

Increased Mortality and Vascular Phenotype in a Knock-In Mouse Model of Retinal Vasculopathy With Cerebral Leukoencephalopathy and Systemic Manifestations

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Background and Purpose—Retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations (RVCL-S) is an autosomal dominant small vessel disease caused by C-terminal frameshift mutations in the *TREX1* gene that encodes the major mammalian 3' to 5' DNA exonuclease. RVCL-S is characterized by vasculopathy, especially in densely vascularized organs, progressive retinopathy, cerebral microvascular disease, white matter lesions, and migraine, but the underlying mechanisms are unknown.

Methods—Homozygous transgenic RVCL-S knock-in mice expressing a truncated Trex1 (three prime repair exonuclease 1) protein (similar to what is seen in patients) and wild-type littermates, of various age groups, were subjected to (1) a survival analysis, (2) in vivo postocclusive reactive hyperemia and ex vivo Mulvany myograph studies to characterize the microvascular and macrovascular reactivity, and (3) experimental stroke after transient middle cerebral artery occlusion with neurological deficit assessment.

Results—The mutant mice show increased mortality starting at midlife ($P=0.03$ with hazard ratio, 3.14 [95% CI, 1.05–9.39]). The mutants also show a vascular phenotype as evidenced by attenuated postocclusive reactive hyperemia responses (across all age groups; $F[1, 65]=5.7$, $P=0.02$) and lower acetylcholine-induced relaxations in aortae (in 20- to 24-month-old mice; RVCL-S knock-in: E_{\max} : $37\pm 8\%$ versus WT: E_{\max} : $65\pm 6\%$, $P=0.01$). A vascular phenotype is also suggested by the increased infarct volume seen in 12- to 14-month-old mutant mice at 24 hours after infarct onset (RVCL-S knock-in: 75.4 ± 2.7 mm³ versus WT: 52.9 ± 5.6 mm³, $P=0.01$).

Conclusions—Homozygous RVCL-S knock-in mice show increased mortality, signs of abnormal vascular function, and increased sensitivity to experimental stroke and can be instrumental to investigate the pathology seen in patients with RVCL-S.

Visual Overview—An online [visual overview](#) is available for this article. (*Stroke*. 2020;51:300-307. DOI: 10.1161/STROKEAHA.119.025176.)

Key Words: ischemic stroke ■ magnetic resonance imaging ■ vasculature

Retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations (RVCL-S) is an autosomal dominant disorder caused by C-terminal frameshift mutations in the *TREX1* gene,¹ which encodes the ubiquitously expressed Trex1 (three prime repair exonuclease 1) protein.^{1,2} The mechanisms underlying the cause of RVCL-S, however, are unknown. RVCL-S is a systemic progressive small vessel disease with middle-age severe symptomatology and decreased life expectancy, characterized by vasculopathy in various organs, most pronounced in retina, kidney, and brain, with white matter lesions and intracerebral mass lesions.³⁻⁶

Patients with RVCL-S report more often migraine (generally without aura) and have increased risk for ischemic stroke, both suggesting vascular involvement. Pathological studies revealed thicker endothelial cells with increased vesicles and coarse cytoplasm, and thicker, multi-laminated, basement membranes of endothelial cells.⁵ Hence it has been hypothesized that the endothelium of small vessels, particularly in highly vascularized organs, is affected in RVCL-S. Endothelial dysfunction (impaired endothelium-dependent vasodilation and endothelial cell damage),⁷ which has been proposed as disease mechanism for the vascular phenotype

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seen in stroke and migraine, was also shown to underlie RVCL-S in patients.⁴

Although patients with RVCL-S are heterozygous for the *TREX1* mutation, here we investigated transgenic homozygous RVCL-S knock-in (KI) mutant mice that express a truncated Trex1 protein (similar to what is seen in patients with RVCL-S).^{1,5} The mutant mice express a truncated mouse protein (with the endogenous regulation of gene expression still intact), which is slightly different from the existing humanized RVCL-S mouse model in which the human cDNA bearing the same V235fs mutation was introduced in exon 2 of *TREX1*.⁸ Here we assessed whether homozygous RVCL-S KI mice have pathological features seen in patients with RVCL-S, such as a reduced life expectancy and a vascular phenotype (as assessed by functional vascular measurements and induction of experimental stroke).

Methods

The data that support the findings of this study are available from the corresponding author on reasonable request.

Experimental Animals

Homozygous transgenic RVCL-S KI and wild-type (WT) littermate mice of different age groups (3–6, 12–14, and 20–24 months) were used. As the heterozygous mutants show a normal survival (Figure 1), we focused on a comparison between the homozygous mutant and WT mice. For the generation of the mutant mice, a targeting construct was used by which a frameshift mutation was introduced at amino acid position 235 of the Trex1 protein, resulting in a truncated protein as observed with the most prevalent RVCL-S mutation in patients (Methods and Figure I in the [online-only Data Supplement](#)).^{1,5} Homozygous mutant and WT mice were generated by interbreeding heterozygous mice. For the stroke experiments, only male mice were used. For the in vivo and in vitro vascular characterization, results of both sexes were pooled because no sex difference was detected. Animals were randomized (flip of a coin) for surgical procedure, and researchers were blinded for genotype during experimental and analyses procedures. Animals were housed in a temperature-controlled environment with food and water ad libitum. All animal experiments were approved by the local committee for animal health, ethics and research of the Leiden University Medical Center.

Survival Analysis

Survival in naive WT and homozygous RVCL-S KI mice was assessed in animals who, at 2 months, were randomly assigned to one of the age groups (3–6, 12–14, and 20–24 months) to be included in the experimental stroke studies (n=97). Mice were censored for analysis when they reached the age of inclusion for the stroke experiment. In addition, data of 51 heterozygous animals, which were not used in experiments, were added to the survival analysis.

In Vivo Microvascular Characterization

Microvascular characteristics were assessed in vivo using postocclusive reactive hyperemia (PORH) measurements of the hind leg with laser Doppler flowmetry (PeriFlux System 5000, Perimed, Järfälla-Stockholm, Sweden). Twenty-four hours before dermal blood flow measurements, the hair of the left hind leg was removed with hair removal cream (Veet, ReckittBenckiser, Inc, Berkshire, United Kingdom). On the day of measurement, mice were anesthetized using 4% isoflurane/O₂ ventilation and kept on a heating pad regulated by a rectal thermometer (FHC, Inc, Bowdoin, ME) to maintain body temperature at 36.7±0.3°C. After a 5-minute equilibration time period, 10 minutes of baseline perfusion was recorded. Subsequently, the hind

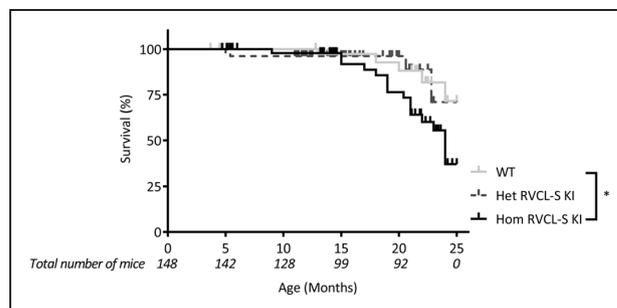


Figure 1. Survival curves from wild-type (WT), heterozygous (Het), and homozygous (Hom) retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations (RVCL-S) knock-in (KI) male mice. At 2 mo, Hom mutant and WT mice were randomly assigned to one of the 3 age groups for experimental stroke surgery. Mice reaching the age of surgical inclusion were censored from the analysis. WT (n=45, gray line), Het RVCL-S KI (n=51, black dotted line), and Hom RVCL-S KI mice (n=52, black solid line). Vertical bars along curves represent censored animals. WT and Het survival curves were not significantly different. * $P < 0.05$ indicates statistical significance between WT and Hom RVCL-S KI mice.

leg circulation was occluded for 2 minutes with a tourniquet. After release of the tourniquet, blood flow was monitored until return to baseline blood flow (maximally 10 minutes). The area under the curve and PORH peak were used as readout measures. After the experimental procedure, mice were euthanized using decapitation, where the aorta was collected for in vitro measurements.

In Vitro Macrovascular Characterization

Macrovascular characterization was performed in vitro by mounting segments of the mouse aorta (inner diameter 0.5–1 mm) in Mulvany myographs with separated 6-mL organ baths⁹ containing carbogenated (5% CO₂ and 95% O₂) Krebs-Henseleit buffer solution (in mM: NaCl 118, KCl 4.7, CaCl₂ 2.5, MgSO₄ 1.2, H₂PO₄ 1.2, NaHCO₃ 25 and glucose 8.3; pH 7.4) at 37°C. Tension was normalized to 90% of the estimated diameter at 100 mmHg of transmural pressure. A reference contractile response of vessel segments was determined with 100 mmol/L KCl. For the acetylcholine and sodium nitroprusside (SNP) concentration-response curves, aorta segments were precontracted with the thromboxane A₂ analog U46619 (10–100 nmol/L). Relaxant effects of acetylcholine (endothelium-dependent) and SNP (endothelium-independent) and contractile effects of 5-hydroxytryptamine were examined by cumulative application of increasing concentrations of drug.

Transient Middle Cerebral Artery Occlusion Model

Mice were anesthetized using isoflurane (3% induction, 1.5% maintenance) in 70% pressurized air and 30% O₂. Carprofen 5 mg/kg, SC (Carproal, 50 mg/mL; AST Farma B.V., Oudewater, the Netherlands) was administered before surgery for pain relief. During surgery, the mouse body temperature was maintained at 36.7±0.3°C as described above. Experimental stroke was induced using the transient middle cerebral artery occlusion (tMCAO) model.¹⁰ A silicone-coated nylon monofilament (7017PK5Re; Docol cooperation, Sharon, MA) was introduced into the internal carotid artery, via an incision in the common carotid artery, to block the middle cerebral artery (MCA) at its origin for 30 minutes. Cerebral blood flow in the MCA territory was measured using laser Doppler flowmetry (PeriFlux System 5000; Perimed) and calculated as %-measure of baseline. In a subset of animals, tcCO₂ was measured (TCM4 CombiM monitor with accompanying software version 3.0; Radiometer Medical ApS, Brønshøj, Denmark) to confirm physiological stability during surgery (data not shown). After surgery, the animal was placed in a temperature-controlled recovery incubator (V1200; Peco Services Ltd, Brough, United Kingdom) maintained at 33°C for 2 hours, with easy access to food and water. Postprocedural observation was performed twice daily.

Magnetic Resonance Imaging

Animals were scanned using a small animal 7T MRI system (Pharmascan, Bruker BioSpin, Ettlingen, Germany) under isoflurane anesthesia. A Multi Slice Multi Echo sequence protocol was run with repetition time/echo time of 4.000 ms/9 ms, 20 echoes, 2 averages, matrix 128×128, field of view of 2.50 cm, bandwidth 59 523.8 Hz, slice thickness of 0.5 mm, and 16 slices (no gap). Quantitative T2 maps were calculated using Paravision 5.1 software (Bruker BioSpin). Scans were performed at 4, 24, and 48 hours after *t*MCAO surgery. An in-house developed automated software pipeline¹¹ was used to calculate infarct sizes. If automated segmentation showed artifacts, for example, due to wrapping, small manual corrections were made, or scans were excluded due to technical failure.

Behavior Analysis

Behavior analyses were performed directly before magnetic resonance imaging measurements, that is, before and after (at 4, 24, 48 hours, and 5 days) *t*MCAO surgery. During behavioral tests, mice were videotaped, and blinded videos were analyzed after completion of the experiments. Neurological deficit scores were measured on a 56-point scale.^{12,13} The score involved general and focal deficits, where 0 reflected no deficits, and 56 reflected the poorest. Categories concerning general appearance and performance included fur (0–2), ears (0–2), eyes (0–4), posture (0–4), spontaneous activity (0–4), and epileptic behavior (0–12). Concerning the focal deficits a score was stated for body asymmetry (0–4), gait (0–4), climbing on a surface inclined at 45° (0–4), circling behavior (0–4), front-limb symmetry (0–4), compulsory circling (0–4), and whisker response to light touch (0–4).

Vessel Anatomy

A subset of mice was used for vessel anatomy analysis.^{14,15} In brief, mice were euthanized using CO₂ and transcardially perfused using 2.5 mL PBS with 50 µL heparin (Heparine natrium 5000 I.U./mL; LEO Pharma B.V., Amsterdam, the Netherlands) and 2.5 mL ink mixture (ratio 1:9, Herlitz stempelfarbe ink and Pelikan Scribto ink; Pelikan Vertriebsgesellschaft mbH & Co. KG, Hannover, Germany) at a rate of 3 mL/min, preheated to 37°C. Brains were removed, photographed, and post-fixed in fresh 4% paraformaldehyde. Vessel anatomy was scored as the number of anastomoses between MCA and anterior cerebral artery, distance between midline, and the anastomotic line at 2, 4, and 6 mm from anterior and diameter of the main vessels of the circle of Willis (MCA, anterior cerebral artery, internal carotid artery, posterior cerebral artery, basilar artery, and posterior communicating artery). Pial anastomoses were defined as the narrowest part of the vessel or halfway between the nearest branch points of the anterior and the MCA, localized by tracing the peripheral branches of these major vessels.

Statistical Analyses

All statistical analyses were performed in SPSS software (SPSS statistics 23, IBM Corporation, Armonk, NY). Survival analyses were performed using the log-rank (Mantel-Cox) test, and Cox univariable regression analyses were done to calculate the hazard ratio. Animals who reached the inclusion age of the experiment were censored in the analyses. The PORH results were expressed as relative values compared with baseline and included the area under the curve (AUC) and the maximal hyperemia response (E_{\max}), expressed as mean±SEM. Vascular relaxant responses to acetylcholine and SNP were expressed as the percentage of contraction induced by 10 to 100 mmol/L U46619. Data for the vascular responses to 5-hydroxytryptamine were expressed as percentage of the contractile response to 100 mmol/L KCl. Both the in vivo and in vitro data were analyzed using univariate ANOVA followed by Bonferroni post hoc tests. Outcome values were expressed as mean±SEM. Infarct volume and neurological deficit scores were compared between genotypes using marginal mixed model analyses (data are shown as estimated marginal means±SD). Survival analyses post-surgery were performed using the log-rank (Mantel-Cox) test. Group comparison concerning vessel diameter and anastomotic distance were analyzed using multivariate ANOVA with

discriminant function analysis post hoc test. Absence of the posterior communicating artery was analyzed using Pearson χ^2 (2-sided Fisher exact) test, and mean pial number of anastomoses were analyzed with a *t* test. $P<0.05$ indicates statistical significance.

Results

Increased Mortality in RVCL-S KI Mice

Homozygous RVCL-S KI mutant mice showed increased mortality compared with WT controls: 16 (31%) mutant versus 4 (9%) WT mice died during the observation period presurgery ($P=0.03$ with hazard ratio, 3.14 [95% CI, 1.05–9.39]). The increased mortality in the mutant mice compared to WT mice was apparent from 9 months of age (Figure 1). Heterozygous mutant mice did not show a reduced lifespan ($P=0.55$ with hazard ratio, 1.25 [95% CI, 0.60–2.59]; Figure 1). Therefore, only homozygous mutants were investigated in subsequent functional experiments. Postmortem examination, although sub-optimal because considerable organ deterioration had occurred at the time of tissue examination, did not reveal a macroscopic cause of death (data not shown). Postmortem examination also did not show signs of brain atrophy, neither in the heterozygous nor the homozygous mutants, but also here the analysis was not reliable due to organ deterioration (data not shown). Finally, basic histological hematoxylin and eosin staining analyses of various organs (eg, eye, kidney, skin, liver, brain) of the homozygous and heterozygous mutant mice did not reveal any obvious abnormalities (data not shown). In contrast with patients with RVCL-S,⁵ we observed no structural abnormalities in endothelial cells or the basement membrane of the mutant mice. It remains unclear whether this is because these disease features are not captured in the mouse model or whether these features need longer time to develop than the 2-year maximal age of a mouse.

Reduced In Vivo Microvascular Reactivity in RVCL-S KI Mice

Compared with WT, the RVCL-S KI mice showed a significantly lower area under the reactive hyperemia curve across all age groups ($F[1, 60]=5.1$, $P=0.03$; Figure 2A). The maximal reactive hyperemia responses, however, were comparable between genotypes (Figure 2B). Baseline dermal blood flow and time to maximum value were also similar between genotypes (data not shown).

Increased In Vitro Macrovascular Reactivity in RVCL-S KI Mice

No significantly different contraction reaction to 100 mmol/L KCl was seen between both genotypes. Instead, the concentration-response curve to acetylcholine showed statistically significantly attenuated relaxant responses in 20- to 24-month-old RVCL-S KI mice compared with WT mice (E_{\max} : 65±6% versus 37±8%, respectively, $P=0.01$), but not in mice of other age groups (Figure 2C). No genotypic difference was observed in the response to SNP or 5-hydroxytryptamine (Figure 2C).

Increased Susceptibility to Experimental Stroke With Worse Outcome in RVCL-S KI Mice

Overall, infarct volume was higher in RVCL-S KI compared with WT mice ($P=0.02$), with a significant effect of age

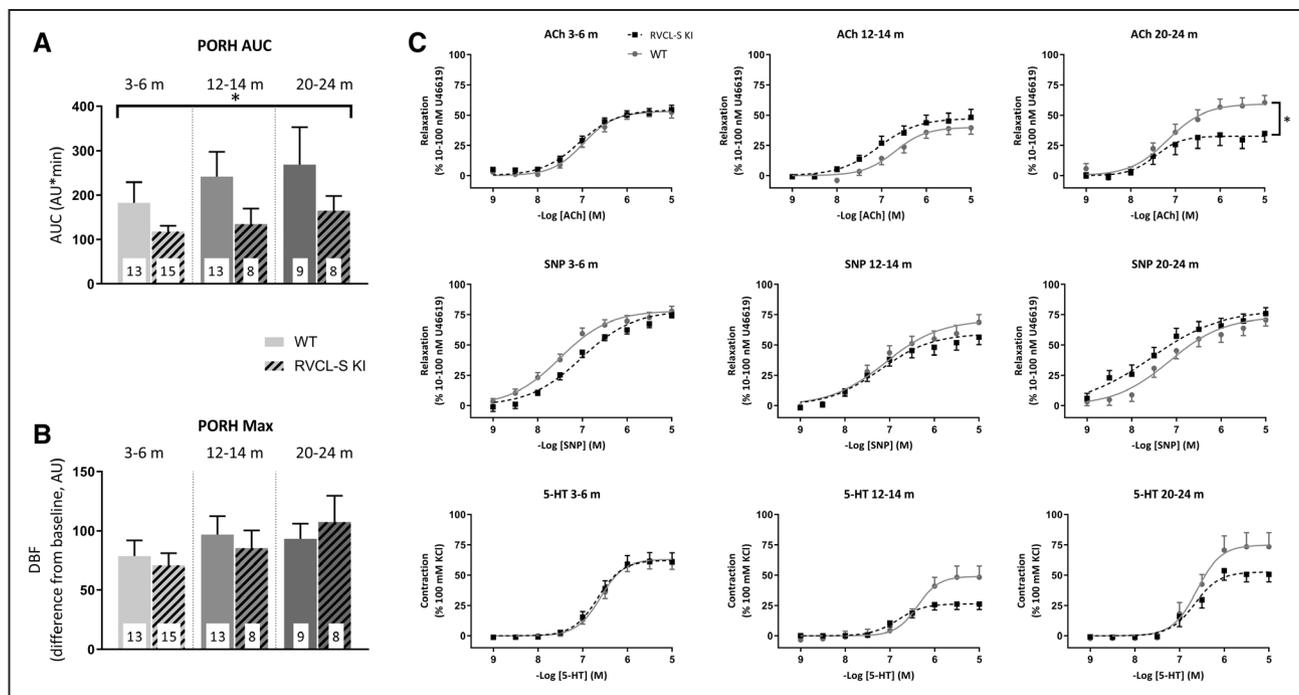


Figure 2. Microvascular and macrovascular characterization of wild-type (WT) and homozygous retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations (RVCL-S) knock-in (KI) mice. In vivo microvascular characterization using postocclusive reactive hyperemia (PORH) measurements of the hind leg with the area under the reactive hyperemia curve (A) and maximal reactive hyperemia (B) responses for WT and RVCL-S KI mice of the 3 age groups (3–6, 12–14, and 20–24 mo). * $P < 0.05$ indicates a statistical significant difference between RVCL-S KI and WT mice across all age groups. Number of animals is indicated within each bar. C, In vitro macrovascular characterization of isolated aortae obtained from WT (gray circles) and homozygous RVCL-S KI (black squares) mice. Panels depict concentration-response curves to acetylcholine (ACh, $n = 7–16$), sodium nitroprusside (SNP, $n = 7–17$), and serotonin (5-hydroxytryptamine [5-HT], $n = 5–15$). * $P < 0.05$ indicates statistical significance for attenuated relaxant responses to ACh in 20- to 24-mo-old RVCL-S KI mice compared with WT mice. AU indicates arbitrary units; AUC, area under the curve; DBF, dermal blood flow; M, molar; m, months; and Max, maximal.

($P = 0.02$) and time after surgery ($P < 0.001$). The effects were mainly driven by the increased infarct volume in RVCL-S KI mice at the age of 12 to 14 months (at 4 hours: RVCL-S KI: 60.4 ± 5.4 versus WT: 39.8 ± 4.8 mm³, $P = 0.007$; 24 hours: RVCL-S KI: 84.3 ± 9.5 versus WT: 52.9 ± 7.8 mm³, $P = 0.01$; 48 hours: RVCL-S KI: 114.0 ± 13.3 versus WT: 58.1 ± 10.7 mm³, $P = 0.03$; Figure 3). There was no significant genotypic difference in cerebral vessel anatomy (Figure II and Table I in the [online-only Data Supplement](#)). The overall neurological deficit score was worse in RVCL-S KI mice compared to WT for all age groups ($P = 0.002$; Figure 4A). This effect was mainly driven by the 12- to 14-month-old group. In that age group neurological deficit scores was worse in RVCL-S KI than WT after surgery for an extended period of time (at 4 hours: RVCL-S KI: 29.3 ± 3.4 versus WT: 11.1 ± 2.9 , $P < 0.001$; 24 hours: RVCL-S KI: 24.9 ± 3.4 versus WT: 9.9 ± 2.7 , $P = 0.001$; 48 hours: RVCL-S KI: 28.5 ± 3.8 versus WT: 10.0 ± 2.6 , $P < 0.001$; 5 days: 24.2 ± 3.7 versus 7.5 ± 2.4 , $P = 0.001$), whereas there was no difference before surgery (presurgery: RVCL-S KI: 0.6 ± 0.4 versus WT: 0.5 ± 0.2 , $P = 0.20$). Mortality within 5 days after stroke onset was also increased in the 12- to 14-month-old group (RVCL-S KI: 7 [8%] versus WT: 4 [25%], $P = 0.04$), an effect also present in the 20- to 24-month-old group (RVCL-S KI: 8 [57%] versus WT: 2 [13%], $P = 0.02$); the young age group only showed a nonsignificant trend (RVCL-S KI: 4 [57%] versus WT: 2 [33%]). There were no differences in basic parameters such as body weight (Table II in the [online-only Data Supplement](#)) or activity levels (determined from

presurgery analyses; Figure 4) between mutant and WT mice that could explain the difference in age-related survival.

Discussion

Here, we investigated whether RVCL-S KI mice, which express a truncated mouse Tbx1 protein that mimics the truncated protein seen in patients with the RVCL-S V235fs mutation,^{1,5} exhibit features in line with the phenotype of patients with RVCL-S. We demonstrated that homozygous RVCL-S KI mice have (1) a shorter life expectancy, (2) a vascular phenotype, as evidenced by abnormal PORH (microvascular characterization; in vivo in hind leg) and acetylcholine-induced relaxant (macrovascular characterization; in vitro in aortic segments) responses, and (3) an increased susceptibility to experimental stroke with a worsened outcome. Except for outcome measures of experimental stroke, the other features can be considered spontaneous phenotypes. Except for signs of abnormal microvascular responses that were already observed from a young age (3–6 months), all other abnormalities were only observed in old mice, that is, in mice of 12 to 14 months and 20 to 24 months. Our study in mice is relevant because it is not understood how vasculopathy, at middle-age, is brought about in patients with RVCL-S and how it is related to the lower life expectancy and increased risk for ischemic stroke.^{3–6} Similar to earlier observations in patients with RVCL-S, our data point to decreased functional endothelial responses in mice. In patients, this is exemplified by reduced flow-mediated dilatation responses,⁴ and in mice

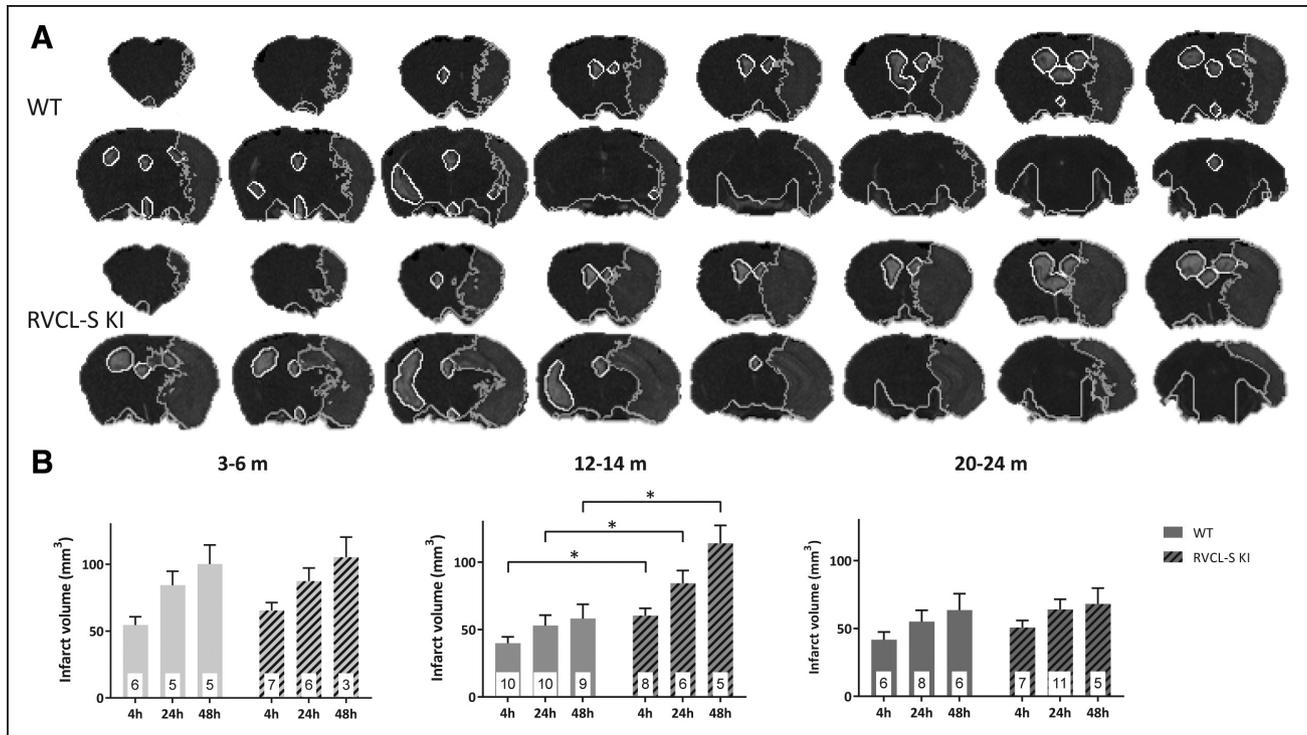


Figure 3. Infarct volume in wild-type (WT) and homozygous retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations (RVCL-S) knock-in (KI) male mice. **A**, Example traces of automated stroke segmentation in T_2 -weighted magnetic resonance imaging (MRI) images at 24 h after infarct induction in 12- to 14-month-old WT and RVCL-S KI mice. **B**, Stroke volumes at 4, 24 and 48 h after infarct induction in WT and RVCL-S KI mice of the 3 age groups (3–6, 12–14, and 20–24 mo; estimated marginal mean \pm SD), * P <0.05 indicates a statistically significant difference between 12- to 14-month-old RVCL-S KI and WT mice for each individual time point. Number of animals is indicated within each bar. Numbers can vary due to death of the animal or technical failure of the MRI scan. h indicates hours; and m, months.

by reduced blood flow after PORH across all investigated ages and by reduced macrovascular relaxant responses to the endothelium-dependent vasodilator acetylcholine.

RVCL-S KI Mice Have a Vascular Phenotype

This study is the first to show a vascular phenotype in RVCL-S KI mice. The results of the *in vivo* PORH experiments point towards a microvascular dysfunction, with a decreased AUC of the PORH but a normal maximal PORH peak. As the effect was seen in all age groups, it shows that a vascular phenotype is developing already before the mortality and increased stroke susceptibility become apparent. The PORH response¹⁶ represents an interplay between multiple vasodilation pathways, such as the endothelium-derived hyperpolarizing factor pathway¹⁷ (which is characterized by the AUC) and the release of neuropeptides from perivascular nerves during the hyperemia peak (which is characterized by the maximal hyperemia response [E_{max}]).¹⁸ Notably, *in vitro* examinations of macrovasculature showed abnormal endothelial function in response to acetylcholine but only in very old RVCL-S KI mice. The decreased relaxant response to acetylcholine seems because of diminished endothelial function, as responses to the endothelium-independent NO donor SNP were normal in RVCL-S KI mice of the same age category. As mentioned above, the *in vivo* PORH AUC data suggest involvement of an affected endothelium-derived hyperpolarizing factor pathway response. Endothelium-derived hyperpolarizing factor acts as vasorelaxant and is known to act in microvessels rather than in larger vessels.^{19,20} Thus, the

results of our *in vitro* macrovascular studies are not in contrast with the microvascular PORH results. Testing the natural ligand 5-hydroxytryptamine²¹ showed no abnormal vasoconstrictive response in RVCL-S KI mice, although concentration-response curves tended to be lower in the 12- to 14- and 20- to 24-month-old groups compared with WT mice.

Increased Ischemic Vulnerability in RVCL-S KI Mice

The vascular dysfunction seen in RVCL-S KI mice may be the underlying cause of increased stroke risk in patients with RVCL-S. Therefore, we investigated whether RVCL-S KI mice had a worse stroke outcome. Although the infarct area is still evolving days to weeks after *tMCAO*, we used 48 hours as final magnetic resonance imaging readout point since infarct size on T_2 -weighted magnetic resonance imaging is maximized at 48 hours post-*tMCAO*.²² An overall increased infarct volume was observed in RVCL-S KI mice, compared with WT, albeit driven by the 12- to 14-month-old group. Given the decreased survival in the very old mutant mice, one can envisage that no genotypic difference in infarct volume was seen in the 20- to 24-month-old group because mutant mice that are most affected had already died before reaching the age of inclusion. Endothelial dysfunction was shown to lead to a procoagulatory, proinflammatory and proliferative state, which predisposes to atherosclerosis and increases stroke risk,⁷ but it remains to be researched whether this mechanism explains the increased infarct size in RVCL-S KI mice. In

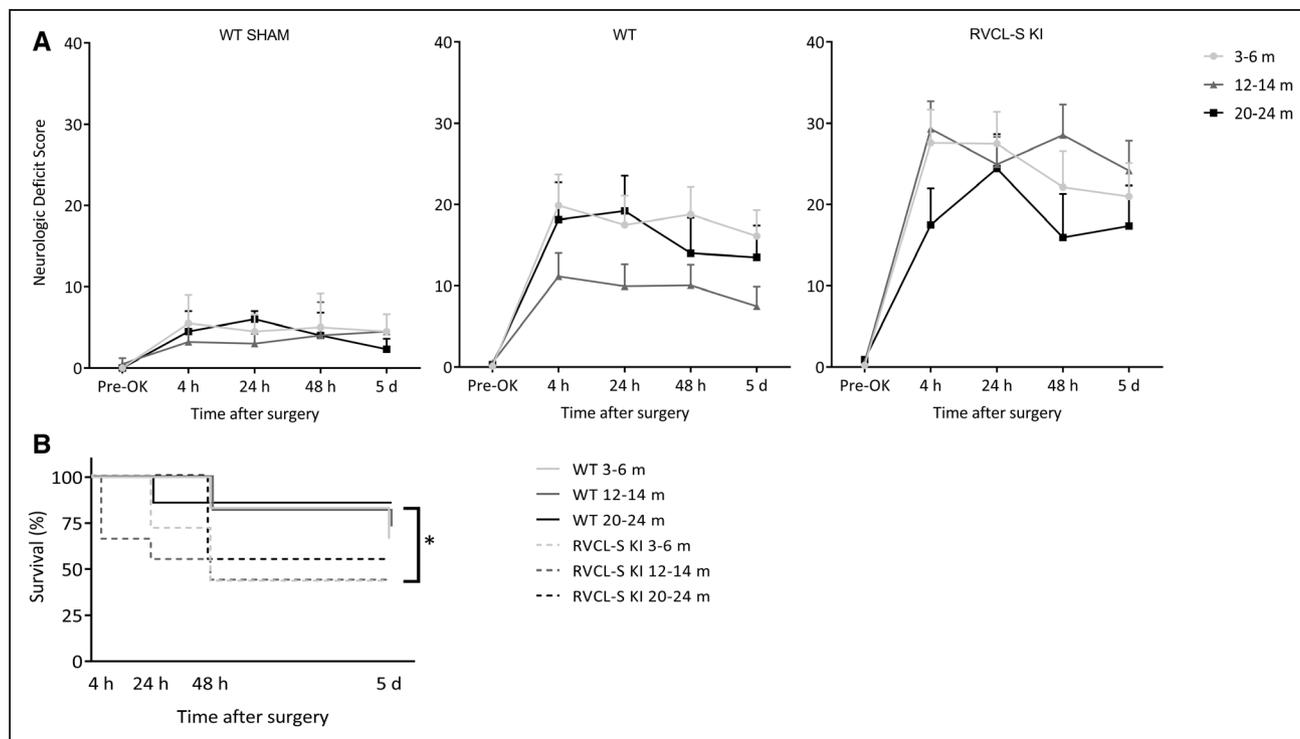


Figure 4. Behavior analysis after infarct induction in wild-type (WT) and homozygous retinal vasculopathy with cerebral leukoencephalopathy and systemic manifestations (RVCL-S) knock-in (KI) male mice. **A**, Neurological deficit score (NDS; estimated marginal mean \pm SD) for WT SHAM surgery and WT and RVCL-S KI post-transient middle cerebral artery occlusion (tMCAO) and **B** survival curves at 4, 24, 48 h and 5 days after tMCAO or SHAM surgery in WT (continuous line) and RVCL-S KI mice (discontinuous line) of the 3 age groups (3–6, 12–14, and 20–24 mo). * $P < 0.05$ indicates a statistically significant difference **(A)** between 12- to 14-mo-old RVCL-S KI and WT mice for NDS and **(B)** between RVCL-S KI and WT mice concerning survival post-surgery. d indicates days; h, hours; and m, months.

future experiments, this should be investigated using electron microscopy for in-depth morphological examination to uncover the possible presence of subtle lesions of the brain endothelium. There remains the, in our view unlikely, possibility that thrombus formation in the vessel after tMCAO may have influenced infarct outcome, as we have no a priori reason to assume that thrombus formation would be different between mutant and WT mice; although this, admittedly, has not been demonstrated, which would require the use of different methodology than used in the present study.

There are other striking differences for the various age groups when considering infarct size and outcome. Not finding a genotypic effect in young mice may be a reflection that the pathology has not advanced enough, although in vivo microvascular reactivity was already abnormal at that age. As alternative explanation, a genotypic effect may be masked by the seemingly unusually large stroke volume in the respective WT mice, as volumes in WT mice from that age analyzed in our lab usually are 5% to 20% smaller (previously published data¹¹).

It is even more surprising that mutant mice of 20–24 months have a worse outcome after tMCAO but no increased infarct size. One plausible explanation may be inclusion bias. Because animals with large infarcts have a higher a priori chance of dying within 48 hours from infarct induction, mice that do survive tend to have smaller infarcts. Apparently, the mutant mice that could be included still exhibit mutation-associated pathology, as reflected by the increased mortality.

Such underlying pathology may also explain why a total of 50% of the 20- to 24-month-old RVCL-S KI mice (compared with 15% of the WT group; Figure 1) had already died before scheduled surgery. Of relevance, WT aged male mice (and rats) have smaller infarct volumes,²³ but worse functional outcome,^{24,25} when compared with young animals. It remains enigmatic why aged male mice have a decreased infarct size.^{26,27} Aged vessels may have undergone morphological and pathological changes,²⁸ such as structural alterations in the vessel wall, atherosclerosis and vessel wall calcification,^{29,30} small vessel disease,³¹ and a reduced impairment of poststroke synaptic plasticity.^{27,32}

RVCL-S Phenotype in Mice Versus Humans

Although the truncated protein product in our KI mouse is very similar to the human canonical TREX1 frameshift product, one always needs to be cautious when extrapolating data from transgenic mouse models to human disease. Although RVCL-S in humans is an autosomal dominant disorder, the presence of 1 mutated allele causes disease; in our experiments, we focused on homozygotes mice instead to increase chances of capturing disease aspects by doubling the genetic load. It is not uncommon to focus investigations on homozygous mutant mice first as the lifespan in mice (maximal ~2 years) is much shorter than in humans, so a mutation has less time to exert its effect.

In humans, RVCL-S is characterized by various structural abnormalities, foremost the vasculopathy of the retina and

other organs, microinfarcts, and white matter lesions.^{5,6} A basic histological analysis of the eye and other organs in our mutant mice did not reveal such abnormalities, even not in mice that were far older than 12 months. This can be regarded as a shortcoming of our model, but this disease feature may not be captured because of the shorter lifespan of a mouse. Not finding histological abnormalities in our mutant mice is in accordance with findings in a previously described RVCL-S mouse model,^{8,33} that also expresses V235fs-truncated, albeit human, Trex1 protein. In that model, DNase activity, a hallmark of Trex1, was maintained in truncated protein.³³ Both heterozygous and homozygous mutants were reported to be grossly normal with no signs of tissue inflammation; the mutant mice did exhibit striking elevations of autoantibodies against non-nuclear antigens in the serum.³³ Of note, the authors did not report the decreased survival phenotype, likely because they did not investigate very old mice. We would consider both RVCL-S mouse models equally relevant and having multiple transgenic models will allow replication in separate models.

RVCL-S is considered a small vessel disease.⁵ Abnormal functionality of the microvasculature, exemplified by the abnormal PORH response, was observed in mutant mice already of young age. Moreover, in the stroke experiments, outcome (as in human ischemic stroke) is influenced by the status of the microvasculature bed. Both types of experiments show that abnormal functionality of the microvasculature is captured in our mouse model. We recently found that functional endothelial responses were decreased in patients with RVCL-S, while microvascular responses to capsaicin were normal (analyzed using flow-mediated vasodilatation).⁴ This points to endothelial dysfunction in RVCL-S, which is in line with our microvascular and macrovascular observations in the homozygous mutant mice. Although flow-mediated vasodilatation responses are generally considered as a measure for macrovascular function, it should be kept in mind that flow-mediated vasodilatation is a physiological response to the increased shear stress during reactive hyperemia and is thus critically dependent on the reactivity of the downstream microvasculature to transient ischemia.

Limitations

As discussed above, our study has several limitations. First, we used homozygous RVCL-S KI mice, although RVCL-S in humans is caused by the presence of only 1 mutated allele, and therefore, translation of the results should be done with caution. Second, several pathological features seen in RVCL-S in humans (ie, vasculopathy of the retina and other organs, microinfarcts, and white matter lesions) were not seen during basic examination in our model. In future experiments, a more in-depth analysis, for example, using electron microscopy, should be done to address this point. Furthermore, because of study design, we used surviving older animals, which probably have a less severe aging and disease phenotype. It remains an enigma why experimental stroke outcomes in the oldest animals are less severe.

In conclusion, we found that the homozygous RVCL-S KI mice exhibit early mortality in line with the reduced life expectancy seen in (heterozygous) patients with RVCL-S.

Moreover, we observed a blunted (cerebral) microvascular response suggestive of endothelial involvement, which seems to occur already in young mutant mice, as well as an abnormal macrovascular response in old mutants. Finally, we observed an increased infarct size and worse outcome in homozygous RVCL-S KI mice in response to experimentally induced stroke, but it is at present unclear how this relates to the phenotype in patients with RVCL-S. Regardless, our RVCL-S KI mouse model seems very promising to help unravel the pathophysiology of RVCL-S.

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