CLINICAL OVERVIEW

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Quality of probiotic products for preterm infants: Contamination and missing strains

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Abstract

Probiotics are effective in reducing necrotising enterocolitis in preterm infants, but routine use is not generally adopted. We describe a safety issue concerning contamination by pathogenic bacteria and missing of labelled strains in a probiotic product widely used in neonatal care. We recommend all centres using probiotics in the care of vulnerable patients to consider product safety checks in addition to the quality reports of manufacturers. Meanwhile, clinicians and manufacturers should collaborate to define quality standards for probiotic products in clinical settings as long as specific international regulations are lacking.

KEYWORDS
contamination, Infloran, necrotising enterocolitis, probiotics, safety

Probiotics can prevent necrotizing enterocolitis (NEC) in preterm infants, reducing the incidence by nearly half in meta-analysis of trials including over 10 000 infants.[1] Still, probiotic use is not generally adopted in all neonatal intensive care units (NICUs), varying from 14% in the United States to 58% in Germany.[2,3] Why is a cost-effective therapy not generally adopted by all neonatal centres worldwide? First, evidence is still lacking in the most optimal treatment strategy regarding the strains and dose.[4] Second, availability of products actually tested for efficacy in clinical trials is limited. Third, reporting of safety data is often lacking or inadequate.[5] One concern is the risk of probiotic sepsis, which has been reported in several preterm infant cases.[6] However, this complication seems to be very rare, not always corresponds to the probiotics in the product, and occurs mainly in infants with additional risk factors compromising gut integrity or in association with congenital anomalies.[3,7-9] On the other hand, probiotic bacteriaemia may have been overlooked in other cases, because anaerobic strains can be difficult to culture from standard single paediatric culture bottles. Fourth, there may be local regulatory issues on a complex product being marketed as either a drug or a food additive with or without medical purpose.[10] For example, after 2009, we faced stringent additional conditions of the Health Inspectorate, after a Dutch probiotic trial in adults with acute pancreatitis showed

increased mortality in the intervention group.[11] Here, we report our experience with another safety issue which has led to ceasing of probiotic administration in our level III NICU: product contamination and missing of labelled strains.

Since 2012, approximately 900 preterm infants were treated with routine probiotics for the prevention of NEC in our level III NICU of the Erasmus MC in Rotterdam. According to local protocol, we treated all infants with a gestational age below 32 weeks or birthweight below 1500 grams starting from day one.[12] We used Infloran® 250 mg capsules, containing a combination of 10^9 CFU *Lactobacillus acidophilus* and 10^9 CFU *Bifidobacterium bifidum* (SIT Laboratorio Farmaceutico, Mede, Italy), which had shown NEC reduction in a double-blind trial.[13] We strictly followed all storage and handling procedures. During the treatment period, we observed a significant decline in the incidence of NEC from 7.8% to 5.1% (OR 0.63 (95% CI 0.42-0.93), P = .02). The NEC reduction was not significantly associated with the introduction of probiotics after adjustment for potential confounders including the interaction with type of feeding (only breast milk, mixed or only formula) and the underlying downtrend in NEC incidence.[12] Because of increasing concerns on the quality and safety of probiotics products used in clinical practice, we did routine quality checks on the product content.

We tested 15 capsules from five different strips, originating from three different lots. The expiry dates ranged from 3-13 months after testing. To detect both aerobic and anaerobic bacteria, fungi and yeasts, we cultured the content on 4 different agar plates for 5 days, according to the following procedures. The full content of probiotic powder of each tested capsule was dissolved in 1 mL BHI broth with Vitamin B6. Then, 10 μL was plated on TSA blood agar (BA), Chocolate agar (CHOC), Mac Conkey agar (McC) and Hemin and Vitamin K1 enriched Brucella Blood Agar plate (BBA).[14] Simultaneously, sterility control of the BHI-VitB6 was set. The BA, CHOC and McC were incubated for 5 days at 35°C and 5% CO2, and the BBA was incubated 5 days anaerobically at 35°C. All grown bacteria were identified by MALDI-TOF (Bruker). In only 1 lot purchased, both labelled probiotic microorganisms were detectable in all capsules. In the other two lots, one or both strains were missing and varying contamination was found, with *Streptococcus oralis*, *Lactococcus garvieae*, *Enterococcus feacalis*, *Enterococcus faecium* and *Lactococcus lactis* (Table 1).

The level III NICU of the Emma Children's Hospital in Amsterdam was about to start with probiotics in preterm infants and independently purchased the same product. As they also found contamination with *S oralis* in lot 4583A, we consider contamination after purchase highly unlikely.

Unfortunately, we were not able to perform additional validation or quantitative PCR, because the disapproved lots were directly returned to the manufacturer according to hospital regulations. By plating 1/100 of dissolved full capsules (that are supposed to contain 10^9 CFU of each labelled probiotic microorganisms), one would ideally expect to culture 10^7 CFUs of each probiotic strain. Based on this estimated detection level, we can assume that the capsules without detectable probiotics contained <100 CFU per capsule.

Even without knowing the exact quantity of contaminants, we question the safety of the contaminated products. The possible clinical consequences of a contaminated probiotic product have dramatically been shown in 2014, by a fatal gastrointestinal mucormycosis

in a preterm infant.[15] We could only find two other reports on contaminated products as a cause of infant sepsis, of whom both have recovered well.[16,17] Pathogenicity of the contaminating microorganisms is difficult to rule out in preterm infants whose gastrointestinal barrier function and immune system are still compromised. Even apparently commensal skin and gastrointestinal bacteria frequently cause sepsis in very preterm infants.[18,19] We detected *L lactis*, which may be harmless as natural exposure is likely common for its widespread use in cheese industry without reports on pathogenicity.[20] *L garviae* is associated with outbreaks of infections in fish. It is a rare cause of infective endocarditis in adults with valvular disease after ingestion of raw seafood, with no reports on infection in children.[21] The human commensals *Streptococcus oralis* (including *Streptococcus pneumoniae)* and *Enterococcus feacalis* and *faecium* are likely of most concern, as these can cause neonatal sepsis in preterm infants.[18]

The tested probiotic product is produced according to general European drug regulations that lack special regulations for probiotic products.[22] In accordance to this and other international Pharmacopeas, microbiological quality of 'non-sterile drugs' is deemed acceptable if *Escherichia coli*, *Staphylococcus aureus* and *Salmonella* species are absent, and if total counts of bile-tolerant Gram-negative bacteria, aerobic microbes and yeast or moulds are below predefined levels of colony forming units (CFU) per gram or mL.[23,24] We suggest that these regulations may not be strict enough when it comes to safety of probiotic use in preterm infants and that redefinition of so-called 'objectionable organisms' for this vulnerable population is needed. From a clinical point of view, an ideal product would contain only the labelled probiotic strains, in an acceptable quantity during shelf life with no contamination of any other microorganism at all.

In response to our report, the manufacturer retested the products and confirmed contamination of lot 4583A and 5145A (personal communication). They did not report on the presence or absence of the labelled probiotic strains in the product. As the product was still considered safe according to the regulations, no recall action was taken by the manufacturer. Of the three lots tested, we only used the uncontaminated lot (4307B) in patient care. When products with missing strains have been administered earlier, this may have been for no benefit and may explain the lack of effect on NEC in our time-series analyses.[12] We have stopped administration of probiotics in our unit until we receive products that pass our own safety and viability checks.

This report has several limitations, as it is just a description of our local clinical findings on a single product, and not part of a prospective quality control project. In retrospect, we should have quantified the contaminants by plate count or qPCR. Considering the sensitive methods used, bacterial enumeration of the contaminants may have been low, which can limit the clinical relevance. Next, additional testing for non-viable probiotic bacteria could have helped to differentiate between issues in the production process versus limited viability during shelf life. However, the product label claims *live* probiotics, by declaring the quantitative amount as CFU per capsule. Viability and purity testing would ideally be done along the chain, from production, storage and shipping to the hospital, pharmacy and patient ward, to

TABLE 1 Microorganisms identified in capsules with label claiming *Lactobacillus acidophilu*s and *Bifidobacterium bifidum*, by MALDI-TOF. Shown are percentages of capsules of the specified lot tested positive for mentioned microorganism

Lot	4307B	4583A	5145A
Capsules tested	N = 4	N = 4	N = 7
Label claimed			
Lactobacillus acidophilus	100%	100%	0%
Bifidobacterium bifidum	100%	75%	0%
Contaminants			
Streptococcus oralis	–	100%	–
Lactococcus garvieae	–	–	100%
Enterococcus feacalis	–	–	57%
Enterococcus faecium	–	–	43%
Lactococcus lactis	–	–	29%

reveal critical steps in the process. However, we consider quality control the responsibility of the manufacturer and regulatory agencies and we lack funding to study this in more detail. How our findings relate to other products and other settings are unclear. Of note, we have no data on the product widely used in Germany, with the same brand name but different content (*Bifidobacterium infantis* instead of *Bifidobacterium bifidum)* or on any other products. However, our findings are not unique. Earlier reports have shown that mismatches between the content and the label claims are common in commercial probiotic products. Reported composition mismatches included incorrect taxonomy, presence of un-labelled species or missing of the labelled (sub) species. Similar to our findings, these content biases were found with pill-to-pill (within a lot) and lot-to-lot variation.[25,26] Another possible shortcoming is that we have no access to detailed quality control findings of the manufacturer. The main strengths of this report include our methods to detect a wide range of potential contaminants. Methods were based on international standards with additional state-of-the-art culture and detection procedures, run in a ISO 15189 certified laboratory. The contamination by *S oralis* was confirmed by an independent laboratory, and the positive findings in the first lot indicate sufficient sensitivity to detect the probiotics.

We aim to call attention to probiotic product quality, as these can threat patient safety in at-risk populations, but may also lead to bias and misinterpretation of clinical trial results. This underlines the need for independent quality control, and for rapid and accurate molecular techniques for identification of (sub) species, such as quantitative real-time PCR, flow cytometry and MALDI-TOF Mass Spectrometry.[24,27] However, also by simple culture techniques, contamination of products may be found.

In conclusion, despite the mass of current literature showing that probiotics can prevent NEC and save many lives, general adoption of probiotic use is hampered by different issues. This report illustrates that quality issues of the products currently available are one of them. We are reluctant to withhold patients a highly effective treatment, but we are also worried about the potential risks of impure products. We understand that quality control of unsterile products is complex and that 100% pure products are difficult to guarantee. As long as regulatory requirements may not be strict enough, we recommend all centres using probiotics in the care of vulnerable patients to consider routine bacterial checks in addition to quality checks of manufacturers to guarantee product safety. The same warning holds true for ongoing and future clinical trials. Meanwhile, clinicians, microbiologists and manufacturers should collaborate to define population-specific quality standards for probiotic products in clinical settings as long as specific international regulations are lacking for at-risk populations.

CONFLICT OF INTEREST

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