Multihospital Outbreak of a Middle East Respiratory Syndrome Coronavirus Deletion Variant, Jordan: A Molecular, Serologic, and Epidemiologic Investigation

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Background. An outbreak of Middle East respiratory syndrome coronavirus (MERS-CoV) in Jordan in 2015 involved a variant virus that acquired distinctive deletions in the accessory open reading frames. We conducted a molecular and seroepidemiologic investigation to describe the deletion variant's transmission patterns and epidemiology.

Methods. We reviewed epidemiologic and medical chart data and analyzed viral genome sequences from respiratory specimens of MERS-CoV cases. In early 2016, sera and standardized interviews were obtained from MERS-CoV cases and their contacts. Sera were evaluated by nucleocapsid and spike protein enzyme immunoassays and microneutralization.

Results. Among 16 cases, 11 (69%) had health care exposure and 5 (31%) were relatives of a known case; 13 (81%) were symptomatic, and 7 (44%) died. Genome sequencing of MERS-CoV from 13 cases revealed 3 transmissible deletions associated with clinical illness during the outbreak. Deletion variant sequences were epidemiologically clustered and linked to a common transmission chain. Interviews and sera were collected from 2 surviving cases, 23 household contacts, and 278 health care contacts; 1 (50%) case, 2 (9%) household contacts, and 3 (1%) health care contacts tested seropositive.

Conclusions. The MERS-CoV deletion variants retained human-to-human transmissibility and caused clinical illness in infected persons despite accumulated mutations. Serology suggested limited transmission beyond that detected during the initial outbreak investigation.

Keywords. coronavirus; emerging infectious disease; genome deletion; Jordan; MERS-CoV; Middle East respiratory syndrome; outbreak investigation; sero-epidemiology.

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Middle East respiratory syndrome coronavirus (MERS-CoV) causes acute respiratory illness that can progress rapidly to respiratory failure and death in approximately 35%-40% of reported laboratory-confirmed cases [1, 2]. The first known cases of MERS-CoV occurred during an outbreak of severe acute respiratory infections in Zarqa, Jordan, during March–April 2012 [3, 4]. Since then, new cases and clusters of MERS-CoV infections continue to occur within the Arabian Peninsula and have been exported to other countries around the world [5].

During August–October 2015, a MERS-CoV outbreak with 16 laboratory-confirmed cases occurred in hospitals in Amman and Zarqa, Jordan. An outbreak investigation was conducted by the Jordan Ministry of Health (JMoH), followed by a World Health Organization review in September 2015. Sequencing performed by Lamers et al. [6] on samples from 13 outbreak cases found the virus to be associated with a novel recombinant clade that predominated in Saudi Arabia in 2015 [7] and was exported to South Korea and China [8]. Importantly, genetic analysis also revealed 2 unique mutations resulting from substantial deletion events—a 48-nucleotide (nt) in-frame deletion in accessory open reading frame (ORF) 4a present in sequences from 13 outbreak cases and an additional 9-nt in-frame deletion in ORF3 in a subset of case specimens collected later in the outbreak—leading the authors to conclude that all cases were infected following a single introduction of the virus [6]. We conducted an investigation that included additional viral genome and serologic analysis to describe the transmission patterns and epidemiology of MERS-CoV deletion variants during the 2015 Jordan outbreak.

METHODS

The JMoH, US Centers for Disease Control and Prevention (CDC), and Eastern Mediterranean Public Health Network collaboratively conducted a follow-up investigation of the 2015 MERS-CoV outbreak in Jordan during March–April 2016.
We reviewed outbreak investigation records, conducted key informant interviews with clinicians and infection control staff at the affected hospitals, and reviewed medical charts of confirmed cases. A confirmed MERS case met the World Health Organization case definition [9] and was laboratory-confirmed at the Jordan Central Public Health Laboratory using MERS-CoV N2 and/or N3 real-time reverse transcription polymerase chain reaction (rRT-PCR) assays [10]. A case was considered health-care–associated if they were a patient, visitor, or health care personnel (HCP) at a health care facility during the 14 days before symptom onset (exposure period), and if exposure to a known MERS case occurred exclusively in this setting [11].

To identify previously undetected MERS-CoV infections among contacts of confirmed cases, we interviewed and collected sera from health care and household contacts of confirmed MERS cases from the 2015 outbreak. Additionally, we interviewed and collected sera from available surviving cases. Contacts were identified using contact tracing lists developed by JMoH and the affected hospitals during the initial outbreak investigation. All health care contacts included in the investigation were health care personnel; all household contacts were relatives of confirmed cases, although they did not necessarily live under the same roof. Standardized interview forms were completed, which included questions about types of exposure to confirmed cases, symptoms after exposure, occupation, and testing for MERS-CoV at the time of the outbreak.

**Laboratory Methods**

**Genome Sequencing and Phylogenetic Analysis**

MERS-CoV rRT-PCR-positive nasopharyngeal swabs, sputum, tracheal aspirate, or bronchoalveolar lavage samples from 15 of the 16 confirmed cases collected during August–September 2015 were stored at −80°C and shipped to the CDC for molecular analysis. Sample aliquots (200 µL) were extracted on a NucliSens EasyMAG (BioMerieux), and 100 µL of total nucleic acid was recovered. The specimen extracts were retested by MERS-CoV N2 and/or N3 rRT-PCR assays [10], and genome sequencing was performed on confirmed positive samples with sufficient viral load using previously described primer sets and protocol [7].

Nucleotide sequences were aligned using Clustal X, ver. 1.83, implemented in BioEdit, ver. 7.2.5. Phylogenetic analyses were performed using MrBayes, ver. 3.2.6 [12], under a GTR model of nucleotide substitution with 4 categories of γ-distributed rate heterogeneity and a proportion of invariant sites (GTR + 4 + I).

**Seroogy**

Serum samples collected during March–April 2016 were shipped to the CDC for MERS-CoV–specific serum antibody testing using MERS-CoV nucleocapsid (N) and spike (S) indirect enzyme-linked immunosorbent assay (ELISA) and microneutralization (MNT) assays [4, 13, 14]. Sera were considered potentially positive when the optical density (OD) values were at or above the screening 0.15 cutoff value for N-ELISA or 0.1 for S-ELISA. Any serum potentially positive by N- or S-ELISA at a screening dilution of 1:400 was subsequently titered with serial 4-fold dilutions (1:100–1:6400) with OD cutoffs of 0.3 for the N and 0.2 for the S-ELISAs to determine end point titer and tested by MNT for confirmation. Neutralizing antibodies to MERS-CoV were detected by MNT assay performed in Biosafety Level 3 (BSL-3) containment, as previously described [4]. The neutralization titer was defined as the reciprocal of the highest serum dilution that completely inhibited the live MERS-CoV–mediated cytopathic effect in Vero cells in at least 1 of the 3 triplicate wells. N- and S-ELISAs were considered positive with reciprocal end point titers of 400 or greater. MNT were considered positive with reciprocal titers of 20 or greater. MERS-CoV seropositivity was defined as having ≥2 of 3 positive assays, including N-ELISA, S-ELISA, and MNT, or positive by MNT alone [14]. Sera indeterminate seropositivity was defined as S-ELISA positive, but N-ELISA and MNT negative.

**Ethics**

This investigation was classified by the CDC as a public health response to an emerging disease outbreak, and Jordanian Institutional Review Board approval was obtained. Verbal consent was obtained in Arabic.

**RESULTS**

During August–October 2015, 16 laboratory-confirmed MERS-CoV patients were identified in Jordan. These MERS cases received care at 9 hospitals in Jordan, with case clusters associated with 3: hospitals A, B, and C. The median patient age was 55 years (10 months–78 years), 9 (56%) were male, and 13 (81%) were symptomatic (Table 1). Seven (44%) patients died, all of whom had underlying medical conditions. Eleven (69%) cases were health care associated, and 5 (31%) were relatives of a known case. Health care–associated cases included 6 hospital patients, 3 visitors, and 2 HCP.

**Molecular Epidemiology**

Respiratory specimens from 15 (94%) of the MERS-CoV–positive patients were received by the CDC and confirmed positive by rRT-PCR; 13 specimens representing 11 symptomatic and 2 asymptomatic cases had sufficient viral RNA for genome sequencing. Phylogenetic analysis of the 13 nearly complete genomes (GenBank accession No. MF741825-MF741837) and 240 previously published MERS-CoV genomes showed well-supported clustering of the Jordan outbreak sequences within a previously identified clade defined by a signature recombination (Figure 1) [7, 8]. Jordan outbreak sequences showed the closest similarity with sequences obtained from 4 temporally concurrent but epidemiologically unrelated human MERS cases from Riyadh, Saudi Arabia, in August 2015 (GenBank accession Nos. KU851860–851862 and KU851864), which did not possess any deletion mutations [15].

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**Table 1.** Characteristics of MERS-CoV patients in Jordan during the 2015 outbreak.

<table>
<thead>
<tr>
<th>Patient</th>
<th>Sex</th>
<th>Age (yrs)</th>
<th>Underlying Condition</th>
<th>Symptoms</th>
<th>Outcome</th>
<th>Clinical Setting</th>
</tr>
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<tr>
<td>1</td>
<td>M</td>
<td>60</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital A</td>
</tr>
<tr>
<td>2</td>
<td>M</td>
<td>78</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital A</td>
</tr>
<tr>
<td>3</td>
<td>F</td>
<td>55</td>
<td>Diabetes, COPD</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital B</td>
</tr>
<tr>
<td>4</td>
<td>M</td>
<td>50</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital C</td>
</tr>
<tr>
<td>5</td>
<td>F</td>
<td>40</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital A</td>
</tr>
<tr>
<td>6</td>
<td>M</td>
<td>65</td>
<td>Diabetes, COPD</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital B</td>
</tr>
<tr>
<td>7</td>
<td>M</td>
<td>70</td>
<td>Diabetes, COPD</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital C</td>
</tr>
<tr>
<td>8</td>
<td>M</td>
<td>55</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital A</td>
</tr>
<tr>
<td>9</td>
<td>M</td>
<td>60</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital B</td>
</tr>
<tr>
<td>10</td>
<td>F</td>
<td>45</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital C</td>
</tr>
<tr>
<td>11</td>
<td>M</td>
<td>55</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital A</td>
</tr>
<tr>
<td>12</td>
<td>M</td>
<td>65</td>
<td>Diabetes, COPD</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital B</td>
</tr>
<tr>
<td>13</td>
<td>F</td>
<td>50</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital C</td>
</tr>
<tr>
<td>14</td>
<td>M</td>
<td>70</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital A</td>
</tr>
<tr>
<td>15</td>
<td>M</td>
<td>60</td>
<td>Diabetes, COPD</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital B</td>
</tr>
<tr>
<td>16</td>
<td>M</td>
<td>55</td>
<td>Diabetes</td>
<td>Fever</td>
<td>Died</td>
<td>Hospital C</td>
</tr>
</tbody>
</table>
We confirmed the 48-nt (16-aa) in-frame deletion in ORF4a present in all Jordan sequences as well as the tandem 9-nt (3-aa) in-frame deletion in ORF3 in sequences from 5 patients (cases 8, 12, 13, 14, 15) obtained later in the outbreak [6]. Additionally, we identified a third 726-nt out-of-frame deletion in ORF4b and ORF5 (genome position 26339–27064 based on accession number JX869059.2) (Supplementary Table 1) from 2 patients (cases 3, 11). In ORF4a, ORF3, and ORF4b, 5 deletions were all identified in 2 or more epidemiologically linked patients, indicating successful virus transmission. We also identified 5 additional unique deletions ranging in size from 978 to 1438 nt variably involving ORF1a, ORF3, ORF4a, ORF4b, and ORF5 that were only present as mixed variants in individual patients (cases 2, 5, 6, 14) without evidence of transmission (Figure 2).

The viral transmission patterns deduced independently through the epidemiologic investigation corresponded to the grouping of cases by deletions observed in their viral genomes (Figure 3). Three hospital clusters were identified; the earliest cases occurred at hospital A, and index cases for the hospital B and C clusters both had epidemiologic links to hospital A. The 2 cases with the earliest dates of illness onset (cases 1 and 2) were both present at the same coronary care unit (CCU) in hospital A on a single day (case 1 as a patient and case 2 as a visitor); however, no direct contact was documented between them. Before the 2015 outbreak involving the MERS-CoV deletion variants, the last identified MERS case in Jordan had been 8 months earlier, in December 2014. In the 2015 outbreak, cases 1 and 2 had both recently traveled to Jordan from locations on the Arabian Peninsula (Jeddah, Saudi Arabia, and Kuwait City, Kuwait, respectively). Interestingly, viruses genetically similar to the viruses detected in cases 1 and 2 in Jordan, but without deletions, were concurrently circulating in Riyadh, Saudi Arabia, during the period corresponding to the earliest cases in Jordan (August 2015) [15]. Neither case 1 nor case 2 reported contact with a known MERS-CoV patient, camels, or a health care setting outside of Jordan during their exposure period. A review of previous CCU admissions did not reveal other likely index case candidates. Genome sequence analysis could not definitively identify the antecedent virus. Despite an extensive investigation, epidemiologic evidence did not definitively determine which of these 2 cases may have been the index case. Subsequently, 3 patients (cases 3, 5, and 6) and 1 visitor (case 8) at hospital A were identified as cases. Cases 5 and 6 had been admitted in the CCU at hospital A MERS before symptom onset. No cases were identified among HCP at hospital A, including from the CCU. The wife of case 3 tested positive (case 11) shortly after case 3 died; a 726-nt deletion in the ORF4b and ORF5 genes was shared between this couple.

The hospital B cluster index case was case 2, who visited a relative at hospital A and subsequently became ill with respiratory symptoms within 2 days of his last exposure to hospital A. Case 2 was admitted to hospital B with severe respiratory illness and tested positive for MERS-CoV. His sister, case 4, was not symptomatic and was tested as a contact. Case 7 was a patient at hospital B during the period that case 2 was admitted. Additionally, 1 patient admitted at hospital B during the outbreak period became symptomatic with acute respiratory symptoms 10 days after discharge and was diagnosed...
with MERS-CoV after returning home to Saudi Arabia. She was not counted among the 16 Jordanian cases. The patient had returned to Jeddah, Saudi Arabia, just before onset of respiratory symptoms and was subsequently hospitalized there, transmitting MERS-CoV to an HCP in Jeddah. This case was detected during an investigation of otherwise unexplained hospital-associated MERS-CoV in Saudi Arabia [16] and could represent export of the deletion variant virus from Jordan to Saudi Arabia; however, sequences were not available for confirmation.

The case introducing the deletion variant to hospital C by evidence of onset time (case 8) had visited his daughter at hospital A during the period when MERS-CoV patients were admitted at hospital A. He became ill approximately 5–6 days after visiting hospital A and was admitted at hospital C for care. Subsequently, 1 patient (case 14), 1 visitor (case 13), and 2 health care personnel (cases 12 and 15) at hospital C were identified with MERS-CoV infection. The 9-nt deletion in the ORF3 gene was detected among all identified cases linked to the hospital C cluster. The visit of case 8 to hospital A corresponded

Figure 1. Phylogenetic tree constructed from near complete Middle East respiratory syndrome coronavirus (MERS-CoV) genome sequences obtained from the 13 Jordan cases, indicated by solid-colored circles, and 240 previously published genome sequences in GenBank using the program MrBayes v3.2.6 [12] under a general time-reversible (GTR) model of nucleotide (nt) substitution with 4 categories of $\gamma$-distributed rate heterogeneity and a proportion of invariant sites (GTR + $4 + I$). Clade-credibility values $\geq 70\%$ are indicated at selected nodes. Circle colors correspond to transmitted deletion mutations: blue indicates 48-nt in-frame deletion in ORF4a; purple indicates 48-nt in-frame deletion in ORF4a and tandem 9-nt in-frame deletion in ORF3; and yellow indicates 48-nt in-frame deletion in ORF4a and tandem 726-nt out-of-frame deletion in ORF4b and 5. Case numbers assigned by the Jordan Ministry of Health are indicated within the circles. The scale bar shows the genetic distance as nt substitutions per site. For case 5, 2 sequence variants were detected: (i) a sequence with the ORF4a deletion only and (ii) a sequence with the ORF3 deletion and a tandem deletion in ORF3, 4a and 4b. Only the sequence with the ORF4a deletion is presented.
temporally with the hospitalization of case 5 at hospital A, whose specimen contained 2 genome sequence variants, 1 with the ORF4a deletion only and a second with the 9-nt ORF3 deletion noted above and a downstream 1042-nt out-of-frame deletion involving ORF3, 4a, and 4b.

**Seroepidemiologic Investigation**

Interviews and sera were obtained from 301 contacts of the 16 confirmed MERS-CoV cases, including 278 health care and 23 household contacts, as well as 2 surviving confirmed cases (Supplementary Table 2). Of the 278 health care contacts, 3 (1%) were seropositive and 4 (1%) were serologically indeterminate. Seropositive HCP included a nurse, a respiratory therapist, and a radiology clerk. Serologically indeterminate HCP included 2 respiratory therapists, a nurse, and a housekeeper (Table 2). The seropositive respiratory therapist reported fever and cough within 14 days of exposure to a MERS-CoV patient; all others did not report symptoms. The seropositive radiology clerk and symptomatic respiratory therapist tested negative for MERS-CoV by rRT-PCR during the initial outbreak investigation.

Seropositive and serologically indeterminate HCP were detected from 3 hospitals: B, C, and D (Table 2). Of the 58 hospital A HCP tested, none were seropositive or serologically indeterminate. At hospitals B and C, health care–associated transmission was identified at the time of the outbreak; at hospital D, where 2 serologically indeterminate HCP were detected, health care–associated transmission had not been documented. At hospital C, the investigation revealed a seropositive respiratory therapist who cared for both case 8 hospital C (index case) and case 14, who later developed symptoms after hospital discharge. The 2 cases briefly overlapped on the same floor, and case 8 immediately preceded case 14 in a particular radiology suite; however, no direct contact between the patients was reported. Case 15, a nurse at hospital C, also cared for both case 8 and case 14, but her suspected onset followed the exposure periods of these cases.

Of the 23 household contacts, 2 (9%) were seropositive and an additional 2 (9%) were serologically indeterminate. Reported household contact included living in the same household as a confirmed MERS-CoV case, caring for the case at home, and visiting the case in the hospital (Table 2). One seropositive household contact had fever and respiratory symptoms within 14 days of contact with a confirmed case. All household contacts tested negative for MERS-CoV by rRT-PCR during the initial investigation.

Of the 2 surviving confirmed MERS-CoV patients tested, 1 (50%) was seropositive by MNT (case 12) and 1 was seronegative (case 15). Case 12 was a nurse with no comorbidities who had severe illness requiring hospitalization. Case 15 was a nurse with no comorbidities who was documented as asymptomatic.
DISCUSSION

The MERS-CoV ORF 4a deletion variant and additional mutations observed over the course of this outbreak, including deletions in ORFs 3, 4b, and 5, were epidemiologically clustered in time and space, suggesting a single transmission chain and demonstrating human-to-human transmissibility of accumulated deletions during the outbreak. Although all outbreak specimens exhibited the ORF4a deletion, an additional deletion in ORF3, which was observed later in the outbreak first reported by Lamers et al. [6], was predominantly associated with a cluster at a single hospital. Additionally, we observed transmission of a unique 726nt deletion spanning ORF4b and ORF5 between a husband and wife as well as detection of multiple other deletion variants within the same hosts. The overall epidemiology of this outbreak, including prominent transmission among healthcare contacts, a less pronounced role of household transmission, a high case fatality rate (44%), presence of comorbidities among fatal cases, and retrospective evidence of asymptomatic infections, is all consistent with previous descriptions of hospital outbreaks, despite the distinctive genomic variation we observed [17, 18]. The genomic variability of this MERS-CoV deletion virus and the accumulation of genetic mutations observed as the outbreak continued, as well as information from other recent reports [19, 20], highlight the critical importance of ongoing contemporaneous molecular and epidemiologic surveillance for the persistent emergence of MERS-CoV.

This outbreak was unique in having many deletions detected from multiple patients, the accumulation of deletion mutations over time, and that these deletion variants retained transmissibility and virulence. The observed deletion mutations allowed us to link all outbreak patients to a single originating virus (ORF4a) and revealed some degree of transmission directionality in the outbreak. We observed multiple deletion mutations of varying size mostly distributed among accessory ORFs of the Jordan outbreak viruses. Deletion variants of MERS-CoV present as mixed infections with wild-type virus involving the ORF5, and E protein genes [20] and spike protein gene [19] have been previously reported from patients.

MERS-CoV has 5 group-specific ORFs (3, 4a, 4b, 5, and 8b) that encode accessory proteins whose functions are not
fully characterized [21]. Although certain ORFs have been shown to be nonessential for virus replication in vitro [22], evidence exists that they may play a role in modulation of the host innate immune response [23–25]. For example, ORF 4a, ORF 4b, and ORF 5 proteins have been shown to be potent interferon antagonists [23]. Using an ORF3-5 deleted MERS-CoV infectious clone, Baric et al. observed increased stimulation of the interferon response in vitro and reduced viral replication and pathogenesis in a mouse model, compared with a wild-type virus [26]. Although deletions in accessory ORFs hypothetically could modulate the host response during infection, no apparent changes in pathogenicity were observed during our investigation; variant viruses with deletions in ORF 4a, ORFs 3, and 4a, and ORFs 4a, 4b, 5 each resulted in symptomatic illness and death in some cases. The ORF 4a deletion we observed is in-frame and located within the dsRNA-binding domain. The impact on the functionality of the protein with the deletion is unknown and warrants further study. Notably, deletions in accessory ORFs were also observed in SARS-CoV after its introduction to human hosts, and viruses containing a specific ORF 8 deletion were predominant later in the SARS epidemic [25, 27]. Viral genomic changes may have arisen due to selective pressures exerted by the human host, or due to the lack of such pressures on genes that are required specifically for sustained replication in camels, as suggested by Lamers et al. [6].

The combination of serologic and epidemiologic data improved our understanding of viral transmission patterns, including a seropositive HCP that may have been the link between 2 patient cases. We found that the majority of seropositive or indeterminate HCP were either respiratory therapists or nurses, demonstrating risks for these occupations in outbreak settings [28]. We also identified a nonclinical HCP (radiology clerk) who was seropositive. Infection among nonclinical HCP has been less commonly reported; however, these findings highlight the importance of including nonclinical HCP in contact tracing and monitoring efforts and the need to further evaluate specific health care activities associated with risk of MERS-CoV transmission.

Previous studies on MERS-CoV antibody kinetics suggest that serologic investigations might underestimate mild and asymptomatic infections [29]. Individuals with mild or asymptomatic infection appear to less reliably develop a detectable antibody response after infection, although antibodies have been observed to persist for nearly 3 years after infection in even young persons without significant comorbidities [13, 29–31]. In our seroepidemiologic investigation, we found that case 15, a mildly ill, rRT-PCR- and sequence-confirmed MERS-CoV-infected patient, may have been the source of infection in 2 other patients, including a seropositive HCP that may have been the link between 2 patient cases.
case at the time of the outbreak, tested seronegative approximately 7 months later. Conversely, we identified individuals who reported no symptoms and tested MERS-CoV negative during outbreak contact tracing but were subsequently seropositive. Several exposed individuals had detectable anti-MERS-CoV S-serum antibodies by ELISA, including 1 who exhibited acute respiratory symptoms but did not have detectable neutralizing serum antibodies (serologic result classified as indeterminate). These individuals may have developed anti-S antibodies to non-neutralizing epitopes, neutralizing antibodies may have decayed to below detectable levels at the time we collected the sera (~8 months after infection), or they may have never mounted robust antibody responses during the initial acute infection. The natural history of the immune response to MERS-CoV infection as related to disease severity needs further investigation. The role of mildly ill or asymptomatic individuals in transmission of MERS-CoV remains a particularly critical question in health care settings where patients with underlying conditions might be at increased risk for infection.

Direct epidemiologic links between some health care–associated cases could not be established. In addition to potentially unrecognized human cases, indirect contact or fomite transmission in this outbreak also remains a possibility. Although contamination of environmental surfaces is a suspected mode of transmission for some respiratory viruses in certain settings [32], this has not been confirmed as a route of transmission for MERS-CoV. Key informant interviews with hospital infection control officers and clinicians working during the outbreak, as well as hygienic observations during the investigation, suggest that patients admitted before MERS-CoV was recognized could have contaminated shared hospital equipment with transmissible virus (specifically, an x-ray table and portable echocardiogram machine). Viable MERS-CoV has been recovered from surfaces after 48 hours at 20°C and 40% relative humidity [33], suggesting plausibility that MERS-CoV survives on indoor surfaces in a climate-controlled setting. Viral contamination of commonly handled surfaces in a hospital setting was reported during the 2015 South Korea outbreak [34, 35]. The importance of fomite transmission compared with other transmission routes in nosocomial outbreaks of MERS-CoV remains uncertain but could have important implications for infection prevention and control guidance.

Our investigation had several limitations. The seroepidemiologic study was conducted 6–8 months after the outbreak, and seropositive individuals having antibody titers wane below detectable levels might have been missed. Additionally, we were conservative in the interpretation of indeterminate serology results as this result could represent cross-reactivity or a partial or waning immune response. We did not capture all health care and household contacts in our seroepidemiologic investigation, and we are unable to assess whether those captured were similar to those not captured. An overall limitation of the MERS-CoV outbreak investigation is that RT-PCR testing of contacts might miss cases depending on the timing of nasopharyngeal swab swab collection. The Sanger sequencing method used in our study may not have detected sequence variants (small deletions or insertions or single base polymorphisms) present at low levels in the samples and therefore likely underestimates the complexity of the virus populations present.

In conclusion, we found that the epidemiology of a MERS-CoV deletion variant did not appear to deviate substantially from previous MERS-CoV outbreaks that did not exhibit these mutations. Our investigation revealed that the MERS cases and clusters detected in multiple hospitals were all linked to a common transmission chain with a single introduction event. The juxtaposition of molecular, serologic, and epidemiologic data revealed that the ORF4a genetic deletion and additional deletions observed over the course of the outbreak were epidemiologically clustered, yielding an important approach for using molecular information to support epidemiological findings and strengthen our understanding of MERS-CoV transmission. Close monitoring of MERS-CoV genomic variation [19] and the epidemiology of infection remains critical amidst continued reporting of sporadic cases and outbreaks of MERS-CoV on the Arabian Peninsula.

Supplementary Data
Supplementary materials are available at Open Forum Infectious Diseases online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

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Disclaimer. The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the US Centers for Disease Control and Prevention.

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