Mechanisms of putative IGF-I receptor resistance in active acromegaly

Joseph A.M.J.L. Janssen

Department of Internal Medicine, Division of Endocrinology, Erasmus MC, Rotterdam, the Netherlands

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ABSTRACT

Acromegaly is a disease characterized by overproduction of growth hormone (GH). As a consequence of excessive GH secretion, circulating insulin-like growth factor-I (IGF-I) is elevated in active (untreated) acromegaly. IGF-I is often used as a marker of disease activity and growth hormone status in acromegaly. Although IGF-I can directly improve insulin sensitivity and glucose uptake in muscles, the excessive GH secretion in active acromegaly frequently leads to insulin resistance, glucose intolerance and even diabetes. In this review, evidence will be discussed that in active acromegaly chronically elevated IGF-I, insulin and soluble Klotho (S-Klotho) levels play a pathophysiological role in the development of IGF-I receptor (IGF-IR) resistance. It is postulated that as soon as circulating IGF-I, insulin and S-Klotho rise above a certain level the IGF-IR becomes relatively resistant to actions of IGF-I. The development of a degree of IGF-IR resistance for metabolic actions may help to explain why in active acromegaly diabetogenic effects of GH predominate and are not completely counteracted and neutralized by elevated circulating levels of IGF-I. Further studies are necessary in order to support this hypothesis.

1. The GH-IGF-I axis in healthy subjects

In healthy subjects GH is released from somatotroph cells of the anterior pituitary in a pulsatile fashion and activates the growth hormone receptor (GHR) which is present at the cellular surface of target tissues such as liver, muscle, adipose tissue, bone and kidney [36]. Release of GH is primarily regulated by growth hormone-releasing hormone (GHRH; positive regulation), and somatostatin (negative regulation) [36]. GH produced by the pituitary is the main regulator of circulating insulin-like growth factor-I (IGF-I), which is particularly produced by the liver [36,56]. A complicated system of short and long feedback loops negatively regulates GH secretion in the healthy state [36] (Fig. 1). It has been found that IGF-I itself might exert negative feedback on GH secretion at the level of the hypothalamus and at anterior pituitary [5,7] (Fig. 1). Pituitary cells contain specific receptors for IGF-I [45]. IGF-I inhibits basal and GHRH-stimulated GH release from cultured pituitary cells after 24 h of incubation [5]. Furthermore, IGF-I stimulates the release of somatostatin from hypothalamic tissue [3]. GH signaling is initiated by binding of GH to the GHR. One GH molecule binds to two GHR molecules that exist as preformed homodimers. After GH binding to the GHR, the intracellular domains of the GHR dimer undergo rotation, which brings together the two intracellular domains of the GHR [14]. A number of signaling proteins and pathways activated by GH have been identified, including Janus Kinases (JAKs), signal transducers and activators of transcription (Stats), the mitogen activated protein kinase (MAPK) pathway, and the phosphatidylinositol 3'-kinase (PI3K) pathway [33]. Although these signal transduction pathways have been well characterized, the manner by which GH activates these pathways and the downstream signals induced by these pathways are not completely understood [33]. The suppressor of cytokine signaling (SOCS) family of proteins plays an important role in the negative regulation of GH signaling [33]. Other important mechanisms whereby GH signaling is thought to be negatively regulated is through protein tyrosine phosphatases (PTPs) and ubiquitin-dependent GHR endocytosis [33]. Precise regulation of GH signaling is important for the proper maintenance of body growth and metabolism.

2. The IGF-I receptor and IGF-I receptor resistance

The IGF-IR is a heterotetrametric transmembrane protein composed of two α and two β subunits. Binding of IGF-I to the alpha subunit of the IGF-IR is followed by autophosphorylation of tyrosine residues on one β subunit. The activated receptor recruits then phosphorylated intracellularly substrates as the insulin receptor substrate proteins (IRSs) and SH2-containing collagen-related proteins (SHC). Tyrosine phosphorylation of the IRSs in turn activates then the phosphatidylinositol 3-kinase (PI3K-Akt) pathway, which is predominantly involved in metabolic actions (Fig. 2). Tyrosine phosphorylation of SHC induces downstream signaling activation through the Ras/Raf/MEK/Erk
pathway which is predominantly involved in mitogenic (growth-promoting) actions (Fig. 2).

IGF-IR resistance has been previously reported in a variety of clinical conditions. Some rare conditions are associated with genetic IGF-I resistance [24]. Disease states such as diabetes, severe insulin resistance, chronic renal insufficiency and HIV infection may cause acquired IGF-IR resistance [24]. Acquired IGF-IR resistance may present at the pre-receptor level through decreased bioavailability of IGF-I (for example by circulating inhibitors of IGF-I action) [24]. It is also possible that IGF-IR resistance is caused by defects of the IGF-IR self (by decreased number and/or affinity for IGF-I) or by post-receptor defects in the intracellular signaling pathways of the IGF-IR [24]. IGF-IR resistance may have different effects in various cells and organs, depending on the cell-predominant enzymatic machinery. In addition, in the setting of resistance by defects of the IGF-IR self, one would expect that circulating IGF-I levels are elevated if classical negative feedback regulation is operative [24].

3. Metabolic effects of growth hormone and insulin-like growth factor-I

Metabolic effects of GH and IGF-I are essentially different. Whereas GH leads to a decreased glucose uptake, elevated insulin secretion, reduced insulin sensitivity and an increased lipolysis, IGF-I leads to an increased glucose uptake, a reduced insulin secretion, enhanced insulin sensitivity and neutral effects on lipolysis (Table 1). In healthy subjects metabolic effects of GH and IGF-I are well in balance whereas in active (untreated) acromegaly insulin antagonistic and diabetogenic effects of GH on glucose metabolism predominate over IGF-I (Table 2). As a direct consequence active acromegaly eventually may progress to impaired glucose tolerance and diabetes by inducing a decreased glucose uptake and an increased glycogenolysis and gluconeogenesis (Table 2).
gland and hypothalamus would be ineffective, resulting in elevated GH levels and, subsequently, further elevation of IGF-I levels [24].

In this paper we discuss the possibility that in (untreated) active acromegaly chronically elevated IGF-I, insulin and S-Klotho levels induce a degree of IGF-IR resistance: we hypothesize that as soon as circulating IGF-I, insulin and S-Klotho levels rise above a certain level (desensitization threshold), the IGF-IR becomes functionally resistant to actions of IGF-I. When this hypothesis is correct, elevated circulating IGF-I, insulin and S-Klotho levels may be drivers of (acquired) IGF-IR resistance in active acromegaly. In the paragraphs that follow, I will present further data and arguments to support this hypothesis.

6. Arguments for IGF-IR resistance in acromegaly

6.1. IGF-I and IGF-IR expression

In several animal models it has been found that IGF-IR expression in muscle is negatively correlated to IGF-I levels, at least at the mRNA level: bGH giant mice are mice with transgenic overexpression of bovine GH. They showed twofold higher GH levels as well as high hepatic IGF-I expression and serum IGF-I levels relative to controls but low mRNA expression of the IGF-IR [42]. In contrast, dwarf GHR −/− mice, mice which lack the GHR showed the opposite picture: the diminished GH signaling led to very low IGF-I levels and increases expression of the IGF-IR [42]. Furthermore, GHR and IGF-IR mRNA expression measured by real-time RT-PCR was significantly lower in human somatotroph tumors of acromegaly patients than in normal pituitary tissue [28]. In addition, immunostaining showed that the reduced expression of mRNA for both the IGF-R in somatotroph tumors was also reflected in lower protein expression of the IGF-IR [20]. Diminution of IGF-IR auto phosphorylation following chronic stimulation by high IGF-I levels may be a mechanism that enables the body to avoid overstimulation by IGF-I [20]. It has been reported that high IGF-I itself rapidly downregulates the IGF-IR expression following binding to the IGF-IR [11,48]. Chronically elevated IGF-I levels in active acromegaly may be thus in part due the fact that less IGF-I is cleared from the circulation by receptor-mediated mechanism. Moreover, high IGF-I levels may change post-receptor signaling of the IGF-IR. Chronic IGF-I stimulation may induce both internalization of the IGF-IR to the lysosome and degradation of insulin receptor substrates (IRS) proteins, thereby the downregulating the Akt (=protein kinase B) pathway,

![Fig. 2. Signaling pathways of the IGF-I receptor activated by IGF-I. Binding of IGF-I to the α subunit of the IGF-I receptor stimulates phosphorylation of tyrosine residues of the β subunit which then induces phosphorylation of the Insulin Receptor Substrates (IRS 1–4) and SHC, which are docking proteins for activating of the phosphatidylinositol 3-kinase (PI3K-Akt) pathway or the Ras/Raf/MEK/Erk pathway, respectively. The PI3K-Akt pathway is predominantly involved in metabolic actions (glucose uptake, insulin sensitivity) whereas Ras/Raf/MEK/Erk pathway is predominantly involved in mitogenic effects (proliferation, growth).](image)

Table 1

<table>
<thead>
<tr>
<th>Metabolic effects of Growth Hormone (GH) and Insulin-like Growth Factor-I (IGF-I).</th>
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<tbody>
<tr>
<td></td>
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<tr>
<td>Glucose uptake            ↓       ↑</td>
</tr>
<tr>
<td>Insulin secretion         ↑       ↓</td>
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<tr>
<td>Insulin sensitivity       ↓       ↑</td>
</tr>
<tr>
<td>Lipolysis               ↑/↓     No effect</td>
</tr>
<tr>
<td>Protein synthesis        ↑/↓     No effect or ↑</td>
</tr>
<tr>
<td>Protein breakdown        ↑/↓     No effect</td>
</tr>
<tr>
<td>Net protein balance       ↑/↓     No effect or ↑</td>
</tr>
</tbody>
</table>

↑ Stimulating.
↓ Inhibiting.

Table 2

In healthy subjects metabolic effects of growth hormone (GH) and insulin-like growth factor-I (IGF-I) are well in balance whereas in active (untreated) acromegaly insulin antagonistic effects of GH on glucose metabolism predominate over IGF-I.

<table>
<thead>
<tr>
<th>Healthy subjects</th>
<th>Active acromegaly</th>
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<tbody>
<tr>
<td>GH levels</td>
<td>Normal</td>
</tr>
<tr>
<td>IGF-I levels</td>
<td>Normal</td>
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<tr>
<td>Negative feedback of IGF-I on GH secretion</td>
<td>Normal</td>
</tr>
<tr>
<td>Insulin levels</td>
<td>Normal</td>
</tr>
<tr>
<td>glucose uptake</td>
<td>Normal</td>
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<tr>
<td>Gluconeogenesis</td>
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<tr>
<th>Tyrosine phosphorylation</th>
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<tr>
<td>Ras-MAPK pathway</td>
</tr>
<tr>
<td>MetabolicActions</td>
</tr>
<tr>
<td>IGF-I Receptor</td>
</tr>
<tr>
<td>Pi 3-kinase pathway</td>
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which - as above discussed- is predominantly involved in metabolic actions [20]. Recently it has been suggested that depletion of insulin receptor substrate-1 (IRS-1) by prolonged IGF-I stimulation accelerates IGF-IR endocytosis and thereby induces a shift from sustained to transient Akt signaling [20]. In the presence of a degree of IGF-IR resistance of the pituitary (and hypothalamus), the IGF-I-induced feedback inhibition of GH secretion is ineffective, leading to increased circulating GH and IGF-I levels. The high circulating IGF-I levels have also reduced effects in muscles and other tissues through the (IGF-I-induced) (relative) IGF-IR resistance. As a consequence the IGF-I-induced feedback inhibition of GH secretion at the pituitary level is reduced, leading to elevations in GH and IGF-I. In addition, the increased circulating IGF-I levels have reduced effects in muscles and other tissues through the increased GH secretion, hyperglycemia and hyperinsulinemia (Fig. 3B).

6.2. Insulin and IGF-I receptor signaling

Hyperinsulinemia and insulin resistance are important metabolic hallmarks of acromegaly and they are strongly related to disease
activity [58]. Stimulation of the insulin receptor by insulin leads to a decrease in intracellular IRS-1 levels [13]. Since IGF-I can also signal through IRS-1, this may contribute to impaired IGF-I-stimulated AKT pathway [13]. In addition, it has been found that insulin after binding to the insulin receptor may induce heterologous desensitization of the signaling of the IGF-IR by downregulating β-arrestin-1 and inhibition of IGF-I-stimulated MAP kinase phosphorylation [13]. Thus hyperinsulinemia may reset the responsiveness of the IGF-IR and this may be another mechanism responsible for (some degree of) IGF-IR resistance at the post-receptor level in active acromegaly. It has been further suggested that serine phosphorylation of IRS-1 is another mechanism of the body to inhibit excess insulin- and IGF-I-mediated Akt1 signaling in active acromegaly [12]. In favor of this latter possibility, it has been found that elevated insulin and IGF-I in bGH giant mice increase serine phosphorylation of IRS-1 in skeletal muscles and it has been suggested that this may act to inhibit exaggerated glycolytic muscle growth in environments of chronic GH/IGF-I excess [12].

It is thought that hyperinsulinemia, insulin resistance, glucose intolerance and type 2 diabetes in active acromegaly are induced by the prolonged exposure to high GH. For sure, hypersecretion of GH is an important mediator of hyperinsulinemia and insulin resistance. Chronic GH excess acts at several levels to block insulin actions and this induces hyperglycaemia by increasing endogenous glucose production and decreasing peripheral glucose disposal in muscle [9]. Chronic GH excess may induce a reduction in the number of insulin receptors at the cell surface and an impairment of the kinase activity of the insulin receptors [3,16]. Although the GH receptor lacks intrinsic tyrosine kinase activity, it stimulates the activity of JAK-2, a tyrosine kinase that associates with the cytoplasmic domain of the activated GH receptor. It has been suggested that activation of JAK-2 might result in tyrosine phosphorylation of IRS-1 and possibly other proteins in the insulin signaling pathway and thereby account for the insulin-like actions of GH [51,61]. Chronic GH excess may lead to an increased phosphorylation status of the insulin receptor and IRS-1 so that the insulin receptor and IRS-1 are maximally activated in the resting state [17,18]. The basal association of phosphatidylinositol3-kinase (PI-3 kinase) with IRS-1, as well as PI-3 kinase activity, is also increased to maximal levels. The maximal activation of the insulin receptor under basal conditions has the effect of making the insulin receptor insensitive for further stimulation by insulin [3,16]. As a consequence chronic exposure to GH excess is associated with a decreased response to insulin stimulation in terms of IRS-1 phosphorylation and PI-3 kinase activity in skeletal muscle [16]. In the adipocytes chronic GH treatment causes insulin resistance by uncoupling activation of PI-3 kinase and its downstream signals [53]. In the liver GH excess leads to chronic activation of the IR/IRS-1/PI-3 kinase pathway, thus blocking insulin-induced activation [16]. Thus despite hyperinsulinemia peripheral glucose uptake in muscles and other insulin-dependent tissues is reduced whereas also the ability of insulin to suppress gluconeogenesis is significantly reduced [9]. A similar mechanism may be active for elevated circulating IGF-I levels and the IGF-IR.

6.3. IGF-1 receptor and Klotho

Klotho is a transmembrane protein mainly expressed in the kidneys, choroid plexus and various endocrine-related tissues including the pituitary [46]. Klotho is an essential cofactor for the binding of fibroblast growth factor 23 to its receptor, serving as a major regulator of phosphate metabolism [30]. It is also a potent inhibitor of the insulin and IGF-I signaling [1,31,60]. In addition to its local effects Klotho can be cleaved by two specific proteases (ADAM 10 and 17), shed and released from the cell membrane to act as a circulating hormone (Soluble Klotho (S-Klotho)) [29]. Insulin has been shown to activate both of these proteases [8]. Currently GH and IGF-I are the biochemical markers used to confirm active acromegaly and also for assessing the activity of acromegaly [57]. However, recent data show that serum levels of S-Klotho levels in acromegaly depend on GH to a comparable extent as IGF-I. Excessive GH secretion results in markedly increased circulating S-klotho levels in (untreated) active acromegaly [41] and they decrease rapidly following adenoma removal suggesting a causal relation between GH-producing adenoma and high serum levels of S-Klotho in active acromegaly [41,52]. Therefore circulating S-Klotho has been introduced as a novel biomarker for the activity of GH-producing adenoma [41]. Elevated circulating S-Klotho is most likely of renal origin, but the exact mechanism by which excess GH results in increased release of kidney-derived S-klotho has not yet been elucidated [49]. It has been further found that S-Klotho can inhibit the activation of the IGF-IR in a dose-dependent manner [31,59]. Thus elevated S-Klotho levels in active acromegaly may contribute to (some degree of) IGF-IR resistance [57]. As a consequence the IGF-I-induced feedback inhibition of GH secretion at the pituitary (or hypothalamus) level is reduced, leading to elevations in GH and IGF-1 (Fig. 3C). Interestingly, S-Klotho is a protein that has been reported also to inhibit IGF-IR and insulin receptor signaling by inhibiting tyrosine phosphorylation of both receptors and their downstream signaling proteins (i.e. IRS) [31,60] (Fig. 4). Thus due to S-Klotho-induced IGF-IR resistance elevated circulating IGF-I levels in active acromegaly have reduced effects in muscles and other tissues.

6.4. Hyperglycaemia and IGF-IR resistance

Chronic hyperglycaemia probably plays also a role in the development of IGF-IR resistance in active acromegaly. In cells exposed in vitro to normal glucose, stimulation of the IGF-IR and its kinase results in phosphorylation IRS-1 [38]. As above discussed, this is an important adaptor protein that couples IGF-I stimulation of the IGF-IR to downstream activation of the PI-3 kinase signaling pathway. However, following exposure to hyperglycaemia it has been reported that IGF-I responsiveness was reduced and IRS-1 was down regulated [38].

7. GH hypersecretion and functional IGF-IR resistance in active acromegaly

Persistent GH hypersecretion in active acromegaly together with highly elevated IGF-I implies impaired negative feedback by IGF-I. In support of this, it has been reported that negative feedback regulation is greatly attenuated in active acromegaly [23]. The possibility that loss of
IGF-I feedback at the level of the pituitary/hypothalamus plays a pathophysiological role in GH hypersecretion in acromegaly is supported by in vitro data [23]. Furthermore, it has been suggested that a decrease in number of IGF-IRs on somatotroph adenoma cells contributes to GH hypersecretion in acromegaly or that alternatively, post-receptor abnormalities play a role in IGF-IR resistance [23]. Development of a degree of IGF-IR resistance (mediated by elevated IGF-I, insulin and S-Klotho levels) in active acromegaly may function as a safety valve of the body that counters overstimulation by IGF-I of the IGF-IR in order to avoid too large changes of the homeostasis [20].

8. Elevated IGF-I levels, IGF-IR resistance and effects on metabolism in active acromegaly

As discussed above, compared to GH, IGF-I has divergent effects on carbohydrate and lipid metabolism [37]. Most of the established metabolic effects of GH may be classified as insulin-antagonistic: for example, gluconeogenesis and lipolysis are stimulated by GH [21]. Most metabolic effects of IGF-I can be classified as insulin-like [37]. For example, IGF-I increases glucose uptake of muscles and inhibits gluconeogenesis. In addition, IGF-I can improve insulin sensitivity [9]. In fact, GH antagonizes the effects of IGF-I on glucose metabolism [37]. Although IGF-I has been found to be 0.05–0.1 times as potent as insulin in terms of stimulating glucose transport in vitro and inducing hypoglycemia, in vivo its concentration is in healthy subjects approximately 100 times higher compared to that of insulin [34,43,62]. In active (untreated) acromegaly circulating IGF-I levels are usually >2 SD above the upper limit of the normal range. Despite these elevated IGF-I levels and the insulin-like effects of IGF-I, active acromegaly is characterized by hyperinsulinemia, insulin resistance, high plasma glucose concentrations and type 2 diabetes. However, in subjects without acromegaly a potent glucose lowering effect is typically observed after recombinant IGF-I administration, with improved insulin sensitivity and lowering of insulin concentrations [37]. Even though this effect may be (partly) mediated through stimulation of the insulin receptor, experimental mice knockout of the insulin receptor also show a potent glucose lowering effect of IGF-I, indicative that the hypoglycemic effect of IGF-I is mediated at least in part directly through the IGF-IR [15,37]. In addition, chronic IGF-I stimulation improves insulin sensitivity of skeletal muscle and adipose tissue resulting in increased glucose transport, increased glycogenesis and decreased lipolysis [22]. Administration of IGF-I to subjects with type 2 diabetes results in a marked enhancement of insulin sensitivity and a clear improvement of diabetic control [10,39,40]. When Clemmons et al. previously studied patients with active acromegaly and severe insulin resistance, they observed -as expected- an improvement of insulin sensitivity and glucose control after administration of the GH antagonist pegvisomant [44]. Although insulin resistance that ensues acromegaly was at least partly improved by lowering GH hypersecretion in this latter study, interestingly, a more marked improvement in insulin sensitivity and glucose control was observed when IGF-I was added to pegvisomant [44]. This finding does not only suggest that IGF-I may have insulin-sensitizing actions on top of and independent of its ability to suppress GH secretion, but it also suggests that in untreated acromegaly endogenous markedly elevated circulating IGF-I levels are insufficient to neutralize GH-mediated effects on insulin resistance [9]. Development of a degree of IGF-IR resistance -initiated and sustained by high circulating IGF-I, insulin and S-Klotho levels- could explain why in active acromegaly even IGF-I levels >2SD above the normal range are not able and sufficient to overcome the effects of GH on metabolism and to normalize insulin sensitivity and glucose tolerance. When chronically elevated IGF-I, insulin and soluble Klotho play a pathophysiological role in the development of IGF-IR resistance in active acromegaly, sensitivity of the IGF-IR will be improved by removing elevated circulating IGF-I, insulin and soluble Klotho levels. Normalization of circulating IGF-I is more frequently achieved by monotherapy with the GH antagonist (pegvisomant) than by somatostatin analogues [19,55]. Moreover, pegvisomant ameliorates all aspects of glucose metabolism, while somatostatin analogues sometimes increase plasma glucose levels [58]. At first glance this suggests, theoretically at least, that medical therapy with pegvisomant is better able to reverse IGF-IR resistance in active acromegaly than somatostatin analogues. However, further studies are needed to elucidate the exact role of pegvisomant and somatostatin analogues on reversal of IGF-IR resistance in active acromegaly.

In conclusion, active acromegaly is characterized by increased GH secretion, elevated circulating IGF-I, insulin and S-Klotho levels. In this paper the hypothesis is born that in active acromegaly chronically elevated IGF-I, insulin and/or S-Klotho levels initiate and sustain a degree of IGF-IR resistance. The development of a degree of IGF-IR resistance (especially for metabolic functions of the IGF-IR) may help to explain why in active acromegaly GH-mediated effects on glucose metabolism predominate over IGF-I and are not prevented and counter-regulated by elevated circulating levels of IGF-I. Further studies are necessary to support this hypothesis.

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Data availability statement

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Declaration of Competing Interest

The author has no competing interests to declare.

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