



Fetal phthalates and bisphenols and childhood lipid and glucose metabolism. A population-based prospective cohort study



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ABSTRACT

Background and aims: Fetal exposure to endocrine disruptors such as phthalates and bisphenols may lead to developmental metabolic adaptations. We examined associations of maternal phthalate and bisphenol urine concentrations during pregnancy with lipids, insulin, and glucose concentrations at school age.

Methods: In a population-based, prospective cohort study among 757 mother–child pairs, we measured maternal phthalate and bisphenol urine concentrations in first, second and third trimester of pregnancy. We measured non-fasting lipids, glucose and insulin blood concentrations of their children at a mean age of 9.7 (standard deviation 0.2) years. Analyses were performed for boys and girls separately.

Results: An interquartile range (IQR) higher natural log transformed third trimester maternal urine phthalic acid concentration was associated with a 0.20 (95% confidence interval (CI) 0.07–0.34) standard deviation score (SDS) higher triglycerides concentration among boys. Maternal bisphenol urine concentrations were not associated with non-fasting lipid concentrations during childhood. An IQR higher natural log transformed second trimester maternal high molecular weight phthalates (HMWP) and di-2-ethylhexylphthalate (DEHP) urine concentration were associated with a 0.19 (95% CI 0.31–0.07) respectively 0.18 (95% CI 0.31–0.06) SDS lower glucose concentration among boys. An IQR higher natural log transformed third trimester maternal bisphenol F urine concentration was associated with a 0.22 (95% CI 0.35–0.09) SDS lower non-fasting insulin concentration among boys.

Conclusions: Our results suggest potential persisting sex specific effects of fetal exposure to phthalates and bisphenols on childhood lipid concentrations and glucose metabolism. Future studies are needed for replication and exploring underlying mechanisms.

1. Introduction

Endocrine-disrupting chemicals (EDCs), such as phthalates and bisphenols, are widely used in food packaging, household products and

medical devices (Ye et al., 2008; Russo et al., 2019; Schettler, 2006). Fetal life may be a specific critical period for the possible effects of phthalates and bisphenols because they pass the placenta (Nahar et al., 2015; Mose et al., 2007). The mechanisms by which phthalates and

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Table 1
Urinary concentrations of phthalates and bisphenols during pregnancy, stratified in boys and girls.

	First trimester		Second trimester		Third trimester	
	Median (25th-75th percentile)		Median (25th-75th percentile)		Median (25th-75th percentile)	
	Boys	Girls	Boys	Girls	Boys	Girls
Phthalic Acid (PA) (nmol/L)	343.1 (179.4–645.5)	365.1 (185.0–779.4)	903.8 (372.0–1658.6)	832.3 (337.8–1687.7)	368.7 (188.9–778.0)	419.5 (219.9–889.9)
Low-molecular-weight phthalates (LMWP) (nmol/L)	1094.8 (477.8–2653.5)	1102.0 (400.8–3118.0)	520.5 (228.1–1281.0)	573.8 (221.4–1659.0)	921.1 (372.6–2244.8)	1008.3 (410.7–2773.1)
Monomethylphthalate (mMP) (nmol/L)	28.7 (15.1–54.5)	30.2 (15.3–55.7)	17.9 (9.9–33.6)	19.1 (9.3–35.4)	19.4 (10.2–37.8)	24.8 (11.6–52.7)
Monoethylphthalate (mEP) (nmol/L)	677.8 (210.3–2329.3)	710.8 (208.1–2574.9)	315.4 (112.6–970.4)	376.9 (124.0–1409.1)	581.0 (205.4–1818.1)	658.1 (219.9–2378.1)
Mono-isobutylphthalate (mIBP) (nmol/L)	101.5 (45.9–217.6)	98.1 (43.0–191.3)	36.5 (19.3–78.6)	39.9 (18.8–78.8)	69.3 (39.7–138.3)	84.2 (43.8–174.4)
Mono-n-butylphthalate (mBP) (nmol/L)	78.5 (32.5–150.0)	72.6 (32.7–140.2)	43.0 (23.5–82.7)	41.3 (22.3–85.6)	52.9 (27.2–97.2)	56.9 (28.0–117.1)
High molecular-weight phthalates (HMWP) (nmol/L)	217.3 (111.0–414.9)	218.4 (115.8–385.5)	131.8 (73.7–242.3)	121.9 (67.8–221.4)	156.7 (92.5–268.9)	178.2 (97.4–318.1)
Monobenzylphthalate (mBzBP) (nmol/L)	22.0 (8.7–47.9)	22.4 (9.4–47.8)	19.6 (7.7–37.5)	10.8 (8.2–42.5)	10.8 (3.9–23.7)	12.8 (4.1–25.2)
Mono-hexylphthalate (mHxP) (nmol/L)	1.0 (0.4–2.2)	0.9 (0.3–1.9)	NA	NA	NA	NA
Mono-2-heptylphthalate (mHpP) (nmol/L)	2.1 (0.8–5.3)	2.1 (0.8–5.5)	NA	NA	NA	NA
Monocyclohexyl-phthalate (mCHP) (nmol/L)	0.1 (0.1–0.1)	0.1 (0.1–0.1)	NA	NA	NA	NA
Di-2-ethylhexylphthalate (DEHP) (nmol/L)	171.9 (89.8–315.1)	174.2 (89.9–300.7)	96.8 (49.9–184.2)	89.4 (48.2–164.9)	132.3 (75.3–225.9)	146.8 (80.3–274.1)
Mono-(2-ethyl-5-carboxy-pentyl)phthalate (mECP) (nmol/L)	51.6 (25.9–100.0)	51.2 (27.1–99.4)	33.8 (17.4–64.9)	30.6 (17.8–58.6)	54.1 (28.9–103.7)	59.8 (30.4–110.3)
Mono-(2-ethyl-5-hydroxy-hexyl)phthalate (mEHHP) (nmol/L)	41.4 (20.8–79.9)	43.5 (20.5–75.0)	19.2 (10.0–37.1)	17.8 (9.2–34.2)	31.0 (17.1–59.4)	39.4 (18.2–75.2)
Mono-(2-ethyl-5-oxohexyl)phthalate (mEOHP) (nmol/L)	27.0 (12.1–55.6)	26.5 (12.5–49.5)	25.2 (12.4–57.5)	22.0 (10.6–52.5)	22.0 (13.3–42.3)	26.9 (14.0–50.0)
Mono-[(2-carboxymethyl)-hexyl] phthalate (mCMHP) (nmol/L)	44.2 (24.3–67.8)	46.9 (24.5–83.2)	12.51 (6.8–24.2)	12.3 (6.9–22.9)	9.7 (6.0–19.7)	12.4 (5.7–22.5)
Di-n-octylphthalate (DNOP)	5.7 (3.2–11.2)	6.4 (2.9–10.7)	3.5 (2.0–6.8)	3.3 (1.9–6.1)	6.6 (3.7–12.1)	6.9 (3.8–13.2)
Mono(3-carboxypropyl)- phthalate (mCPP) (nmol/L)	5.7 (3.2–11.2)	6.4 (2.9–10.7)	3.5 (2.0–6.8)	3.3 (1.9–6.1)	6.6 (3.7–12.1)	6.9 (3.8–13.2)
Bisphenols (nmol/L)	9.7 (3.6–21.7)	8.7 (3.4–21.6)	6.0 (2.8–12.5)	5.8 (2.9–13.5)	8.4 (3.8–17.0)	10.0 (4.5–19.3)
Bisphenol A (BPA) (nmol/L)	5.0 (1.0–12.4)	4.6 (1.0–15.0)	5.7 (2.6–11.6)	5.1 (2.5–11.8)	5.9 (2.5–11.0)	7.0 (2.8–13.2)
Bisphenol S (BPS) (nmol/L)	0.8 (0.1–3.0)	0.6 (0.1–2.3)	0.1 (0.1–0.5)	0.1 (0.1–0.4)	NA	NA
Bisphenol F (BPF) (nmol/L)	0.6 (0.6–1.7)	0.6 (0.6–2.4)	NA	NA	0.6 (0.6–2.6)	0.6 (0.6–1.3)

Values represent medians (25th-75th percentiles). Absolute urinary concentration of the grouped exposures (in nmol/L urine) and individual exposures (in nmol/L urine) with concentrations below the limit of detection imputed as limit of detection/square root of 2. Only values that are included in the calculation of the grouped exposures are included in this table.

NA: not applicable due to > 80% of concentrations below limit of detection.

bisphenols may affect fetal development are by stimulating estrogen and inhibiting androgen receptors, activating peroxisome proliferator-activated receptors (PPARs) or retinoid X receptors (RXRs), and changing the fetal transcriptome (Mattison et al., 2014). Thus far, cross-sectional studies in adult populations showed inconsistent results for the associations of phthalates and bisphenols with several metabolic diseases in humans, such as obesity, hypertension and diabetes (Ranciere et al., 2015; Golestanzadeh et al., 2019; James-Todd et al., 2012).

Currently, only limited prospective studies on the associations of fetal exposure to phthalates or bisphenols with metabolic diseases are available and they showed no clear association of fetal exposure to phthalates or bisphenols with childhood lipid and glucose metabolism (Vafeiadi et al., 2016, 2018; Perng et al., 2017; Watkins et al., 2016). In a Greek cohort study, first trimester bisphenol A (BPA) urine concentrations were not associated with non-fasting lipid concentrations at 4 years, while third trimester phthalates urine concentrations were negatively associated with non-fasting high density lipoprotein (HDL)-

cholesterol concentrations at 4 years, but not with cholesterol concentrations or metabolic outcomes at 6 years (Vafeiadi et al., 2016, 2018). In another study among 227 mother-child pairs from Mexico, no consistent associations were found for pregnancy-averaged maternal urine BPA or phthalate concentrations with lipid profiles among 8–14-year-old children (Perng et al., 2017). Sex- and pubertal status-dependent associations were observed of third trimester phthalate and BPA exposures with C-peptide and fasting glucose at 8–14 years (Watkins et al., 2016). Although the previous studies explored sex-specific associations, the study populations were smaller than 300 subjects and thus might have been underpowered.

We hypothesized that fetal exposure to phthalates and bisphenols leads to fetal metabolic adaptations, which persistently affect glucose and lipid metabolism. We assessed the sex-specific associations of maternal phthalate and bisphenol urine concentrations in first, second and third trimester of pregnancy with non-fasting lipids, glucose and insulin concentrations in their children at the age of 10 years.

2. Materials and methods

2.1. Study design

This study was embedded in the Generation R Study, a population-based prospective cohort study from early fetal life onwards in Rotterdam, the Netherlands (Jaddoe et al., 2012). Phthalate and bisphenol concentrations were measured among a subgroup of 1,405 mothers, whose singleton children also participated in postnatal studies. We excluded mothers without information on phthalate and bisphenol urine concentrations for at least one time point in pregnancy and whose children had no measurement of metabolic risk factors at 10 years. The population for analysis comprises 757 mother–child pairs (the specific sample per outcome is shown in Supplemental Fig. 1). The study protocol conforms to the ethical guidelines of the 1975 Declaration of Helsinki and has been approved by the Medical Ethical Committee of the Erasmus MC, University Medical Centre in Rotterdam. Written informed consent was obtained from all participants.

2.2. Maternal phthalate and bisphenol urine concentrations

Phthalate and bisphenol concentrations were measured in spot urine samples obtained from each woman at three time points during pregnancy (median 12.9 weeks of gestation (25th–75th percentiles 12.1–14.6); median 20.4 weeks of gestation (25th–75th percentiles 19.9–20.9); median 30.2 weeks of gestation (25th–75th percentiles 29.9–30.8)). These periods were considered as first, second and third trimester. Urine samples were collected between February 2004 and July 2005. The analyses of phthalate, bisphenol and creatinine concentrations were performed at the Wadsworth Center, New York State Department of Health, Albany, New York, USA, using previously described methods (Philips et al., 2018). Urine biomarkers for exposure to phthalate metabolites were grouped according to their molecular weight and parent phthalates into low molecular weight phthalates (LMWP) and high molecular weight phthalates (HMWP), which includes subgroups of di-2-ethylhexylphthalate (DEHP) and di-n-octylphthalate (DNOP) metabolites. Phthalic acid (PA) was analyzed separately as a proxy for total phthalate exposure. Bisphenols A (BPA), S (BPS) and F (BPF) were grouped and used as proxy for total bisphenol exposure. Weighted molar sums were calculated for the different groups of phthalates and bisphenols. Individual phthalates and bisphenols were included in the groups if < 80% of their concentrations at that time point was below the limit of detection (LOD). All concentrations below the LOD were substituted by LOD divided by the square root of 2 ($LOD/\sqrt{2}$) (Hornung, 1990). The descriptive statistics of the individual and grouped phthalates and bisphenols investigated are shown for boys and girls in Table 1 and for the total group in Supplemental Table S1. The intraclass correlation coefficients between the grouped natural log-transformed phthalates and bisphenols across pregnancy were assessed using a single measurement, absolute agreement and two-way mixed effects model and varied between 0.06 and 0.35 (Supplemental Table S1). We also assessed the Pearson's correlation coefficients between all natural log-transformed creatinine-corrected phthalates and bisphenols which showed that the overall correlations were low-moderate, especially between different trimesters and different groups (Supplemental Table S2). To account for urinary dilution, urine concentrations of phthalates and bisphenols were converted to $\mu\text{mol/g}$ creatinine for the metabolite groups. To reduce the potential for exposure misclassification due to temporal variability, we calculated the overall mean exposure during pregnancy by summing the first, second and third trimester phthalate and bisphenol urine concentrations and dividing that by the three time points.

2.3. Childhood metabolic risk factors

As described previously, children were invited to visit our research center around the age of 10 years (Kooijman et al., 2016). We obtained non-fasting venous blood samples and measured total cholesterol, HDL-cholesterol, triglycerides, glucose and insulin concentrations. Total cholesterol, HDL-cholesterol, triglycerides and glucose concentrations were measured on the Cobas 8000 analyzer using the c702 module. Insulin was measured with electrochemiluminescence immunoassay (ECLIA) on the E411 module (Roche, Almere, the Netherlands) (Kruithof et al., 2014). Low-density lipoprotein (LDL)-cholesterol was calculated according to the Friedewald formula (Onyenekwu et al., 2014). For the assessment of the risk of adverse cardio-metabolic risk factors, children were divided into groups based on their concentrations of triglycerides, HDL-cholesterol, LDL-cholesterol and insulin. The 75th percentile was used as the cut-off for triglycerides (≥ 1.3000 mmol/l), LDL-cholesterol (≥ 2.6927 mmol/l) and insulin (≥ 310.0500 pmol/l) and the 25th percentile was used as the cut-off for HDL-cholesterol (≤ 1.2300 mmol/l).

2.4. Covariates

Information on maternal characteristics, including age, ethnicity, pre-pregnancy body mass index, use of folic acid supplementation, educational level, parity, and maternal smoking habits and alcohol consumption (specifically in first, second and third trimester of pregnancy or during pregnancy) and maternal diet were obtained from questionnaires during pregnancy. To assess maternal diet, a previously developed food-based diet quality score was used, reflecting adherence to national dietary guidelines (Nguyen et al., 2017). Child's sex was obtained from midwife and hospital records at birth. Weight and height were measured during the visit at our research center to calculate the child's body mass index (BMI), which was then standardized for age and sex. A simplified Directed Acyclic Graph (DAG) showing the hypothesized relationship between fetal exposure to phthalates and bisphenols, childhood metabolic risk factors and the covariates is presented in Supplemental Fig. 2.

2.5. Statistical analysis

For all analyses, maternal phthalate and bisphenol urine concentrations were natural log-transformed to reduce variability and account for right skewness of the distribution and further standardized by the interquartile range (IQR) to ease the interpretation of effect sizes. We also natural log-transformed the non-normally distributed childhood metabolic risk factors (triglycerides and insulin) and constructed standard-deviation scores (SDS) [(observed value - mean)/SD] of the sample distribution of all outcomes to enable comparisons of effect sizes. Participants were compared with non-participants on all exposures and covariates by using Chi-square tests, student's t-tests and Mann-Whitney U tests, when applicable.

We assessed linearity of the associations of maternal phthalate and bisphenol urine concentrations (per time point and overall mean during pregnancy) with childhood metabolic risk factors by assessing the residuals of the regression in a P-P plot, assessing homoscedasticity and multicollinearity and used linear regression models if the residuals were normally distributed, the residuals were homoscedastic and if the variance inflation factors were below 5.00. To examine the independent associations of maternal first, second and third trimester phthalate and bisphenol urine concentrations with childhood metabolic risk factors, we created a mutually adjusted model by simultaneously including in the model the exposures at all three time points during pregnancy, after testing whether there were no issues with multicollinearity

(correlations < 0.70) (Supplemental Table S1). We used logistic regression to assess the associations of maternal urine phthalate or bisphenol concentrations with being in the highest risk-category.

First, models were adjusted for child's age and sex only (basic model). Potential confounders were identified based on the graphical criteria for confounding by visualizing a DAG and then we included those in the models that changed the effect estimates > 10% for at least one of the outcomes (Santos et al., 2019). The inclusion of childhood BMI in the model marginally decreased the effect estimates and we only present the fully adjusted models including childhood BMI. As a sensitivity analysis, we performed the analyses of insulin with glucose included in the model because non-fasting samples were used and the time since the last meal was unknown. Based on our hypothesis of sex-specific effects and based on the influence of folic acid on the metabolism of phthalates and bisphenols, we tested for statistical interaction of child's sex. We also tested for statistical interaction of folic acid supplement use, since folic acid can influence methylation and one of the proposed mechanisms by which phthalates and bisphenols exert their influence could be through changes in DNA methylation (Gules et al., 2019; Dolinoy et al., 2007; Pauwels et al., 2017). We found statistically significant interactions (p -value < 0.10) for child's sex only and presented all results for boys and girls separately.

To correct for multiple hypothesis testing, each p -value was compared with a threshold defined as 0.05 divided by the effective number of independent tests estimated based on the correlation structure between the exposures (p -value threshold of 0.0098) (Li et al., 2012). To maintain statistical power and reduce bias related to missing data on covariates, we performed multiple imputation according to the Markov Chain Monte Carlo method. The percentage of missing values for covariates ranged from 0 to 22.7%. Covariates were used as predictor variables and imputed when necessary, while metabolic risk factors were used as predictor variables only. In addition, birth weight, gestational age at birth, blood pressure at 10 years old and heart frequency at 10 years old were used as predictor variables. Ten imputed datasets were created and no substantial differences were found between the original and imputed datasets. We present results based on pooled imputed datasets. All statistical analyses were performed using the Statistical Package of Social Sciences version 25.0 for Windows (SPSS Inc., Chicago, IL, USA).

3. Results

3.1. Participant characteristics

Table 2 shows participant characteristics for the total group and for boys and girls separately. Non-response analyses showed that non-participants tended to have higher urine concentrations of some phthalates, were younger, lower educated and less likely to consume alcohol during pregnancy (Supplemental Tables S3 and S4).

3.2. Maternal phthalate urine concentrations and childhood lipid profiles

Table 3 shows that among boys an IQR increase in natural log-transformed third trimester maternal urine phthalic acid concentrations was associated with 0.20 (95% Confidence Interval (CI) 0.07; 0.34) SDS higher natural log transformed triglyceride concentrations (basic models are presented in Supplemental Table S5). This association was also present in the mutually adjusted model (Supplemental Table S6). The associations of higher overall mean maternal DEHP and overall mean maternal DNOP urine concentrations with higher triglycerides concentrations among boys did not remain significant after multiple testing adjustment (Table 3 and Supplemental Table S7). Similarly, the associations among girls of higher second trimester maternal phthalic

acid urine concentrations with lower triglyceride concentrations in both the adjusted and mutually adjusted model and the association of higher third trimester maternal DNOP urine concentrations with higher natural log transformed triglyceride concentrations did not remain significant after multiple testing adjustment (Table 3 and Supplemental Table S6). Maternal phthalate urine concentrations were not associated with cholesterol concentrations among boys or girls.

3.3. Maternal bisphenol urine concentrations and childhood lipid profiles

Among boys, maternal bisphenol urine concentrations were not associated with childhood total or LDL cholesterol concentrations (Supplemental Table S8). The associations of higher first and third trimester maternal BPF urine concentrations with higher HDL-cholesterol and triglycerides concentrations, respectively, among boys did not remain significant after correction for multiple testing (Table 4). Higher natural log-transformed overall mean maternal bisphenol and BPA urine concentrations were associated with lower natural log transformed triglyceride concentrations among girls, although these associations did not remain significant after correction for multiple testing (Supplemental Table S7). Maternal bisphenol urine concentrations were not associated with total or HDL-, or LDL-cholesterol concentrations among girls (Table 4). No associations were found in the mutually adjusted model (Supplemental Table S9).

3.4. Maternal phthalate urine concentrations and childhood glucose metabolism

Among boys, after adjustment for confounders, an IQR higher second trimester and overall mean maternal HMWP and DEHP urine concentrations was associated with a 0.19 (95% CI 0.31–0.07), 0.18 (95% CI 0.30–0.06), 0.18 (95% CI 0.31–0.06), 0.19 (95% CI 0.31–0.07) SDS lower glucose concentrations (Table 5 and Supplemental Table S10), basic models are presented in Supplemental Table S11). In the mutually adjusted model, these associations were attenuated towards non-significance (Supplemental Table S12). Maternal phthalate urine concentrations were not associated with insulin concentrations during childhood (Table 5). When including glucose concentrations in the model, maternal phthalate urine concentrations were again not associated with insulin concentrations during childhood (Supplemental Table S13).

3.5. Maternal bisphenol urine concentrations and childhood glucose metabolism

Among boys, an IQR higher natural log-transformed third trimester maternal BPF urine concentrations was associated with 0.22 (95% CI 0.35; 0.09) and 0.19 (95% CI 0.32; 0.05) SDS lower natural log transformed insulin and glucose concentrations (Table 6, basic models are presented in Supplemental Table S14). The association of third trimester maternal BPF urine concentrations with glucose concentrations and with insulin concentrations when corrected for glucose concentrations did not remain after multiple testing adjustment (Supplemental Table S15). The associations of higher first trimester maternal BPS urine concentrations with lower natural log transformed insulin and glucose concentrations among boys did not remain after adjustment for multiple testing (Table 6). In the fully adjusted and mutually adjusted model, among boys higher third trimester total bisphenol maternal urine concentrations was associated with lower natural log transformed insulin concentrations, although these associations did not remain after correction for multiple testing (Table 6 and Supplemental Table S16). Among girls, maternal bisphenol urine concentrations were not associated with insulin or glucose concentrations (Table 6).

Table 2
Characteristics of mothers and their children.

	Total group n = 757	Boys n = 382 (50.5%)	Girls n = 375 (49.5%)
Maternal characteristics			
Age at enrolment, mean (SD) (years)	31.0 (4.6)	31.1 (4.5)	30.9 (4.7)
Parity, n (%)			
Nullipara	464 (61.6%)	232 (60.9%)	232 (62.4%)
Multipara	289 (38.4%)	149 (39.1%)	140 (37.6%)
Ethnicity, n (%)			
European	478 (63.6%)	245 (64.5%)	233 (62.6%)
Non-European	274 (36.4%)	135 (35.5%)	139 (37.4%)
Education, n (%)			
Low	52 (7.1%)	26 (7.0%)	26 (7.2%)
Middle	286 (39.0%)	141 (37.8%)	145 (40.2%)
High	396 (54.0%)	206 (55.2%)	190 (52.6%)
Pre-pregnancy BMI, median (95% range) (kg/m ²)	22.7 (18.6–34.7)	22.6 (18.5–34.4)	22.8 (18.6–35.1)
Maternal diet quality score, mean (SD) ^a	7.9 (1.5)	7.9 (1.6)	7.9 (1.4)
Folic acid supplementation, n (%), yes	503 (82.2%)	259 (83.8%)	244 (80.5%)
Smoking during pregnancy, n (%), yes	165 (23.9%)	81 (23.6%)	84 (24.1%)
First trimester, n (%), yes	104 (20.7%)	67 (20.1%)	73 (21.4%)
Second trimester, n (%), yes	75 (11.3%)	35 (10.4%)	40 (12.3%)
Third trimester, n (%), yes	68 (10.5%)	36 (10.7%)	32 (10.2%)
Alcohol consumption during pregnancy (any), n (%), yes	419 (61.2%)	221 (64.8%)	198 (57.6%)
First trimester, n (%), yes	361 (53.6%)	194 (57.7%)	167 (49.4%)
Second trimester, n (%), yes	251 (38.1%)	133 (40.2%)	118 (36.1%)
Third trimester, n (%), yes	256 (39.7%)	147 (44.3%)	109 (34.8%)
Child characteristics			
Age, mean (SD) (years)	9.7 (0.2)	9.7 (0.3)	9.7 (0.2)
BMI, median (95% range) (kg/m ²)	16.8 (14.0–25.0)	16.6 (13.9–24.1)	17.1 (14.0–25.5)
Triglycerides, median (95% range) (mmol/L)	1.0 (0.4–2.5)	0.9 (0.4–2.6)	1.0 (0.5–2.3)
Total cholesterol, mean (SD) (mmol/L)	4.3 (0.6)	4.2 (0.6)	4.4 (0.6)
HDL cholesterol, mean (SD) (mmol/L)	1.5 (0.3)	1.5 (0.3)	1.4 (0.3)
LDL cholesterol, mean (SD) (mmol/L)	2.3 (0.6)	2.2 (0.6)	2.4 (0.6)
Insulin, median (95% range) (pmol/L)	198.9 (37.5–671.0)	193.3 (31.5–571.0)	207.4 (38.5–804.6)
Glucose, mean (SD) (mmol/L)	5.5 (0.9)	5.5 (0.9)	5.4 (0.9)

Values represent mean (SD), median (95% range) or number of subjects (valid %). ^aAs based on the food frequency questionnaire filled out in early pregnancy.

3.6. Maternal phthalate and bisphenol urine concentrations and risk of adverse cardio-metabolic risk factors

An IQR increase in the natural log transformed third trimester maternal phthalic acid concentrations was associated with an increased triglycerides concentration among boys (Odds Ratio (OR) 1.67 (95% CI 1.20; 2.31)) (Supplemental Table S17). An IQR increase in the natural log transformed third trimester maternal DNOP concentrations was associated with high triglycerides concentrations among girls (OR 1.62 (95% CI 1.21; 2.18)). Among boys, an IQR increase in the natural log transformed third trimester maternal total bisphenol urine concentrations was associated with lower insulin concentrations (OR 0.64 (95% CI 0.46; 0.90)) (Supplemental Table S18).

4. Discussion

4.1. Main findings

In this population-based prospective cohort study, we observed that higher third trimester fetal exposure to phthalic acid is associated with higher non-fasting triglycerides concentrations among boys, while we observed no association of fetal exposure to phthalates with other lipid concentrations during childhood. Maternal bisphenol urine concentrations were not associated with non-fasting lipid concentrations during childhood. Fetal exposure to phthalates was not associated with non-fasting insulin concentrations at 10 years old. Higher second trimester and overall mean HMWP and DEHP urine concentrations were associated with lower non-fasting glucose concentrations among boys. Higher third trimester maternal urine BPF concentrations were also associated with lower non-fasting insulin concentrations among boys.

4.2. Interpretation of main findings

Exposure to phthalates and bisphenols is ubiquitous. Previous animal and cross-sectional human studies have suggested associations of higher exposure to phthalates and bisphenols with adverse lipid and glucose metabolism (Li et al., 2014; Lee et al., 2016). However, results from the few prospective studies in humans are inconsistent (Vafeiadi et al., 2016, 2018; Perng et al., 2017; Watkins et al., 2016). Fetal exposure to endocrine disruptors such as phthalates and bisphenols may lead to developmental metabolic adaptations, which could have long term consequences. In the current study, we examined the associations of maternal urine phthalate and bisphenol concentrations during pregnancy with non-fasting lipids, glucose, and insulin concentrations at school age.

The associations of fetal phthalate or bisphenol exposure with lipid concentrations during childhood have been studied in three prospective human studies (Vafeiadi et al., 2016, 2018; Perng et al., 2017). In a study among 260 mother–child pairs, maternal third trimester urine monoethylphthalate (mEP) concentrations were inversely associated with non-fasting HDL-cholesterol concentrations at 4 years, but not at 6 years (Vafeiadi et al., 2018). No associations were found between maternal phthalate concentrations and total cholesterol concentrations in childhood (Vafeiadi et al., 2018). Among 235 mothers from the same cohort, no associations were observed for maternal urine BPA concentrations with non-fasting blood concentrations of total cholesterol and HDL-cholesterol among 4-year-old children (Vafeiadi et al., 2016). In another study among 227 mother–child pairs, no consistent associations were observed for the pregnancy-averaged maternal urine phthalate and BPA concentrations with fasting triglycerides, total cholesterol, HDL-cholesterol and calculated LDL-cholesterol among 8–14-year-old children (Perng et al., 2017). In the present study, higher

Table 3
Associations of maternal urinary phthalate concentration during pregnancy with childhood lipid profile at 10 years stratified for boys and girls.

Measures of lipid profile at 10 years (in standard deviation scores, 95% confidence interval)													
Exposure	Trimester	Triglycerides			Total cholesterol			HDL cholesterol			LDL cholesterol		
		Boys (n = 381)	Girls (n = 375)	p-value for inter-action	Boys (n = 382)	Girls (n = 375)	p-value for inter-action	Boys (n = 380)	Girls (n = 375)	p-value for inter-action	Boys (n = 379)	Girls (n = 375)	p-value for inter-action
PA	First trimester	-0.01 (-0.14; 0.11)	0.04 (-0.08; 0.16)	0.57	-0.01 (-0.13; 0.100)	0.04 (-0.09; 0.17)	0.55	0.00 (-0.12; 0.13)	0.05 (-0.07; 0.17)	0.42	-0.02 (-0.14; 0.09)	0.01 (-0.12; 0.14)	0.73
	Second trimester	0.06 (-0.10; 0.21)	-0.14 (-0.27; -0.00)*	0.06#	-0.01 (-0.15; 0.13)	-0.04 (-0.19; 0.11)	0.72	0.01 (-0.15; 0.16)	0.06 (-0.08; 0.19)	0.42	-0.02 (-0.16; 0.12)	-0.02 (-0.17; 0.12)	0.81
	Third trimester	0.20 (0.07; 0.34)†	0.06 (-0.07; 0.19)	0.12	0.05 (-0.08; 0.17)	0.07 (-0.08; 0.21)	0.87	-0.11 (-0.24; 0.03)	-0.06 (-0.19; 0.07)	0.42	0.04 (-0.09; 0.16)	0.08 (-0.06; 0.22)	0.75
LMWP	First trimester	-0.11 (-0.25; 0.02)	0.05 (-0.08; 0.18)	0.09#	-0.02 (-0.14; 0.11)	0.09 (-0.05; 0.23)	0.44	0.08 (-0.05; 0.21)	0.04 (-0.09; 0.16)	0.79	-0.05 (-0.17; 0.07)	0.06 (-0.08; 0.20)	0.36
	Second trimester	0.05 (-0.10; 0.20)	-0.11 (-0.25; 0.03)	0.18	-0.05 (-0.19; 0.09)	0.04 (-0.12; 0.19)	0.45	0.08 (-0.07; 0.22)	0.05 (-0.09; 0.18)	0.87	-0.11 (-0.25; 0.03)	0.06 (-0.09; 0.21)	0.14
	Third trimester	0.15 (-0.01; 0.30)	0.06 (-0.09; 0.21)	0.44	0.01 (-0.14; 0.16)	0.07 (-0.10; 0.23)	0.70	-0.09 (-0.24; 0.07)	-0.03 (-0.17; 0.12)	0.43	0.03 (-0.12; 0.17)	0.06 (-0.10; 0.23)	0.87
HMWP	First trimester	0.08 (-0.04; 0.20)	0.01 (-0.12; 0.13)	0.35	0.04 (-0.07; 0.15)	-0.00 (-0.14; 0.13)	0.56	0.04 (-0.13; 0.11)	0.02 (-0.10; 0.14)	0.46	0.00 (-0.11; 0.11)	-0.01 (-0.14; 0.13)	0.80
	Second trimester	0.07 (-0.06; 0.20)	-0.10 (-0.22; 0.02)	0.05#	-0.01 (-0.13; 0.11)	-0.08 (-0.21; 0.05)	0.43	-0.01 (-0.14; 0.12)	0.00 (-0.12; 0.12)	0.70	-0.02 (-0.14; 0.10)	-0.06 (-0.19; 0.07)	0.57
	Third trimester	0.12 (-0.01; 0.25)	0.03 (-0.08; 0.13)	0.18	-0.03 (-0.15; 0.09)	-0.02 (-0.14; 0.09)	1.00	-0.01 (-0.14; 0.11)	0.05 (-0.05; 0.15)	0.43	-0.08 (-0.20; 0.04)	-0.07 (-0.18; 0.04)	0.95
DEHP	First trimester	0.09 (-0.03; 0.22)	0.01 (-0.12; 0.13)	0.28	0.07 (-0.05; 0.18)	-0.01 (-0.14; 0.13)	0.39	-0.01 (-0.13; 0.11)	0.02 (-0.11; 0.14)	0.55	0.03 (-0.09; 0.14)	-0.01 (-0.14; 0.13)	0.64
	Second trimester	0.10 (-0.04; 0.23)	-0.11 (-0.24; 0.02)	0.04*	-0.00 (-0.13; 0.12)	-0.10 (-0.24; 0.04)	0.30	-0.01 (-0.14; 0.12)	0.00 (-0.13; 0.13)	0.81	-0.02 (-0.14; 0.10)	-0.08 (-0.22; 0.06)	0.53
	Third trimester	0.12 (-0.01; 0.25)	0.02 (-0.09; 0.12)	0.17	0.01 (-0.11; 0.13)	-0.03 (-0.15; 0.09)	0.61	-0.01 (-0.13; 0.12)	0.05 (-0.05; 0.16)	0.50	-0.04 (-0.16; 0.08)	-0.08 (-0.19; 0.03)	0.60
DNOP	First trimester	0.11 (-0.00; 0.22)	0.00 (-0.11; 0.11)	0.13	0.03 (-0.08; 0.13)	0.02 (-0.10; 0.14)	0.82	-0.02 (-0.12; 0.09)	0.01 (-0.10; 0.12)	0.63	-0.02 (-0.12; 0.08)	0.02 (-0.10; 0.14)	0.70
	Second trimester	0.10 (-0.04; 0.23)	-0.01 (-0.13; 0.11)	0.24	-0.04 (-0.16; 0.08)	0.00 (-0.13; 0.13)	0.66	-0.06 (-0.19; 0.06)	-0.04 (-0.15; 0.08)	0.77	-0.04 (-0.16; 0.08)	0.02 (-0.11; 0.15)	0.51
	Third trimester	0.04 (-0.08; 0.17)	0.12 (0.01; 0.24)*	0.43	-0.09 (-0.20; 0.03)	0.04 (-0.09; 0.16)	0.13	-0.05 (-0.17; 0.08)	-0.07 (-0.18; 0.05)	0.78	-0.10 (-0.22; 0.02)	0.02 (-0.10; 0.15)	0.13

Values are regression coefficients (95% confidence interval) from linear regression models that reflect the difference in cardiometabolic risk factors in SDS for an interquartile range increase in each natural log-transformed phthalate (in $\mu\text{mol/g}$ creatinine). Triglycerides are natural log transformed. Model includes child's age and sex- and age-standardized body mass index and maternal age, education, parity, ethnicity, pregnancy body mass index, folic acid supplement use, maternal diet quality score, alcohol consumption and smoking habits (specifically in early, mid and late pregnancy). P-values for interaction of sex are presented. #p-value < 0.10; * p-value < 0.05; † Significant after correction for multiple testing (p-value threshold of 0.0098).

DEHP, di-2-ethylhexylphthalate; DNOP, di-n-octylphthalate; HMWP, high molecular weight phthalate; LMWP, low molecular weight phthalate; PA, phthalic acid; SDS, standard deviation scores.

Table 4
Associations of maternal urinary bisphenol concentration during pregnancy with childhood lipid profile at 10 years stratified for boys and girls.
Measures of lipid profile at 10 years (in standard deviation scores, 95% confidence interval)

Exposure	Trimester	Triglycerides			Total cholesterol			HDL cholesterol			LDL cholesterol		
		Boys (n = 381)	Girls (n = 375)	p-value for inter-action	Boys (n = 382)	Girls (n = 375)	p-value for inter-action	Boys (n = 380)	Girls (n = 375)	p-value for inter-action	Boys (n = 379)	Girls (n = 375)	p-value for inter-action
BP	First trimester	-0.04 (-0.19; 0.11)	-0.09 (-0.21; 0.03)	0.60	0.00 (-0.14; 0.14)	-0.03 (-0.16; 0.10)	0.68	0.08 (-0.07; 0.22)	0.05 (-0.07; 0.16)	0.72	-0.02 (-0.15; 0.12)	-0.02 (-0.15; 0.11)	0.95
	Second trimester	0.06 (-0.07; 0.19)	-0.09 (-0.22; 0.04)	0.11	-0.00 (-0.12; 0.12)	-0.02 (-0.16; 0.12)	0.89	0.00 (-0.13; 0.13)	0.05 (-0.08; 0.17)	0.56	-0.03 (-0.15; 0.09)	-0.02 (-0.15; 0.12)	0.89
	Third trimester	0.03 (-0.10; 0.16)	-0.04 (-0.17; 0.09)	0.38	0.01 (-0.11; 0.13)	-0.05 (-0.19; 0.09)	0.56	0.03 (-0.10; 0.15)	0.04 (-0.09; 0.16)	0.89	-0.00 (-0.12; 0.11)	-0.06 (-0.20; 0.08)	0.71
BPA	First trimester	0.01 (-0.14; 0.16)	-0.10 (-0.22; 0.02)	0.23	-0.03 (-0.17; 0.11)	0.03 (-0.10; 0.17)	0.65	0.05 (-0.10; 0.20)	0.07 (-0.05; 0.19)	0.88	-0.06 (-0.20; 0.08)	0.04 (-0.09; 0.17)	0.30
	Second trimester	0.06 (-0.07; 0.19)	-0.09 (-0.22; 0.03)	0.11	-0.01 (-0.12; 0.11)	-0.01 (-0.15; 0.13)	0.99	-0.01 (-0.14; 0.11)	0.06 (-0.07; 0.18)	0.36	-0.02 (-0.14; 0.10)	-0.01 (-0.15; 0.12)	0.94
	Third trimester	0.03 (-0.10; 0.15)	-0.05 (-0.18; 0.07)	0.35	0.04 (-0.08; 0.15)	-0.06 (-0.20; 0.07)	0.25	0.03 (-0.09; 0.15)	0.00 (-0.12; 0.12)	0.59	0.03 (-0.09; 0.14)	-0.05 (-0.18; 0.08)	0.47
BPS	First trimester	0.05 (-0.07; 0.18)	0.02 (-0.09; 0.13)	0.67	0.06 (-0.06; 0.17)	-0.09 (-0.21; 0.03)	0.09 [#]	-0.10 (-0.22; 0.02)	-0.01 (-0.12; 0.11)	0.21	0.11 (-0.01; 0.22)	-0.10 (-0.22; 0.02)	0.02 [*]
	Second trimester	0.04 (-0.10; 0.17)	-0.01 (-0.12; 0.11)	0.68	0.04 (-0.09; 0.16)	-0.01 (-0.14; 0.13)	0.78	0.09 (-0.04; 0.22)	-0.03 (-0.15; 0.09)	0.09 [#]	-0.05 (-0.18; 0.07)	0.03 (-0.10; 0.15)	0.24
	Third trimester	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
BPF	First trimester	-0.08 (-0.23; 0.07)	-0.06 (-0.19; 0.07)	0.78	0.08 (-0.06; 0.21)	0.02 (-0.12; 0.16)	0.81	0.18 (0.04; 0.32) [*]	0.02 (-0.11; 0.15)	0.14	0.01 (-0.13; 0.14)	0.04 (-0.10; 0.18)	0.56
	Second trimester	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
	Third trimester	0.15 (0.0; 0.29) [*]	0.03 (-0.10; 0.17)	0.22	0.04 (-0.09; 0.17)	0.03 (-0.11; 0.18)	0.74	-0.02 (-0.16; 0.12)	-0.01 (-0.14; 0.12)	0.94	-0.01 (-0.14; 0.11)	0.03 (-0.11; 0.17)	0.45

Values are regression coefficients (95% confidence interval) from linear regression models that reflect the difference in cardiometabolic risk factors in SDS for an interquartile range increase in each natural log-transformed bisphenol (in μmol/g creatinine). Triglycerids are natural log transformed. Model includes child's age and sex- and age-standardized body mass index and maternal age, education, parity, ethnicity, pregnancy body mass index, folic acid supplement use, maternal diet quality score, alcohol consumption and smoking habits (specifically in early, mid and late pregnancy). P-values for interaction of child's sex are presented.

[#] p-value < 0.10; * p-value < 0.05; † Significant after correction for multiple testing (p-value threshold of 0.0098).

BP, bisphenols; BPA, bisphenol A; BPF, bisphenol F; BPS, bisphenol S; NA: not applicable due to > 80% of concentrations below limit of detection; SDS, standard deviation scores.

Table 5
Associations of maternal urinary phthalate concentration during pregnancy with childhood glucose metabolism at 10 years stratified for boys and girls.
Measures of glucose metabolism at 10 years (in standard deviation scores, 95% confidence interval)

Exposure	Trimester	Insulin			Glucose		
		Boys (n = 380)	Girls (n = 373)	p-value for interaction	Boys (n = 382)	Girls (n = 374)	p-value for interaction
PA	First trimester	-0.02 (-0.14; 0.10)	0.02 (-0.11; 0.15)	0.49	-0.05 (-0.17; 0.07)	0.04 (-0.10; 0.17)	0.55
	Second trimester	-0.06 (-0.21; 0.08)	-0.01 (-0.15; 0.14)	0.48	-0.15 (-0.30; -0.01)*	0.02 (-0.13; 0.16)	0.15
	Third trimester	0.01 (-0.12; 0.14)	0.03 (-0.11; 0.16)	0.86	-0.03 (-0.16; 0.10)	-0.02 (-0.16; 0.12)	0.85
LMWP	First trimester	0.02 (-0.11; 0.14)	0.05 (-0.09; 0.19)	0.53	-0.01 (-0.14; 0.12)	-0.08 (-0.22; 0.05)	0.30
	Second trimester	0.05 (-0.09; 0.19)	-0.04 (-0.18; 0.11)	0.60	0.02 (-0.13; 0.16)	-0.05 (-0.20; 0.10)	0.43
	Third trimester	0.00 (-0.15; 0.15)	0.05 (-0.11; 0.21)	0.61	-0.03 (-0.18; 0.12)	-0.04 (-0.21; 0.12)	0.64
HMWP	First trimester	-0.02 (-0.13; 0.09)	0.06 (-0.07; 0.19)	0.29	-0.11 (-0.22; 0.00)	0.01 (-0.13; 0.14)	0.25
	Second trimester	-0.10 (-0.22; 0.02)	0.02 (-0.11; 0.15)	0.08*	-0.19 (-0.31; -0.07)†	0.03 (-0.10; 0.16)	0.02*
	Third trimester	-0.08 (-0.20; 0.04)	0.02 (-0.09; 0.13)	0.25	-0.10 (-0.22; 0.03)	0.03 (-0.09; 0.14)	0.22
DEHP	First trimester	-0.03 (-0.14; 0.09)	0.06 (-0.08; 0.19)	0.33	-0.11 (-0.23; 0.00)	0.01 (-0.13; 0.14)	0.24
	Second trimester	-0.11 (-0.24; 0.02)	0.01 (-0.12; 0.15)	0.12	-0.18 (-0.31; -0.06)†	0.04 (-0.10; 0.18)	0.02*
	Third trimester	-0.09 (-0.21; 0.03)	0.02 (-0.09; 0.13)	0.24	-0.10 (-0.23; 0.02)	0.03 (-0.08; 0.15)	0.19
DNOP	First trimester	-0.02 (-0.12; 0.09)	0.03 (-0.09; 0.15)	0.52	-0.07 (-0.18; 0.03)	0.05 (-0.07; 0.18)	0.15
	Second trimester	-0.04 (-0.16; 0.08)	0.02 (-0.11; 0.14)	0.42	-0.09 (-0.22; 0.03)	0.05 (-0.08; 0.18)	0.14
	Third trimester	-0.02 (-0.14; 0.10)	0.03 (-0.09; 0.15)	0.55	-0.08 (-0.20; 0.04)	0.06 (-0.06; 0.19)	0.10

Values are regression coefficients (95% confidence interval) from linear regression models that reflect the difference in glucose metabolism in SDS for an interquartile range increase in each natural log-transformed phthalate (in $\mu\text{mol/g}$ creatinine). Insulin levels are natural log transformed. Model includes child's age and age- and sex-standardized body mass index and maternal age, education, parity, ethnicity, pre-pregnancy body mass index, folic acid supplement use, maternal diet quality score, alcohol consumption and smoking habits (specifically in early, mid and late pregnancy). P-values for interaction of sex are presented.

* $p\text{-value} < 0.10$; * $p\text{-value} < 0.05$; † Significant after correction for multiple testing ($p\text{-value}$ threshold of 0.0098).

DEHP, di-2-ethylhexylphthalate; DNOP, di-n-octylphthalate; HMWP, high molecular weight phthalate; LMWP, low molecular weight phthalate; PA, phthalic acid; SDS, standard deviation scores.

Table 6
Associations of maternal urinary bisphenol concentration during pregnancy with childhood glucose metabolism at 10 years stratified for boys and girls.

Exposure	Trimester	Measures of glucose metabolism at 10 years (in standard deviation scores, 95% confidence interval)					
		Insulin			Glucose		
		Boys (n = 380)	Girls (n = 373)	p-value for interaction	Boys (n = 382)	Girls (n = 374)	p-value for interaction
BP	First trimester	0.02 (-0.12; 0.16)	-0.03 (-0.16; 0.10)	0.44	0.00 (-0.14; 0.14)	-0.03 (-0.16; 0.10)	0.73
	Second trimester	-0.00 (-0.12; 0.12)	-0.02 (-0.15; 0.12)	0.93	-0.01 (-0.14; 0.11)	0.07 (-0.07; 0.21)	0.49
	Third trimester	-0.16 (-0.28; -0.04)*	-0.07 (-0.20; 0.07)	0.35	-0.03 (-0.15; 0.09)	-0.07 (-0.21; 0.07)	0.61
BPA	First trimester	0.08 (-0.06; 0.22)	-0.05 (-0.18; 0.08)	0.14	0.00 (-0.14; 0.14)	-0.01 (-0.14; 0.12)	0.89
	Second trimester	-0.00 (-0.12; 0.12)	-0.00 (-0.14; 0.13)	0.92	-0.02 (-0.15; 0.10)	0.07 (-0.07; 0.21)	0.41
	Third trimester	-0.10 (-0.21; 0.02)	-0.08 (-0.21; 0.05)	0.83	0.04 (-0.07; 0.16)	-0.12 (-0.25; 0.01)	0.05*
BPS	First trimester	-0.13 (-0.25; -0.02)*	-0.01 (-0.13; 0.11)	0.19	-0.15 (-0.26; -0.03)*	-0.02 (-0.14; 0.10)	0.12
	Second trimester	-0.05 (-0.18; 0.08)	-0.03 (-0.16; 0.09)	0.98	-0.06 (-0.19; 0.07)	-0.02 (-0.11; 0.15)	0.33
	Third trimester	NA	NA	NA	NA	NA	NA
BPF	First trimester	0.06 (-0.08; 0.20)	-0.02 (-0.15; 0.12)	0.38	0.07 (-0.07; 0.21)	-0.05 (-0.19; 0.10)	0.24
	Second trimester	NA	NA	NA	NA	NA	NA
	Third trimester	-0.22 (-0.35; -0.09)†	-0.05 (-0.19; 0.09)	0.15	-0.19 (-0.32; -0.05)*	0.05 (-0.10; 0.19)	0.01*

Values are regression coefficients (95% confidence interval) from linear regression models that reflect the difference in glucose metabolism in SDS for an interquartile range increase in each natural log-transformed bisphenol (in $\mu\text{mol/g}$ creatinine). Insulin levels are natural log transformed. Model includes child's age and age- and sex-standardized body mass index and maternal age, education, parity, ethnicity, pre-pregnancy body mass index, folic acid supplement use, maternal diet quality score, alcohol consumption and smoking habits (specifically in early, mid and late pregnancy). P-values for interaction of child's sex are presented. #p-value < 0.10; * p-value < 0.05; † Significant after correction for multiple testing (p-value threshold of 0.0098).

BP, bisphenols; BPA, bisphenol A; BPF, bisphenol F; BPS, bisphenol S; NA: not applicable due to > 80% of concentrations below limit of detection; SDS, standard deviation scores.

third trimester maternal urine phthalic acid concentrations were associated with higher non-fasting triglycerides concentrations among boys. Since phthalic acid is a common metabolite of all phthalates, the higher effect estimates for phthalic acid than for HMWP or DEHP metabolites could be explained by effects of metabolites of newer phthalates which were not measured separately but are included in the phthalic acid measurement. The only other study that investigated non-fasting triglycerides concentrations after fetal exposure to phthalates in humans did not find any associations among all children and among boys and girls separately, which could be due to a power issue (Vafeiadi et al., 2018). The association of higher maternal urine BPA concentrations with lower triglycerides concentrations among girls in the present study was not in line with our hypothesis, which was mainly based on the earlier reported association of higher BPA concentrations with increased risk of cardiovascular disease (Lang et al., 2008). However, our observation was in line with results from a recent meta-analysis of cross-sectional human studies suggesting that higher BPA concentrations were associated with lower triglycerides (Dunder et al., 2019). The associations of overall mean BPA concentrations with triglycerides in our study have to be interpreted with caution, due to the fact that they did not remain significant after correction for multiple testing and could thus be a chance finding. In the current study, no other associations of bisphenols with lipid concentrations were found. No previous longitudinal human studies looked into the association of these newer bisphenols such as BPS or BPF with lipid concentrations during childhood. Animal studies that investigated these associations found varying effects for specific bisphenols (Meng et al., 2018). These differences in associations could suggest distinct underlying mechanisms for different bisphenols and this urges further investigation of the newer bisphenols or trimester specific effects.

The associations of fetal exposure to bisphenols with childhood glucose metabolism markers have also been studied in multiple animal studies, which in general showed tendencies for associations of higher fetal exposure to BPA with diabetogenic effects (Bano et al., 2019; Taylor et al., 2018). In humans, cross-sectional studies have also shown an association between higher exposure to several phthalates and diabetes (James-Todd et al., 2012). To our knowledge, among humans this association has only been studied in one prospective study (Watkins et al., 2016). In that study, a few sex- and pubertal status-dependent associations were observed of third trimester phthalate and BPA exposures with C-peptide, a marker of insulin secretion, and fasting glucose among 219 children at 8–14 years old (Watkins et al., 2016). More specifically, higher maternal total dibutylphthalate (DBP) and mono(3-carboxypropyl)-phthalate (mCPP) urine concentrations were associated with lower fasting glucose concentrations among pubertal boys, while higher maternal mEP urine concentrations were associated with lower C-peptide index among prepubertal girls (Watkins et al., 2016). No associations of fetal BPA with childhood metabolic outcomes were reported (Watkins et al., 2016). In the current study, we observed that fetal exposure to HMWP and DEHP was associated with non-fasting glucose concentrations. This is partially in line with the previous study, that found that higher phthalate exposure was associated with lower glucose and insulin concentrations. The absence of an association of BPA with glucose and insulin concentrations in the current study is in line with the previous study (Watkins et al., 2016). Among boys, we observed an association between third trimester maternal BPF concentrations with lower insulin and glucose concentrations and between third trimester maternal BPA concentrations with lower insulin concentrations after correction for glucose. It is important to note that the association of higher third trimester maternal BPF concentrations with lower glucose concentrations among boys and the association of higher third trimester maternal BPA concentrations with lower insulin concentrations after correction for glucose among boys did not remain significant after correction for multiple testing and could thus be a chance finding, while it is in line with the lower insulin concentrations that were found. We cannot compare these association with previous

studies, as these effects of BPF were not previously assessed in humans. These results suggest that fetal exposure to bisphenols could be associated with lower non-fasting glucose and insulin concentrations later in life, however, these results need to be replicated with fasting glucose and insulin levels to more accurately assess insulin sensitivity.

In conclusion, our results add to the growing body of evidence that fetal exposure of specific phthalates and bisphenols could influence later metabolic health in humans. We cannot draw conclusions about causality due to the observational nature of this study. The effect estimates observed in this study are small, but they are of interest from a public health perspective because of the widespread use of bisphenols and phthalates.

4.3. Possible underlying mechanisms

From animal studies it has been suggested that phthalates, via the PPAR α -receptor, can increase HDL-cholesterol concentrations and decrease production of triglycerides by influencing lipid oxidation and fatty acid synthesis and can increase the uptake of triglycerides (Hayashi et al., 2011; Colin et al., 2013; Laplante et al., 2007). It is not clear whether these mechanisms are as important in humans as they are in animals. The discrepancy between the findings in human studies and animal studies might thus be based on an interspecies difference of unknown origin. This could also explain the absence of effect of most studied phthalates on lipid concentrations observed in this study. On the other hand, laboratory studies have found that phthalate metabolites increase cytokine production and thus can lead to an adverse inflammatory environment (Jepsen et al., 2004). This could be the reason that higher exposure to phthalic acid was found to be associated with higher triglycerides concentrations among boys in this study. Specific phthalates have also been shown to affect insulin signaling (Rajagopal et al., 2019).

It has been found that BPA could affect fatty acid and glucose metabolism and it is thought that exposure to BPA alters the glucose-stimulated insulin response (Ji et al., 2019; Stahlhut et al., 2018; Alonso-Magdalena et al., 2006). However, in this study fetal exposure to BPA was not associated with lipid concentrations or glucose metabolism at 10 years old. In contrast to a previous study that suggested that BPF exposure could lead to hyperglycemia in zebrafish, we found that higher BPF concentrations during pregnancy were associated with lower insulin concentrations and not with higher glucose concentrations in children at 10 years old (Zhao et al., 2018).

Both phthalates and bisphenol A influence epigenetic regulatory mechanisms, which may need considerable time to lead to measurable changes in circulating biomarkers and this timing issue could possibly explain the discrepancies (Martinez-Ibarra et al., 2019; Somme et al., 2009; Goodrich et al., 2016). It has even been suggested that exposure to bisphenols influences health in subsequent generations (Bansal et al., 2019; Susiarjo et al., 2015). The relationship between exposure to endocrine disruptors and metabolic disturbances is potentially sex-specific (Miura et al., 2019). This could be due to sex-specific differences in PPAR-activity of phthalates and the estrogenic effect of BPA (Jalouli et al., 2003; Huang and Chen, 2017). In animals, epigenetic reprogramming is also considered a potential molecular mechanism that might underlie the sex-specific associations of the prenatal exposure to endocrine disruptors with metabolic outcomes later in life (McCabe et al., 2017; Neier et al., 2019). Further studies are needed to elucidate the underlying mechanisms of endocrine disruptors on lipid and glucose metabolism in humans specifically.

4.4. Methodological considerations

An important strength of this study is the population-based cohort design from fetal life onward, with repeated measurements of maternal phthalate and bisphenol concentrations. We also measured childhood lipid and glucose concentrations in a large number of mother-child

pairs. Selection bias due to selective loss to follow-up would be of concern in any study if the associations of prenatal phthalate and bisphenol concentrations with childhood lipid and glucose metabolism are different between participants and non-participants. This seems unlikely, but cannot be excluded with certainty, because the reported differences in ethnicity could influence diet and lifestyle choices which could influence the associations. Furthermore, we measured maternal phthalate and bisphenol urine concentrations once per trimester. It is possible that one urine sample per trimester does not accurately represent the concentration during the whole trimester because of the reported short biological half-lives of phthalates and bisphenols (Mattison et al., 2014; Braun et al., 2013). However, it has been found that a single urine sample reflects phthalate exposure up to three months (Hauser et al., 2004). Variability has also been reported to be biomarker specific, with strong correlations for LMWP metabolites and reasonable correlations for BPA and DEHP metabolites (Braun et al., 2012; Mahalingaiah et al., 2008). In this study, we found moderate variability for phthalates and high variability for bisphenols during pregnancy. We also used the overall mean to partly correct for the remaining variability. These associations were comparable to the trimester-specific associations. The main limitation of using a linear regression approach is that it does not take into account the association of the investigated exposure with other exposures. This could provide unreliable results if the exposures are highly correlated, which the exposures are not (Supplemental Table S2). We used non-fasting blood samples collected at different times during the day depending on the time of visit. This could have led to non-differential misclassification of children and consequently an underestimation of the associations. However, previous studies in adults have shown that non-fasting blood lipid concentrations can accurately predict increased risks of cardiovascular events later in life and that semi-fasting insulin resistance is moderately correlated with fasting values (Langsted and Nordestgaard, 2019; Hancox and Landhuis, 2011). We also assessed the association of phthalates and bisphenols with insulin, independent of glucose, to partially correct for the varying times since last meal and found comparable results. However, these results need to be replicated with non-fasting lipid, insulin and glucose concentrations in children. Future studies could also assess the effects of childhood exposure to phthalates and bisphenols on metabolic outcomes. Finally, we collected information on many potential confounders, although, as in any observational study, residual confounding due to unmeasured variables might remain an issue.

5. Conclusion

Results from our population-based prospective cohort study suggest persisting effects of fetal exposure to phthalates and bisphenols on childhood lipid concentrations which could be sex specific. Future studies are needed for replication and exploring underlying mechanisms.

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CRediT authorship contribution statement

Chalana M. Sol: Methodology, Formal analysis, Writing - original draft. **Susana Santos:** Methodology, Formal analysis, Writing - original draft. **Liesbeth Duijts:** Writing - review & editing. **Alexandros G. Asimakopoulos:** Investigation, Writing - review & editing. **Maria-Pilar Martinez-Moral:** Investigation, Writing - review & editing. **Kurunthachalam Kannan:** Investigation, Writing - review & editing. **Vincent W.V. Jaddoe:** Conceptualization, Resources, Supervision, Funding acquisition, Writing - review & editing. **Leonardo Trasande:** Conceptualization, Resources, Supervision, Funding acquisition, Writing - review & editing.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.envint.2020.106063>.

References

- Alonso-Magdalena, P., Morimoto, S., Ripoll, C., et al., 2006. The estrogenic effect of bisphenol A disrupts pancreatic beta-cell function in vivo and induces insulin resistance. *Environ. Health Perspect.* 114, 106–112.
- Bano, U., Memon, S., Shahani, M.Y., et al., 2019. Epigenetic effects of in utero bisphenol A administration: Diabetogenic and atherogenic changes in mice offspring. *Iran J. Basic Med. Sci.* 22, 521–528.
- Bansal, A., Li, C., Xin, F., et al., 2019. Transgenerational effects of maternal bisphenol A exposure on offspring metabolic health. *J. Dev. Orig. Health Dis.* 10, 164–175.
- Braun, J.M., Smith, K.W., Williams, P.L., et al., 2012. Variability of urinary phthalate metabolite and bisphenol A concentrations before and during pregnancy. *Environ. Health Perspect.* 120, 739–745.
- Braun, J.M., Sathyanarayana, S., Hauser, R., 2013. Phthalate exposure and children's health. *Curr. Opin. Pediatr.* 25, 247–254.
- Colin, S., Briand, O., Touche, V., et al., 2013. Activation of intestinal peroxisome proliferator-activated receptor- α increases high-density lipoprotein production. *Eur. Heart J.* 34, 2566–2574.
- Dolinoy, D.C., Huang, D., Jirtle, R.L., 2007. Maternal nutrient supplementation counteracts bisphenol A-induced DNA hypomethylation in early development. *Proc. Natl. Acad. Sci. U. S. A.* 104, 13056–13061.
- Dunder, L., Lejonklou, M.H., Lind, P.M., et al., 2019. Urinary bisphenol A and serum lipids: a meta-analysis of six NHANES examination cycles (2003–2014). *J. Epidemiol. Community Health* 73, 1012–1019.
- Golestanzadeh, M., Riahi, R., Kelishadi, R., 2019. Association of exposure to phthalates with cardiometabolic risk factors in children and adolescents: a systematic review and meta-analysis. *Environ. Sci. Pollut. Res. Int.*
- Goodrich, J.M., Dolinoy, D.C., Sanchez, B.N., et al., 2016. Adolescent epigenetic profiles and environmental exposures from early life through peri-adolescence. *Environ. Epigenet* 2:dvw018.
- Gules, O., Yildiz, M., Naseer, Z., et al., 2019. Effects of folic acid on testicular toxicity induced by bisphenol-A in male Wistar rats. *Biotech. Histochem.* 94, 26–35.
- Hancox, R.J., Landhuis, C.E., 2011. Correlation between measures of insulin resistance in fasting and non-fasting blood. *Diabetol. Metab. Syndr.* 3, 23.
- Hauser, R., Meeker, J.D., Park, S., et al., 2004. Temporal variability of urinary phthalate metabolite levels in men of reproductive age. *Environ. Health Perspect.* 112, 1734–1740.
- Hayashi, Y., Ito, Y., Yamagishi, N., et al., 2011. Hepatic peroxisome proliferator-activated receptor α may have an important role in the toxic effects of di(2-ethylhexyl) phthalate on offspring of mice. *Toxicology* 289, 1–10.
- Hornung, R.W., 1990. RL Estimation of average concentration in the presence of non-detectable values. *Appl. Occup. Environ. Hyg.* 5, 46–51.
- Huang, Q., Chen, Q., 2017. Mediating roles of PPARs in the effects of environmental chemicals on sex steroids. *PPAR Res.* 2017, 3203161.
- Jaddoe, V.W., van Duijn, C.M., Franco, O.H., et al., 2012. The Generation R Study: design and cohort update 2012. *Eur. J. Epidemiol.* 27, 739–756.
- Jalouli, M., Carlsson, L., Ameen, C., et al., 2003. Sex difference in hepatic peroxisome proliferator-activated receptor α expression: influence of pituitary and gonadal hormones. *Endocrinology* 144, 101–109.
- James-Todd, T., Stahlhut, R., Meeker, J.D., et al., 2012. Urinary phthalate metabolite concentrations and diabetes among women in the National Health and Nutrition Examination Survey (NHANES) 2001–2008. *Environ. Health Perspect.* 120, 1307–1313.
- Jepsen, K.F., Abildtrup, A., Larsen, S.T., 2004. Monophthalates promote IL-6 and IL-8 production in the human epithelial cell line A549. *Toxicol. In Vitro* 18, 265–269.
- Ji, H., Song, N., Ren, J., et al., 2019. Metabonomics reveals bisphenol A affects fatty acid and glucose metabolism through activation of LXR in the liver of male mice. *Sci. Total Environ.* 703.
- Kooijman, M.N., Kruijthof, C.J., van Duijn, C.M., et al., 2016. The Generation R Study: design and cohort update 2017. *Eur. J. Epidemiol.* 31, 1243–1264.
- Kruijthof, C.J., Kooijman, M.N., van Duijn, C.M., et al., 2014. The Generation R Study: Biobank update 2015. *Eur. J. Epidemiol.* 29, 911–927.
- Lang, I.A., Galloway, T.S., Scarlett, A., et al., 2008. Association of urinary bisphenol A concentration with medical disorders and laboratory abnormalities in adults. *JAMA* 300, 1303–1310.
- Langsted, A., Nordestgaard, B.G., 2019. Nonfasting versus fasting lipid profile for cardiovascular risk prediction. *Pathology* 51, 131–141.
- Laplante, M., Festuccia, W.T., Soucy, G., et al., 2007. Involvement of adipose tissues in the early hypolipidemic action of PPAR γ agonism in the rat. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 292, R1408–R1417.
- Lee, K.I., Chiang, C.W., Lin, H.C., et al., 2016. Maternal exposure to di(2-ethylhexyl) phthalate exposure deregulates blood pressure, adiposity, cholesterol metabolism and social interaction in mouse offspring. *Arch. Toxicol.* 90, 1211–1224.
- Li, G., Chang, H., Xia, W., et al., 2014. F0 maternal BPA exposure induced glucose intolerance of F2 generation through DNA methylation change in Gck. *Toxicol. Lett.* 228, 192–199.
- Li, M.X., Yeung, J.M., Cherny, S.S., et al., 2012. Evaluating the effective numbers of independent tests and significant p-value thresholds in commercial genotyping arrays and public imputation reference datasets. *Hum. Genet.* 131, 747–756.
- Mahalingaiah, S., Meeker, J.D., Pearson, K.R., et al., 2008. Temporal variability and predictors of urinary bisphenol A concentrations in men and women. *Environ. Health Perspect.* 116, 173–178.
- Martinez-Ibarra, A., Martinez-Razo, L.D., Vazquez-Martinez, E.R., et al., 2019. Unhealthy levels of phthalates and bisphenol A in Mexican pregnant women with gestational diabetes and its association to altered expression of miRNAs involved with metabolic disease. *Int. J. Mol. Sci.* 20.
- Mattison, D.R., Karyakina, N., Goodman, M., et al., 2014. Pharmacokinetics of selected exogenous and endogenous estrogens: a review of the data and identification of knowledge gaps. *Crit. Rev. Toxicol.* 44, 696–724.
- McCabe, C., Anderson, O.S., Montrose, L., et al., 2017. Sexually dimorphic effects of early-life exposures to endocrine disruptors: sex-specific epigenetic reprogramming as a potential mechanism. *Curr. Environ. Health Rep.* 4, 426–438.
- Meng, Z., Wang, D., Yan, S., et al., 2018. Effects of perinatal exposure to BPA and its alternatives (BPS, BPF and BPAF) on hepatic lipid and glucose homeostasis in female mice adolescent offspring. *Chemosphere* 212, 297–306.
- Miura, R., Araki, A., Minatoya, M., et al., 2019. An epigenome-wide analysis of cord blood DNA methylation reveals sex-specific effect of exposure to bisphenol A. *Sci. Rep.* 9, 12369.
- Mose, T., Knudsen, L.E., Hedegaard, M., et al., 2007. Transplacental transfer of mono-methyl phthalate and mono(2-ethylhexyl) phthalate in a human placenta perfusion system. *Int. J. Toxicol.* 26, 221–229.
- Nahar, M.S., Liao, C., Kannan, K., et al., 2015. In utero bisphenol A concentration, metabolism, and global DNA methylation across matched placenta, kidney, and liver in the human fetus. *Chemosphere* 124, 54–60.
- Neier, K., Cheatham, D., Bedrosian, L.D., et al., 2019. Longitudinal metabolic impacts of perinatal exposure to phthalates and phthalate mixtures in mice. *Endocrinology* 160, 1613–1630.
- Nguyen, A.N., de Barse, L.M., Tiemeier, H., et al., 2017. Maternal history of eating disorders: Diet quality during pregnancy and infant feeding. *Appetite* 109, 108–114.
- Onyenekwu, C.P., Hoffmann, M., Smit, F., et al., 2014. Comparison of LDL-cholesterol estimate using the Friedewald formula and the newly proposed de Cordova formula with a directly measured LDL-cholesterol in a healthy South African population. *Ann. Clin. Biochem.* 51, 672–679.
- Pauwels, S., Ghosh, M., Duca, R.C., et al., 2017. Maternal intake of methyl-group donors affects DNA methylation of metabolic genes in infants. *Clin. Epigenetics* 9, 16.
- Perng, W., Watkins, D.J., Cantoral, A., et al., 2017. Exposure to phthalates is associated with lipid profile in peripubertal Mexican youth. *Environ. Res.* 154, 311–317.
- Philips, E.M., Jaddoe, V.W.V., Asimakopoulos, A.G., et al., 2018. Bisphenol and phthalate concentrations and its determinants among pregnant women in a population-based cohort in the Netherlands, 2004–5. *Environ. Res.* 161, 562–572.
- Rajagopal, G., Bhaskaran, R.S., Karunde, B., 2019. Maternal di-(2-ethylhexyl) phthalate exposure alters hepatic insulin signal transduction and glucoregulatory events in rat F1 male offspring. *J. Appl. Toxicol.* 39, 751–763.
- Ranciere, F., Lyons, J.G., Loh, V.H., et al., 2015. Bisphenol A and the risk of cardiometabolic disorders: a systematic review with meta-analysis of the epidemiological evidence. *Environ. Health* 14, 46.
- Russo, G., Barbato, F., Mita, D.G., et al., 2019. Occurrence of Bisphenol A and its analogues in some foodstuff marketed in Europe. *Food Chem. Toxicol.* 131.
- Santos, S., Zugna, D., Pizzi, C., et al., 2019. Sources of confounding in life course epidemiology. *J. Dev. Orig. Health Dis.* 10, 299–305.
- Schettler, T., 2006. Human exposure to phthalates via consumer products. *Int. J. Androl.*

- 29, 134–139 discussion 181–135.
- Somm, E., Schwitzgebel, V.M., Toulotte, A., et al., 2009. Perinatal exposure to bisphenol a alters early adipogenesis in the rat. *Environ. Health Perspect.* 117, 1549–1555.
- Stahlhut, R.W., Myers, J.P., Taylor, J.A., et al., 2018. Experimental BPA exposure and glucose-stimulated insulin response in adult men and women. *J. Endocr. Soc.* 2, 1173–1187.
- Susiarjo, M., Xin, F., Bansal, A., et al., 2015. Bisphenol a exposure disrupts metabolic health across multiple generations in the mouse. *Endocrinology* 156, 2049–2058.
- Taylor, J.A., Sommerfeld-Sager, J.M., Meng, C.X., et al., 2018. Reduced body weight at weaning followed by increased post-weaning growth rate interacts with part-per-trillion fetal serum concentrations of bisphenol A (BPA) to impair glucose tolerance in male mice. *PLoS ONE* 13.
- Vafeiadi, M., Roumeliotaki, T., Myridakis, A., et al., 2016. Association of early life exposure to bisphenol A with obesity and cardiometabolic traits in childhood. *Environ. Res.* 146, 379–387.
- Vafeiadi, M., Myridakis, A., Roumeliotaki, T., et al., 2018. Association of early life exposure to phthalates with obesity and cardiometabolic traits in childhood: sex specific associations. *Front. Public Health* 6, 327.
- Watkins, D.J., Peterson, K.E., Ferguson, K.K., et al., 2016. Relating phthalate and BPA exposure to metabolism in peripubescence: the role of exposure timing, sex, and puberty. *J. Clin. Endocrinol. Metab.* 101, 79–88.
- Ye, X., Pierik, F.H., Hauser, R., et al., 2008. Urinary metabolite concentrations of organophosphorous pesticides, bisphenol A, and phthalates among pregnant women in Rotterdam, the Netherlands: the Generation R study. *Environ. Res.* 108, 260–267.
- Zhao, F., Wang, H., Wei, P., et al., 2018. Impairment of bisphenol F on the glucose metabolism of zebrafish larvae. *Ecotoxicol. Environ. Saf.* 165, 386–392.