44 human CLL samples (15 non-stereotypic U-CLL, 14 stereotypic U-CLL, 15 M-CLL, **Supplementary Table 5**)

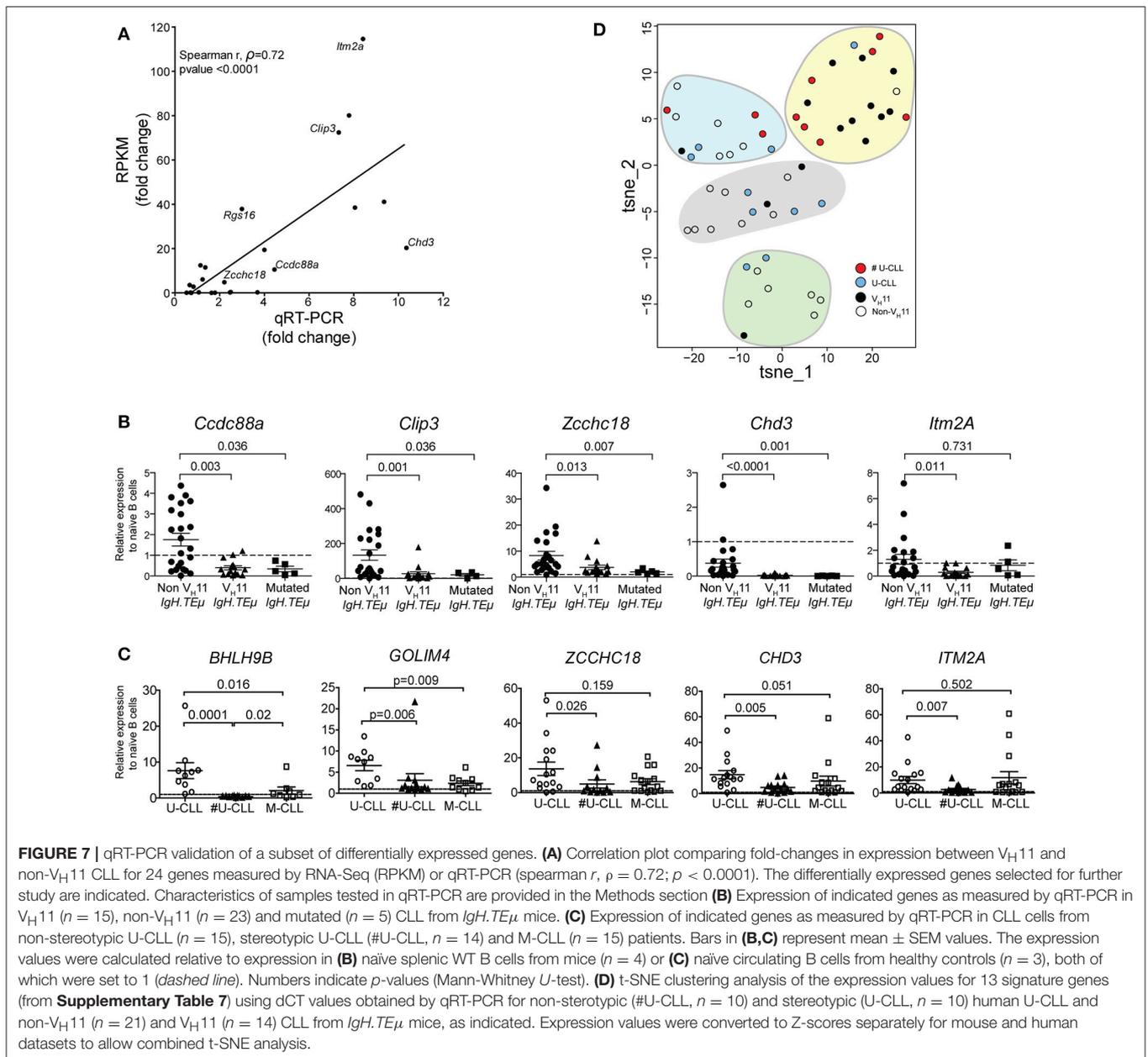


by qRT-PCR. Hereby, 6/13 (~46%) genes (*CCDC88A*, *CLIP3*, *ZCCHC18*, *CHD3*, *ITM2A*, *GOLIM4*) were significantly higher expressed in all three CLL subsets than in naïve B-cells from healthy individuals, suggesting a role for these genes in CLL (**Figure 7C**, **Supplementary Table 6**). Expression of *CLIP3* was significantly higher in non-stereotypic than M-CLL. Expression of *ZCCHC18*, *CHD3*, *GOLIM4*, *BHLH9B*, and *ITM2A* was significantly higher in non-stereotypic U-CLL compared to stereotypic U-CLL (**Figure 7C**).

To compute any parallel between stereotypic and heterogeneous U-CLL from patients and *IgH.TE μ* mice, we performed t-SNE clustering analysis on expression values for the 13 signature genes (**Figure 7D**, **Supplementary Table 6**).

We used dCT values obtained by qRT-PCR for non-stereotypic (#U-CLL, $n = 10$) and stereotypic (U-CLL, $n = 10$) U-CLL as well as for non- V_H11 ($n = 21$) and V_H11 ($n = 14$) CLL from *IgH.TE μ* mice. Interestingly, 7/10 stereotypic U-CLL clustered with 10/14 V_H11 CLL (**Figure 7D**). Conversely, non-stereotypic human U-CLL and mouse non- V_H11 CLL showed a more heterogeneous distribution into several clusters largely devoid of stereotypic human U-CLL or mouse V_H11 CLL.

Taken together, we conclude that differences in the expression of these signature genes in heterogeneous U-CLL, stereotyped U-CLL and M-CLL were partly overlapping between human CLL and the corresponding CLL subgroups in our *IgH.TE μ* CLL mouse model.



DISCUSSION

In this report, we investigated the role of antigenic pressure and BCR signaling thresholds on clonal selection of CLL cells in the *IgH.TE μ* CLL mouse model. We found that U-CLL tumors that develop in these mice can be classified into two different groups based on their *IghV* usage. The stereotypic V_H11 -2/ $V_{\kappa 14}$ -126 CLL subset recognized the PtC self-antigen, developed independently of T cell help or GC formation and represented a somewhat more aggressive type of CLL. Proportions of $V_H11/V_{\kappa 14}$ -expressing CLL were increased in the absence of functional germinal centers in *IgH.TE μ* mice deficient for CD40L or activation-induced cytidine deaminase. Conversely, *in vivo*

T cell-dependent immunization decreased the proportions of $V_H11/V_{\kappa 14}$ -expressing CLL. Mice were immunized at 10–12 weeks of age, with a secondary immunization at 15–17 weeks of age. In a proportion of mice at these time points, CLL cells become detectable in peripheral blood (**Figure 1**). In our immunization model the onset or frequency of CLL was not altered, but we cannot exclude that there will be effects on CLL onset or disease progression when immunizations are performed at a different age.

Consistent with the observed effects of defective germinal center function or robust T-cell dependent immunization on V_H usage in CLL, PCA of a gene signature comprised of 148 genes differentially expressed between V_H11 and

non- V_{H11} CLL revealed that V_{H11} and non- V_{H11} CLL clustered with BCR-stimulated and anti-CD40-stimulated B cells, respectively.

The unmutated V_{H11} CLL cells parallel B-1 cells, because these also have a restricted BCR repertoire, may recognize autoantigens including PtC, and produce natural IgM antibodies in the absence of T cell co-stimulation (12). In concordance, it was recently shown that peritoneal $CD5^+$ B-1 cells generated early during fetal or neonatal development, increase in number over time and can progress into CLL in aged mice (47, 48). Interestingly, CLL development in these mice was linked to the expression of a restricted BCR repertoire ($V_{H}Q52/V_{H}\kappa9$ or $V_{H}3609/V_{H}\kappa21$, reactive toward non-muscle myosin-IIA or Thy-1, respectively) independent of CD40 signaling. Hereby, expression of the $E\mu$ -*TCL1* transgene enhanced aggressiveness of the disease.

Non- V_{H11} CLL, on the other hand, consisted of tumors with heterogeneous *IghV/IglV* expression and CDR3 length, lacking affinity for PtC. Although these tumors were T-cell dependent, strongly reduced in the absence of functional GCs, their BCRs were not hypermutated (<3%). This is in line with findings in human U-CLL, indicating that U-CLL cells can recognize both TD and TI autoantigens that have relocated to the external cell surface during apoptosis (11, 13, 14). Our observations are also consistent with gene expression profiling studies suggesting that U-CLL reflect memory B cells (49). In contrast, more recent transcriptome analyses revealed that U-CLL resemble mature pre-GC $CD5^+CD27^-$ B cells, while M-CLL resembles a distinct, previously unrecognized, $CD5^+CD27^+$ post-GC B cell subset (18). Our findings imply that in mice unmutated CLL can be derived from (i) T cell-independent B-1 cells (e.g., PtC-recognizing V_{H11} -2/ $V_{H}\kappa14$ -126) or (ii) from B cells that recognize their antigen in the presence of cognate T-cell help and are activated without SHM. This latter group of T cell-dependent unmutated CLL displayed an expression signature, as defined by 13 genes including the *CCDC88A-CLIP3-ZCCHC18-CHD3-ITM2A* module, that is not only different from TI unmutated CLL, but also from mutated CLL in the *IgH.TE μ* mouse model. Moreover, we found evidence that this expression signature may be partly associated with non-stereotypic human U-CLL, suggesting that the development of human U-CLL can also be TD. Such TD U-CLL may derive from B cells involved in an extra-follicular response or alternatively may be related to auto-antibody producing B cells in mice that were shown to recognize TD antigens, mount a rapid IgM response and enter GCs, but do not develop into IgG-expressing plasma cells (50, 51). Although our data suggest a role for T-cell help in human non-stereotypic U-CLL pathophysiology, further investigation is required to translate our findings to human disease. Such studies should include expression profiling of (1) large CLL patient cohorts containing a wide range of stereotypic and non-stereotypic U-CLL samples and (2) activated B cells that received various stimulations including anti-CD40.

Gene expression profiling revealed a set of genes that distinguish V_{H11} from non- V_{H11} CLL and are similarly regulated in BCR or CD40-stimulated cells, respectively. This observation probably reflects differences in supporting external

cues: pathways induced by interleukin or growth factor-mediated signaling were specifically upregulated in non- V_{H11} CLL. These include the regulator of G-protein signaling 16, *Rgs16*, which is upregulated in autoimmune B cells of BXD2 mice and enhances GC formation by the canonical NF- κ B pathway, signifying the post-GC origin of non- V_{H11} CLL (52, 53). Second, the actin-binding protein *Ccdc88a*, which plays a role in cytoskeletal remodeling and cell migration following activation of Akt downstream of EGFR (54) and can also enhance Akt signaling (42, 55). Third, integral membrane protein 2A (*Itm2a*) is a type II integral membrane protein that has been associated with an enhanced GATA3-mediated regulatory network in B ALL (56). *Chd3* encodes a chromatin remodeler with unexplored function in lymphocytes.

On the other hand, Wnt-associated genes were specifically upregulated in V_{H11} tumors, which is interesting because the BTK-inhibitor ibrutinib restrains Wnt signaling in CLL (57). Although the function of several other upregulated genes is currently unknown, *Zcchc18* has been associated with a CLL-specific transcriptomic signature (42) and *Clip3* was differentially regulated in a CLL patient undergoing spontaneous regression (58). Notably, many gene sets or pathways were active in both CLL subsets, including high expression levels of MET receptor tyrosine kinase, which prolongs CLL cell survival through STAT3 and AKT phosphorylation (40, 59). This could contribute to the enhanced constitutive activation of the p-Akt/p-S6 pathway in *IgH.TE μ* CLL as reported previously (23, 24). Additionally, genes involved in KRAS signaling were highly expressed in both CLL subsets, consistent with its essential role in B cell lymphopoiesis (60), particularly for B-1 cells recognizing PtC (61).

Our data also indicated that availability of T cell help and GC formation did not affect tumor incidence or onset. In contrast, the finding of a significantly earlier CLL incidence of mainly the non- V_{H11} type in *IgH.TE μ .Siglec-G $^{-/-}$* and *IgH.TE μ .E-Btk-2* mice suggests that BCR signaling thresholds are a key factor in determining CLL disease course. Yet, the appearance of V_{H11} CLL in these mouse lines may indicate a substantial selective advantage of these clones, because in *Siglec-G $^{-/-}$* and *E-Btk-2* transgenic mice the frequency of PtC-recognizing cells within the B-1 cell population is very low (28, 36).

In conclusion, we found that the formation of a major subset of unmutated CLL in *IgH.TE μ* mice is dependent on T cell signals. Our findings therefore provide a mechanistic explanation for the role of B-cell intrinsic factors, in particular BCR signaling, as well as extrinsic factors such as T cell help and support from the tumor microenvironment, in shaping the repertoire of CLL in mice. These findings are of potential clinical relevance, because B-cell extrinsic signals may reflect effective targets for novel therapeutic strategies in CLL patients.

AUTHOR CONTRIBUTIONS

SPS designed the research studies, performed experiments, analyzed the data, and wrote the manuscript. MdB, SP, RM, and MdA performed experiments and analyzed the data. RS analyzed RNA sequencing data and contributed to writing

the manuscript. AL and LN contributed to the research design and the writing of the manuscript. RH contributed to the research design and the writing of the manuscript and supervised the study. All co-authors approved the final manuscript.

FUNDING

These studies were partly supported by the Dutch Cancer Society (KWF 2014-6564), the Association for International Cancer Research (10-562) to RH, NWO (to SPS) and a NWO Veni Fellowship (Grant No. 91617114) to RS.

REFERENCES

- Hallek M. Chronic lymphocytic leukemia: 2015 Update on diagnosis, risk stratification, and treatment. *Am J Hematol.* (2015) 90:446–60. doi: 10.1002/ajh.23979
- Stamatopoulos K, Agathangelidis A, Rosenquist R, Ghia P. Antigen receptor stereotypy in chronic lymphocytic leukemia. *Leukemia* (2017) 31:282–91. doi: 10.1038/leu.2016.322
- Damle RN, Wasil T, Fais F, Ghiotto F, Valetto A, Allen SL, et al. Ig V gene mutation status and CD38 expression as novel prognostic indicators in chronic lymphocytic leukemia. *Blood* (1999) 94:1840–7.
- Hamblin TJ, Davis Z, Gardiner A, Oscier DG, Stevenson FK. Unmutated Ig V(H) genes are associated with a more aggressive form of chronic lymphocytic leukemia. *Blood* (1999) 94:1848–54.
- Mertens D, Stilgenbauer S. Prognostic and predictive factors in patients with chronic lymphocytic leukemia: relevant in the era of novel treatment approaches? *J Clin Oncol.* (2014) 32:869–72. doi: 10.1200/JCO.2013.53.8421
- Pal Singh S, Dammeijer F, Hendriks RW. Role of Bruton's tyrosine kinase in B cells and malignancies. *Mol Cancer* (2018) 17:57. doi: 10.1186/s12943-018-0779-z
- Byrd JC, Furman RR, Coutre SE, Flinn IW, Burger JA, Blum KA, et al. Targeting BTK with ibrutinib in relapsed chronic lymphocytic leukemia. *N Engl J Med.* (2013) 369:32–42. doi: 10.1056/NEJMoa1215637
- Byrd JC, Harrington B, O'Brien S, Jones JA, Schuh A, Devereux S, et al. Acalabrutinib (ACP-196) in relapsed chronic lymphocytic leukemia. *N Engl J Med.* (2016) 374:323–32. doi: 10.1056/NEJMoa1509981
- Dühren-von Minden M, Uebelhart R, Schneider D, Wossning T, Bach MP, Buchner M, et al. Chronic lymphocytic leukaemia is driven by antigen-independent cell-autonomous signalling. *Nature* (2012) 489:309–12. doi: 10.1038/nature11309
- Herve M, Xu K, Ng YS, Wardemann H, Albesiano E, Messmer BT, et al. Unmutated and mutated chronic lymphocytic leukemias derive from self-reactive B cell precursors despite expressing different antibody reactivity. *J Clin Invest.* (2005) 115:1636–43. doi: 10.1172/JCI24387
- Lanemo Myhrinder A, Hellqvist E, Sidorova E, Soderberg A, Baxendale H, Dahle C, et al. A new perspective: molecular motifs on oxidized LDL, apoptotic cells, and bacteria are targets for chronic lymphocytic leukemia antibodies. *Blood* (2008) 111:3838–48. doi: 10.1182/blood-2007-11-125450
- Baumgarth N. B-1 cell heterogeneity and the regulation of natural and antigen-induced IgM production. *Front Immunol.* (2016) 7:324. doi: 10.3389/fimmu.2016.010.3389/fimmu.2016.00324
- Catera R, Silverman GJ, Hatzi K, Seiler T, Didier S, Zhang L, et al. Chronic lymphocytic leukemia cells recognize conserved epitopes associated with apoptosis and oxidation. *Mol Med.* (2008) 14:665–74. doi: 10.2119/2008-00102
- Chu CC, Catera R, Hatzi K, Yan XJ, Zhang L, Wang XB, et al. Chronic lymphocytic leukemia antibodies with a common stereotypic rearrangement recognize nonmuscle myosin heavy chain IIA. *Blood* (2008) 112:5122–9. doi: 10.1182/blood-2008-06-162024
- Mockridge CI, Potter KN, Wheatley I, Neville LA, Packham G, Stevenson FK. Reversible anergy of sIgM-mediated signaling in the two subsets of

ACKNOWLEDGMENTS

We thank Laurens P. Kil, Alice F. Muggen, Melanie Lukkes and Catarina da Graça (all Erasmus MC) and Arnon P. Kater (Amsterdam Medical Center, Amsterdam, The Netherlands) for assistance and advice.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2018.01996/full#supplementary-material>

- CLL defined by VH-gene mutational status. *Blood* (2007) 109:4424–31. doi: 10.1182/blood-2006-11-056648
- Muzio M, Apollonio B, Scielzo C, Frenquelli M, Vandoni I, Boussiotis V, et al. Constitutive activation of distinct BCR-signaling pathways in a subset of CLL patients: a molecular signature of anergy. *Blood* (2008) 112:188–95. doi: 10.1182/blood-2007-09-111344
- Garcia-Munoz R, Galiacho VR, Llorente L. Immunological aspects in chronic lymphocytic leukemia (CLL) development. *Ann Hematol.* (2012) 91:981–96. doi: 10.1007/s00277-012-1460-z
- Seifert M, Sellmann L, Bloehdorn J, Wein F, Stilgenbauer S, Durig J, et al. Cellular origin and pathophysiology of chronic lymphocytic leukemia. *J Exp Med.* (2012) 209:2183–98. doi: 10.1084/jem.20120833
- Chiorazzi N, Rai KR, Ferrarini M. Chronic lymphocytic leukemia. *N Engl J Med.* (2005) 352:804–15. doi: 10.1056/NEJMra041720
- Simonetti G, Bertilaccio MT, Ghia P, Klein U. Mouse models in the study of chronic lymphocytic leukemia pathogenesis and therapy. *Blood* (2014) 124:1010–9. doi: 10.1182/blood-2014-05-577122
- Chen SS, Batliwalla F, Holodick NE, Yan XJ, Yancopoulos S, Croce CM, et al. Autoantigen can promote progression to a more aggressive T-CLL leukemia by selecting variants with enhanced B-cell receptor signaling. *Proc Natl Acad Sci USA.* (2013) 110:E1500–7. doi: 10.1073/pnas.1300616110
- ter Brugge PJ, Ta VB, de Bruijn MJ, Keijzers G, Maas A, van Gent DC, et al. A mouse model for chronic lymphocytic leukemia based on expression of the SV40 large T antigen. *Blood* (2009) 114:119–27. doi: 10.1182/blood-2009-01-198937
- Kil LP, de Bruijn MJ, van Hulst JA, Langerak AW, Yuvaraj S, Hendriks RW. Bruton's tyrosine kinase mediated signaling enhances leukemogenesis in a mouse model for chronic lymphocytic leukemia. *Am J Blood Res.* (2013) 3:71–83.
- Singh SP, Pillai SY, de Bruijn MJW, Stadhouders R, Corneth OBJ, van den Ham HJ, et al. Cell lines generated from a chronic lymphocytic leukemia mouse model exhibit constitutive Btk and Akt signaling. *Oncotarget* (2017) 8:71981–95. doi: 10.18632/oncotarget.18234
- Renshaw BR, Fanslow WC III, Armitage RJ, Campbell KA, Liggitt D, Wright B, et al. Humoral immune responses in CD40 ligand-deficient mice. *J Exp Med.* (1994) 180:1889–900. doi: 10.1084/jem.180.5.1889
- Muramatsu M, Kinoshita K, Fagarasan S, Yamada S, Shinkai Y, Honjo T. Class switch recombination and hypermutation require activation-induced cytidine deaminase (AID), a potential RNA editing enzyme. *Cell* (2000) 102:553–63. doi: 10.1016/S0092-8674(00)00078-7
- Hoffmann A, Kerr S, Jellusova J, Zhang J, Weisel F, Wellmann U, et al. Siglec-G is a B1 cell-inhibitory receptor that controls expansion and calcium signaling of the B1 cell population. *Nat Immunol.* (2007) 8:695–704. doi: 10.1038/ni1480
- Kersseboom R, Kil L, Flierman R, van der Zee M, Dingjan GM, Middendorp S, et al. Constitutive activation of Bruton's tyrosine kinase induces the formation of autoreactive IgM plasma cells. *Eur J Immunol.* (2010) 40:2643–54. doi: 10.1002/eji.201040521
- Kil LP, de Bruijn MJ, van Nimwegen M, Corneth OB, van Hamburg JP, Dingjan GM, et al. Btk levels set the threshold for B-cell activation and negative selection of autoreactive B cells in mice. *Blood* (2012) 119:3744–56. doi: 10.1182/blood-2011-12-397919

30. Robinson MD, McCarthy DJ, Smyth GK. edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics* (2010) 26:139–40. doi: 10.1093/bioinformatics/btp616
31. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol.* (2014) 15:550. doi: 10.1186/s13059-014-0550-8
32. Saldanha AJ. Java Treeview—extensible visualization of microarray data. *Bioinformatics* (2004) 20:3246–8. doi: 10.1093/bioinformatics/bth349
33. Wohner M, Tagoh H, Bilic I, Jaritz M, Poliakova DK, Fischer M, et al. Molecular functions of the transcription factors E2A and E2-2 in controlling germinal center B cell and plasma cell development. *J Exp Med.* (2016) 213:1201–21. doi: 10.1084/jem.20152002
34. Xu J, Foy TM, Laman JD, Elliott EA, Dunn JJ, Waldschmidt TJ, et al. Mice deficient for the CD40 ligand. *Immunity* (1994) 1:423–31. doi: 10.1016/1074-7613(94)90073-6
35. Park H, Wahl MI, Afar DE, Turck CW, Rawlings DJ, Tam C, et al. Regulation of Btk function by a major autophosphorylation site within the SH3 domain. *Immunity* (1996) 4:515–25. doi: 10.1016/S1074-7613(00)80417-3
36. Jellusova J, Duber S, Guckel E, Binder CJ, Weiss S, Voll R, et al. Siglec-G regulates B1 cell survival and selection. *J Immunol.* (2010) 185:3277–84. doi: 10.4049/jimmunol.1001792
37. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci USA.* (2005) 102:15545–50. doi: 10.1073/pnas.0506580102
38. Huttmann A, Klein-Hitpass L, Thomale J, Deenen R, Carpinteiro A, Nuckel H, et al. Gene expression signatures separate B-cell chronic lymphocytic leukaemia prognostic subgroups defined by ZAP-70 and CD38 expression status. *Leukemia* (2006) 20:1774–82. doi: 10.1038/sj.leu.2404363
39. Wang J, Coombes KR, Highsmith WE, Keating MJ, Abruzzo LV. Differences in gene expression between B-cell chronic lymphocytic leukemia and normal B cells: a meta-analysis of three microarray studies. *Bioinformatics* (2004) 20:3166–78. doi: 10.1093/bioinformatics/bth381
40. Giannoni P, Scaglione S, Quarto R, Narcisi R, Parodi M, Balleari E, et al. An interaction between hepatocyte growth factor and its receptor (c-MET) prolongs the survival of chronic lymphocytic leukemic cells through STAT3 phosphorylation: a potential role of mesenchymal cells in the disease. *Haematologica* (2011) 96:1015–23. doi: 10.3324/haematol.2010.029736
41. Kasar S, Kim J, Impropo R, Tiao G, Polak P, Haradhvala N, et al. Whole-genome sequencing reveals activation-induced cytidine deaminase signatures during indolent chronic lymphocytic leukaemia evolution. *Nat Commun.* (2015) 6:8866. doi: 10.1038/ncomms9866
42. Chadeau-Hyam M, Vermeulen RC, Hebls DG, Castagne R, Campanella G, Portengen L, et al. Prediagnostic transcriptomic markers of Chronic lymphocytic leukemia reveal perturbations 10 years before diagnosis. *Ann Oncol.* (2014) 25:1065–72. doi: 10.1093/annonc/mdu056
43. Bartolome-Izquierdo N, de Yébenes VG, Alvarez-Prado AF, Mur SM, Lopez Del Olmo JA, Roa S, et al. miR-28 regulates the germinal center reaction and blocks tumor growth in preclinical models of non-Hodgkin lymphoma. *Blood* (2017) 129:2408–19. doi: 10.1182/blood-2016-08-731166
44. Pottier N, Paugh SW, Ding C, Pei D, Yang W, Das S, et al. Promoter polymorphisms in the beta-2 adrenergic receptor are associated with drug-induced gene expression changes and response in acute lymphoblastic leukemia. *Clin Pharmacol Ther.* (2010) 88:854–61. doi: 10.1038/clpt.2010.212
45. Sitthi-Amorn J, Herrington B, Megason G, Pullen J, Gordon C, Hogan S, et al. Transcriptome analysis of minimal residual disease in subtypes of pediatric B cell acute lymphoblastic leukemia. *Clin Med Insights Oncol.* (2015) 9:51–60. doi: 10.4137/CMO.S1704910.4137/CMO.S17049
46. Camos M, Esteve J, Jares P, Colomer D, Rozman M, Villamor N, et al. Gene expression profiling of acute myeloid leukemia with translocation t(8;16)(p11;p13) and MYST3-CREBBP rearrangement reveals a distinctive signature with a specific pattern of HOX gene expression. *Cancer Res.* (2006) 66:6947–54. doi: 10.1158/0008-5472.CAN-05-4601
47. Hayakawa K, Formica AM, Colombo MJ, Shinton SA, Brill-Dashoff J, Morse Iii HC, et al. Loss of a chromosomal region with synteny to human 13q14 occurs in mouse chronic lymphocytic leukemia that originates from early-generated B-1 B cells. *Leukemia* (2016) 30:1510–9. doi: 10.1038/leu.2016.61
48. Hayakawa K, Formica AM, Brill-Dashoff J, Shinton SA, Ichikawa D, Zhou Y, et al. Early generated B1 B cells with restricted BCRs become chronic lymphocytic leukemia with continued c-Myc and low Bmf expression. *J Exp Med.* (2016) 213:3007–24. doi: 10.1084/jem.20160712
49. Klein U, Tu Y, Stolovitzky GA, Mattioli M, Cattoretti G, Husson H, et al. Gene expression profiling of B cell chronic lymphocytic leukemia reveals a homogeneous phenotype related to memory B cells. *J Exp Med.* (2001) 194:1625–38. doi: 10.1084/jem.194.11.1625
50. Matejuk A, Beardall M, Xu Y, Tian Q, Phillips D, Alabyev B, et al. Exclusion of natural autoantibody-producing B cells from IgG memory B cell compartment during T cell-dependent immune responses. *J Immunol.* (2009) 182:7634–43. doi: 10.4049/jimmunol.0801562
51. Zenz T, Mertens D, Kuppers R, Dohner H, Stilgenbauer S. From pathogenesis to treatment of chronic lymphocytic leukaemia. *Nat Rev Cancer* (2010) 10:37–50. doi: 10.1038/nrc2764
52. Xie S, Li J, Wang JH, Wu Q, Yang P, Hsu HC, et al. IL-17 activates the canonical NF-kappaB signaling pathway in autoimmune B cells of BXD2 mice to upregulate the expression of regulators of G-protein signaling 16. *J Immunol.* (2010) 184:2289–96. doi: 10.4049/jimmunol.0903133
53. Hsu HC, Yang P, Wang J, Wu Q, Myers R, Chen J, et al. Interleukin 17-producing T helper cells and interleukin 17 orchestrate autoreactive germinal center development in autoimmune BXD2 mice. *Nat Immunol.* (2008) 9:166–75. doi: 10.1038/ni1552
54. Enomoto A, Ping J, Takahashi M. Girdin, a novel actin-binding protein, and its family of proteins possess versatile functions in the Akt and Wnt signaling pathways. *Ann N Y Acad Sci.* (2006) 1086:169–84. doi: 10.1196/annals.1377.016
55. Mayer RL, Schwarzmeier JD, Gerner MC, Bileck A, Mader JC, Meier-Menches SM, et al. Proteomics and metabolomics identify molecular mechanisms of aging potentially predisposing for chronic lymphocytic leukemia. *Mol Cell Proteomics* (2017) 17:290–303. doi: 10.1074/mcp.RA117.000425
56. Hou Q, Liao F, Zhang S, Zhang D, Zhang Y, Zhou X, et al. Regulatory network of GATA3 in pediatric acute lymphoblastic leukemia. *Oncotarget* (2017) 8:36040–53. doi: 10.18632/oncotarget.16424
57. Li PP, Lu K, Geng LY, Zhou XX, Li XY, Wang X. Bruton's tyrosine kinase inhibitor restrains Wnt signaling in chronic lymphocytic leukemia. *Mol Med Rep.* (2016) 13:4934–8. doi: 10.3892/mmr.2016.5111
58. Del Giudice I, Chiaretti S, Tavolaro S, De Propriis MS, Maggio R, Mancini F, et al. Spontaneous regression of chronic lymphocytic leukemia: clinical and biologic features of 9 cases. *Blood* (2009) 114:638–46. doi: 10.1182/blood-2008-12-196568
59. Eksioğlu-Demiralp E, Akdeniz T, Bayik M. Aberrant expression of c-met and HGF/c-met pathway provides survival advantage in B-chronic lymphocytic leukemia. *Cytometry B Clin Cytom.* (2011) 80:1–7. doi: 10.1002/cyto.b.20553
60. Chen Y, Zheng Y, You X, Yu M, Fu G, Su X, et al. Kras is critical for B cell lymphopoiesis. *J Immunol.* (2016) 196:1678–85. doi: 10.4049/jimmunol.1502112
61. Guo B, Rothstein TL. RasGRP1 is an essential signaling molecule for development of B1a cells with autoantigen receptors. *J Immunol.* (2016) 196:2583–90. doi: 10.4049/jimmunol.1502132

Conflict of Interest Statement: SP was affiliated to Erasmus MC Rotterdam whilst this study was completed. After finishing the work, he founded the company EpiExpressions.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Pal Singh, de Bruijn, de Almeida, Meijers, Nitschke, Langerak, Pillai, Stadhouders and Hendriks. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.