Body Transformation in Life after Tumor: Long-term Consequences for Endocrinology, Metabolism and Bone

With a special focus on craniopharyngioma

S.S. van Santen

Body Transformation in Life after Tumor: Long-term Consequences for Endocrinology, Metabolism and Bone

With a special focus on craniopharyngioma

S.S. van Santen

Financial support by the Dutch Heart Foundation for the publication of this thesis is gratefully acknowledged.

Furthermore, financial support for printing of this thesis was generously provided by:
ACE Pharmaceuticals
Chipsoft
GOODLIFE Pharma B.V.
Medical Images/ISM
Pfizer B.V.
Erasmus University Rotterdam
Stichting Research Klinische Endocrinologie

The studies described in this thesis were conducted at the Erasmus University Medical Center, Rotterdam, the Netherlands, and supported by the Princess Máxima Center for Pediatric Oncology, Utrecht, the Netherlands.

Cover design: Annika Geurtsen
Medical illustrations: Ana G.G. de Lima
Lay-out and design: Annika Geurtsen
Printing: ProefschriftMaken II www.proefschriftmaken.nl

ISBN: 978-94-6423-557-9

Body Transformation in Life after Tumor: Long-term Consequences for Endocrinology, Metabolism and Bone

With a special focus on craniopharyngioma

Lichaamstransformatie in het leven na een tumor: langetermijnconsequenties voor endocrinologie, metabolisme en botgezondheid

met bijzondere aandacht voor het craniopharyngioom

Thesis

to obtain the degree of Doctor from the
Erasmus University Rotterdam
by command of the
rector magnificus

Prof. dr. A.L. Bredenoord

and in accordance with the decision of the Doctorate Board.
The public defence shall be held on
Wednesday 15th of December 15.30 hrs

by
Selveta Sanne van Santen
born in Dordrecht

Erasmus University Rotterdam

Promotiecommissie

Promotoren:	Prof. dr. A.J. van der Lelij Prof. dr. M.M. van den Heuvel-Eibrink
Overige leden:	Prof. dr. E.F.C. van Rossum Prof. dr. A.M. Pereira Arias Dr. A.J.P.E. Vincent
Copromotor:	Dr. S.J.C.M.M. Neggers

Table of Contents

1

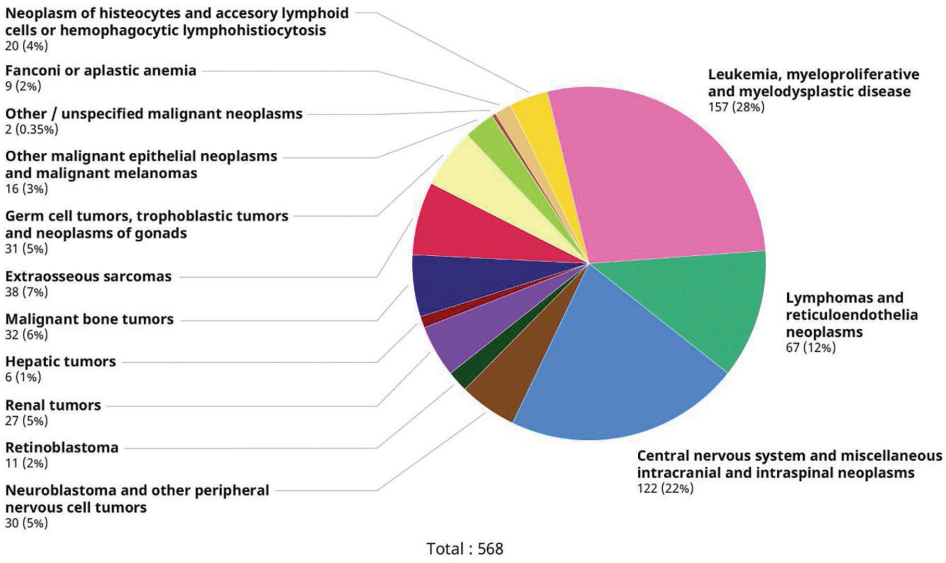
General Introduction

Cancer: an extensive health problem

Two out of five individuals develop cancer in their lives.[1] In the Netherlands, it is the leading cause of death in men and shares the first place with cardiovascular disease in women.[2] One out of three men and one out of four women die due to this devastating disease.[2] This impressive number underlines that cancer is a major burden and concern in modern society. The incidence of cancer has grown over the years: in 2016, 17.2 million cases of cancer were diagnosed worldwide, and 8.9 million people died of it.[3] It costed 213.2 million disability adjusted life years (DALYs) in only one year.[3] Although the average yearly incidence rates for cancer increased between 2006 and 2016 in most countries, the death rates relatively decreased.[3]

Cancer is not solely a disease of adults or elderly, but can occur in children as well. Cancer in children differs from cancer in adults in cancer type, causes of late death and survival rates. Although childhood cancer represents only a small part of all cancers (~1%),[4] it is still a very important cause of death in their age category, only exceeded by congenital anomalies and accidents.[5] Globally, estimates for the yearly incidence of cancer in the age group 0-14 years range from 163,284 to 184,856 individuals.[4] In the Netherlands, around 600 children are newly diagnosed with cancer yearly.[6] Childhood cancer is a very heterogeneous disease.[7] Common childhood cancer types are leukemias, lymphomas, and immature tumors, i.e. blastomas as well as soft tissue sarcomas, epithelial or germ cell tumors (Figure 1), whereas adults with cancer suffer mostly from more differentiated forms, such as carcinomas and chronic leukemias (Figure 2).[4,6,8] Current cancer treatment often combines multiple treatment modalities such as surgery, chemotherapy, stem cell transplantation, radiotherapy, and novel strategies such as immunotherapy.[9] Not all tumors (or 'neoplasms') are necessarily malignant or considered to be cancer, but benign tumors can cause problems as well.

Figure 1 Types of childhood cancer



Mean yearly incidence of different types of childhood cancer in the Netherlands according to the SKION basal registration.[6]

Survival after tumors

While the number of deaths due to a neoplasm has increased over time in adults, it decreased in children (Figure 3).[5] The 5-year survival rate in adults with cancer is approximately 54%,[10] while children have a much better prognosis with 80% survival.[1] Survival rates in children improved due to better care and the treatment modalities mentioned above.[7,9,11] This has led to a large group of between 300,000 and 500,000 living childhood cancer survivors in Europe alone.[11,12] However, childhood cancer survivors remain at increased risk of death their entire life, especially after intensive treatment.[13,14] Their late overall cumulative mortality is 13-30% at 30-40 years from diagnosis.[15-20] The main cause of late death after childhood cancer is recurrence of the original cancer, especially during the first 15 years post-diagnosis; other frequent causes are subsequent neoplasms, cardiac and pulmonary death.[16-18,20-27] Survivors of brain tumors seem even more at risk of premature death: the cumulative mortality of pediatric medulloblastoma survivors was 23% after only 15 years.[28,29] Benign neoplasms can be associated with

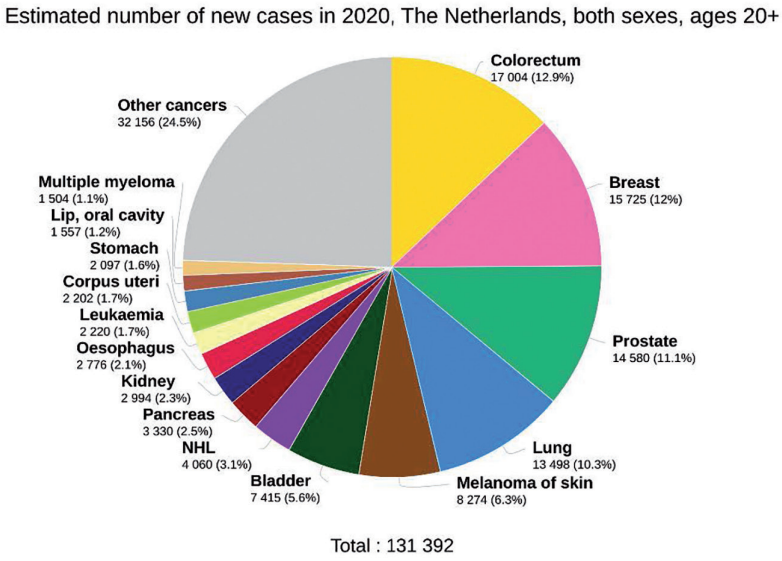
decreased survival rates as well. For instance, after craniopharyngioma, a benign brain tumor after which only 66-85% of the patients survive after 20 years.[30-32] Another example of benign brain tumors are pituitary adenomas, of which the 10-year survival rate is estimated at 69% in men and 76% in women.[33]

Chronic diseases

Next to survival, quality of life is an important consideration. In the whole world, one in three adults has multiple chronic conditions.[34] In high-income countries, the most important conditions leading to decrease in DALYs are non-communicable diseases, including ischemic heart disease, stroke, diabetes, lung cancer, and pain in the neck and back.[34,35] Chronic diseases form a serious challenge, as they are a burden on the individual as well the economy.[36] This underlines the importance of preventing of these diseases if possible, especially in high risk groups. Also, approximately 313 million of surgeries are performed worldwide each year, which can be accompanied with complications leading to death after surgery in 8% of the cases.[37-39] Postoperative complications are globally the third largest contributor to mortality.[39] Postoperative complications after major surgery in general is associated with lower survival.[40]

A group at high risk for chronic diseases are childhood cancer survivors; nearly all (95-99%) suffer from a chronic health condition at the age of 50.[41,42] The surgery, chemotherapeutics and radiotherapy that survivors owe their life to, have long-term consequences.[18,42-45] Any organ system may be impaired in childhood cancer survivors,[42,46] but endocrine conditions are one of the most common adverse health outcomes, and occur in 62% of survivors.[42] Not only childhood cancer survivors are affected by endocrine conditions; they are highly prevalent in patients with benign brain tumors, patients with a primary auto-immune deficiency, those with obesity and in women (with menopause being an endocrine condition that naturally occurs with advancing age).[47-53] Endocrine conditions can cause metabolic alterations.[54] This contributes to an increased risk of cardiovascular disease[9,55-62] and stroke[63-65] (Table 1).[42,46,49]

Figure 2 Types of adulthood cancer[8]

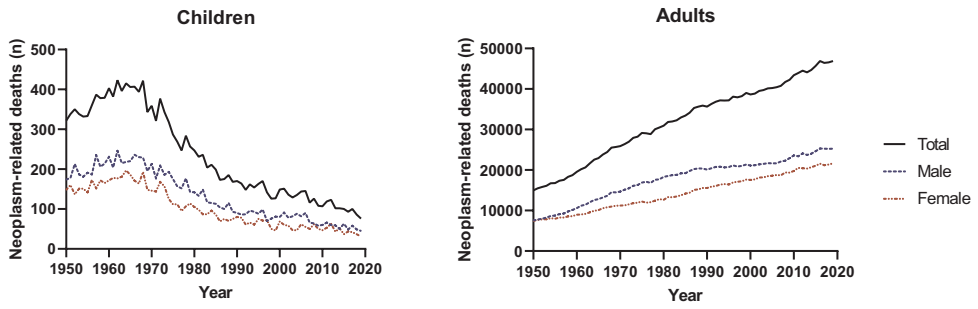


Endocrinology

Endocrinology is very important for homeostasis (or stable equilibrium in physiological processes) in the body. It is the study of hormones, which were classically defined as a chemical messenger that spreads from cell to cell along the bloodstream.[66] The first structures of hormones were discovered in 1849.[66] The masters of the endocrine system are the hypothalamus and pituitary gland. Figure 4 gives an illustration of a healthy pituitary gland, as imaged by magnetic resonance imaging (MRI), while Figure 5 illustrates the general anatomy. In general, the hypothalamus produces hormones that stimulate the pituitary, and the pituitary stimulates other endocrine glands such as the thyroid, reproductive organs, breasts, liver, kidney and adrenals, to produce hormones as well.[67] These hormones in turn provide negative feedback towards pituitary and hypothalamus.[67] Endocrine glands regulate a variety of functions like growth, bone health, metabolism and blood glucose control, and sexual and reproductive functions.[68-70] These glands make necessary changes to the internal environment in response to the external environment.[68-70] It is a system that continuously aims to maintain the balance. As the final hormone cleverly provides its

own negative feedback, the stimulus for producing this final hormone diminishes. Thereafter the negative feedback diminishes as well, leading to an increase of this hormone back again: a balance is established.

Figure 3 Neoplasm-related deaths in children and adults in the Netherlands (1950-2019)



Source: CBS.[5] Yearly cumulative neoplasm related deaths in children (0-20 years) and adults (20 years or older) in the Netherlands.

An example of a hormone system for which careful balancing is needed, is the regulation of the blood glucose. The pancreas produces the blood glucose-lowering hormone insulin, and the hormone glucagon, which in turn increases blood glucose levels. The most important endocrine organ for blood glucose regulation is the pancreas, which produces two key hormones: insulin (through the beta-cells in the pancreatic islets) and glucagon (through the alpha-cells).[71,72] Insulin decreases blood glucose levels after food intake, while glucagon increases glucose in the circulation by increasing take up of glucose stored in the form of glycogen in the liver (glycogenolysis), new-formed glucose from other components (gluconeogenesis) by the liver and possibly release of glycerol and fatty acids from triacylglycerol of white adipose tissue (lipolysis).[71,73,74]

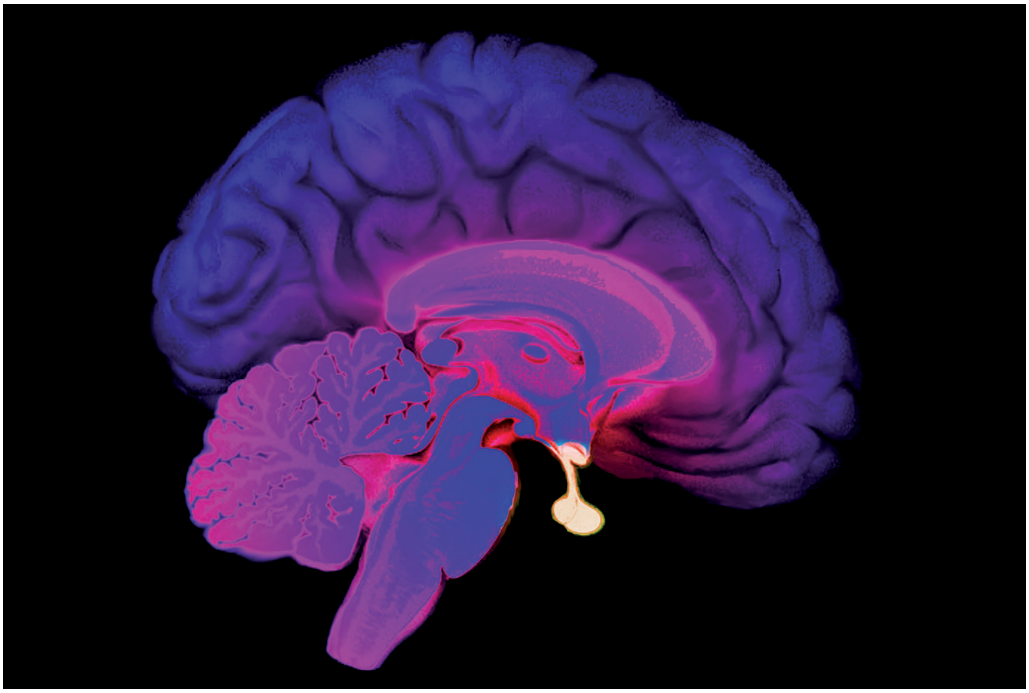
Table 1 Endocrine and metabolic late effects after childhood cancer[258,284-286]

Organ system	Potential complication	Risk factor
Hypothalamic-pituitary axis	Pituitary deficiencies	Total body irradiation Neurosurgery Cranial radiotherapy
	· Growth hormone	Immune checkpoint inhibitors
	· Follicle-stimulating hormone and luteinizing hormone	Immune checkpoint inhibitors
	· Adrenocorticoid stimulating hormone	Immune checkpoint inhibitors
	· Thyroid stimulating hormone	Immune checkpoint inhibitors
	· ADH	
	Delayed puberty	Pelvic radiotherapy, glucocorticoids
	Excess pituitary hormones Precocious puberty Hyperprolactinemia	Cranial radiotherapy
Hypothalamus	Hypothalamic obesity	Tumor in its surroundings Neurosurgery Cranial radiotherapy
	Underweight (diencephalic syndrome)	Damage to AgRP-expressing neurons
	Alterations in body temperature	Tumor in its surroundings Neurosurgery Cranial radiotherapy
Thyroid	Primary hyperthyroidism	Radiotherapy on the head, neck, cervical spine, whole spine or total body Iodine-131-metaiodobenzyl-guanidine (I-131-MIBG) Tyrosine kinase inhibitors Immune checkpoint inhibitors
	Primary hypothyroidism	Radiotherapy on the head, neck, cervical spine or whole spine Tyrosine kinase inhibitors Immune checkpoint inhibitors
	Benign and malignant thyroid nodules	I-131-MIBG

	Auto-immune thyroiditis	Hematopoietic stem cell transplantation Immune checkpoint inhibitors Interferon
Adrenals	Adrenal insufficiency	Direct insult due to tumor expansion or surgery Glucocorticoids (transient)
Gonads	♀ Gonadal dysfunction (premature ovarian insufficiency)	Alkylating agents Heavy metals Radiotherapy on the abdomen, pelvis, sacral or whole spine, or total body I-131-MIBG Bevacizumab
	♂ Leydig and germ cell dysfunction	Alkylating agents Radiotherapy to the pelvis or total body
Bone	Impaired final height/short stature	Growth hormone deficiency Glucocorticoids Radiotherapy on the spine, chest, abdomen or total body Retinoic acid Hedgehog pathway inhibitors
	Low bone mineral density/ Osteoporosis	Glucocorticoids Methotrexate
Metabolism	Obesity	Hypothalamic damage Hypopituitarism Cranial radiotherapy and total body irradiation Glucocorticoids Surgery
	Diabetes mellitus	Radiotherapy on the abdomen or total body Alkylating agents
	The metabolic syndrome	Radiotherapy to the head, abdomen and total body

Alkylating agents is a drug category containing busulfan, carmustine, chlorambucil, cyclophosphamide, dacarbazine, ifosfamide, lomustine, mechlorethamine, melphalan, procarbazine, temozolomide, thiotepa. Examples of applied heavy metals are carboplatin and cisplatin. Examples of tyrosine kinase inhibitors are imatinib, sorafenib and sunitinib. An immune checkpointinhibitor-anti-CTLA4 monoclonal antibody is ipilimumab.[30,231,285-293]

Figure 4 The Pituitary



3D MRI, midsagittal section, showing a normal brain, highlighting the pituitary gland and the pituitary stalk (yellow). By: medicalimages.com.

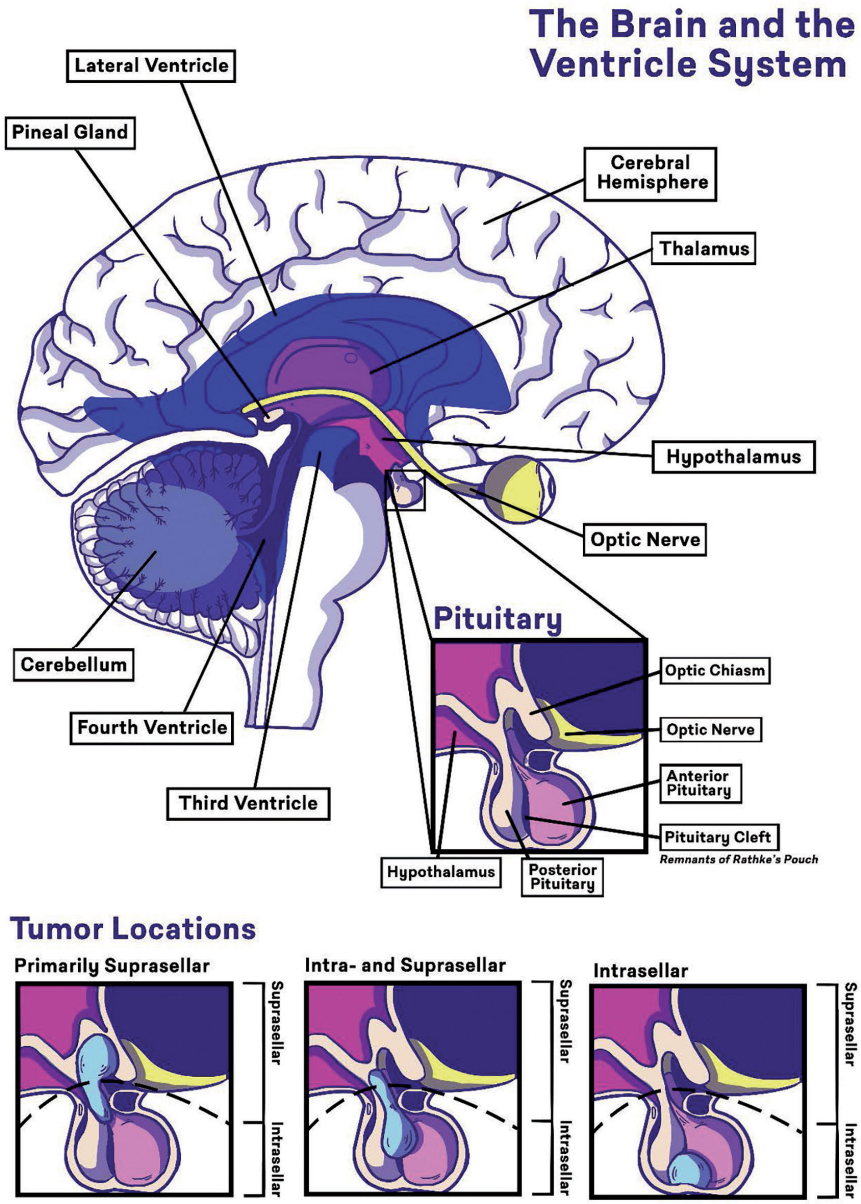
White adipose tissue is most known for its ability of energy storage, but is potentially the largest endocrine organ in humans as well.[75,76] White adipose produces the hormones leptin and adiponectin.[76] Leptin inhibits food intake by sending anorexigenic signals to the brain.[76] Adiponectin is important for insulin sensitivity and lipid metabolism, and has anti-atherogenic properties.[77] In case of excessive nutrition, fat accumulates excessively. This is called obesity; a disease in which adipose tissue dysfunctions and health may be impaired.[75] Obesity occurs quite regularly in the general population: in 2019, 13% of all Dutch citizens were obese.[78] People with a history of childhood cancer are at even higher risk of obesity; it is 3.8 times higher in survivors than in the general population.[60] Obesity is associated with diseases such as type 2 diabetes mellitus, non-alcoholic fatty liver disease, cancer and cerebro- and cardiovascular disease.[75,79,80] Endocrine alterations occur

frequently in case of obesity.[81,82] Obese subjects have high leptin levels while the exerted anorexigenic effect is lowered (leptin resistance).[76] On the other hand, adiponectin levels are decreased and therefore the impact on anti-atherogenicity and insulin sensitivity is lowered as well.[77] The combined presence of too much accumulated fat (adiposity), decreased insulin sensitivity (insulin resistance), abnormal serum lipid levels (dyslipidemia) and high blood pressure (hypertension) is called the metabolic syndrome.[79,83-99] The metabolic syndrome quintuples type 2 diabetes risk, and doubles the risk of cardio- and cerebrovascular disease.[79,83-99]

The embryonic development of the hypothalamus and pituitary

As mentioned, the hypothalamus and pituitary are the master glands of the endocrine system. The pituitary consists of an anterior and a posterior part, the adenohypophysis and neurohypophysis.[100] The anterior and posterior lobe have distinct functions and are derived from different tissues in embryonal development (Figure 6).[100,101] During embryogenesis, the neurohypophysis and endocrine hypothalamus are derived from neural ectoderm, while the adenohypophysis is derived from an invagination of oral ectoderm, called Rathke's pouch.[100,102,103] The neurohypophysis stores and secretes antidiuretic hormone and oxytocin, hormones which are produced in the hypothalamus.[100] The adenohypophysis, or anterior pituitary, has lactotropic, somatotropic, gonadotropic, corticotropic and thyrotropic cells which produce the hormones prolactin, growth hormone, luteinizing hormone (LH) and follicle stimulating hormone (FSH), adrenocorticotropic hormone (ACTH) and thyroid stimulating hormone (TSH), respectively.[100]

Figure 5 Normal anatomy of the brain, ventricle system and pituitary, and possible locations for pituitary tumors

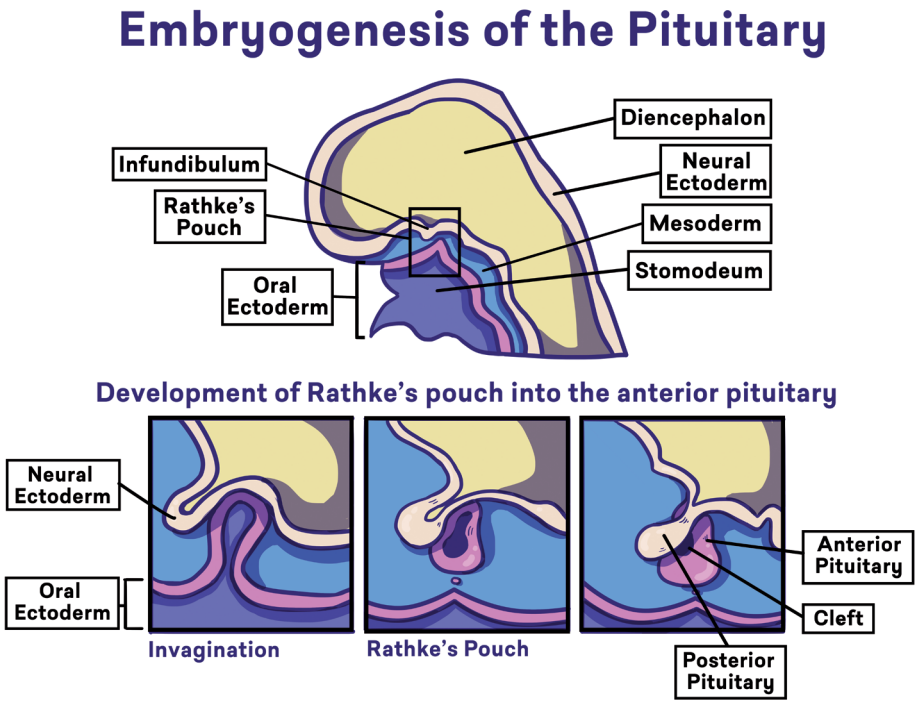


By: Ana G.G. de Lima.

Functions of the posterior pituitary

Antidiuretic hormone, or AVP is released after a stimulus by increasing osmolality and inhibits diuresis. This is established by stimulation of thirst, and acting on G-coupled V2 receptors in the kidneys leading to increased water reabsorption.[104] When oxytocin is released from the posterior pituitary, it stimulates the epididymal and uterine muscle contraction, and stimulates the nipples for breastfeeding.[105,106] Furthermore, oxytocin is important for social functioning, as it enhances sociality and influences behavior regarding sexual contact and aggressive attacks.[105,107]

Figure 6 Embryogenesis of the pituitary gland



The anterior pituitary is developed from neural ectoderm, and the posterior pituitary from oral ectoderm. By: Ana G.G. de Lima.

Functions of the anterior pituitary

The hypothalamo-prolactin axis

The hypothalamo-prolactin axis is predominantly inhibitory, as opposed to the other axis.[108] Dopaminergic neurons in the hypothalamus inhibit the lactotropic cells from producing prolactin, a hormone named after its main function, which is promoting lactation.[108] Prolactin increases dopamine secretion and therefore provides its own negative feedback.[108] During lactation, a physiological state of hyperprolactinemia occurs due to a loss of sensitivity of this feedback system, and suckling causes a further release of prolactin.[108,109] Prolactin has several other functions next to lactation in the maternal adaptation to pregnancy, which relate to maternal behavior, bone metabolism, lipid metabolism, neurogenesis, glucose homeostasis, appetite regulation, reproduction, stress responses and oxytocin secretion.[108,110-115]

Hypothalamo-pituitary-somatotropic axis

Growth hormone is mostly known for the properties of enhancing linear growth but is also essential for its metabolic properties and for increasing bone mass.[69,116,117] Growth hormone-releasing hormone (GHRH) from the hypothalamus stimulates the anterior pituitary to release growth hormone.[69,118,119] This in turn stimulates production of insulin-like growth factors (IGFs).[69,118,119] Negative feedback is provided from somatostatin produced in the hypothalamus, ghrelin (the 'hunger hormone') produced in the stomach, estrogens and androgens from the gonadal organs, and from growth hormone and IGF themselves.[69,118,119] Growth hormone is released in a pulsatile manner; factors that stimulate release are hypoglycemia and stress, while hyperglycemia suppresses its release.[69] The liver is stimulated to produce insulin-like growth factor 1 (IGF-1), but the relationship between levels of growth hormone and IGF-1 are not linear: many other factors influence IGF-1 concentrations, such as age, puberty status and BMI.[69,120] When aging, growth hormone and IGF-1 levels decline until the levels are very low at the age of 60 years and older.[121] IGF-1 and growth hormone have slightly different

metabolic abilities: growth hormone leads to elevated insulin and glucose levels and reduced insulin sensitivity, while IGF-1 reduces insulin and improves insulin sensitivity.[122] IGF-1 and growth hormone both enhance the energy expenditure and lower protein oxidation.[122] As the secretion of growth hormone is stimulated by exercise and inhibited by food intake, measuring a randomly taken growth hormone measurement is unreliable to evaluate either growth hormone deficiency or oversecretion.[69] Instead, growth hormone deficiencies are evaluated by tests which stimulate growth hormone production, such as the insulin tolerance test, GHRH or intravenous arginine infusion test.[69] When growth hormone is replaced after a deficiency, this causes an increase in lean body mass, potentially an increase in bone mineral density, and decrease in fat mass.[123-125] If excessive growth hormone secretion is expected (acromegaly), this can be evaluated with an oral glucose tolerance test.[126]

The hypothalamic-pituitary-gonadal axis

The hypothalamic-pituitary-gonadal axis has important reproductive functions and influences bone mineralization.[127] It relies on the gonadotrophs LH and FSH in the pituitary, which are stimulated by hypothalamic GnRH in a pulsatile way.

In females of reproductive age, LH stimulates production of the pre-hormone androstenedione by the theca cells in the ovaries.[104,128] FSH stimulates ovarian follicular growth in the first place.[104,128] Furthermore, FSH stimulates the granulosa cells to generate progesterone, and estrogen derived from androstenedione through conversion with aromatase enzymes.[104,128] Estrogen has important functions. It is involved in multiple organ systems including reproduction and bone maturation, and it protects against bone mass loss and cardiovascular disease.[129-131] Estrogen levels may rise towards excessive levels in case of obesity, as androstenedione levels are increased 3 to 4-fold.[132] Consequently, the target organs such as breasts and ovaries are stimulated, which unfortunately gives a risk of developing neoplasias.[132] After the reproductive age, around the ages of 46-52 years, the

menopause occurs.[133,134] The natural course of life in women leads to this final menstrual period, which is a form of low gonadal hormones (hypogonadism) combined with high FSH levels.[135] The final menstrual period is defined as 12 months of amenorrhea and FSH levels being two standard deviations higher than the mean level of women of peak reproductive age.[134-136] The progressive decline of estrogen levels is accompanied with a decline in bone mass and increase in percentual fat mass, especially at the abdominal/visceral region.[134,137]

In males, LH stimulates release of testosterone by the Leydig cells in the testes, while FSH stimulates production of (androgen-binding) protein and initiation of spermatogenesis by the Sertoli cells in the testes.[104,138,139] In case of obesity, testosterone production does not change, but sex-hormone binding proteins are decreased, which leads to increased clearance rates and low levels of circulating testosterone.[132] Testosterone levels correlate with higher lean (muscle) mass and lower fat mass in men.[140]

The hypothalamic-pituitary-adrenal axis

The hypothalamic-pituitary-adrenal (HPA-) axis is important for stress-responses.[141] Corticostropin-releasing hormone (CRH) is synthesized in the hypothalamus and stimulates the anterior pituitary to produce ACTH in a pulsatile manner.[104,142,143] ACTH thereafter stimulates the zona fasciculata in the cortex of the adrenals to release the glucocorticoid cortisol, the 'stress hormone'.[104,142,143] Cortisol signals negative feedback towards the hypothalamus and pituitary.[142] Activation of the HPA-axis leads to decreased activity of the growth hormone and gonadal axis.[144-146]

Cortisol can affect almost all organ systems, such as the cardiovascular, immune and respiratory system, and induce an inflammatory and/or stress response.[147,148] In case of a continuous threat of internal or external stressors to the body, at first the sympathetic nervous system is activated, inducing the 'flight or flight' response with release of catecholamines (i.e. epinephrine) by the adrenals.[149]

If the stressor remains, the HPA-axis is activated, and cortisol induces catabolic mechanisms.[150,151] This especially aims to maintain energy substrates in the circulation at high enough levels to provide the body with energy in these flight-or-flight conditions, especially the brain.[151,152] These energy substrates can be fatty acids, amino acids and glucose.[152] Cortisol stimulates the liver's gluconeogenesis and glycogenolysis, and inhibits glycogen synthesis, inducing higher glucose levels.[151,153,154] Cortisol causes the muscles to decrease their glucose uptake due to insulin resistance, which further increases circulating glucose levels.[152,154,155] Furthermore, the muscles are stimulated by cortisol to liberate amino acids by increased breakdown of proteins (proteolysis). In fat tissue, lipolysis is enhanced, freeing fatty acids.[152,154] Cortisol increases the activity catecholamines and increases leptin concentrations.[152,154,156]

Cortisol release is characterized by a circadian rhythm, as the amplitude of ACTH and cortisol release is altered during the day: it is at the highest peak at 8 AM and low between 8 PM and 2 AM.[104,143,157] Physical or physiological stress may increase ACTH and cortisol production, while exogenous glucocorticoids decrease CRH release and therefore impair ACTH and cortisol release as well, as a negative feedback.[104,158] Low cortisol levels (hypocortisolism) can be caused by primary adrenal insufficiency, such as Addison's disease (due to an autoimmune response),[159] or secondary adrenal insufficiency, which is caused by low levels of ACTH.[154] Hypocortisolism can be life threatening and present with hypotension and shock.[154,159] Hypocortisolism can be tested with early morning cortisol, (low dose) ACTH test, insulin tolerance test or metapyrone test.[159,160] High cortisol levels (hypercortisolism or Cushing syndrome) can occur due to a tumor on the adrenal gland or in the pituitary.[152,161] Endogenous hypercortisolism can be diagnosed by determining late night salivary cortisol or urine free cortisol, and by the low-dose dexamethasone suppression test.[150,161] Chronic hypercortisolism causes an increased amount of visceral or abdominal adipose tissue and reduced bone mineral density.[152,156]

The hypothalamus-pituitary-thyroid axis

Thyroid hormones are highly important for metabolic pathways, and cause a general increase in basal energy expenditure.[162] Furthermore, thyroid hormones are important in brain and skeletal development in children, and bone maintenance in adults.[163,164] The hypothalamus-pituitary-thyroid axis starts with the release of thyrotropin-releasing hormone (TRH) in the hypothalamus, stimulating the pituitary to produce thyrotropin (TSH).[165] Further on the axis, TSH stimulates the thyroid to produce thyroxine (T4) and triiodothyronine (T3). T3 is a biologically active thyroid hormone, and the thyroid receptor's main ligand due a 10-fold higher affinity with T3 than T4.[166] T3 is predominantly produced in peripheral tissues, especially the liver and kidney, after conversion from T4; only <20% is produced by the thyroid itself.[166-168] T3 and T4 give a negative feedback impulse to the pituitary and hypothalamus.[165] Low thyroid hormone levels (hypothyroidism) are associated with gain in body weight, constipation, bradycardia, decreased gluconeogenesis, increased concentrations of atherogenic lipoproteins, hyperlipidemia and decreased bone turnover.[162-164,169] High levels (hyperthyroidism) on the other hand are associated with weight loss, reduced cholesterol levels, increased lipolysis, gluconeogenesis, diarrhea, muscle weakness and accelerated bone loss or fracture risk.[163,164,169]

The thyroid axis is clearly influenced by other pituitary axes. Start of growth hormone replacement therapy may unmask a disguised central hypothyroidism, as growth hormone gives an increase in serum free T3 and a decrease in T4, suggesting the rising IGF-1 levels stimulate the conversion from T4 to T3.[167] Dopamine (agonists) and glucocorticoids inhibit TSH secretion.[167] Gonadal hormones have a minor influence on the axis; estradiol treatment increases the number of TRH receptors, and testosterone analog fluoxymesterone shows a decreased response of the pituitary on THR in hypogonadal men.[167,170-172]

Endocrine alterations due to tumors – too high, too low

The pituitary axis may be altered by many factors, and tumors and related treatment is one of them. An increased hormone production can be directly caused by tumors in endocrine organs or ectopic tumors.[173]

Decreased pituitary hormones (hypopituitarism) are most often caused by tumors in the hypothalamo-pituitary region (in ~65% of the cases).[174] Radiotherapy for treatment of cancer is also an important risk factor for endocrine conditions.[42] When endocrine organs such as the hypothalamus, pituitary, thyroid and reproductive organs lie in the irradiation area, this may lead to a lowered hormone production.[46,175] Approximately 40% of patients irradiated for brain tumors which were not near the hypothalamus or pituitary, develop hypopituitarism as well.[174,176,177] Surgical removal of endocrine tissue can cause endocrine deficiencies too,[178] particularly after pituitary surgery.[179] Endocrine conditions are an important cause for need of hospitalization in childhood cancer survivors.[180]

Pituitary tumors

Tumors do not necessarily have to be malignant to cause major health problems in the short as well as the long term. A benign tumor at an unfortunate location can have serious adverse outcomes. Tumors in the pituitary region may lead to increased or decreased secretion of hormones, or oppress other structures in the brain and impair their function. The most common pituitary (and even intracranial) tumors are pituitary adenomas, which have a prevalence of 1 in 1000 persons.[181] The pituitary adenoma is indeed a benign tumor derived from proliferation of anterior pituitary cells, which can cause a hypersecretion of the hormones produced by the specific differentiated cell types (functioning pituitary adenoma) or have a mass effect.[182,183] Of the hormone-secreting adenomas, the lactotroph adenoma (prolactinoma) is most

common (25-41%), with thereafter the somatotroph adenoma (10-15%) and corticotroph adenoma (~10%), while thyrotroph adenomas and gonadotroph adenomas are more rare (<1%).[184] The treatment is dependent on the pattern of hormone secretion. Prolactinomas are primarily treated pharmaceutically with dopamine agonists.[185] Other pituitary adenomas are intervened with by surgical removal, and if necessary additional pharmaceutical treatment with somatostatin analogues, dopamine agonists, growth hormone receptor antagonist pegvisomant in somatotroph adenomas (acromegaly), and sometimes radiotherapy.[184] Other entities which are enlisted in the differential diagnosis of neoplasms occurring in the pituitary region, are hemangioblastomas, eosinophilic granulomas, dermoid/epidermoid or arachnoid cysts, optic nerve gliomas, lipomas, meningiomas, teratomas, germinomas, chordomas or chondrosarcomas, hamartomas, Rathke's cleft cysts and craniopharyngiomas.[186] The Rathke's cleft cyst, which is a diverticulum in Rathke's pouch,[52,187,188] is characterized by lower rates of visual impairment, hypothalamic damage and endocrine deficiencies compared to craniopharyngioma.[30] The craniopharyngioma is more rare than the pituitary adenoma, and has an incidence of 0.5-2.5 new cases per million persons per year.[30,189,190] This tumor is more often associated with serious adverse outcomes.[30] Craniopharyngiomas induce the highest mortality among all pituitary tumors.[191]

Craniopharyngioma – epidemiology, presentation and diagnosis

Craniopharyngiomas are brain tumors with a low histological malignancy (WHO grade I), which still show aggressive clinical behavior.[30,192] Although they are considered benign, they are closely situated to important neurovascular structures, such as the hypothalamus, pituitary, carotid arteries and optic nerves.[30,193] The function of these structures can be impaired, as the tumor does not respect its boundaries, and treatment may cause additional damage.[30] Craniopharyngioma can occur at any age, but the distribution is bimodal, with peak incidences between the ages 5-15 and 45-60 years (Figure 7).[30,190,194] Between 30-50% of the cases are diagnosed

during childhood or adolescence.[30,194,195] Craniopharyngiomas account for 5-11% of intracranial tumors in children and are therefore the most common non-neuroepithelial intracerebral neoplasm in this age category.[30,196-198] Children most frequently present with endocrine deficiencies leading to growth retardation and polyuria due to a declined release of ADH (central diabetes insipidus); another important feature is visual impairment.[30] In adults, central diabetes insipidus and visual impairment can manifest as well; other common signs and symptoms are headaches and other signs of high intracranial pressure, hypothalamic dysfunction (i.e. dysregulation in body temperature and body weight) and endocrine deficiencies.[30,199-201] The tumor can consist of a solid part with calcifications and/or a cystic part.[202] The location of a craniopharyngioma is most often described as either intrasellar or suprasellar.[30] For this designation, the sella turcica is used as a reference. The sella turcica is a depression in the sphenoid bone which contains the pituitary gland.[30] Most craniopharyngiomas (75%) originate suprasellarly and extend to the intrasellar region, while 20% remains fully suprasellar, and only a small number (5%) remains entirely intrasellar.[203] Approximately half of the tumors originate from the third ventricle floor[30] with high risk of damaging the hypothalamus.[204] The preferred imaging modalities to support the diagnosis and prepare for neurosurgery are magnetic resonance imaging with and without gadolinium contrast enhancement, and computed tomography for evaluation of calcifications.[205]

Adamantinomatous and papillary craniopharyngioma

There are two distinct histopathological or morphologic subtypes: the adamantinomatous (ACP) and papillary craniopharyngioma (PCP) (Table 2 and Figure 8).[206] The papillary subtype represents only a small proportion of all craniopharyngiomas (~10%).[30,207] ACP and PCP differ in their pathogenesis, genetic make-up and age distribution.[30] Both craniopharyngioma types develop from remnants of the embryological development of the hypothalamus and pituitary and can therefore occur along the hypothalamic-pituitary axis.[30] ACPs

are derived from remnants of Rathke's pouch, the craniopharyngeal duct epithelium.[30,193,208,209] PCPs develop from metaplasia of squamous epithelial cells which are remnants of the part of the stomodeum that contributed to the buccal mucosa.[209] The bimodal age distribution for craniopharyngioma[30,190] relates to these subtypes. The adamantinomatous subtype can occur in children and adults, while the papillary subtype almost exclusively occurs in adults.[30,210] Regarding genetic makeup: ACPs carry somatic *CTNNB1* clonal mutations in in 75-96% of the cases, a gene in exon 3 which regulates beta-catenin stability.[211-216] Nuclear beta-catenin accumulation is an important characteristic in adamantinomatous craniopharyngioma, and is absent in PCP and Rathke's cleft cysts.[217] PCPs are very frequently (in ~95% associated with somatic *BRAF*^{V600E} clonal mutations.[214,216] However, *BRAF* and *CTNNB1* mutations can occur simultaneously, too.[218]

Treatment of craniopharyngioma

Craniopharyngiomas are mainly treated with neurosurgery, frequently followed by radiotherapy in case of residual tumor tissue.[219,220] The goal of craniopharyngioma treatment is to achieve tumor control with as little neurovascular damage as possible, in order not to aggravate symptoms and keep tumor- and treatment related comorbidities to a minimum (described in a summary of Figure 9).[221] The treatment of choice is dependent on tumor- and patient characteristics, and on the experience and expertise of the treating center.[220]

Surgical treatment may have urgent indications such as raised intracranial pressure or vision loss.[30] Neurosurgery can be performed in the form of gross total resection or as limited surgery, such as partial resection.[30] The surgical approach depends on precise tumor location and size; predominantly intrasellar tumors can often be operated transsphenoidally.[30] Transcranial resection is performed by either pterional, subfrontal, transcallosal or transcortical-transventricular route (Figure 10).[220] Higher morbidity rates are observed in those who are initially treated with cyst aspiration or who need pterional surgery, and furthermore when patients become ill at a young age, or have

symptoms of intracranial hypertension, compared with patients without these characteristics.[193,199]

Table 2 Comparison of adamantinomatous and papillary craniopharyngioma

	Adamantinomatous subtype	**Papillary subtype**
Origin	Embryogenetic theory: Embryonic ectodermal epithelial remnants of the craniopharyngeal duct or Rathke's pouch. The duct and pouch are derived from the stomodeum	Metaplastic theory: Metaplasia of adenohypophyseal cells in the pars tuberalis of the adenohypophysis (remnants of the part of the stomadeum that contributed to the buccal mucosa), which results in formation of squamous cell nests
Age group	Presentation in children and adults: bimodal incidence peaks at 5-15 and 45-60 years	Presentation in adults: incidence peak between 40 and 55 years
Genetic aberrations	Somatic *CTNNB1* mutations Mostly point mutations within exon 3	Somatic *BRAF*^{V600E}mutations
Mutation rate	Low: ~15 non-synonymous mutations per megabase	Low: ~15 non-synonymous mutations per megabase
Pathway	Increase beta-catenin stability by overactivation WNT pathway	Mitogen-activated protein kinase (MAPK) signaling pathway activation in the basal cells surrounding the fibrovascular cores, which are structures containing blood vessels and stroma surrounded by well-defined lining epithelium that supports tumor growth. They also express high main effectors of MAPK (phosphorylated extracellular signal-regulated kinase 1 and 2) and SOX2

Important symptoms	Visual impairment Hypopituitarism Headache	Headache Hypothalamic dysfunction Psychiatric changes
MRI descriptions	Cauliflower-like shape ~90% calcifications ~90% cysts with cholesterol-rich oily fluid ~90% enhancement	"Fibrovascular cores lined by non-keratizing squamour epithelium"[30]
Pathological features	"Distinctive epithelium that forms stellate regiculum, wet keratin and basal palisades"[30]	"Fibrovascular cores lined by non-keratinizing squamous epithelium"[30]
Immunohistochemical features	Sporadic cells with nucleocytoplasmatic accumulation of beta-catenin in single cells throughout the tumor or in cell clusters; most tumor cells have normal membranous beta-catenin expression. Beta-catenin accumulating cells often located at the base of finger-like protrusions of tumor epithelium that invade surrounding tissues	No beta-catenin accumulating cells
Inflammatory mediators	High concentrations of cytokines IL-1β, IL-6, IL-8, CXCL1, IL-10, IL-18, TNF in cystic fluid, and high expression of receptors and coreceptors on the tumor: compatible with inflammasome activation. High expression of immunosuppressive factors IL-10, indoleamine-pyrrole 2,3-dioxygenase, galectin-1, programmed death ligand 1 and its receptor: promote escape of immune surveillance	Not investigated except for expression of PD-L1 in basal cells
Potential novel targeted therapies	Being developed; relate to IL-6, PD1/PD-L1, MEK, IDO-1, but not yet applied	*BRAF* inhibitors (dabrafenib, bemurafenib) combined with MEK inhibiters (i.e., trametinib), applied scarcely

Adapted from Müller et al. 2019 and Müller et al. 2014.[30,101,197,209,294,295]

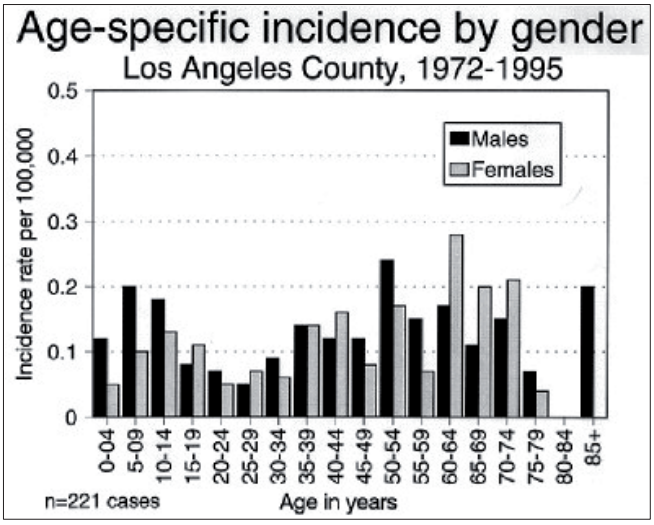
The choice for either gross total resection or subtotal resection remains a topic of discussion.[220] The severity of adherence to the hypothalamus and third ventricle (walls) must be evaluated preoperatively, to anticipate the surgical risk of hypothalamic injury and plan the degree of removal.[30,222,223] Craniopharyngiomas with hypothalamic adhesion generally have the worst outcomes.[30,222] The degree of tumor removal with the intent to achieve tumor control, is generally divided as either gross total resection or partial resection with often postoperative radiotherapy. The choice for limited surgery is made to alleviate or prevent symptoms and optimize dose delivery of postoperative radiotherapy.[30] Radiotherapy can be administered as stereotactic radiosurgery or intracystic application of ^{90}Yttrium, which emits beta energy.[219] Intracystic application of chemotherapeutics such as bleomycin or interferon-alpha are alternative treatment options.[219] If it is decided to apply postoperative radiotherapy, a total dose of 50-54 Gy in fractions of 1.8-2.0 Gy are normally applied.[224] Conventional radiotherapy sends X-ray in different directions towards the tumor to scatter unintended but unavoidable irradiation of non-target tissue, while proton beam therapy deposits a dose along a path that ends in the target.[30] Proton beam therapy therefore reduces the dose in non-target tissue significantly.[30]

Craniopharyngioma: survival, long-term sequelae and comorbidities

As the craniopharyngioma is considered benign, the oncological aggressiveness of the tumor is not the main factor in the prognosis of patients.[30] Moreover, the prognosis is dependent on the location of the tumor and involvement of neurovascular structures, and to treatment-related damage.[30] Overall survival rates are estimated at 54-96% after five years, and 66-85% after 20 years.[30-32] Long-term tumor control rates[30,225] and survival rates are similar between gross total resection and subtotal resection with radiotherapy (reported 5-year survival rates 82-100% versus 85-100% and 5-year recurrence-free survival rates 60-100 versus 72-82%, respectively).[32,226,227] If radiotherapy is not applied after subtotal resection, 5-year survival rates and especially

recurrence-free survival rates are generally lower (5-year overall survival 75-90%; recurrence-free survival 25-47%).[32,226,227] There are no known histopathological characteristics which predict survival in patients with childhood-onset disease.[228] In adulthood-onset patients, calcifications are related to higher recurrence rates and unfavorable outcome.[30,229]

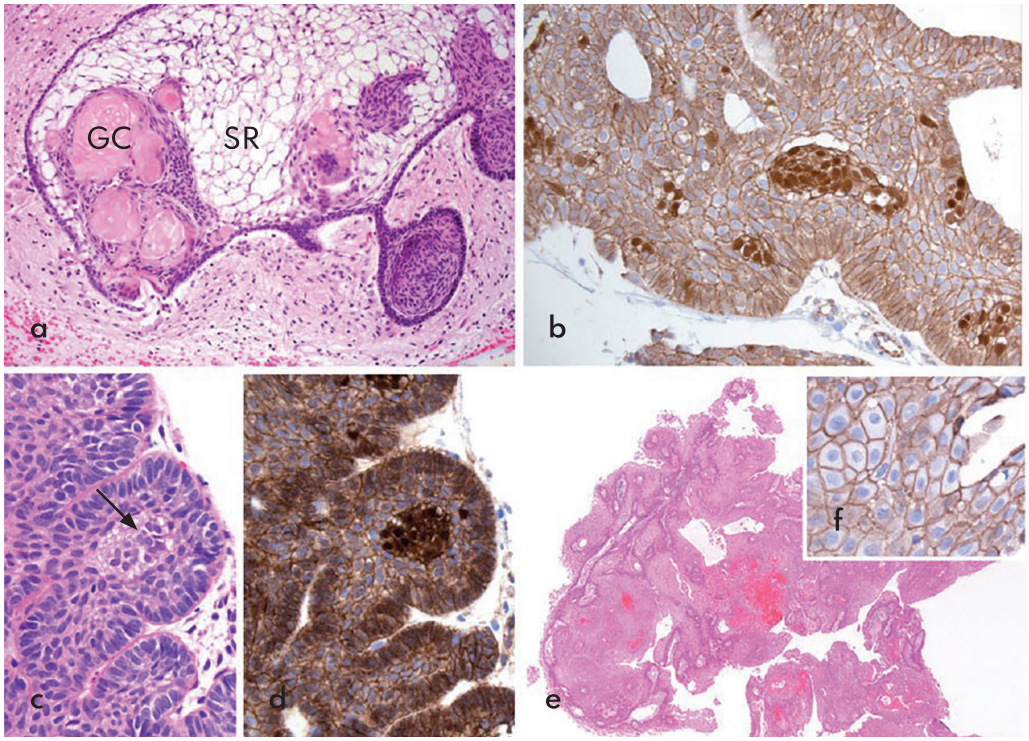
Figure 7 Epidemiology of craniopharyngioma



Graph showing the age-specific incidence of craniopharyngioma derived from the University of Southern California/Los Angeles county CSP data for the years 1972–1995. From: Bunin 1997 (with permission).[194]

Long-term health issues can be derived from either persistently presenting symptoms or occurrence after tumor treatment, and can impair quality of life.[30,201,230-235] The most frequent long-term health issues consist of pituitary hormone deficiencies, visual impairment and hypothalamic damage related obesity (Figure 11).[30]

Figure 8 Histopathology of craniopharyngioma



Adamantinomatous craniopharyngioma (a–d). a Classical appearance with peripheral palisading epithelium (arrow), loose 'stellate reticulum' (SR) and nodules of 'ghost cells' (GC). The tumor is invading brain. b Clusters and individual tumor cells show nuclear b-catenin translocation. Note that in most tumor cells b-catenin remains at the cell membrane. c, d Epithelial nodules (arrows) within the advancing front of the tumor commonly show nuclear b-catenin expression. A papillary craniopharyngioma is characterized by clefted (pseudo-) papillae lacking 'stellate reticulin' and 'ghost cells'. b-catenin remains at the cell membrane (f). See text for details. a, c, e haematoxylin and eosin, b, d, f b-catenin immunohistochemistry (brown reaction product). (a 9200, b, c, d 9400, e 9100, f 9600). From: Larkin et al. 2013 (with permission).[296]

Obesity in patients with craniopharyngioma

Hypothalamic obesity occurs in ~50% of the patients diagnosed with craniopharyngioma (Figure 11).[30,86,199,236,237] Visual impairment and hypothalamic damage are risk factors for the metabolic syndrome (components) in these patients.[86] The metabolic syndrome which occurred in 40% of Dutch craniopharyngioma patients versus 26% in controls.[86] This partially explains their increased risk to develop type 2 diabetes mellitus (standardized incidence ratio [SIR] 4.4) and cerebral infarction (SIR 4.9).[200]

Hypothalamic damage comes with a parasympathetic predominance, causing reduced heart rate and body temperature, and increased sleepiness during the day, resulting in a reduced energy expenditure.[238,239] This contributes the high risk of obesity and is associated with hyperinsulinemia, hypopituitarism, psychosocial disorders and hyperphagia as well.[231] There is currently very little evidence for the efficacy of pharmaceutics or lifestyle changes for the treatment of hypothalamic obesity.[231] In rare cases, hypothalamic damage can express itself at the opposite end of the spectrum, with the diencephalic syndrome (in ~4% of the patients from the German Childhood Craniopharyngioma Registry), which results in cachexia and severe weight loss.[30,240,241]

Body weight is mainly composed of fat mass, muscle mass and bone mass. The pituitary hormones influence body composition, particularly growth hormone.[60,242] As patients with craniopharyngioma have a high risk of pituitary deficiencies (~92%),[86] their body composition may be altered as well. Obesity is often evaluated using anthropomorphic measurements such as body mass index (BMI), which only accounts for total mass, and does not particularly check adiposity. Alternatively, body composition can be evaluated with the dual-energy X-ray absorptiometry (DXA)-scan; this is considered the gold standard for evaluating adiposity if the body composition is potentially altered.[243-245] In childhood cancer survivors, adiposity is underestimated using BMI if compared with DXA-scan derived body fat percentage.[243] In patients with craniopharyngioma, it has not yet been investigated whether obesity rates are correctly evaluated with BMI.

Figure 9 Summary of long-term sequelae of craniopharyngioma

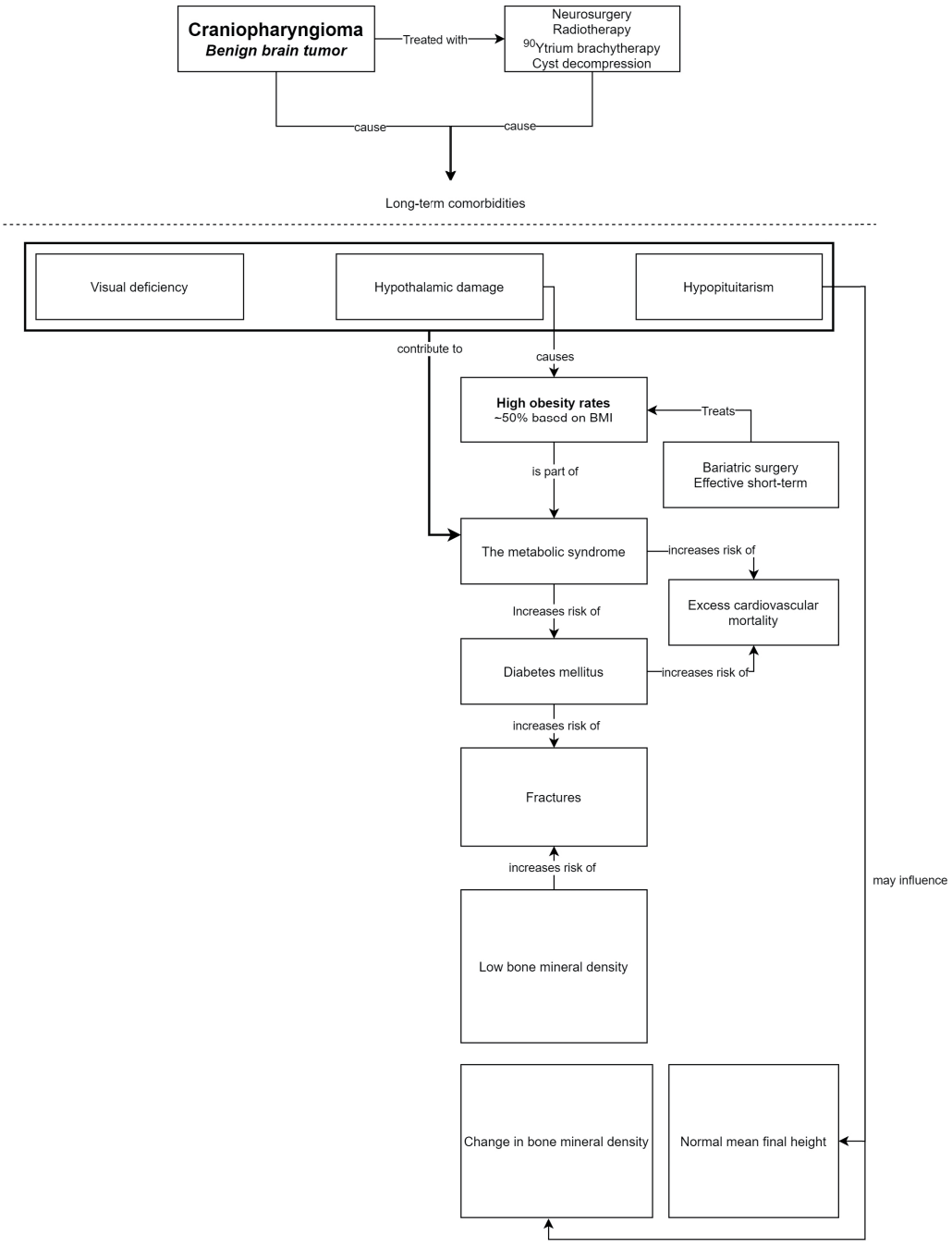
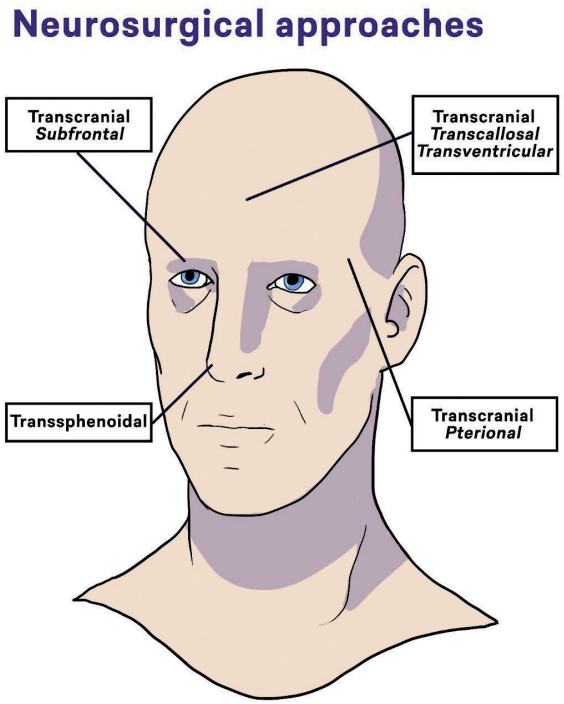


Figure 10 Neurosurgical approaches for treatment of craniopharyngioma



By: Ana G.G. de Lima.

In the general population, severe obesity can be treated with bariatric surgery.[231,246-249] Bariatric surgery appears an effective weight loss strategy in patients with craniopharyngioma as well, even regardless presence of hypothalamic damage, for up to two years past this intervention; longer follow-up data are not yet available.[250]

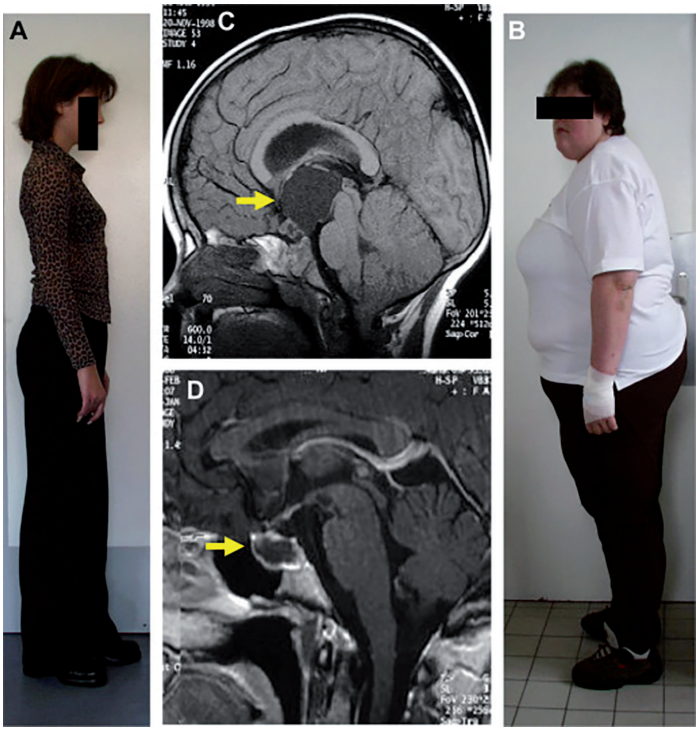
Bone health in patients with craniopharyngioma

The DXA-scan is also the gold standard for evaluation of bone mineral density, to evaluate the presence of osteoporosis.[251] In the general population, bone mass and density reach their peak at a young adult age (20-30 years old) and remain relatively constant until the age of 50, and then decline.[252-254] The bone density values measured by DXA-scan are compared to values of a reference population, expressed

as a T-score or Z-score.[252,253] When bone mass is very low (T-score ≤ -2.5), this is a sign of osteoporosis, which occurs in 0.6% of the general population.[252] Osteoporosis is defined as "a systemic skeletal disease characterized by a reduction in bone mass and qualitative skeletal changes (macro- and microarchitecture, material properties, geometry, and micro-damage) that cause an increase in bone fragility and higher fracture risk."[251] Fractures cause pain and at least temporary, or perhaps even permanent loss of function, may require hospitalization and give reduced DALYs.[255,256] Fragility fractures are mostly located at the vertebral column, proximal side of the humerus and femur, and distal side of the radius.[251] Application of the DXA-scan is already advised in childhood cancer survivors, who do often do not reach a normal peak bone mass, causing low bone mineral density in ~45-51% and increased risk of long-bone fractures.[255,257,258] Risk factors for pathological bone density and fractures in survivors were exposure to glucocorticoids and methotrexate, growth hormone and gonadal hormone deficiency, male sex, smoking status and cranial or abdominal irradiation, while height, weight and attained age were protective factors.[255,258,259] These factors are partially applicable to patients with craniopharyngioma. They could have a high fracture risk as well, due to a combined risk of low bone density and high risk of falling.[260-262] Low bone density is associated to hypopituitarism: growth hormone deficiency gives reduced bone remodeling activity, and both patients with isolated growth hormone deficiency and multiple pituitary deficiencies are at high risk of reduced bone mineral density.[30,260,261,263-266] The growth hormone deficiency can also add to the potential risk of falling, as it causes decreased muscle mass and muscle function,[263] leading to a higher fracture risk.[267] Other potential falling risk factors that occur frequently in patients with craniopharyngioma, are diabetes,[86,268] epilepsy,[32,199,269,270] visual deficiencies[271-273] and neurological conditions.[199,274,275] Some studies have indeed demonstrated a high risk of pathological bone density[276-278] of 53%.[277] Only a few studies investigated fracture rates in patients with craniopharyngioma; some studies[196,279] did indeed find a high fracture risk, but not all.[200] The above described risk factors for fractures and bone density (changes) have barely been investigated in patients with craniopharyngioma. In small cohorts of patients

with craniopharyngioma, risk factors for low bone density were as hypothalamic involvement, low levels of sex steroids or insufficient gonadal hormone replacement.[261,278] It is unknown what factors are important for the change in bone mineral density or fracture risk, what type of fractures occur in patients with a craniopharyngioma, and there are only limited studies performed that regard final height.[235,280-283]

Figure 11 Hypothalamic obesity



Degree of obesity with regard to location of childhood craniopharyngioma. In both patients, craniopharyngioma (as indicated by the arrows on MRI before surgery) could be completely resected. Both patients had complete hypopituitarism after surgery, requiring endocrine substitution of all hypothalamic-pituitary axes. The patient in (B) developed severe obesity because of hypothalamic lesions of suprasellar parts of craniopharyngioma (C). The patient in (A) presented with a small tumor confined to the sellar region (D). After complete resection she kept normal weight without any eating disorders (A). (From Müller HL, Kaatsch P, Warmuth-Metz M, et al. Kraniopharyngeom im Kindesund Jugendalter – Diagnostische und therapeutische Strategien. Monatsschr Kinderheilkd 2003; 151:1056-63;[297] with permission.)

Aims and outline of this thesis

The aim of this thesis is to examine long-term endocrine and metabolic alterations and related risk factors in who suffer(ed) from a tumor, with a special focus on patients with tumors near the pituitary (craniopharyngioma and macro-adenoma). In the current chapter (**Chapter 1**), an introduction is given of important characteristics of childhood cancer, craniopharyngioma and metabolic complications. **Chapter 2** describes a unique case illustrating the potential severity of pituitary disease and treatment issues of a very young boy with acrogigantism. **Chapter 3** reveals results on low bone density and fractures in patients with a craniopharyngioma, risk factors for fractures and achieved final height. In **Chapter 4**, we report the assessment of changes in body composition and bone mineral density over time, including factors influencing changes bone mineral density in patients with a craniopharyngioma. In **Chapter 5**, we show results of a study that aimed to evaluate the added value of applying a dual-energy X-ray absorptiometry scan for the diagnosis of obesity and the metabolic syndrome, and information of related risk factors of the metabolic syndrome in patients with a craniopharyngioma. **Chapter 6** reports as a systematic review on the value of diagnostic and prognostic value of novel biomarkers for metabolic syndrome in childhood cancer survivors. **Chapter 7** shows the comparison of efficacy of bariatric surgery for the treatment of obesity in patients with craniopharyngioma versus controls. **Chapter 8** provides a general discussion of this thesis.

References

1. Howlader N, Noone AM, Krapcho M, et al. SEER Cancer Statistics Review, 1975-2017. National Cancer Institute 2020.
2. OECD, Systems EOoH, Policies. Netherlands: Country Health Profile 2017; 2017.
3. Fitzmaurice C, Akinyemiju TF, Al Lami FH, et al. Global, Regional, and National Cancer Incidence, Mortality, Years of Life Lost, Years Lived With Disability, and Disability-Adjusted Life-Years for 29 Cancer Groups, 1990 to 2016: A Systematic Analysis for the Global Burden of Disease Study. JAMA Oncol 2018; 4(11): 1553-68.
4. Bhakta N, Force LM, Allemani C, et al. Childhood cancer burden: a review of global estimates. Lancet Oncol 2019; 20(1): e42-e53.
5. CBS. Deaths; underlying cause of death (shortlist), sex, age. 18 December 2020 2020. https://opendata.cbs.nl/statline/#/CBS/en/dataset/7052eng/table?ts=1612257968931 (accessed 02-02-2021 2021).
6. SKION. SKION registratie. 28-01-2021 2021. https://www.skion.nl/ (accessed 02-02-2021 2021).
7. Erdmann F, Frederiksen LE, Bonaventure A, et al. Childhood cancer: Survival, treatment modalities, late effects and improvements over time. Cancer Epidemiol 2020: 101733.
8. Ferlay J EM, Lam F, Colombet M, Mery L, Piñeros M, Znaor A, Soerjomataram I, Bray F. Global Cancer Observatory: Cancer Today. 2020. https://gco.iarc.fr/today (accessed 09-04-2021 2021).
9. Hudson MM, Link MP, Simone JV. Milestones in the curability of pediatric cancers. J Clin Oncol 2014; 32(23): 2391-7.
10. Allemani C, Weir HK, Carreira H, et al. Global surveillance of cancer survival 1995-2009: analysis of individual data for 25,676,887 patients from 279 population-based registries in 67 countries (CONCORD-2). Lancet 2015; 385(9972): 977-1010.
11. Winther JF, Kenborg L, Byrne J, et al. Childhood cancer survivor cohorts in Europe. Acta Oncol 2015; 54(5): 655-68.
12. Institute of M, National Research Council National Cancer Policy B. In: Hewitt M, Weiner SL, Simone JV, eds. Childhood Cancer Survivorship: Improving Care and Quality of Life. Washington (DC): National Academies Press (US Copyright 2003 by the National Academy of Sciences. All rights reserved.; 2003.
13. Youlden DR, Walwyn TS, Cohn RJ, Harden HE, Pole JD, Aitken JF. Late mortality from other diseases following childhood cancer in Australia and the impact of intensity of treatment. Pediatr Blood Cancer 2020: e28835.

14. Armstrong GT, Stovall M, Robison LL. Long-term effects of radiation exposure among adult survivors of childhood cancer: results from the childhood cancer survivor study. Radiat Res 2010; 174(6): 840-50.

15. Fidler-Benaoudia MM, Oeffinger KC, Yasui Y, et al. A comparison of late mortality among survivors of childhood cancer in the United States and United Kingdom. J Natl Cancer Inst 2020.

16. Bagnasco F, Caruso S, Andreano A, et al. Late mortality and causes of death among 5-year survivors of childhood cancer diagnosed in the period 1960-1999 and registered in the Italian Off-Therapy Registry. Eur J Cancer 2019; 110: 86-97.

17. Armstrong GT, Liu Q, Yasui Y, et al. Late mortality among 5-year survivors of childhood cancer: a summary from the Childhood Cancer Survivor Study. J Clin Oncol 2009; 27(14): 2328-38.

18. Cardous-Ubbink MC, Heinen RC, Langeveld NE, et al. Long-term cause-specific mortality among five-year survivors of childhood cancer. Pediatr Blood Cancer 2004; 42(7): 563-73.

19. Mertens AC, Liu Q, Neglia JP, et al. Cause-specific late mortality among 5-year survivors of childhood cancer: the Childhood Cancer Survivor Study. J Natl Cancer Inst 2008; 100(19): 1368-79.

20. Marina NM, Liu Q, Donaldson SS, et al. Longitudinal follow-up of adult survivors of Ewing sarcoma: A report from the Childhood Cancer Survivor Study. Cancer 2017; 123(13): 2551-60.

21. Mueller BA, Doody DR, Weiss NS, Chow EJ. Hospitalization and mortality among pediatric cancer survivors: a population-based study. Cancer Causes Control 2018; 29(11): 1047-57.

22. Schindler M, Spycher BD, Ammann RA, Ansari M, Michel G, Kuehni CE. Cause-specific long-term mortality in survivors of childhood cancer in Switzerland: A population-based study. Int J Cancer 2016; 139(2): 322-33.

23. Gibson TM, Robison LL. Impact of Cancer Therapy-Related Exposures on Late Mortality in Childhood Cancer Survivors. Chem Res Toxicol 2015; 28(1): 31-7.

24. Kero AE, Järvelä LS, Arola M, et al. Late mortality among 5-year survivors of early onset cancer: a population-based register study. Int J Cancer 2015; 136(7): 1655-64.

25. Prasad PK, Signorello LB, Friedman DL, Boice JD, Jr., Pukkala E. Long-term non-cancer mortality in pediatric and young adult cancer survivors in Finland. Pediatr Blood Cancer 2012; 58(3): 421-7.

26. Wilson CL, Cohn RJ, Johnston KA, Ashton LJ. Late mortality and second cancers in an Australian cohort of childhood cancer survivors. Med J Aust 2010; 193(5): 258-61.

27. Tukenova M, Diallo I, Hawkins M, et al. Long-term mortality from second malignant neoplasms in 5-year survivors of solid childhood tumors: temporal pattern of risk according to type of treatment. Cancer Epidemiol Biomarkers Prev 2010; 19(3): 707-15.

28. Effinger KE, Stratton KL, Fisher PG, et al. Long-term health and social function in adult survivors of paediatric astrocytoma: A report from the Childhood Cancer Survivor Study. Eur J Cancer 2019; 106: 171-80.

29. Salloum R, Chen Y, Yasui Y, et al. Late Morbidity and Mortality Among Medulloblastoma Survivors Diagnosed Across Three Decades: A Report From the Childhood Cancer Survivor Study. J Clin Oncol 2019; 37(9): 731-40.

30. Muller HL, Merchant TE, Warmuth-Metz M, Martinez-Barbera JP, Puget S. Craniopharyngioma. Nat Rev Dis Primers 2019; 5(1): 75.

31. Pereira AM, Schmid EM, Schutte PJ, et al. High prevalence of long-term cardiovascular, neurological and psychosocial morbidity after treatment for craniopharyngioma. Clin Endocrinol (Oxf) 2005; 62(2): 197-204.

32. Karavitaki N, Brufani C, Warner JT, et al. Craniopharyngiomas in children and adults: systematic analysis of 121 cases with long-term follow-up. Clin Endocrinol (Oxf) 2005; 62(4): 397-409.

33. Nilsson B, Gustavasson-Kadaka E, Bengtsson BA, Jonsson B. Pituitary adenomas in Sweden between 1958 and 1991: incidence, survival, and mortality. J Clin Endocrinol Metab 2000; 85(4): 1420-5.

34. Hajat C, Stein E. The global burden of multiple chronic conditions: A narrative review. Preventive medicine reports 2018; 12: 284-93.

35. GBD C. Global health observatory data repository. Accessed at: https://vizhub.healthdata.org/gbd-compare/. http://apps.who.int/gho/data/node.home (accessed 12-04-2021 2021).

36. Martin CM. Chronic disease and illness care: adding principles of family medicine to address ongoing health system redesign. Canadian family physician Medecin de famille canadien 2007; 53(12): 2086-91.

37. Healey MA, Shackford SR, Osler TM, Rogers FB, Burns E. Complications in surgical patients. Arch Surg 2002; 137(5): 611-7; discussion 7-8.

38. Meara JG, Leather AJ, Hagander L, et al. Global Surgery 2030: evidence and solutions for achieving health, welfare, and economic development. Lancet 2015; 386(9993): 569-624.

39. Nepogodiev D, Martin J, Biccard B, Makupe A, Bhangu A. Global burden of postoperative death. Lancet 2019; 393(10170): 401.

40. Khuri SF, Henderson WG, DePalma RG, Mosca C, Healey NA, Kumbhani DJ. Determinants of long-term survival after major surgery and the adverse effect of postoperative complications. Ann Surg 2005; 242(3): 326-41; discussion 41-3.

41. Bhakta N, Liu Q, Ness KK, et al. The cumulative burden of surviving childhood cancer: an initial report from the St Jude Lifetime Cohort Study (SJLIFE). Lancet 2017; 390(10112): 2569-82.

42. Hudson MM, Ness KK, Gurney JG, et al. Clinical ascertainment of health outcomes among adults treated for childhood cancer. JAMA 2013; 309(22): 2371-81.

43. Marina N. Long-term survivors of childhood cancer. The medical consequences of cure. Pediatr Clin North Am 1997; 44(4): 1021-42.

44. Houghton PJ, Kurmasheva RT. Challenges and Opportunities for Childhood Cancer Drug Development. Pharmacol Rev 2019; 71(4): 671-97.

45. Phelan R, Eissa H, Becktell K, et al. Upfront Therapies and Downstream Effects: Navigating Late Effects in Childhood Cancer Survivors in the Current Era. Curr Oncol Rep 2019; 21(12): 104.

46. Hudson MM, Mulrooney DA, Bowers DC, et al. High-risk populations identified in Childhood Cancer Survivor Study investigations: implications for risk-based surveillance. J Clin Oncol 2009; 27(14): 2405-14.

47. Golden SH, Brown A, Cauley JA, et al. Health disparities in endocrine disorders: biological, clinical, and nonclinical factors--an Endocrine Society scientific statement. J Clin Endocrinol Metab 2012; 97(9): E1579-639.

48. Coopmans EC, Chunharojrith P, Neggers S, et al. Endocrine Disorders Are Prominent Clinical Features in Patients With Primary Antibody Deficiencies. Front Immunol 2019; 10: 2079.

49. Newman CB, Blaha MJ, Boord JB, et al. Lipid Management in Patients with Endocrine Disorders: An Endocrine Society Clinical Practice Guideline. The Journal of Clinical Endocrinology & Metabolism 2020; 105(12): 3613-82.

50. Aghi M, Barker Ii FG. Benign adult brain tumors: an evidence-based medicine review. Prog Neurol Surg 2006; 19: 80-96.

51. Hussain I, Eloy JA, Carmel PW, Liu JK. Molecular oncogenesis of craniopharyngioma: current and future strategies for the development of targeted therapies. J Neurosurg 2013; 119(1): 106-12.

52. Han SJ, Jahangiri A, Aghi MK. Rathke's Cleft Cysts. Pituitary Disorders; 2013: 146-52.

53. Takahashi TA, Johnson KM. Menopause. Med Clin North Am 2015; 99(3): 521-34.

54. Morales JS, Valenzuela PL, Rincón-Castanedo C, Santos-Lozano A, Fiuza-Luces C, Lucia A. Is health status impaired in childhood cancer survivors? A systematic review and meta-analysis. Crit Rev Oncol Hematol 2019; 142: 94-118.

55. Razzouk BI, Rose SR, Hongeng S, et al. Obesity in survivors of childhood acute lymphoblastic leukemia and lymphoma. J Clin Oncol 2007; 25(10): 1183-9.

56. Lipshultz SE, Lipsitz SR, Sallan SE, et al. Chronic progressive cardiac dysfunction years after doxorubicin therapy for childhood acute lymphoblastic leukemia. J Clin Oncol 2005; 23(12): 2629-36.

57. Blanco JG, Sun CL, Landier W, et al. Anthracycline-related cardiomyopathy after childhood cancer: role of polymorphisms in carbonyl reductase genes--a report from the Children's Oncology Group. J Clin Oncol 2012; 30(13): 1415-21.

58. Hudson MM, Rai SN, Nunez C, et al. Noninvasive evaluation of late anthracycline cardiac toxicity in childhood cancer survivors. J Clin Oncol 2007; 25(24): 3635-43.

59. Oeffinger KC, Adams-Huet B, Victor RG, et al. Insulin resistance and risk factors for cardiovascular disease in young adult survivors of childhood acute lymphoblastic leukemia. J Clin Oncol 2009; 27(22): 3698-704.

60. Oeffinger KC, Mertens AC, Sklar CA, et al. Obesity in adult survivors of childhood acute lymphoblastic leukemia: a report from the Childhood Cancer Survivor Study. J Clin Oncol 2003; 21(7): 1359-65.

61. Visscher H, Ross CJ, Rassekh SR, et al. Pharmacogenomic prediction of anthracycline-induced cardiotoxicity in children. J Clin Oncol 2012; 30(13): 1422-8.

62. Devidi M, Buddam A, Dacha S, Rao DS. Atrial Fibrillation and Its Association with Endocrine Disorders. J Atr Fibrillation 2014; 6(5): 959.

63. Bowers DC, Liu Y, Leisenring W, et al. Late-occurring stroke among long-term survivors of childhood leukemia and brain tumors: a report from the Childhood Cancer Survivor Study. J Clin Oncol 2006; 24(33): 5277-82.

64. Bowers DC, McNeil DE, Liu Y, et al. Stroke as a late treatment effect of Hodgkin's Disease: a report from the Childhood Cancer Survivor Study. J Clin Oncol 2005; 23(27): 6508-15.

65. Sheu JJ, Kang JH, Lin HC, Lin HC. Hyperthyroidism and risk of ischemic stroke in young adults: a 5-year follow-up study. Stroke 2010; 41(5): 961-6.

66. Bahadoran Z, Mirmiran P, Azizi F, Ghasemi A. A Brief History of Modern Endocrinology and Definitions of a True Hormone. Endocr Metab Immune Disord Drug Targets 2019; 19(8): 1116-21.

67. Veldhuis JD. Changes in pituitary function with ageing and implications for patient care. Nat Rev Endocrinol 2013; 9(4): 205-15.

68. Dittfeld A, Gwizdek K, Brończyk-Puzoń A. History of endocrinology in the world and in Poland. Pediatr Endocrinol Diabetes Metab 2017; 23(3): 146-51.

69. Nussey S, Whitehead S. Endocrinology: An Integrated Approach. Oxford: BIOS Scientific Publishers Copyright © 2001, BIOS Scientific Publishers Limited.; 2001.

70. Cannarella R, Barbagallo F, Condorelli RA, Aversa A, La Vignera S, Calogero AE. Osteoporosis from an Endocrine Perspective: The Role of Hormonal Changes in the Elderly. Journal of clinical medicine 2019; 8(10): 1564.

71. Röder PV, Wu B, Liu Y, Han W. Pancreatic regulation of glucose homeostasis. Exp Mol Med 2016; 48(3): e219.

72. Göke B. Islet cell function: alpha and beta cells--partners towards normoglycaemia. Int J Clin Pract Suppl 2008; (159): 2-7.

73. Galsgaard KD, Pedersen J, Knop FK, Holst JJ, Wewer Albrechtsen NJ. Glucagon Receptor Signaling and Lipid Metabolism. Frontiers in physiology 2019; 10: 413.

74. Duncan RE, Ahmadian M, Jaworski K, Sarkadi-Nagy E, Sul HS. Regulation of Lipolysis in Adipocytes. Annual Review of Nutrition 2007; 27(1): 79-101.

75. Herold J, Kalucka J. Angiogenesis in Adipose Tissue: The Interplay Between Adipose and Endothelial Cells. Front Physiol 2020; 11: 624903.

76. Coelho M, Oliveira T, Fernandes R. Biochemistry of adipose tissue: an endocrine organ. Archives of medical science : AMS 2013; 9(2): 191-200.

77. Guerre-Millo M. Adipose tissue hormones. J Endocrinol Invest 2002; 25(10): 855-61.

78. (RIVM) RvVeM. Volksgezondheidenzorg.info / Statline. https://statline.rivm.nl/#/RIVM/nl/dataset/50050NED/table?dl=44D7F or https://statline.rivm.nl/#/RIVM/nl/dataset/50050NED/table?ts=1604915266015 / https://statline.rivm.nl/#/RIVM/nl/dataset/50050NED/table?ts=1607007571919 / https://opendata.cbs.nl/statline/#/CBS/nl/dataset/81565NED/table?ts=1613737408375 (accessed 09-11-2020).

79. Pluimakers VG, van Waas M, Neggers S, van den Heuvel-Eibrink MM. Metabolic syndrome as cardiovascular risk factor in childhood cancer survivors. Crit Rev Oncol Hematol 2019; 133: 129-41.

80. Walker SP, Rimm EB, Ascherio A, Kawachi I, Stampfer MJ, Willett WC. Body size and fat distribution as predictors of stroke among US men. Am J Epidemiol 1996; 144(12): 1143-50.

81. Wilding JPH. Endocrine testing in obesity. Eur J Endocrinol 2020; 182(4): C13-c5.

82. van Hulsteijn LT, Pasquali R, Casanueva F, et al. Prevalence of endocrine disorders in obese patients: systematic review and meta-analysis. Eur J Endocrinol 2020; 182(1): 11-21.

83. Kopin L, Lowenstein C. Dyslipidemia. Ann Intern Med 2017; 167(11): Itc81-itc96.

84. Wittcopp C, Conroy R. Metabolic Syndrome in Children and Adolescents. Pediatr Rev 2016; 37(5): 193-202.

85. Kaplan NM. The deadly quartet. Upper-body obesity, glucose intolerance, hypertriglyceridemia, and hypertension. Arch Intern Med 1989; 149(7): 1514-20.

86. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. The metabolic syndrome and its components in 178 patients treated for craniopharyngioma after 16 years of follow-up. Eur J Endocrinol 2017; 178(1): 11-22.

87. Alberti KG, Zimmet PZ. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. Diabet Med 1998; 15(7): 539-53.

88. Balkau B, Charles MA. Comment on the provisional report from the WHO consultation. European Group for the Study of Insulin Resistance (EGIR). Diabet Med 1999; 16(5): 442-3.

89. Executive Summary of The Third Report of The National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, And Treatment of High Blood Cholesterol In Adults (Adult Treatment Panel III). Jama 2001; 285(19): 2486-97.

90. Grundy SM, Cleeman JI, Daniels SR, et al. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute Scientific Statement. Circulation 2005; 112(17): 2735-52.

91. Alberti KG, Zimmet P, Shaw J. Metabolic syndrome--a new world-wide definition. A Consensus Statement from the International Diabetes Federation. Diabet Med 2006; 23(5): 469-80.

92. Alberti KG, Eckel RH, Grundy SM, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation 2009; 120(16): 1640-5.

93. Reaven GM. Banting lecture 1988. Role of insulin resistance in human disease. Diabetes 1988; 37(12): 1595-607.

94. Oda E. Metabolic syndrome: its history, mechanisms, and limitations. Acta Diabetol 2012; 49(2): 89-95.

95. Ju SY, Lee JY, Kim DH. Association of metabolic syndrome and its components with all-cause and cardiovascular mortality in the elderly: A meta-analysis of prospective cohort studies. Medicine (Baltimore) 2017; 96(45): e8491.

96. Alberti G; Zimmet P; Shaw J GS. The IDF consensus worldwide definition of the Metabolic Syndrome. 2006. https://www.idf.org/component/attachments/attachments.html?id=705&task=download (accessed 2019 April 08 2019).

97. Wilson PW, D'Agostino RB, Parise H, Sullivan L, Meigs JB. Metabolic syndrome as a precursor of cardiovascular disease and type 2 diabetes mellitus. Circulation 2005; 112(20): 3066-72.

98. Ford ES. The metabolic syndrome and mortality from cardiovascular disease and all-causes: findings from the National Health and Nutrition Examination Survey II Mortality Study. Atherosclerosis 2004; 173(2): 309-14.

99. Cornier MA, Dabelea D, Hernandez TL, et al. The metabolic syndrome. Endocr Rev 2008; 29(7): 777-822.

100. de Moraes DC, Vaisman M, Conceição FL, Ortiga-Carvalho TM. Pituitary development: a complex, temporal regulated process dependent on specific transcriptional factors. J Endocrinol 2012; 215(2): 239-45.

101. Amar AP, Weiss MH. Pituitary anatomy and physiology. Neurosurg Clin N Am 2003; 14(1): 11-23, v.

102. Treier M, Rosenfeld MG. The hypothalamic-pituitary axis: co-development of two organs. Curr Opin Cell Biol 1996; 8(6): 833-43.

103. Watanabe YG. Effects of brain and mesenchyme upon the cytogenesis of rat adenohypophysis in vitro. I. Differentiation of adrenocorticotropes. Cell Tissue Res 1982; 227(2): 257-66.

104. Cheung CC, Lustig RH. Pituitary development and physiology. Pituitary 2007; 10(4): 335-50.

105. Ito E, Shima R, Yoshioka T. A novel role of oxytocin: Oxytocin-induced well-being in humans. Biophysics and physicobiology 2019; 16: 132-9.

106. Hawker RW, Robertson PA. Oxytocin and lactation. J Clin Endocrinol Metab 1957; 17(3): 448-51.

107. Campbell A. Oxytocin and human social behavior. Pers Soc Psychol Rev 2010; 14(3): 281-95.

108. Grattan DR. 60 YEARS OF NEUROENDOCRINOLOGY: The hypothalamo-prolactin axis. The Journal of endocrinology 2015; 226(2): T101-T22.

109. Grattan DR, Kokay IC. Prolactin: a pleiotropic neuroendocrine hormone. J Neuroendocrinol 2008; 20(6): 752-63.

110. Ben-Jonathan N, Hugo ER, Brandebourg TD, LaPensee CR. Focus on prolactin as a metabolic hormone. Trends Endocrinol Metab 2006; 17(3): 110-6.

111. Carré N, Binart N. Prolactin and adipose tissue. Biochimie 2014; 97: 16-21.

112. Parsons JA, Brelje TC, Sorenson RL. Adaptation of islets of Langerhans to pregnancy: increased islet cell proliferation and insulin secretion correlates with the onset of placental lactogen secretion. Endocrinology 1992; 130(3): 1459-66.

113. Bone AJ, Taylor KW. Mitabolic adaptation to pregnancy shown by increased biosynthesis of insulin in islets of Langerhans isolated from pregnant rat. Nature 1976; 262(5568): 501-2.

114. Sorenson RL, Parsons JA. Insulin secretion in mammosomatotropic tumor-bearing and pregnant rats. A role for lactogens. Diabetes 1985; 34(4): 337-41.

115. Newbern D, Freemark M. Placental hormones and the control of maternal metabolism and fetal growth. Curr Opin Endocrinol Diabetes Obes 2011; 18(6): 409-16.

116. Henry RK. When They're Done Growing, Don't Forget They May Still Need Growth Hormone. Metab Syndr Relat Disord 2021.

117. Vila G, Jørgensen JOL, Luger A, Stalla GK. Insulin Resistance in Patients With Acromegaly. Frontiers in endocrinology 2019; 10: 509.

118. Khatib N, Gaidhane S, Gaidhane AM, et al. Ghrelin: ghrelin as a regulatory Peptide in growth hormone secretion. Journal of clinical and diagnostic research : 2014; 8(8): MC13-MC7.

119. Pradhan G, Samson SL, Sun Y. Ghrelin: much more than a hunger hormone. Curr Opin Clin Nutr Metab Care 2013; 16(6): 619-24.

120. Faupel-Badger JM, Berrigan D, Ballard-Barbash R, Potischman N. Anthropometric correlates of insulin-like growth factor 1 (IGF-1) and IGF binding protein-3 (IGFBP-3) levels by race/ethnicity and gender. Ann Epidemiol 2009; 19(12): 841-9.

121. Junnila RK, List EO, Berryman DE, Murrey JW, Kopchick JJ. The GH/IGF-1 axis in ageing and longevity. Nat Rev Endocrinol 2013; 9(6): 366-76.

122. Hussain MA, Schmitz O, Mengel A, et al. Comparison of the effects of growth hormone and insulin-like growth factor I on substrate oxidation and on insulin sensitivity in growth hormone-deficient humans. The Journal of clinical investigation 1994; 94(3): 1126-33.

123. Momozono A, Hayashi A, Takano K, Shichiri M. The effectiveness of growth hormone replacement on energy expenditure and body composition in patients with adult growth hormone deficiency. Endocr J 2020.

124. van Bunderen CC, Meijer RI, Lips P, Kramer MH, Serné EH, Drent ML. Titrating Growth Hormone Dose to High-Normal IGF-1 Levels Has Beneficial Effects on Body Fat Distribution and Microcirculatory Function Despite Causing Insulin Resistance. Front Endocrinol (Lausanne) 2020; 11: 619173.

125. Xue P, Wang Y, Yang J, Li Y. Effects of Growth Hormone Replacement Therapy on Bone Mineral Density in Growth Hormone Deficient Adults: A Meta-Analysis. International Journal of Endocrinology 2013; 2013: 216107.

126. Zahr R, Fleseriu M. Updates in Diagnosis and Treatment of Acromegaly. European endocrinology 2018; 14(2): 57-61.

127. Stafford DE. Altered hypothalamic-pituitary-ovarian axis function in young female athletes: implications and recommendations for management. Treat Endocrinol 2005; 4(3): 147-54.

128. Adashi EY. Endocrinology of the ovary. Hum Reprod 1994; 9(5): 815-27.

129. Väänänen HK, Härkönen PL. Estrogen and bone metabolism. Maturitas 1996; 23 Suppl: S65-9.

130. Iorga A, Cunningham CM, Moazeni S, Ruffenach G, Umar S, Eghbali M. The protective role of estrogen and estrogen receptors in cardiovascular disease and the controversial use of estrogen therapy. Biology of sex differences 2017; 8(1): 33.

131. Hamilton KJ, Hewitt SC, Arao Y, Korach KS. Estrogen Hormone Biology. Current topics in developmental biology 2017; 125: 109-46.

132. Kirschner MA, Ertel N, Schneider G. Obesity, hormones, and cancer. Cancer Res 1981; 41(9 Pt 2): 3711-7.

133. Schoenaker DA, Jackson CA, Rowlands JV, Mishra GD. Socioeconomic position, lifestyle factors and age at natural menopause: a systematic review and meta-analyses of studies across six continents. Int J Epidemiol 2014; 43(5): 1542-62.

134. Ambikairajah A, Walsh E, Tabatabaei-Jafari H, Cherbuin N. Fat mass changes during menopause: a metaanalysis. Am J Obstet Gynecol 2019; 221(5): 393-409.e50.

135. Su HI, Freeman EW. Hormone changes associated with the menopausal transition. Minerva ginecologica 2009; 61(6): 483-9.

136. Burger HG. Diagnostic role of follicle-stimulating hormone (FSH) measurements during the menopausal transition--an analysis of FSH, oestradiol and inhibin. Eur J Endocrinol 1994; 130(1): 38-42.

137. Heymsfield SB, Gallagher D, Poehlman ET, et al. Menopausal changes in body composition and energy expenditure. Exp Gerontol 1994; 29(3-4): 377-89.

138. McFarland KC, Sprengel R, Phillips HS, et al. Lutropin-choriogonadotropin receptor: an unusual member of the G protein-coupled receptor family. Science 1989; 245(4917): 494-9.

139. Matsumoto AM, Karpas AE, Paulsen CA, Bremner WJ. Reinitiation of sperm production in gonadotropin-suppressed normal men by administration of follicle-stimulating hormone. J Clin Invest 1983; 72(3): 1005-15.

140. Mouser JG, Loprinzi PD, Loenneke JP. The association between physiologic testosterone levels, lean mass, and fat mass in a nationally representative sample of men in the United States. Steroids 2016; 115: 62-6.

141. McEwen BS. Central effects of stress hormones in health and disease: Understanding the protective and damaging effects of stress and stress mediators. European journal of pharmacology 2008; 583(2-3): 174-85.

142. Angelousi A, Margioris AN, Tsatsanis C. ACTH Action on the Adrenals. In: Feingold KR, Anawalt B, Boyce A, et al., eds. Endotext. South Dartmouth (MA): MDText.com, Inc.Copyright © 2000-2021, MDText.com, Inc.; 2000.

143. Ramamoorthy S, Cidlowski JA. Corticosteroids: Mechanisms of Action in Health and Disease. Rheum Dis Clin North Am 2016; 42(1): 15-31, vii.

144. Rosmond R, Dallman MF, Björntorp P. Stress-related cortisol secretion in men: relationships with abdominal obesity and endocrine, metabolic and hemodynamic abnormalities. J Clin Endocrinol Metab 1998; 83(6): 1853-9.

145. Laatikainen TJ. Corticotropin-releasing hormone and opioid peptides in reproduction and stress. Ann Med 1991; 23(5): 489-96.

146. Björntorp P. Visceral obesity: a "civilization syndrome". Obes Res 1993; 1(3): 206-22.

147. Kadmiel M, Cidlowski JA. Glucocorticoid receptor signaling in health and disease. Trends Pharmacol Sci 2013; 34(9): 518-30.

148. Oakley RH, Cidlowski JA. The biology of the glucocorticoid receptor: new signaling mechanisms in health and disease. J Allergy Clin Immunol 2013; 132(5): 1033-44.

149. Hakamata Y, Komi S, Moriguchi Y, et al. Amygdala-centred functional connectivity affects daily cortisol concentrations: a putative link with anxiety. Sci Rep 2017; 7(1): 8313.

150. Lee DY, Kim E, Choi MH. Technical and clinical aspects of cortisol as a biochemical marker of chronic stress. BMB Rep 2015; 48(4): 209-16.

151. Kuo T, McQueen A, Chen TC, Wang JC. Regulation of Glucose Homeostasis by Glucocorticoids. Adv Exp Med Biol 2015; 872: 99-126.

152. Geer EB, Islam J, Buettner C. Mechanisms of glucocorticoid-induced insulin resistance: focus on adipose tissue function and lipid metabolism. Endocrinology and metabolism clinics of North America 2014; 43(1): 75-102.

153. Khani S, Tayek JA. Cortisol increases gluconeogenesis in humans: its role in the metabolic syndrome. Clin Sci (Lond) 2001; 101(6): 739-47.

154. Husebye ES, Pearce SH, Krone NP, Kämpe O. Adrenal insufficiency. Lancet 2021; 397(10274): 613-29.

155. Exton JH. Regulation of gluconeogenesis by glucocorticoids. Monogr Endocrinol 1979; 12: 535-46.

156. Ragnarsson O, Glad CA, Bergthorsdottir R, et al. Body composition and bone mineral density in women with Cushing's syndrome in remission and the association with common genetic variants influencing glucocorticoid sensitivity. Eur J Endocrinol 2015; 172(1): 1-10.

157. Watabe T, Tanaka K, Kumagae M, et al. Diurnal rhythm of plasma immunoreactive corticotropin-releasing factor in normal subjects. Life Sci 1987; 40(17): 1651-5.

158. Engelmann M, Landgraf R, Wotjak CT. The hypothalamic-neurohypophysial system regulates the hypothalamic-pituitary-adrenal axis under stress: an old concept revisited. Front Neuroendocrinol 2004; 25(3-4): 132-49.

159. Michels A, Michels N. Addison disease: early detection and treatment principles. Am Fam Physician 2014; 89(7): 563-8.

160. De Sanctis V, Soliman A, Yassin M, Garofalo P. Cortisol levels in central adrenal insufficiency: light and shade. Pediatr Endocrinol Rev 2015; 12(3): 283-9.

161. Raff H, Carroll T. Cushing's syndrome: from physiological principles to diagnosis and clinical care. J Physiol 2015; 593(3): 493-506.

162. Pucci E, Chiovato L, Pinchera A. Thyroid and lipid metabolism. Int J Obes Relat Metab Disord 2000; 24 Suppl 2: S109-12.

163. Williams GR, Bassett JHD. Thyroid diseases and bone health. J Endocrinol Invest 2018; 41(1): 99-109.

164. Shahid MA, Ashraf MA, Sharma S. Physiology, Thyroid Hormone. StatPearls. Treasure Island (FL): StatPearls Publishing Copyright © 2021, StatPearls Publishing LLC.; 2021.

165. Ortiga-Carvalho TM, Chiamolera MI, Pazos-Moura CC, Wondisford FE. Hypothalamus-Pituitary-Thyroid Axis. Compr Physiol 2016; 6(3): 1387-428.

166. Abdalla SM, Bianco AC. Defending plasma T3 is a biological priority. Clinical endocrinology 2014; 81(5): 633-41.

167. Mariotti S, Beck-Peccoz P. Physiology of the Hypothalamic-Pituitary-Thyroid Axis. In: Feingold KR, Anawalt B, Boyce A, et al., eds. Endotext. South Dartmouth (MA): MDText.com, Inc. Copyright © 2000-2021, MDText.com, Inc.; 2000.

168. Bianco AC, Salvatore D, Gereben B, Berry MJ, Larsen PR. Biochemistry, cellular and molecular biology, and physiological roles of the iodothyronine selenodeiodinases. Endocr Rev 2002; 23(1): 38-89.

169. Mullur R, Liu Y-Y, Brent GA. Thyroid hormone regulation of metabolism. Physiological reviews 2014; 94(2): 355-82.

170. Morley JE, Sawin CT, Carlson HE, Longcope C, Hershman JM. The relationship of androgen to the thyrotropin and prolactin responses to thyrotropin-releasing hormone in hypogonadal and normal men. J Clin Endocrinol Metab 1981; 52(2): 173-6.

171. Faglia G, Beck-Peccoz P, Ferrari C, Ambrosi B, Spada A, Travaglini P. Enhanced plasma thyrotrophin response to thyrotrophin-releasing hormone following oestradiol administration in man. Clin Endocrinol (Oxf) 1973; 2(3): 207-10.

172. Gershengorn MC, Marcus-Samuels BE, Geras E. Estrogens increase the number of thyrotropin-releasing hormone receptors on mammotropic cells in culture. Endocrinology 1979; 105(1): 171-6.

173. Barakat MT, Meeran K, Bloom SR. Neuroendocrine tumours. Endocr Relat Cancer 2004; 11(1): 1-18.

174. Doknić M, Pekić S, Miljić D, et al. Etiology of Hypopituitarism in Adult Patients: The Experience of a Single Center Database in the Serbian Population. International journal of endocrinology 2017; 2017: 6969286.

175. Palmer JD, Hall MD, Mahajan A, Paulino AC, Wolden S, Constine LS. Radiotherapy and Late Effects. Pediatr Clin North Am 2020; 67(6): 1051-67.

176. Darzy KH. Radiation-induced hypopituitarism. Curr Opin Endocrinol Diabetes Obes 2013; 20(4): 342-53.

177. Popovic V, Pekic S, Golubicic I, Doknic M, Dieguez C, Casanueva FF. The impact of cranial irradiation on GH responsiveness to GHRH plus GH-releasing peptide-6. J Clin Endocrinol Metab 2002; 87(5): 2095-9.

178. Engstrom WW, Lennon EJ. The management of endocrine deficiencies of surgical origin. Am J Surg 1960; 99: 580-7.

179. Webb SM, Rigla M, Wägner A, Oliver B, Bartumeus F. Recovery of hypopituitarism after neurosurgical treatment of pituitary adenomas. J Clin Endocrinol Metab 1999; 84(10): 3696-700.

180. Streefkerk N, Tissing WJE, Korevaar JC, et al. A detailed insight in the high risks of hospitalizations in long-term childhood cancer survivors-A Dutch LATER linkage study. PLoS One 2020; 15(5): e0232708.

181. Daly AF, Beckers A. The Epidemiology of Pituitary Adenomas. Endocrinol Metab Clin North Am 2020; 49(3): 347-55.

182. Melmed S. Pituitary tumors. Endocrinology and metabolism clinics of North America 2015; 44(1): 1-9.

183. Fernandez A, Karavitaki N, Wass JA. Prevalence of pituitary adenomas: a community-based, cross-sectional study in Banbury (Oxfordshire, UK). Clin Endocrinol (Oxf) 2010; 72(3): 377-82.

184. Kopczak A, Renner U, Karl Stalla G. Advances in understanding pituitary tumors. F1000prime reports 2014; 6: 5.

185. Casanueva FF, Molitch ME, Schlechte JA, et al. Guidelines of the Pituitary Society for the diagnosis and management of prolactinomas. Clin Endocrinol (Oxf) 2006; 65(2): 265-73.

186. Jagannathan J, Kanter AS, Sheehan JP, Jane JA, Jr., Laws ER, Jr. Benign brain tumors: sellar/parasellar tumors. Neurol Clin 2007; 25(4): 1231-49, xi.

187. Asa SL, Ezzat S. The Endocrine System. Pituitary Disorders; 2013: 1-12.

188. Devin JK. Craniopharyngioma. Pituitary Disorders; 2013: 59-66.

189. Nielsen EH, Feldt-Rasmussen U, Poulsgaard L, et al. Incidence of craniopharyngioma in Denmark (n = 189) and estimated world incidence of craniopharyngioma in children and adults. J Neurooncol 2011; 104(3): 755-63.

190. Zacharia BE, Bruce SS, Goldstein H, Malone HR, Neugut AI, Bruce JN. Incidence, treatment and survival of patients with craniopharyngioma in the surveillance, epidemiology and end results program. Neuro Oncol 2012; 14(8): 1070-8.

191. Erfurth EM. Endocrine aspects and sequel in patients with craniopharyngioma. J Pediatr Endocrinol Metab 2015; 28(1-2): 19-26.

192. Andoniadou CL, Gaston-Massuet C, Reddy R, et al. Identification of novel pathways involved in the pathogenesis of human adamantinomatous craniopharyngioma. Acta Neuropathol 2012; 124(2): 259-71.

193. Gautier A, Godbout A, Grosheny C, et al. Markers of recurrence and long-term morbidity in craniopharyngioma: a systematic analysis of 171 patients. J Clin Endocrinol Metab 2012; 97(4): 1258-67.

194. Bunin GR, Surawicz TS, Witman PA, Preston-Martin S, Davis F, Bruner JM. The descriptive epidemiology of craniopharyngioma. J Neurosurg 1998; 89(4): 547-51.

195. Müller HL. Consequences of craniopharyngioma surgery in children. J Clin Endocrinol Metab 2011; 96(7): 1981-91.

196. Olsson DS, Andersson E, Bryngelsson IL, Nilsson AG, Johannsson G. Excess mortality and morbidity in patients with craniopharyngioma, especially in patients with childhood onset: a population-based study in Sweden. J Clin Endocrinol Metab 2015; 100(2): 467-74.

197. Muller HL. Craniopharyngioma. Handb Clin Neurol 2014; 124: 235-53.

198. Rosemberg S, Fujiwara D. Epidemiology of pediatric tumors of the nervous system according to the WHO 2000 classification: a report of 1,195 cases from a single institution. Childs Nerv Syst 2005; 21(11): 940-4.

199. Wijnen M, van den Heuvel-Eibrink MM, Janssen J, et al. Very long-term sequelae of craniopharyngioma. Eur J Endocrinol 2017; 176(6): 755-67.

200. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. Excess morbidity and mortality in patients with craniopharyngioma: a hospital-based retrospective cohort study. Eur J Endocrinol 2018; 178(1): 95-104.

201. Muller HL, Emser A, Faldum A, et al. Longitudinal study on growth and body mass index before and after diagnosis of childhood craniopharyngioma. J Clin Endocrinol Metab 2004; 89(7): 3298-305.

202. Rossi A, Cama A, Consales A, et al. Neuroimaging of pediatric craniopharyngiomas: a pictorial essay. J Pediatr Endocrinol Metab 2006; 19 Suppl 1: 299-319.

203. Harwood-Nash DC. Neuroimaging of childhood craniopharyngioma. Pediatr Neurosurg 1994; 21 Suppl 1: 2-10.

204. Pascual JM, Carrasco R, Prieto R, Gonzalez-Llanos F, Alvarez F, Roda JM. Craniopharyngioma classification. J Neurosurg 2008; 109(6): 1180-2; author reply 2-3.

205. Buchfelder M, Schlaffer S. Imaging of pituitary pathology. Handb Clin Neurol 2014; 124: 151-66.

206. Adult Craniopharyngiomas: Differences and Lessons from Paediatrics; 2020.

207. Borrill R, Cheesman E, Stivaros S, Kamaly-Asl ID, Gnanalingham K, Kilday JP. Papillary craniopharyngioma in a 4-year-old girl with BRAF V600E mutation: a case report and review of the literature. Childs Nerv Syst 2019; 35(1): 169-73.

208. Goldberg GM, Eshbaugh DE. Squamous cell nests of the pituitary gland as related to the origin of craniopharyngiomas. A study of their presence in the newborn and infants up to age four. Arch Pathol 1960; 70: 293-9.

209. Garnett MR, Puget S, Grill J, Sainte-Rose C. Craniopharyngioma. Orphanet J Rare Dis 2007; 2: 18.

210. Pekmezci M, Louie J, Gupta N, Bloomer MM, Tihan T. Clinicopathological characteristics of adamantinomatous and papillary craniopharyngiomas: University of California, San Francisco experience 1985-2005. Neurosurgery 2010; 67(5): 1341-9; discussion 9.

211. Sekine S, Shibata T, Kokubu A, et al. Craniopharyngiomas of adamantinomatous type harbor beta-catenin gene mutations. Am J Pathol 2002; 161(6): 1997-2001.

212. Apps JR, Carreno G, Gonzalez-Meljem JM, et al. Tumour compartment transcriptomics demonstrates the activation of inflammatory and odontogenic programmes in human adamantinomatous craniopharyngioma and identifies the MAPK/ERK pathway as a novel therapeutic target. Acta Neuropathol 2018; 135(5): 757-77.

213. Buslei R, Nolde M, Hofmann B, et al. Common mutations of beta-catenin in adamantinomatous craniopharyngiomas but not in other tumours originating from the sellar region. Acta Neuropathol 2005; 109(6): 589-97.

214. Brastianos PK, Taylor-Weiner A, Manley PE, et al. Exome sequencing identifies BRAF mutations in papillary craniopharyngiomas. Nat Genet 2014; 46(2): 161-5.

215. Asa SL, Mete O, Ezzat S. Genomics and Epigenomics of Pituitary Tumors: What Do Pathologists Need to Know? Endocr Pathol 2021.

216. Omay SB, Chen YN, Almeida JP, et al. Do craniopharyngioma molecular signatures correlate with clinical characteristics? J Neurosurg 2018; 128(5): 1473-8.

217. Hofmann BM, Kreutzer J, Saeger W, et al. Nuclear beta-catenin accumulation as reliable marker for the differentiation between cystic craniopharyngiomas and rathke cleft cysts: a clinico-pathologic approach. Am J Surg Pathol 2006; 30(12): 1595-603.

218. Larkin SJ, Preda V, Karavitaki N, Grossman A, Ansorge O. BRAF V600E mutations are characteristic for papillary craniopharyngioma and may coexist with CTNNB1-mutated adamantinomatous craniopharyngioma. Acta Neuropathol 2014; 127(6): 927-9.

219. Karavitaki N. Management of craniopharyngiomas. J Endocrinol Invest 2014; 37(3): 219-28.

220. Buchfelder M, Schlaffer S-M, Lin F, Kleindienst A. Surgery for craniopharyngioma. Pituitary 2013; 16(1): 18-25.

221. Muller HL. Childhood craniopharyngioma. Pituitary 2013; 16(1): 56-67.

222. Prieto R, Pascual JM, Rosdolsky M, et al. Craniopharyngioma adherence: a comprehensive topographical categorization and outcome-related risk stratification model based on the methodical examination of 500 tumors. Neurosurg Focus 2016; 41(6): E13.

223. Puget S, Garnett M, Wray A, et al. Pediatric craniopharyngiomas: classification and treatment according to the degree of hypothalamic involvement. J Neurosurg 2007; 106(1 Suppl): 3-12.

224. Iannalfi A, Fragkandrea I, Brock J, Saran F. Radiotherapy in craniopharyngiomas. Clin Oncol (R Coll Radiol) 2013; 25(11): 654-67.

225. Dandurand C, Sepehry AA, Asadi Lari MH, Akagami R, Gooderham P. Adult Craniopharyngioma: Case Series, Systematic Review, and Meta-Analysis. Neurosurgery 2018; 83(4): 631-41.

226. Schoenfeld A, Pekmezci M, Barnes MJ, et al. The superiority of conservative resection and adjuvant radiation for craniopharyngiomas. J Neurooncol 2012; 108(1): 133-9.

227. Rao YJ, Hassanzadeh C, Fischer-Valuck B, et al. Patterns of care and treatment outcomes of patients with Craniopharyngioma in the national cancer database. J Neurooncol 2017; 132(1): 109-17.

228. Moszczynska E, Prokop-Piotrkowska M, Bogusz-Wojcik A, Grajkowska W, Szymanska S, Szalecki M. Ki67 as a prognostic factor of craniopharyngioma's recurrence in paediatric population. Childs Nerv Syst 2020.

229. Cheng J, Shao Q, Pan Z, You J. Analysis and Long-Term Follow-Up of the Surgical Treatment of Children With Craniopharyngioma. J Craniofac Surg 2016; 27(8): e763-e6.

230. Heinks K, Boekhoff S, Hoffmann A, et al. Quality of life and growth after childhood craniopharyngioma: results of the multinational trial KRANIOPHARYNGEOM 2007. Endocrine 2018; 59(2): 364-72.

231. van Iersel L, Brokke KE, Adan RAH, Bulthuis LCM, van den Akker ELT, van Santen HM. Pathophysiology and Individualized Treatment of Hypothalamic Obesity Following Craniopharyngioma and Other Suprasellar Tumors: A Systematic Review. Endocr Rev 2019; 40(1): 193-235.

232. Bogusz A, Boekhoff S, Warmuth-Metz M, Calaminus G, Eveslage M, Müller HL. Posterior hypothalamus-sparing surgery improves outcome after childhood craniopharyngioma. Endocr Connect 2019; 8(5): 481-92.

233. Müller HL. The Diagnosis and Treatment of Craniopharyngioma. Neuroendocrinology 2020; 110(9-10): 753-66.

234. Merchant TE, Kiehna EN, Sanford RA, et al. Craniopharyngioma: the St. Jude Children's Research Hospital experience 1984-2001. Int J Radiat Oncol Biol Phys 2002; 53(3): 533-42.

235. Kendall-Taylor P, Jonsson PJ, Abs R, et al. The clinical, metabolic and endocrine features and the quality of life in adults with childhood-onset craniopharyngioma compared with adult-onset craniopharyngioma. Eur J Endocrinol 2005; 152(4): 557-67.

236. Roth CL, Gebhardt U, Müller HL. Appetite-regulating hormone changes in patients with craniopharyngioma. Obesity (Silver Spring) 2011; 19(1): 36-42.

237. Muller HL, Gebhardt U, Etavard-Gorris N, et al. Prognosis and sequela in patients with childhood craniopharyngioma -- results of HIT-ENDO and update on KRANIOPHARYNGEOM 2000. Klin Padiatr 2004; 216(6): 343-8.

238. Lustig RH. Hypothalamic obesity after craniopharyngioma: mechanisms, diagnosis, and treatment. Frontiers in endocrinology 2011; 2: 60.

239. Roth CL. Hypothalamic obesity in patients with craniopharyngioma: profound changes of several weight regulatory circuits. Frontiers in endocrinology 2011; 2: 49-.

240. Kilday JP, Bartels U, Huang A, et al. Favorable survival and metabolic outcome for children with diencephalic syndrome using a radiation-sparing approach. J Neurooncol 2014; 116(1): 195-204.

241. Hoffmann A, Gebhardt U, Sterkenburg AS, Warmuth-Metz M, Müller HL. Diencephalic syndrome in childhood craniopharyngioma--results of German multicenter studies on 485 long-term survivors of childhood craniopharyngioma. J Clin Endocrinol Metab 2014; 99(11): 3972-7.

242. Green DM, Cox CL, Zhu L, et al. Risk factors for obesity in adult survivors of childhood cancer: a report from the Childhood Cancer Survivor Study. J Clin Oncol 2012; 30(3): 246-55.

243. Blijdorp K, van den Heuvel-Eibrink MM, Pieters R, et al. Obesity is underestimated using body mass index and waist-hip ratio in long-term adult survivors of childhood cancer. PLoS One 2012; 7(8): e43269.

244. van Santen SS, Olsson DS, Hammarstrand C, et al. Diagnosing Metabolic Syndrome in Craniopharyngioma Patients: Body Composition versus BMI. Eur J Endocrinol 2019.

245. Perreault L. Determining body composition in adults. July 10, 2017 2017. https://www-uptodate-com.eur.idm.oclc.org/contents/determining-body-composition-in-adults?search=body%20composition%20adults&source=search_result&selectedTitle=1~116&usage_type=default&display_rank=1#H3702267332 (accessed July 4th, 2018 2018).

246. Gu L, Huang X, Li S, et al. A meta-analysis of the medium- and long-term effects of laparoscopic sleeve gastrectomy and laparoscopic Roux-en-Y gastric bypass. BMC Surg 2020; 20(1): 30.

247. Arterburn D, Wellman R, Emiliano A, et al. Comparative Effectiveness and Safety of Bariatric Procedures for Weight Loss: A PCORnet Cohort Study. Ann Intern Med 2018; 169(11): 741-50.

248. Rao RS. Bariatric surgery and the central nervous system. Obes Surg 2012; 22(6): 967-78.

249. Trotta M, Da Broi J, Salerno A, Testa RM, Marinari GM. Sleeve gastrectomy leads to easy management of hormone replacement therapy and good weight loss in patients treated for craniopharyngioma. Updates Surg 2017; 69(1): 95-9.

250. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. Efficacy and safety of bariatric surgery for craniopharyngioma-related hypothalamic obesity: a matched case-control study with 2 years of follow-up. Int J Obes (Lond) 2017; 41(2): 210-6.

251. Nuti R, Brandi ML, Checchia G, et al. Guidelines for the management of osteoporosis and fragility fractures. Internal and emergency medicine 2019; 14(1): 85-102.

252. Kanis JA. Diagnosis of osteoporosis and assessment of fracture risk. Lancet 2002; 359(9321): 1929-36.

253. Latoch E, Konstantynowicz J, Krawczuk-Rybak M, Panasiuk A, Muszyńska-Rosłan K. A long-term trajectory of bone mineral density in childhood cancer survivors after discontinuation of treatment: retrospective cohort study. Arch Osteoporos 2021; 16(1): 45.

254. Baxter-Jones AD, Faulkner RA, Forwood MR, Mirwald RL, Bailey DA. Bone mineral accrual from 8 to 30 years of age: an estimation of peak bone mass. J Bone Miner Res 2011; 26(8): 1729-39.

255. van Atteveld JE, Pluijm SMF, Ness KK, et al. Prediction of Low and Very Low Bone Mineral Density Among Adult Survivors of Childhood Cancer. J Clin Oncol 2019: Jco1801917.

256. Johnell O, Kanis JA. An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. Osteoporos Int 2006; 17(12): 1726-33.

257. Bloomhardt HM, Sint K, Ross WL, et al. Severity of reduced bone mineral density and risk of fractures in long-term survivors of childhood leukemia and lymphoma undergoing guideline-recommended surveillance for bone health. Cancer 2020; 126(1): 202-10.

258. Chemaitilly W, Sklar CA. Endocrine complications in long-term survivors of childhood cancers. Endocr Relat Cancer 2010; 17(3): R141-59.

259. Isaksson S, Bogefors K, Åkesson K, et al. Low bone mineral density is associated with hypogonadism and cranial irradiation in male childhood cancer survivors. Osteoporosis International 2020; 31(7): 1261-72.

260. Erfurth EM. Uncertainties in endocrine substitution therapy for central endocrine insufficiencies: growth hormone deficiency. Handb Clin Neurol 2014; 124: 407-16.

261. Okinaga H, Matsuno A, Okazaki R. High risk of osteopenia and bone derangement in postsurgical patients with craniopharyngiomas, pituitary adenomas and other parasellar lesions. Endocr J 2005; 52(6): 751-6.

262. Muller HL, Schneider P, Bueb K, et al. Volumetric bone mineral density in patients with childhood craniopharyngioma. Exp Clin Endocrinol Diabetes 2003; 111(3): 168-73.

263. Feldt-Rasmussen U, Klose M. Adult Growth Hormone Deficiency Clinical Management. In: Feingold KR, Anawalt B, Boyce A, et al., eds. Endotext. South Dartmouth (MA): MDText.com, Inc. Copyright © 2000-2021, MDText.com, Inc.; 2000.

264. Lamas Oliveira C. Metabolic consequences of craniopharyingioma and their management. Endocrinol Nutr 2013; 60(9): 529-34.

265. Maiter D, Abs R, Johannsson G, et al. Baseline characteristics and response to GH replacement of hypopituitary patients previously irradiated for pituitary adenoma or craniopharyngioma: data from the Pfizer International Metabolic Database. Eur J Endocrinol 2006; 155(2): 253-60.

266. Wuster C, Abs R, Bengtsson BA, et al. The influence of growth hormone deficiency, growth hormone replacement therapy, and other aspects of hypopituitarism on fracture rate and bone mineral density. J Bone Miner Res 2001; 16(2): 398-405.

267. Vestergaard P, Jørgensen JO, Hagen C, et al. Fracture risk is increased in patients with GH deficiency or untreated prolactinomas--a case-control study. Clin Endocrinol (Oxf) 2002; 56(2): 159-67.

268. Mitchell A, Larsson SC, Fall T, Melhus H, Michaëlsson K, Byberg L. Fasting glucose, bone area and bone mineral density: a Mendelian randomisation study. Diabetologia 2021.

269. Asadi-Pooya AA, Tomson T. A reappraisal of injuries and accidents in people with epilepsy. Curr Opin Neurol 2021; 34(2): 182-7.

270. Elavarasi A, Parihar J, Kapoor S, et al. Seizure-related injuries in inadequately treated epilepsy patients: A case-control study. Seizure 2020; 83: 17-20.

271. Nuijts MA, Veldhuis N, Stegeman I, et al. Visual functions in children with craniopharyngioma at diagnosis: A systematic review. PLoS One 2020; 15(10): e0240016.

272. Choi HG, Lee JK, Lee MJ, Park B, Sim S, Lee SM. Blindness increases the risk for hip fracture and vertebral fracture but not the risk for distal radius fracture: a longitudinal follow-up study using a national sample cohort. Osteoporos Int 2020; 31(12): 2345-54.

273. Hamedani AG, VanderBeek BL, Willis AW. Blindness and Visual Impairment in the Medicare Population: Disparities and Association with Hip Fracture and Neuropsychiatric Outcomes. Ophthalmic Epidemiol 2019; 26(4): 279-85.

274. Jantzi M, Maher AC, Ioannidis G, Hirdes JP, Giangregorio LM, Papaioannou A. Individuals with neurological diseases are at increased risk of fractures within 180 days of admission to long-term care in Ontario. Age and ageing 2015; 44(2): 252-7.

275. Poole KE, Reeve J, Warburton EA. Falls, fractures, and osteoporosis after stroke: time to think about protection? Stroke 2002; 33(5): 1432-6.

276. Zhao Y, Gan X, Luo P, et al. The risk of osteopenia in premenopausal women with various sellar tumors. Gynecol Endocrinol 2012; 28(12): 945-8.

277. Andereggen L, Hess B, Andres R, et al. A ten-year follow-up study of treatment outcome of craniopharyngiomas. Swiss Med Wkly 2018; 148: w14521.

278. Holmer H, Popovic V, Ekman B, Follin C, Siversson AB, Erfurth EM. Hypothalamic involvement and insufficient sex steroid supplementation are associated with low bone mineral density in women with childhood onset craniopharyngioma. Eur J Endocrinol 2011; 165(1): 25-31.

279. Verhelst J, Kendall-Taylor P, Erfurth EM, et al. Baseline characteristics and response to 2 years of growth hormone (GH) replacement of hypopituitary patients with GH deficiency due to adult-onset craniopharyngioma in comparison with patients with nonfunctioning pituitary adenoma: data from KIMS (Pfizer International Metabolic Database). J Clin Endocrinol Metab 2005; 90(8): 4636-43.

280. Geffner M, Lundberg M, Koltowska-Haggstrom M, et al. Changes in height, weight, and body mass index in children with craniopharyngioma after three years of growth hormone therapy: analysis of KIGS (Pfizer International Growth Database). J Clin Endocrinol Metab 2004; 89(11): 5435-40.

281. Jensen BL, Jensen KE, Kastrup KW, Pedersen SA, Wagner A. Final height and craniofacial development after surgical resection of craniopharyngioma. J Craniofac Genet Dev Biol 1997; 17(4): 190-7.

282. Price DA, Wilton P, Jonsson P, et al. Efficacy and safety of growth hormone treatment in children with prior craniopharyngioma: an analysis of the Pharmacia and Upjohn International Growth Database (KIGS) from 1988 to 1996. Horm Res 1998; 49(2): 91-7.

283. Burns EC, Tanner JM, Preece MA, Cameron N. Growth hormone treatment in children with craniopharyngioma: final growth status. Clin Endocrinol (Oxf) 1981; 14(6): 587-95.

284. Rose SR, Horne VE, Howell J, et al. Late endocrine effects of childhood cancer. Nat Rev Endocrinol 2016; 12(6): 319-36.

285. George SA, Effinger KE, Meacham LR. Endocrine Sequelae in Childhood Cancer Survivors. Endocrinol Metab Clin North Am 2020; 49(4): 565-87.

286. van Santen HM, Chemaitilly W, Meacham LR, Tonorezos ES, Mostoufi-Moab S. Endocrine Health in Childhood Cancer Survivors. Pediatr Clin North Am 2020; 67(6): 1171-86.

287. Chemaitilly W, Sklar CA. Childhood Cancer Treatments and Associated Endocrine Late Effects: A Concise Guide for the Pediatric Endocrinologist. Horm Res Paediatr 2019; 91(2): 74-82.

288. DeVile CJ, Grant DB, Hayward RD, Stanhope R. Growth and endocrine sequelae of craniopharyngioma. Arch Dis Child 1996; 75(2): 108-14.

289. Fjalldal S, Follin C, Gabery S, et al. Detailed assessment of hypothalamic damage in craniopharyngioma patients with obesity. Int J Obes (Lond) 2019; 43(3): 533-44.

290. Holmer H, Ekman B, Bjork J, et al. Hypothalamic involvement predicts cardiovascular risk in adults with childhood onset craniopharyngioma on long-term GH therapy. Eur J Endocrinol 2009; 161(5): 671-9.

291. Follin C, Erfurth EM. Long-Term Effect of Cranial Radiotherapy on Pituitary-Hypothalamus Area in Childhood Acute Lymphoblastic Leukemia Survivors. Curr Treat Options Oncol 2016; 17(9): 50.

292. Krashes MJ, Koda S, Ye C, et al. Rapid, reversible activation of AgRP neurons drives feeding behavior in mice. The Journal of clinical investigation 2011; 121(4): 1424-8.

293. Thompson CJ, Costello RW, Crowley RK. Management of hypothalamic disease in patients with craniopharyngioma. Clin Endocrinol (Oxf) 2019; 90(4): 506-16.

294. Erfurth EM. Diagnosis, Background, and Treatment of Hypothalamic Damage in Craniopharyngioma. Neuroendocrinology 2020; 110(9-10): 767-79.

295. Whelan R, Hengartner A, Folzenlogen Z, Prince E, Hankinson TC. Adamantinomatous craniopharyngioma in the molecular age and the potential of targeted therapies: a review. Child's Nervous System 2020; 36(8): 1635-42.

296. Larkin SJ, Ansorge O. Pathology and pathogenesis of craniopharyngiomas. Pituitary 2013; 16(1): 9-17.

297. Müller H, Kaatsch R, Warmuth-Metz M, Flentje M, Sörensen N. Kraniopharyngeom im Kindes- und Jugendalter: Diagnostische und therapeutische Strategien. Monatsschrift Kinderheilkunde 2003; 151: 1056-63.

2

Complicated Clinical Course in Incipient Gigantism Due To A Treatment Resistant, *AIP*-mutated Pediatric Somatotropinoma

Selveta S. van Santen,[1,2] Adrian F. Daly,[3]
Michael Buchfelder,[4] Roland Coras,[5] Yining Zhao,[4]
Albert Beckers,[3] Aart J. van der Lely,[1] Leo J. Hofland,[1]
Rutger K. Balvers,[6] Marry M. van den Heuvel-Eibrink,[6]
Sebastian J. C. M. M. Neggers.[1,2]

[1]Department of Medicine, Endocrinology; Erasmus Medical Center, Rotterdam, The Netherlands;
[2]Princess Máxima Center for Paediatric Oncology, Utrecht, The Netherlands;
[3]Department of Endocrinology, Liège University Hospital Centre, Liège University, Liège, Belgium;
[4]Department of Neurosurgery; University Hospital Erlangen, Erlangen, Germany;
[5]Department of Pathology; University Hospital Erlangen, Erlangen, Germany;
[6]Department of Medicine, Neurosurgery; Erasmus Medical Center, Rotterdam, The Netherlands.

Submitted at AACE Clinical Case Reports

Running title: Incipient gigantism with *AIP* mutation

Abbreviations: *AIP* = aryl hydrocarbon-receptor interacting protein; FSH = follicle stimulating hormone; fT4 = free T4/thyroxine; GH = growth hormone; IGF-1 = Insulin-like Growth Factor 1; LH = luteinizing hormone; PA = pituitary adenoma; standardized deviation score = SDS; SSA = somatostatin receptor analogue; somatostatin receptors = SSTRs; TSH = thyroid stimulating hormone.

Highlights:

- This 7-year-old male patient had accelerated growth, headaches and a high IGF-1
- Pituitary macro-adenoma occurred due to a novel truncating *AIP* mutation, p.Tyr202*
- Multiple treatment modalities were needed to achieve tumoral/biochemical control
- In vitro tissue response to SSA may better predict in vivo response than SSTR immunohistochemistry
- The patient will likely need radiotherapy due to continued tumor growth

Abstract

Objective: To describe the clinical course and treatment challenges in a patient with a pituitary adenoma due to a novel *aryl hydrocarbon-receptor interacting protein* (*AIP*) gene mutation.

Methods: A 7-year-old boy with headache, visual field defects and accelerated growth following failure to thrive.

Results: A growth hormone (GH)- and prolactin-secreting pituitary macroadenoma was identified by magnetic resonance imaging (MRI). The patient underwent three neurosurgical procedures, with every time apparent total resection of the lesion, but the tumor recurred. There were no pituitary insufficiencies, but GH levels remained elevated. A loss of somatostatin receptor 5 was observed between the second and third tumor resections. *In vitro*, no effect on tumoral GH release by pasireotide and pasireotide/cabergoline was seen. *In vivo*, hormonal/tumoral control could not be achieved with lanreotide or pasireotide, while IGF-1 levels could be reduced with the GH receptor blocker pegvisomant. Genetic analysis revealed a novel germline *AIP* mutation: p.Tyr202* (pathogenic; class 4).

Conclusions: A 7-year-old boy with incipient acrogigantism presented with a pathological *AIP* mutation. In this case, *in vitro* response of tumor tissue to somatostatin may better predict tumoral *in vivo* responses of somatostatin analogues than somatostatin receptor immunohistochemistry. The clinical course was complex, as the responses to surgeries and multiple systemic therapies to control hormone levels and tumor growth were unsatisfactory.

Keywords: acromegaly; gigantism; *AIP* mutation; pituitary adenoma; macroadenoma; somatotropinoma

Introduction

Pituitary adenomas (PAs) are among the most frequent intracranial tumors with a prevalence of 1 per 1000 clinically-relevant adenomas in adults.[1] PAs produce signs and symptoms due to abnormal hormonal secretion or mass effects of the tumor itself on local structures. Most cases occur sporadically, but 5% have a familial background,[1,2] the most common being familial isolated pituitary adenomas (FIPA). In FIPA, ~15-30% of cases are associated with pathological germline variants in the *aryl hydrocarbon receptor-interacting protein* (*AIP*) gene, a tumor suppressor gene located on chromosome 11q13.[2-6] Germline *AIP* mutations are particularly associated with growth hormone- (GH) and/or prolactin-secreting pituitary adenomas,[3-6] which occur at a younger age than non-mutated counterparts and comprise an important proportion of somatotropinomas in pediatric/adolescent populations.[7] Up to 21% of pediatric macroadenomas show *AIP* mutations.[2] Patients with *AIP* mutations are often male, have an aggressive clinical phenotype and have large invasive tumors.[2,5,7] *AIP* mutations are the most frequent genetic cause of pituitary gigantism (29%).[8] The median age at presentation in *AIP*-mutated pituitary adenomas is during mid-to-late adolescence up to early adulthood.[9] Herein, we report a case of a recurrent somatotropinoma in a young patient with a novel familial germline *AIP* mutation.

Case report

A 7-year-old boy was hospitalized for evaluation of multiple progressive complaints over the past two years, including frontal headache, fatigue, tics, leg pain, nocturnal sweating, constipation and poor food intake. He was born with a normal birthweight/height following an unremarkable pregnancy and his family history was normal. The growth curve showed normal growth until the age of three, followed by a marked decrease to about -2SDS at the age of six (Figure 1). Thereafter, his growth increased rapidly compared to Dutch national standards. His parents were of modest stature for Dutch standards (midparental height 1.74 m). A sellar tumor was discovered and was initially suspected to be

craniopharyngioma (Figure 2). However, the sella turcica was enlarged. Laboratory analysis showed no signs of pituitary insufficiencies (TSH 1.07 mU/l, fT4 20.2 pmol/l, afternoon cortisol 270 nmol/l). Prolactin level was slightly elevated (0.5 nmol/l). LH and FSH were undetectable, and there were no signs of precocious puberty. However, his IGF-I level was clearly elevated with 56.5 nmol/L (SDS +3.24). Hence, the tumor was diagnosed as a GH-secreting macroadenoma. Over time, growth rate further accelerated (Figure 1), in parallel to a rise in IGF-1 level (71.2 nmol/l; +4.25 SDS), with random GH level of 30.8 mcg/l. Prolactin reached a maximum level of 0.75 nmol/l and he complained of vomiting and loss of appetite. GH levels were not suppressed during an oral glucose tolerance test (lowest GH value of 26.7 mcg/l). Treatment with lanreotide 120 mg four-weekly was initiated, which did neither result in a biochemical response (IGF-I 76.5 nmol/l, GH 28.4 mcg/l), nor in inhibition of tumor growth after four months/injections. Lanreotide was switched to pasireotide LAR 60 mg four-weekly. One month after switching, the first transsphenoidal surgery was performed, because of symptomatic compression of the optic chiasma (onset of bitemporal field defects), and headaches unresponsive to somatostatin receptor analogues (SSAs). Two months postneurosurgery, treatment resulted in a modest effect with a decrease of IGF-I to 70.3 nmol/l, of GH to 23.4 mcg/l. After three months of pasireotide, it was replaced with a weekly dose of 40 mg (GH receptor antagonist) pegvisomant. IGF-I levels dropped to 34.1 nmol/L (0.87 SDS)(GH 48.6 mcg/l). Unfortunately, after one month of pegvisomant, severe headaches returned, and bitemporal hemianopsia reoccurred due to tumor progression (Figure 2). Pegvisomant was stopped and a second transsphenoidal resection followed (Figure 2). The histopathological-anatomical report revealed a pituitary adenoma staining positive for GH and negative for prolactin (Figure 3). One month after the second transsphenoidal surgery, IGF-I level declined to 29.3 (0.4 SDS), GH level to 2.7 mcg/L and prolactin from 0.60 to 0.38 nmol/l after surgery without systemic treatment. Five months after surgery, the headaches returned and an MRI one month thereafter showed a small remnant lateral of the right internal carotid artery (Figure 2). IGF-I level increased again to +2.7 SDS. A third transsphenoidal surgery was performed, leading to normalization of

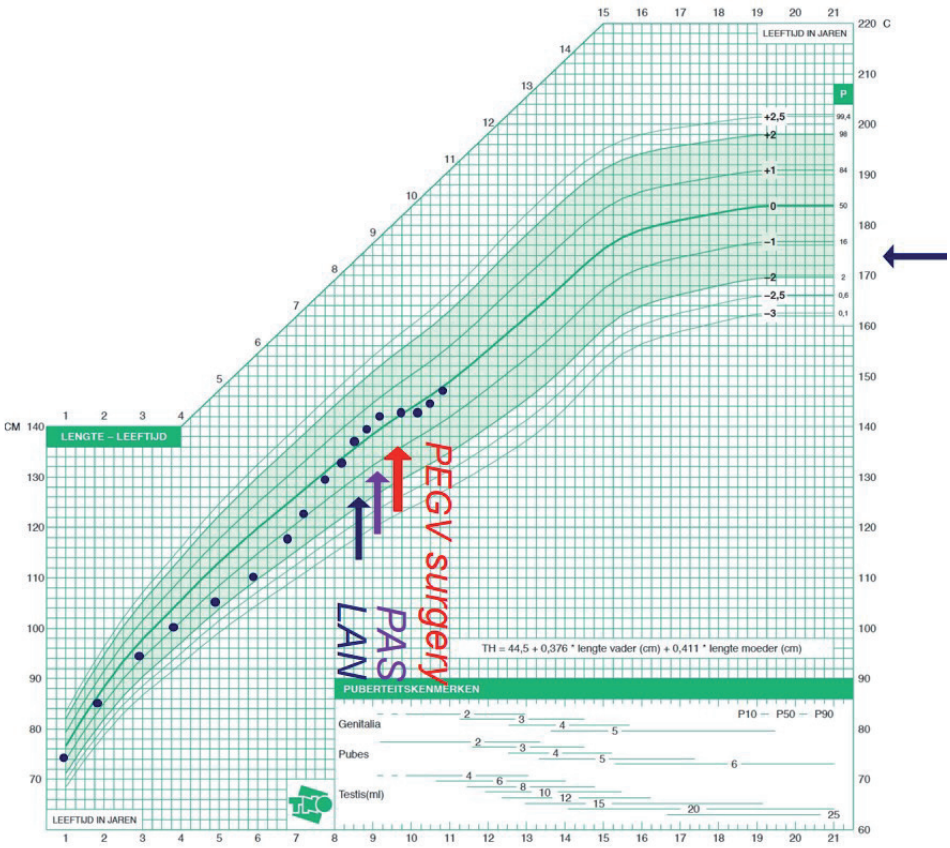
GH and IGF-I levels. Due to the presentation with a macroadenoma at a young age germline genetic testing for sequence variants and deletions in *AIP* and Multiple Endocrine Neoplasia type 1 was performed. A novel heterozygotic truncating variant in the *AIP* gene was discovered (p.Tyr202*) which was accompanied by a second missense variant (p.Pro232 Leu), both paternally inherited. Screening by MRI and hormone evaluation of his 37-year-old father was negative.

Histopathological analysis revealed a loss of SSTR5 expression between the second and third operations (Figure 3). In vitro characterization of the first surgery showed no significant response to coincubation with pasireotide or pasireotide with cabergoline (Figure 4). Other compounds could not be tested due to the limited amount of available tissue.

Discussion

This case report describes a complicated *AIP*-mutation-related somatotropinoma leading to accelerated longitudinal growth which presented at a young age of 7 years.[10] Acro-gigantism can occur with increased growth velocity in young children, even without extreme height compared to age- and sex-matched references. In our patient, the increased growth rate occurred after a period of a relatively decreased longitudinal growth, so that in this case the incipient gigantism was somewhat masked on the growth chart. He is an example of how the totality of growth characteristics is important when assessing children with aberrant growth. Patients with *AIP*-mutation-bearing somatotropinomas require multiple therapeutic modalities applied together than patients with non-*AIP* adenomas.[7] Somatotropinomas are primarily treated with (transsphenoidal) neurosurgery, SSAs, dopamine agonists or GH receptor antagonists.[11] In a large series, patients with *AIP* mutations required more frequently multiple surgeries than patients without *AIP* mutations (22 vs. 6%).[7] Disease control occurred in 27% of 96 *AIP* mutation-positive somatotropinoma patients, versus 81% in non-*AIP* acromegaly patients.[7] Overall, in acromegaly long-acting SSAs can achieve biochemical normalization of GH and IGF-I in 50-60% and often

Figure 1 Growth chart of the patient with incipient gigantism



The initially normal growth of the patient had been declining from three to six years of age, but then deflected markedly upwards. He was diagnosed at the age of 7 years. The blue arrow corresponds with treatment with lanreotide ("LAN"); the purple arrow corresponds with treatment of pasireotide ("PAS") and the red arrow ("PEGV surgery") with pegvisomant and surgery.

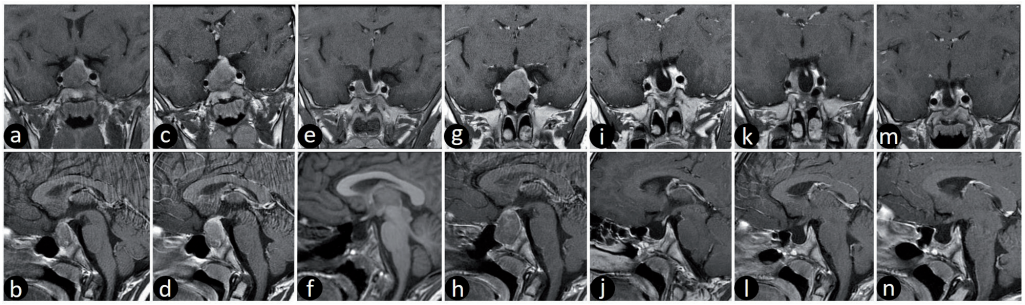
lead to tumor shrinkage.[11-14] Patients with *AIP* mutations had, however, significantly less tumor shrinkage and lower hormonal responses to first-generation SSAs.[7] Patients with *AIP*-mutation-bearing somatotropinomas require more therapeutic modalities than patients with non-mutated adenomas.[7] Somatotropinomas are primarily treated with (transsphenoidal) neurosurgery, SSAs, dopamine agonists or pegvisomant.[11] In a large series, patients with *AIP* mutations required

more frequent multiple surgeries (22 vs. 6%) than patients without *AIP* mutations.[7] Disease control occurred in 27% of 96 *AIP* mutation-positive somatotropinoma patients, versus 81% in non-*AIP* acromegaly patients.[7] Overall, in acromegaly long-acting SSAs can achieve biochemical normalization of GH and IGF-I in 50-60% and often lead to tumor shrinkage.[11-14] Patients with *AIP* mutations had, however, significantly less tumor shrinkage[7] and lower hormonal responses to first-generation SSAs.[7]

SSAs act via somatostatin receptors (SSTRs1-5).[6] The first-generation SSAs octreotide and lanreotide have the highest affinity for SSTR2, and have a low affinity to SSTR3 and SSTR5,[6] while the second-generation SSA pasireotide has the highest affinity for SSTR5, followed by SSTR2, SSTR3 and SSTR1.[15] As reported previously, pasireotide resistance is more related to SSTR2 expression than to SSTR5 in the general acromegalic population.[16,17] SSA resistance may occur if the tumor is lacking SSTR2.[6] Daly et al. recently reported two *AIP*-mutated acromegaly patients with resistance to first-generation SSA, in which pasireotide treatment led to marked tumor shrinkage and persistent hormonal control.[7] In one case, very low to absent SSTR2 levels were seen and the efficacy of pasireotide must have been through other SSTRs like SSTR5.[7]

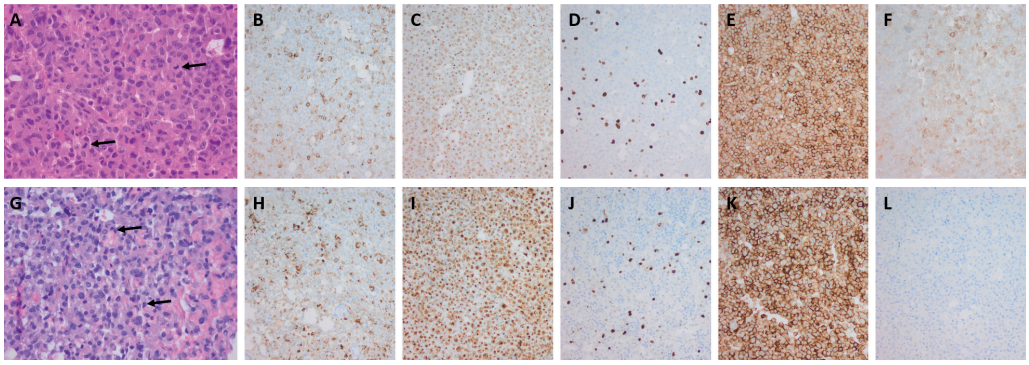
Our case illustrates that in *AIP* mutated somatotropinoma, resistance to first-generation SSAs is not always accompanied by pasireotide responsiveness as was observed previously.[18] The resistance probably relates (partially), to the low SSTR5 expression, since SSTR2 expression remained present. Nevertheless the signaling via SSTR2 may be affected while leaving receptor expression unaffected. Possible factors in this phenomenon include ZAC1 and miR-34a, which both influence SSTR2 signaling.[11,19] In these cases, it may be preferable to test in vitro response of tumor tissue assessed by decreases in GH secretion.[17] Given the lack of tumor size control with first and second generation SSAs and the unresectable remnant, our patient is facing radiotherapy at the age of ten, and will require intensive (endocrinological) follow-up, although no pituitary deficiencies have occurred to date. If needed, excessive GH actions can be controlled by pegvisomant as was shown before, albeit with high vigilance for tumor regrowth.

Figure 2 Sequential magnetic resonance imaging over the clinical course of the patient between 2018 and 2020



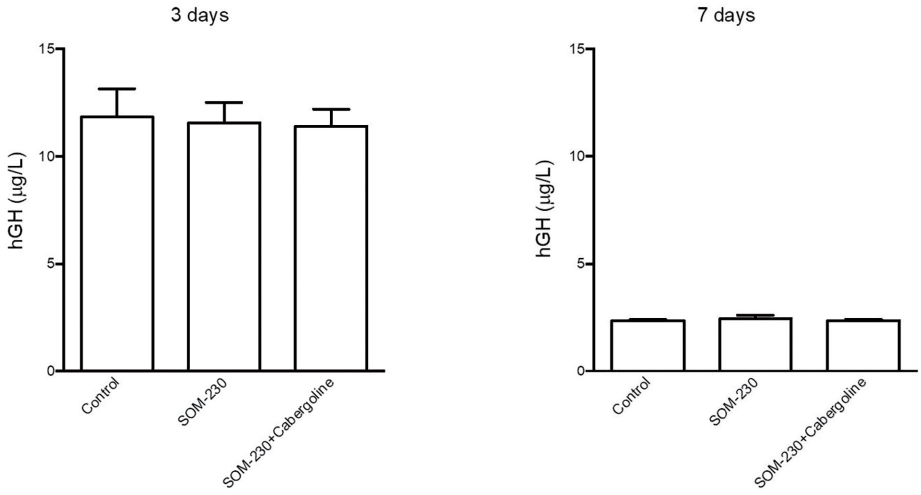
Contrast-enhanced T1-weighted sequences in coronal (**a, c, e, g, l, k, m**) and sagittal (**b, d, f, h, j, l, n**) planes were chosen and corrected for grey scale and magnification. At clinical presentation (**a, b**), before the first operation (**c,d**), delayed postoperatively after the first operation (**e, f**), before the second operation (**g, h**), delayed postoperatively after the second operation (**i, j**), before the third operation (**k, l**), delayed postoperatively after the third operation (**m, n**).

Figure 3 Histopathological features of the tumor at the second and third surgery



A-F: second surgery; **G-L:** third surgery.
A & G: H&E staining show a pituitary adenoma with interspersed mitoses in both surgeries (black arrows).
B & H: GH expression. **C & I:** PanCK immunohistochemistry shows only a few fibrous bodies in both specimens.
In both specimen there is increased proliferation activity (Ki67 staining in **D & J**). Tissue from both surgeries with homogeneous expression of SSTR2 (E & K), while SSTR5 is moderately expressed in the specimen of the first surgery (**F**) and absent in the tissue of the second surgery (**L**).

Figure 4 Experiment to determine *in vitro* sensitivity of tissue to pasireotide and cabergoline



Tumor tissue did not respond to pasireotide (SOM-230) or pasireotide with cabergoline in vitro: there was no significant change in growth hormone production after three days (A) or seven days (B) of exposure.

Conclusion

This unique case of an incipient gigantism in a 7-year old child with a novel *AIP* mutation, p.Tyr202*, was associated with a highly-resistant somatotropinoma. In general, typical features of *AIP* mutation-related PAs were his male sex, young age, macro-adenoma, excessive GH secretion and treatment resistance. This case shows the importance of closely monitoring growth velocity next to height SDS in children. Although previous literature suggests a favorable response to pasireotide in some patients with *AIP* mutations and acromegaly,[18] pasireotide had only limited effect in our patient, possibly related to decreasing SSTR5 expression of the tumor. *In vitro* compound-related GH suppression in cultured tumor tissue may predict *in vivo* treatment response better than assessing SSTR. This patient had tumor size increase despite three neurosurgeries and all available systemic medical therapies. Unfortunately, he would likely need radiotherapy soon before entering puberty, which will have significant impact on the development of his skull and brain. Genetic testing including aberrations of the entire *AIP* gene should be advocated in all patients with GH-secreting pituitary adenomas occurring in childhood and/or in patients with established or incipient pituitary gigantism.[20]

References

1. Daly AF, Beckers A. The Epidemiology of Pituitary Adenomas. Endocrinology and metabolism clinics of North America. 2020;49:347-355.
2. Tichomirowa MA, Barlier A, Daly AF, et al. High prevalence of AIP gene mutations following focused screening in young patients with sporadic pituitary macroadenomas. European journal of endocrinology. 2011;165:509-515.
3. Vierimaa O, Georgitsi M, Lehtonen R, et al. Pituitary adenoma predisposition caused by germline mutations in the AIP gene. Science (New York, NY). 2006;312:1228-1230.
4. Igreja S, Chahal HS, King P, et al. Characterization of aryl hydrocarbon receptor interacting protein (AIP) mutations in familial isolated pituitary adenoma families. Human mutation. 2010;31:950-960.
5. Oriola J, Lucas T, Halperin I, et al. Germline mutations of AIP gene in somatotropinomas resistant to somatostatin analogues. European journal of endocrinology. 2013;168:9-13.
6. Gadelha MR, Kasuki L, Korbonits M. Novel pathway for somatostatin analogs in patients with acromegaly. Trends in endocrinology and metabolism: TEM. 2013;24:238-246.
7. Daly AF, Tichomirowa MA, Petrossians P, et al. Clinical characteristics and therapeutic responses in patients with germ-line AIP mutations and pituitary adenomas: an international collaborative study. The Journal of clinical endocrinology and metabolism. 2010;95:E373-383.
8. Rostomyan L, Daly AF, Petrossians P, et al. Clinical and genetic characterization of pituitary gigantism: an international collaborative study in 208 patients. Endocrine-related cancer. 2015;22:745-757.
9. Joshi K, Daly AF, Beckers A, Zacharin M. Resistant Paediatric Somatotropinomas due to AIP Mutations: Role of Pegvisomant. Hormone research in paediatrics. 2018;90:196-202.
10. Caimari F, Hernández-Ramírez LC, Dang MN, et al. Risk category system to identify pituitary adenoma patients with AIP mutations. Journal of medical genetics. 2018;55:254-260.
11. Theodoropoulou M, Tichomirowa MA, Sievers C, et al. Tumor ZAC1 expression is associated with the response to somatostatin analog therapy in patients with acromegaly. International journal of cancer. 2009;125:2122-2126.

12. Freda PU, Katznelson L, van der Lely AJ, Reyes CM, Zhao S, Rabinowitz D. Long-acting somatostatin analog therapy of acromegaly: a meta-analysis. The Journal of clinical endocrinology and metabolism. 2005;90:4465-4473.

13. van der Lely AJ, Hutson RK, Trainer PJ, et al. Long-term treatment of acromegaly with pegvisomant, a growth hormone receptor antagonist. Lancet (London, England). 2001;358:1754-1759.

14. Bevan JS. Clinical review: The antitumoral effects of somatostatin analog therapy in acromegaly. The Journal of clinical endocrinology and metabolism. 2005;90:1856-1863.

15. Bruns C, Lewis I, Briner U, Meno-Tetang G, Weckbecker G. SOM230: a novel somatostatin peptidomimetic with broad somatotropin release inhibiting factor (SRIF) receptor binding and a unique antisecretory profile. European journal of endocrinology. 2002;146:707-716.

16. Muhammad A, Coopmans EC, Gatto F, et al. Pasireotide Responsiveness in Acromegaly Is Mainly Driven by Somatostatin Receptor Subtype 2 Expression. The Journal of clinical endocrinology and metabolism. 2019;104:915-924.

17. Hofland LJ, van der Hoek J, van Koetsveld PM, et al. The novel somatostatin analog SOM230 is a potent inhibitor of hormone release by growth hormone- and prolactin-secreting pituitary adenomas in vitro. The Journal of clinical endocrinology and metabolism. 2004;89:1577-1585.

18. Daly AF, Rostomyan L, Betea D, et al. AIP-mutated acromegaly resistant to first-generation somatostatin analogs: long-term control with pasireotide LAR in two patients. Endocrine connections. 2019;8:367-377.

19. Bogner EM, Daly AF, Gulde S, et al. miR-34a is upregulated in AIP-mutated somatotropinomas and promotes octreotide resistance. International journal of cancer. 2020;147:3523-3538.

20. Korbonits M, Kumar AV. AIP Familial Isolated Pituitary Adenomas. In: Adam MP, Ardinger HH, Pagon RA, et al., eds. GeneReviews(®). Seattle (WA): University of Washington, Seattle Copyright © 1993-2020, University of Washington, Seattle. GeneReviews is a registered trademark of the University of Washington, Seattle. All rights reserved. 1993.

3

Fractures, Bone Mineral Density, and Final Height in Craniopharyngioma Patients with A Follow-up of 16 Years

Selveta S. van Santen,[1,2,3] Daniel S. Olsson,[4,5]
Marry M. van den Heuvel-Eibrink,[2,3] Mark Wijnen,[1,2]
Casper Hammarstrand,[4,5] Joseph A. M. J. L. Janssen,[1]
Gudmundur Johannsson,[4] Aart J. van der Lely,[1]
Sebastian J. C. M. M. Neggers.[1,2,3]

[1]Department of Medicine, Endocrinology; Erasmus Medical Center,
Rotterdam, The Netherlands;
[2]Department of Paediatric Oncology/Haematology,
Erasmus MC – Sophia Children's Hospital, Rotterdam, The Netherlands;
[3]Princess Máxima Center for Paediatric Oncology,
Utrecht, The Netherlands;
[4]Department of Medicine, Endocrinology;
Sahlgrenska University Hospital, Gothenburg, Sweden;
[5]Department of Internal Medicine and Clinical Nutrition, Institute of Medicine,
Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden.

The Journal of Clinical Endocrinology & Metabolism

Abbreviations: AO, adulthood onset; BMD, bone mineral density; BMI, body mass index; CO, childhood onset; CP, craniopharyngioma patient; DXA, dual-energy X-ray absorptiometry; GHD, growth hormone deficiency; GHRT, GH replacement therapy; OR, odds ratio; SDS, standardized deviation score.

Précis: Male craniopharyngioma patients are at higher risk of fractures than females, while they receive less often medication to increase BMD. A T-score ≤-2.5 or Z-score ≤-2.0 measured using DXA does not predict risk of fracture in patients with CP.

Abstract

Context: Pituitary hormonal deficiencies in patients with craniopharyngioma may impair their bone health.

Objective: To investigate bone health in patients with craniopharyngioma.

Design: Retrospective cross-sectional study.

Setting: Dutch and Swedish referral centers.

Patients: Patients with craniopharyngioma (n=177) with available data on bone health after a median follow-up of 16 years (range 1-62) were included [106 (60%) Dutch, 93 (53%) male, 84 (48%) childhood-onset disease].

Main outcome measures: Fractures, DXA-derived bone mineral density (BMD), and final height were evaluated. Low BMD was defined as T- or Z-score ≤-1, very low BMD as ≤-2.5 or ≤-2.0, respectively.

Results: Fractures occurred in 31 patients (18%) and were more frequent in men than in women (26% vs. 8%, $P = 0.002$). Mean BMD was normal [Z-score total body 0.1 (range -4.1- 3.5)] but T or Z-score ≤-1 occurred in 47 (50%) patients and T-score ≤-2.5 or Z-score ≤-2.0 in 22 (24%) patients. Men received less often treatment for low BMD than women (7% vs. 18%, $P = 0.02$). Female sex (OR 0.3, $P = 0.004$) and surgery (OR 0.2, $P = 0.01$) were both independent protective factors for fractures, while anti-epileptic medication was a risk factor (OR 3.6, $P = 0.03$); whereas T-score ≤-2.5 or Z-score ≤-2.0 was not (OR 2.1, $P = 0.21$). Mean final height was normal and did not differ between men and women, or adulthood and childhood-onset patients.

Conclusions: Men with craniopharyngioma are at higher risk than women for fractures. In patients with craniopharyngioma, a very low BMD (T-score ≤-2.5 or Z-score ≤-2.0) seems not to be a good predictor for fracture risk.

Keywords: craniopharyngioma, bone health, fractures, bone mineral density, final height

Introduction

Craniopharyngioma is a rare tumour in the pituitary region with peak incidences in the age categories 5-9 and 40-44 years.[1] Age of onset correlates highly with tumour histology: the adamantinomatous subtype occurs mainly in childhood, and in adulthood, while the papillary subtype almost exclusively occurs at adult age.[2] The pathogenesis of the two subtypes differs as well, as adamantinomatous craniopharyngioma is driven by somatic mutations in *CTNNB1*, while the papillary subtypes harbour *BRAF*^{V600E} mutations.[2] Craniopharyngioma patients (CP) require extensive long-term follow-up, since the survival rate is relatively high and the patients often suffer from multiple co-morbidities.[1,3] Several comorbidities could give CP an increased risk of fractures, osteoporosis and reduced growth: CP are at risk of pituitary insufficiency and/or possibly non-physiological replacement therapy,[3-9] visual deficiency,[10] late puberty induction, hypothalamic damage and limited physical activity.[11-14] In addition, chronic neurological disease is associated with clumsiness and high incidence of falls.[15,16] Moreover, anti-epileptic therapy may even contribute more to the lifetime fracture risk than the seizures themselves.[17] All these factors may influence bone health and could result in osteoporosis. Osteoporosis is a systemic disease characterized by three elements: loss of bone mass, deterioration of the microarchitecture of the bone, and an increased fracture risk.[18] Fractures may cause chronic pain, disability and need for rehabilitation.[18]

Despite the knowledge of these frequent occurring comorbidities, studies addressing fracture risk and osteoporosis or bone mineral density (BMD) in CP are scarce. Previously found frequencies of fractures were higher compared to matched populations and lie between 15 to 25%.[19-21] Olsson et al. described an increased risk of fractures with a standardized incidence rate (SIR) of 2.1 in a large cohort of Swedish CP.[1] No studies have yet extensively investigated risk factors for fractures in CP.

In foregoing research regarding BMD, very few adulthood onset (AO) patients were included: osteoporotic/osteopenic BMD values were found in 6 out of 10, and 5 out of 6 cases, respectively.[22,23] The

studies regarding childhood onset (CO) CP are slightly larger but led to different conclusions regarding the influence of gender on the risk of fracture;[11,24] Müller et al. found that men were at risk for lower volumetric BMD (n=61),[24] while Holmer et al. found a low BMD in women (n=39).[11]

Only a few studies have addressed final height, mainly in CO CP. In these studies, CO CP reach slightly below average or normal final height.[19,25-33]

In summary, information on bone health in CP is scarce and contradictory. Also, factors predicting increased risk of fractures in patients with CP have not yet been studied. In order to improve long-term care and increase knowledge of possible risk factors of impaired bone health in CP, we investigated bone fractures, BMD, and final height in a large CP cohort.

Design and methods

In this retrospective cross-sectional study, CO and AO CP treated at the Erasmus University Medical Center (Rotterdam, the Netherlands)/Sophia Children's Hospital (Rotterdam, the Netherlands) and Sahlgrenska University Hospital (Gothenburg, Sweden) were included. Data were collected as previously described.[10,34] Subjects were included if data were available on fractures, Dual-Energy X-ray Absorptiometry (DXA)-scan results for BMD, or on final height.

Data on fractures were acquired as described before;[10] data on fractures were included from 1987 until April 2019. Data regarding fractures from the Dutch patients concerns both inpatient as well as outpatient data; for the Swedish patients, data until 2014 concerns only inpatient data. If only the year of fracture was known, the first of January was used as date to calculate age at fracture. Medication to improve BMD was defined as current or past use of bisphosphonates, vitamin D or calcium. Gonadal axis replacement therapy or other hormonal replacement therapy was not included since the indication of treating

pituitary deficiencies was broader than only improving BMD and not solely for preservation of bone health. The definition of hypothalamic damage was described before: it concerns tumour- and/or treatment-related injury to the hypothalamus and/or third ventricle as visualized by neuroimaging.[34] The research proposal was accepted by the Ethical Review Board of the Erasmus MC and the Regional Ethical Review Board of the Gothenborg University; all patients gave informed consent.

All types of DXA-scanners were accepted in the analysis and include Lunar DPXL, Lunar iDXA and Lunar Prodigy. A list of accepted DXA-scanners is shown in Table 1.[35] For the current study, low BMD was defined as T- or Z-score below -1. Osteoporosis was defined as T-score below -2.5 or Z-score below -2. Osteopenia was defined as T- or Z-score between -1 and -2.5 or -2, respectively. If it is not specified in the text whether low BMD, osteopenia or osteoporosis are based on T- or Z-score, or on which site, it is found to exceed the limit of either T- or Z-score, and is measured at the location of femur neck, L2-L4, and/or total body.[36] If multiple DXA-scans were performed, the most recent was chosen. The decision to perform a DXA-scan was made clinically by the physician who was treating the patient.

Final height was defined as the highest measured value, after the age of 18 years. Standardized deviation scores (SDS) or final height for Dutch subjects were calculated based on requested data with references from the Dutch National Statistics ("Centraal Bureau voor Statistiek")[37] based on age, sex and year of measurement (available on request). If final height was measured before 1981 or after 2017, references of 1981 or 2017 respectively were used in Dutch subjects. Final height SDS of Swedish subjects was calculated with references of the same sex compared with young adults with birth year matched as closely as possible.[38-40]

Statistical analysis

Statistical analysis was performed with IBM SPSS Statistics for Windows, Version 24.0 and Graphpad Version 8.01. Data are presented as mean and standard deviation (SD) unless stated otherwise. Significance was accepted if $P \leq 0.05$. For normally distributed data, an unpaired T-test was used for group comparisons, while a Mann-Whitney U was used for nonnormally distributed data. Comparisons of proportions were conducted using either the $\chi 2$ test or the Fisher's exact test as indicated; in case of comparison of two related dichotomous variables, McNemar's test was applied. To identify influencing factors of low BMD at last DXA-scan and fractures, a univariable logistic regression model was primarily performed; only significant predictors ($P < 0.05$) were entered in the multiple logistic regression analysis thereafter. If the number of events allowed it, the multiple logistic regression model was refined by adding variables with *P*-values < 0.20. If more variables were significant in univariable analysis than could be entered in multivariable model due to the size of the cohort, the most significant variables were selected in multivariable analysis. If the options were comparable, the variable contributing to the best discrimination and highest Nagelkerke's R^2 was chosen. The dependent variables were studied with forced entry. For fractures, a Cox Regression model was established with the apparent main predictors from the logistic regression model. The assumption of proportional hazards was verified by a goodness of fit test with Shoenfeld residuals and time and checking significance of time-dependent covariates. The Wilcoxon rank sum test was used to compare nonnormally distributed variables and the Student's t-test was used for normally distributed variables. A two-way ANOVA was applied for comparisons of multiple groups. The incidence rate was calculated as total incidence of fractures divided by the cumulative follow-up years and multiplied by 1000 to calculate the fracture rate per 1000 person-years.

Table 1 Included Dual-Energy X-ray Absorptiometry scanners that were used in this craniopharyngioma cohort

	First DXA-scan (n=117)	Last DXA-scan (n=95)
Unknown/other	10 (9)	8 (8)
Idexa Gothenborg University Hospital	0 (0)	19 (20)
DPXL total	61 (52)	14 (15)
DPXL Erasmus MC	35 (30)	12 (13)
DPXL Gothenborg University Hospital	26 (22)	2 (2)
Prodigy total	46 (39)	54 (57)
Prodigy (1) Erasmus MC	6 (5)	9 (10)
Prodigy (2) Erasmus MC	6 (5)	6 (6)
Prodigy Gothenborg University Hospital	34 (29)	39 (41)

DXA-scanners included in the article. Data is given as N (%). Prodigy (1) is used at the dept. of Nuclear Medicine in the Erasmus MC; Prodigy (2) at the dept. of Internal Medicine. Total includes all scanners of the same type in both centers.

Results

Baseline characteristics

Baseline characteristics of the 177 CP are depicted in Table 2 and Table 3.[41] Data were available on DXA-scan results for 117 patients and on final height for 171 patients. Patients with a DXA-scan available had a higher percentage of diabetes insipidus (68% vs. 51%, $P = 0.03$), of growth hormone deficiency (GHD) (93% vs. 72%, $P < 0.001$) and of TSH deficiency (95% vs. 86%, $P = 0.06$) than patients with no DXA-scan, and less often epilepsy (13% vs. 27%, $P = 0.03$) or a hydrocephalus (23% vs. 40%, $P = 0.03$). There was no difference in fractures (18% vs.

18%, $P = 0.95$). The cohort consisted of 93 AO CP (53%). There were 106 Dutch (60%) and 71 Swedish (40%) patients included; 93 patients were men (53%) and 15 patients (8%) used medication for epilepsy. The median age at last follow-up was 45 years (range 15-92). Median follow-up time was 16 years (range 1-62). Mean body mass index (BMI) was 31 ± 7 kg/m^2 (range 17 – 60) in all patients at last DXA-scan (or, if DXA-scans at adult age were unavailable, last available BMI) and was higher in women than in men (30 ± 6 vs. 33 ± 8 kg/m^2, $P = 0.02$). CO CP had more often received radiotherapy as tumour treatment (66% vs. 38%, $P < 0.001$), had a higher occurrence of hydrocephalus (38% vs. 17%, $P = 0.002$) and epilepsy (26% vs. 12%, $P = 0.01$) than AO CP. In addition, CO CP had more often a GHD (92% vs. 79%, $P = 0.02$) and had higher occurrence of diabetes insipidus (77% vs. 52%, $P < 0.001$). Sex hormone replacement therapy was used in (125 patients) 94% of women aged 50 years or younger and men with gonadal axis failure. In two patients, presence of GHD was unknown. In the GHD group, CO CP were more often using growth hormone replacement therapy (GHRT) than AO CP (87% vs. 74%, $P = 0.04$). Five patients (3%) underwent bariatric surgery (1 gastric sleeve, 4 gastric bypass); all of them were female. Of the 10 patients who were not treated with surgery, five had Yttrium, one was initially treated with RT, one had cyst aspiration, and the others were closely followed with imaging. The non-surgery group had significant lower frequency of DI (10% vs. 67% in the entire cohort, $P < 0.001$), TSH deficiency (50% vs. 95%, $P < 0.001$) and trend towards lower visual deficiencies (50% vs. 78%, $P = 0.09$). Furthermore, BMI was not significantly different ($P = 0.34$). The tumour was described as stable during follow-up of these patients.

Fractures

Fractures occurred in 31 CP (18%). The fracture rate was 5.8 fractures per 1000 person years. Mean age at last follow-up of patients who had fractures was 52.0 ± 14.5 vs. 45.5 ± 18.5 years in patients who had no fractures ($P = 0.05$). Mean follow-up time was 16.8 ± 10.0 years of patients who had fractures vs. 18.6 ± 13.7 of patients who had no fractures ($P = 0.60$, n=135). Seven of the patients with fractures had

osteopenia (70%); four had osteoporosis (40%) (at last DXA-scan). Eight patients suffered from multiple fractures (26%). The mean number of fractures per patient were 1.5 ± 1.1 (range 1-6). The fracture types are described in Table 4.

Fractures occurred more often in men than women (26% vs. 8%, $P = 0.002$) and men were less often treated with medication to improve BMD (7% vs. 18%, $P = 0.02$). A trend was seen towards a higher proportion of fractures in AO CP than in CO CP (23% vs. 12%, $P = 0.06$). There was no difference in percentage of patients with fractures between Dutch and Swedish patients (18% vs. 17%, $P = 0.86$).

Table 2 Baseline characteristics of the included CP survivors

		All survivors (n=177)	Males (n=93)	Females (n=84)	*P*-value
Age at last follow-up (years)[a]		45 (15-92)	45 (16-92)	45 (15-82)	0.83
Age at presentation (years)[a]		23 (0-79)	26 (0-79)	22 (4-73)	0.92
(Female/corresponding) gender (n [%])		84 (48%)	93 (53%)	84 (47%)	0.50
Childhood onset disease (n [%])		84 (48%)	44 (46%)	40 (48%)	0.97
Tumor location at presentation (n [%])	Intrasellar	6 (4%)	5 (5%)	1 (1%)	0.22
	Suprasellar	69 (41%)	35 (52%)	34 (44%)	0.57
	Intra-/ suprasellar	95 (56%)	52 (57%)	43 (55%)	0.86
Craniopharyngioma treatment	Surgery only	77 (44%)	42 (46%)	35 (42%)	0.55
	Radiation only	2 (1%)	1 (1%)	1 (1%)	1.00
	Surgery and radiation	88 (50%)	44 (48%)	44 (52%)	0.59
	^{90}Yttrium brachytherapy	23 (13%)	8 (9%)	15 (18%)	0.09
Pituitary deficiencies	GH deficiency[b]	149 (85%)/ 120 (81%)	78 (86%)	71 (85%)	0.83

	TSH deficiency[b]	162 (92%)/ 161 (99%)	88 (95%)	74 (88%)	0.12
	Hypogonadism[b]	155 (88%)/ 125 (94%)	84 (90%)	71 (85%)	0.24
	Corticotrophic deficiency[b]	146 (83%)/ 142(99%)	81 (87%)	65 (77%)	0.09
	ADH deficiency[b]	113 (64%)/ 111 (98%)	61 (66%)	52 (61%)	0.61
Medical history	Recurrence/ progression	67 (39%)	33 (36%)	34 (41%)	0.53
	Hypothalamic damage	63 (40%)	35 (42%)	28 (38%)	0.67
	Hydrocephalus ever	47 (27%)	22 (24%)	25 (30%)	0.38
	Visual impairment	125 (77%)	67 (41%)	58 (36%)	0.50
	Epilepsy	33 (19%)	18 (19%)	15 (18%)	0.80
	Diabetes mellitus	26 (15%)	11 (12%)	15 (18%)	0.26
	BMD medication	21 (12%)	6 (7%)	15 (18%)	**0.02**

In 7 patients, tumor location at presentation was unknown. Bold values represent significant differences between males and females ($P \leq 0.05$). [a]Median (range). [b]All/using replacement therapy. Abbreviations: ADH, antidiuretic hormone; BMD medication, bone mineral density-increasing medication.

Table 3 Baseline characteristics of the included CP survivors: adulthood onset vs. childhood onset

	Childhood onset CP (n=84)	**Adulthood onset CP (n=93)**	***P*-value**
Age at last follow-up (years)[a]	32 (15-74)	57 (25-92)	**<0.001**
Age at presentation (years)[a]	10 (0-17)	41 (18-79)	**<0.001**
Female gender (n [%])	40 (48%)	44 (47%)	0.97
Tumor location at presentation (n [%])			
Intrasellar	4 (5%)	2 (2%)	0.42
Suprasellar	29 (36%)	40 (44%)	0.28
Intra-/suprasellar	47 (59%)	48 (53%)	0.48
Craniopharyngioma treatment			
Surgery	79 (96%)	86 (93%)	0.34
Radiation	55 (66%)	35 (38%)	**<0.001**
^{90}Yttrium brachytherapy	12 (15%)	10 (11%)	0.42
Pituitary hormonal deficiencies			
GH deficiency	77 (92%)	72 (79%)	**0.02**
TSH deficiency	78 (92%)	84 (90%)	0.55
Hypogonadotrophic hypogonadism	71 (85%)	84 (90%)	0.24
Corticotrophic deficiency	68 (81%)	78 (84%)	0.61
ADH deficiency	65 (77%)	48 (52%)	**<0.001**

Medical history			
Recurrence/progression	34 (42%)	33 (36%)	0.45
Hypothalamic damage	33 (45%)	30 (36%)	0.23
Hydrocephalus ever	31 (38%)	16 (17%)	**0.002**
Visual impairment	60 (78%)	65 (76%)	0.72
Epilepsy*	22 (26%)	11 (12%)	**0.01**
Diabetes mellitus	9 (11%)	17 (18%)	0.16
BMD medication	12 (15%)	9 (10%)	0.31
Myocardial infarction	1 (1%)	10 (11%)	**0.01**

[a]Median (range). ADH = antidiuretic hormone; BMD medication = bone mineral density increasing medication; CP = craniopharyngioma; GH = growth hormone; N = number; TSH = thyroid stimulating hormone. *Anti-epileptic medication was comparable (8 (10%) vs. 7 (8%), *P* = 0.60).

Using a univariable logistic regression model investigating fractures, female sex (OR 0.3, *P* = 0.004) and previous surgery (OR 0.2, *P* = 0.01) were protective factors, while medication for epilepsy was identified as a risk factor (OR 3.6, *P* = 0.03) (Table 5). Osteoporosis was not a significant independent factor (OR 2.1, *P* = 0.21); for osteopenia, a trend was shown (OR 2.6, *P* = 0.09). There were 10 patients who never underwent surgery, of which 5 had a fracture; 26 out of 165 patients who did have surgery had a fracture. A trend was shown for radiotherapy as well, being protective to fractures (OR 0.5, *P* = 0.06). When these independent variables were applied in the multivariable Cox regression model, the following hazard ratio's were obtained: for sex 0.4 (95% CI 0.2-0.8, *P* = 0.02), for surgery 0.3 (0.1-0.8, *P* = 0.02) and for anti-epileptic medication use 2.7 (1.1-6.7, *P* = 0.03) (-2LL Chi^2 <0.001, Omnibus Chi^2 0.001). Fracture free survival curves are depicted in Figure 1.

Low BMD values based on either T- or Z-scores was found in 11 patients (65%) with fractures as opposed to 32 patients (42%) without fractures in

their history (P = 0.08) (Table 6).[41] Patients who had never been treated with surgery (n=10) suffered from more fractures than patients who had been operated (50% vs. 16%, P = 0.006). A trend for more fractures was seen in patients who had never received radiotherapy compared to patients treated with radiotherapy (23% vs. 12%, P = 0.06).

Table 4 Frequencies and site of fractures

Location	Male (all)	Male (Dutch/ Swedish)	Female (all)	Female (Dutch/ Swedish)	All subjects	Dutch/ Swedish
Unknown*	**2 (5%)**	**1 (5%) / 1 (6%)**	**0 (0%)**	**0 (0%) / 0 (0%)**	**2 (4%)**	**1 (4%) / 1 (5%)**
Femur	2 (5%)	1 (5%) / 1 (6%)	0 (0%)	0 (0%) / 0 (0%)	2 (4%)	1 (4%) / 1 (5%)
Radius	1 (3%)	0 (0%) / 1 (6%)	1 (11%)	0 (0%) / 1 (33%)	2 (4%)	0 (0%) / 2 (10%)
Tibia	2 (5%)	1 (5%) / 1 (6%)	1 (11%)	0 (0%) / 1 (33%)	3 (7%)	1 (4%) / 1 (5%)
Fibula	2 (5%)	0 (0%) / 2 (12%)	1 (11%)	1 (17%) / 0 (0%)	3 (7%)	1 (%) / 2 (10%)
Lower leg (unspecified)	2 (5%)	2 (10%) / 0 (0%)	0 (0%)	0 (0%) / 0 (0%)	2 (4%)	2 (8%) / 0 (0%)
Humerus	1 (3%)	0 (0%) / 1 (6%)	2 (22%)	1 (17%) / 1 (33%)	3 (7%)	1 (4%) / 2 (10%)
Ulna	1 (3%)	1 (5%) / 0 (0%)	1 (11%)	1 (17%) / 0 (0%)	2 (4%)	2 (8%) / 0 (0%)
Clavicle	2 (5%)	1 (5%) / 1 (6%)	0 (0%)	0 (0%) / 0 (0%)	2 (4%)	1 (4%) / 1 (5%)
Cervical vertebrae	3 (8%)	0 (0%) / 3 (18%)	0 (0%)	0 (0%) / 0 (0%)	3 (7%)	0 (0%) / 3 (15%)
Lumbar vertebrae	1 (3%)	1 (5%) / 0 (0%)	1 (11%)	1 (17%) / 0 (0%)	2 (4%)	2 (8%) / 0 (0%)
Long pipebones/ vertebrae	**17 (46%)**	**7 (35%) / 10 (59%)**	**7 (78%)**	**4 (67%) / 3 (100%)**	**24 (52%)**	**11 (42%) / 13 (65%)**
Rib	3 (8%)	2 (10%) / 1 (6%)	0 (0%)	0 (0%) / 0 (0%)	3 (7%)	2 (8%) / 1 (5%)
Pelvis	1 (3%)	0 (0%) / 1 (6%)	0 (0%)	0 (0%) / 0 (0%)	1 (2%)	0 (0%) / 1 (5%)

Facial bones/ nose	4 (11%)	3 (15%) / 1 (6%)	0 (0%)	0 (0%) / 0 (0%)	4 (9%)	3 (12%) / 1 (5%)
Hand	5 (14%)	4 (20%) / 1 (6%)	1 (11%)	1 (17%) / 0 (0%)	6 (13%)	5 (19%) / 1 (5%)
Foot (calcaneus/ toe/ unspecified)	5 (14%)	3 (15%) / 2 (12%)	1 (11%)	1 (17%) / 0 (0%)	6 (13%)	4 (15%) / 2 (10%)
Other	**18 (49%)**	**12 (60%) / 6 (%)**	**2 (22%)**	**2 (33%) / 0 (0%)**	**20 (43%)**	**14 (52%) / 6 (30%)**
Total	**37 (100%)**	**20 (54%) / 17 (46%)**	**9 (100%)**	**6 (67%) / 3 (33%)**	**46 (100%)**	**26 (100%) / 20 (100%)**

Fracture locations of all fractures and its frequencies. Data is presented as n (% of all fractures in this category); 80% of all fractures occurred in males and 57% in the Dutch cohort. Percentages of categories unknown, long pipebones/vertebrae and other fractures may not add up to 100% because of rounding issues. *Data was lost during transition of electronic patient files.

Bone mineral density

Information on BMD is shown in Table 7. Mean age at last DXA-scan is 44.8 ± 18.4 years. Overall mean BMD T- and Z-scores at last DXA-scan were in the normal range: for example, for total body, femur neck and L2-L4 the Z-scores were 0.1 ± 1.5 (range -4.1 – 3.5), -0.1 ± 1.3 (range -2.7 – 4.7) and 0.0 ± 2.0 (range -3.5 – 6.8), respectively (Table 7). However, low BMD in any of these sites was reported in 47 patients (50%) (while an SDS of -1 should correspond to a percentage of 16%). Scatter plots of BMD T- and Z-scores are shown in Figure 2, showing a wide spread of SDS values, also in patients with fractures. Osteoporosis occurred in 22 patients (24%) (Table 7). AO CP had a lower BMD of the femur (neck) than CO CP (0.92 ± 0.15 vs. 1.09 ± 0.28, $P = 0.01$) (Table 7) and a lower mean T-score of the femur (neck) (-0.8 ± 1.2 vs. 0.2 ± 2.1, $P = 0.03$), but comparable Z-scores. A trend was found for lower Z-score of the femur neck in males than in females (-0.4 ± 1.2 vs. 0.2 ± 1.4, $P = 0.08$). Of the patients with osteoporosis, 19% has hypothalamic damage, as opposed to 36% of patients without osteoporosis ($P = 0.15$).

First, a univariable analysis for very low BMD/osteoporosis was established: (borderline) significant contributors were age at DXA-scan (OR 1.032 (1.004-1.063), *P* = 0.03), obstructive sleep apnoea syndrome (OR 2.9 (1.0-8.3), *P* = 0.05), and hydrocortisone dose (OR 1.1 (1.0-1.2), *P* = 0.06. After establishing a multivariable analysis, age (OR 1.03 (95% CI 1.00-1.06, *P* = 0.03) was identified as a contributing independent prognostic factor (Table 8).[41] Hypothalamic damage did not contribute significantly to osteoporosis, nor did it contribute in a multivariable model for osteopenia when adjusted for age, medication to improve BMD and GHRT (Table 8: OR 1.6, *P* = 0.49, model not significant).

Final height

CP had a final height SDS within the normal range (Table 7). There were no differences in final height SDS between males and females (-0.3 ± 1.2 vs. -0.3 ± 1.2, *P* = 0.82) or between CO and AO CP (-0.3 ± 1.4 vs. -0.3 ± 1.0, *P* = 0.90).

Table 5 Univariable and multivariable logistic regression analysis for determinants of fractures in CP survivors

	Univariable analysis		Multivariable analysis (Nagelkerke R^2 =0.18)	
Variables	OR (95% CI)	*P*	OR (95% CI)	*P*
Age at presentation	1.0 (0.99-1.0)	0.17		
Female sex	**0.3 (0.1-0.6)**	**0.004**	**0.3 (0.1-0.7)**	**0.004**
Adulthood onset disease	0.2 (1.0-4.9)	0.07		
Swedish cohort	0.9 (0.4-2.1)	0.86		
Surgery	**0.2 (0.1-0.7)**	**0.01**	**0.1 (0.0-0.6)**	**0.009**
Radiotherapy	0.5 (0.2-1.0)	0.06		
Hypothalamic damage	0.9 (0.4-2.0)	0.79		
Visual impairment	1.9 (0.6-5.9)	0.26		
Medication for epilepsy	**3.6 (1.2-11.0)**	**0.03**	3.0 (0.9-10.0)	0.07
Growth hormone deficiency	2.9 (0.6-13.0)	0.16		
GHRT	1.7 (0.7-4.3)	0.25		
Adrenal axis deficiency	0.9 (0.3-2.3)	0.77		
Hydrocortisone dose	1.0 (0.9-1.1)	0.95		
Gonadal axis deficiency*	2.3 (0.5-10.4)	0.28		
TSH deficiency	1.4 (0.3-6.6)	0.66		
Diabetes insipidus	1.2 (0.5-2.8)	0.62		
BMI	0.9 (0.9-1.0)	0.07		
Osteopenia	2.6 (0.9-7.7)	0.09		
Osteoporosis	2.1 (0.7-6.4)	0.21		
BMD increasing medication	2.1 (0.7-5.8)	0.17		

Age at last follow-up not included (violation of assumption of linearity of logit). Bold values represent significant variables ($P \leq 0.05$). *Replacement therapy OR 1.6 (95%CI 0.7-4.0, $P = 0.28$) / after exclusion of postmenopausal females OR 1.3 (95%CI 0.4-4.2, $P = 0.62$). Hosmer Lemeshow $P = 0.81$, Omnibus $Chi^2 < 0.001$. No subjects were excluded. Receiver operating characteristic area under the curve 0.72. Abbreviations: BMD, bone mineral density; BMI, body mass index; GHRT, GH replacement therapy.

Table 6 Characteristics of survivors with and without fractures

		Fracture in history (n=31)	No fracture in history (n=146)	*P*-value
Baseline characteristics				
Age (years) at last FU		52.0 (14.5)	45.5 (18.5)	0.05
Female sex		7 (23%)	77 (53%)	**0.002**
Childhood onset disease		10 (12%)	74 (51%)	0.06
Tumor treatment				
Surgery		26 (84%)	139 (97%)	**0.02**
Radiotherapy		11 (36%)	79 (54%)	0.06
Yttrium brachytherapy		3 (10%)	19 (14%)	0.77
Comorbidities				
Recurrence or progression		10 (33%)	57 (40%)	0.52
Visual impairment		23 (85%)	102 (75%)	0.25
Epilepsy		9 (29%)	24 (16%)	0.10
Hydrocephalus		6 (19%)	41 (29%)	0.29
Hypothalamic damage		11 (38%)	52 (41%)	0.79
Bariatric surgery		0 (0%)	5 (3%)	0.29
Diabetes mellitus		4 (13%)	22 (15%	1.00
Growth hormone deficiency		29 (94%)	120 (83%)	0.18
TSH deficiency		29 (94%)	133 (91%)	1.00
LH/FSH deficiency		29 (94%)	126 (86%)	0.38
ACTH deficiency		25 (81%)	121 (83%)	0.77
Diabetes insipidus		21 (68%)	92 (63%)	0.62
Medication				
Hydrocortisone dose		21.0 (4.9)	21.1 (6.4)	0.57
Ever BMD increasing medication		8 (26%)	25 (17%)	0.28
Gonadal axis replacement		24 (77%)	99 (68%)	0.29
Anti-epileptic treatment		6 (19%)	9 (6%)	**0.03**

DXA-scan results			
Age (years)	48.0 (15.6)	44.1 (19.0)	0.39
Height (cm)	171.8 (11.7)	171.6 (19.0)	0.60
Weight (kg)	93.6 (21.2)	91.9 (19.5)	0.80
BMI (kg/m^2)	31.5 (5.2)	31.3 (5.8)	0.65
Total body T-score	0.30 (1.5)	0.12 (1.7)	0.72
Total body Z-score	-0.2 (1.4)	0.1 (1.5)	0.46
Femur T-score	-0.2 (1.4)	-0.1 (1.6)	0.81
Femur Z-score	-0.3 (1.4)	-0.2 (1.4)	0.70
Femur neck T-score	-0.7 (1.1)	-0.3 (1.7)	0.85
Femur neck Z-score	-0.4 (1.0)	0.0 (1.4)	0.53
Lumbar spine (L2-L4) T-score	0.1 (2.5)	0.1 (1.9)	0.68
Lumbar spine (L2-L4) Z-score	0.0 (2.6)	0.0 (1.8)	0.93
Bone health			
Low BMD (T-score)	9 (53%)	30 (42%)	0.40
Low BMD (Z-score)	9 (53%)	32 (42%)	0.42
Low BMD (T/Z-score)	11 (65%)	37 (48%)	0.21
Osteopenia (T-score)	6 (35%)	22 (31%)	0.73
Osteopenia (Z-score)	9 (53%)	26 (34%)	0.15
Osteopenia (T/Z-score)	11 (65%)	32 (42%)	0.08
Osteoporosis (T-score)	5 (29%)	13 (19%)	0.32
Osteoporosis (Z-score)	3 (18%)	12 (16%)	0.85
Osteoporosis (T/Z-score)	6 (35%)	16 (21%)	0.20
Final height	173.1 (11.0)	172.1 (9.9)	0.41
Final height SDS	-0.6 (1.3)	-0.3 (1.1)	0.13

Abbreviations: BMD = Bone Mineral Density; BMI = Body Mass Index; DXA = Dual-Energy X-ray Absorptiometry; FU = follow-up; SDS = standardized deviations scores. Data are presented as mean (SD) or n (%). BMI is measured at last DXA-scan or at last follow-up. BMD results are of the last DXA-scan. Bone health corresponds with grouped variables of low BMD/osteopenia/osteoporosis at measured at femur neck, L2-L4 or total body. N in the first row represents the total number of patients of which data is available on this topic. Not all data is available in all cases. Pituitary deficiencies are evaluated at last follow-up.

Table 7 Bone mineral density and final height of the included CP survivors

		All		Male		Female		
		Mean (SD)	n	Mean (SD)	n	Mean (SD)	n	*P*-value
Age at last DXA[a]		44.8 (18.4)	95	47.1 (19.2)	47	42.6 (17.4)	48	0.26
BMI at last DXA		31.3 (5.3)	84	30.9 (4.8)	43	31.8 (5.8)	41	0.41
BMD	Total body	1.20 (0.16)	78	1.25 (0.15)	43	1.14 (0.15)	35	**0.002**
	Femur	1.04 (0.22)	65	1.00 (0.20)	32	1.07 (0.23)	33	0.26
	Femur neck	0.99 (0.22)	34	0.95 (0.20)	31	1.03 (0.24)	33	0.12
	L2-L4	1.24 (0.25)	76	1.32 (0.27)	37	1.17 (0.21)	39	**0.01**
T-score	Total body	0.5 (1.7)	79	0.5 (1.8)	44	0.5 (1.6)	35	0.85
	Femur	-0.1 (1.5)	63	-0.4 (1.5)	30	0.1 (1.6)	33	0.22
	Femur neck	-0.4 (1.6)	64	-0.8 (1.5)	31	-0.1 (1.6)	33	0.09
	L2-L4	0.1 (2.0)	73	0.5 (2.1)	36	-0.2 (1.8)	37	0.10
Z-score	Total body	0.1 (1.5)	84	0.0 (1.6)	45	0.2 (1.3)	39	0.57
	Femur	0.0 (1.4)	65	-0.3 (1.3)	32	0.2 (1.4)	33	0.17
	Femur neck	-0.1 (1.3)	64	-0.4 (1.2)	31	0.2 (1.4)	33	0.08
	L2-L4	0.0 (2.0)	75	0.3 (2.2)	36	-0.3 (1.7)	39	0.18
Final height		172.3 (10.1)	171	178.1 (8.0)	91	165.6 (8.0)	80	**<0.001**
Final height SDS		-0.3 (1.2)	171	-0.3 (1.2)	91	-0.3 (1.2)	80	0.82
Low BMD		47 (50%)	94	25 (53%)	47	22 (47%)	47	0.54
Osteoporosis		22 (23%)	94	13 (28%)	47	9 (19%)	47	0.33
Osteopenia		43 (46%)	94	21 (45%)	47	22 (47%)	47	0.84

Age is given in years. BMD (T- and Z-scores) are given of the first DXA scan. For the raw BMD and BMI values, children were excluded. The minimum age at first DXA scan is 6 years. Data are given as mean (SD); proportions are given as n (%). Low BMD is a T- or Z-score below -1.Osteoporosis is a T-score below -2.5 or a Z-score below -2. Osteopenia is a T-score

		CO CP		AO CP		
		Mean (SD)	n	Mean (SD)	n	*P*-value
Age at last DXA[a]		32.9 (14.3)	45	55.6 (14.7)	50	**<0.001**
BMI at last DXA		31.1 (5.1)	37	31.6 (5.5)	47	0.69
BMD	Total body	1.19 (0.17)	37	1.21 (0.15)	47	0.72
	Femur	1.11 (0.26)	27	0.98 (0.16)	38	**0.03**
	Femur neck	1.09 (0.28)	25	0.92 (0.15)	39	**0.01**
	L2-L4	1.22 (0.21)	34	1.26 (0.28)	42	0.53
T-score	Total body	0.4 (1.7)	37	0.5 (1.7)	42	0.87
	Femur	0.4 (1.9)	24	-0.4 (1.2)	39	0.05
	Femur neck	0.2 (2.1)	24	-0.8 (1.2)	40	**0.03**
	L2-L4	-0.1 (1.6)	31	0.3 (2.3)	42	0.46
Z-score	Total body	-0.1 (1.6)	41	0.2 (1.4)	43	0.39
	Femur	0.2 (1.7)	26	-0.2 (1.1)	39	0.29
	Femur neck	0.2 (1.8)	24	-0.3 (1.0)	40	0.20
	L2-L4	-0.4 (1.6)	33	0.3 (2.2)	42	0.10
Final height		172.9 (11.2)	80	171.7 (9.0)	91	0.41
Final height SDS		-0.3 (1.4)	80	-0.3 (1.0)	91	0.90
Low BMD		21 (49%)	43	26 (51%)	51	0.84
Osteoporosis		8 (19%)	43	14 (28%)	51	0.31
Osteopenia		18 (42%)	43	25 (49%)	51	0.49

between -1 and -2.5 or a Z-score between -1 and -2. This is evaluated at 3 locations: L2-L4, femur neck, and total body. Patients were also included if data were available at less than 3 locations. Bold values represent significant differences between groups ($P \leq 0.05$). Abbreviations: AO, adulthood onset; BF%, body fat percentage; BMD, bone mineral density (in g/cm^2); CO, childhood onset; CP, craniopharyngioma patient; DXA = dual-energy X-ray absorptiometry scan; L2-L4 = located at lumbar vertebral body 2–4; SDS = standardized deviation scores. [a]n = 95.

Table 8 Univariable and multivariable logistic regression analysis for determinants of bone health in CP survivors

	T- or Z-score ≤-1			
	Univariable analysis		Multivariable analysis	
Variables	OR (95% CI)	*P*	OR (95% CI)	*P*
Age at DXA-scan	**1.04 (1.0-1.1)**	**0.005**	1.0 (1.0-1.1)	0.08
BMI	0.98 (0.9-1.1)	0.60		
Weight	1.0 (1.0-1.0)	0.37		
Diabetes mellitus	0.8 (0.2-2.7)	0.69		
Hydrocortisone dose	1.0 (1.0-1.1)	0.43		
Childhood onset disease	1.2 (0.5-2.7)	0.69		
Female sex	0.8 (0.4-1.9)	0.68		
Hydrocephalus	1.05 (0.5-2.4)	0.92		
Hypothalamic damage	**0.4 (0.2-1.0)**	**0.04**	1.6 (0.4-5.5)	0.49
Medication for epilepsy	1.3 (0.3-6.2)	0.74		
Growth hormone deficiency**	0.2 (0.0-2.3)	0.22		
GHRT	0.4 (0.11-1.3)	0.11	10.4 (0.9-125.8)	0.07
Adrenal axis deficiency	1.2 (0.4-3.7)	0.71		
Gonadal axis deficiency	1.7 (0.4-6.3)	0.46		
TSH deficiency	1.0 (0.1-7.7)	0.97		

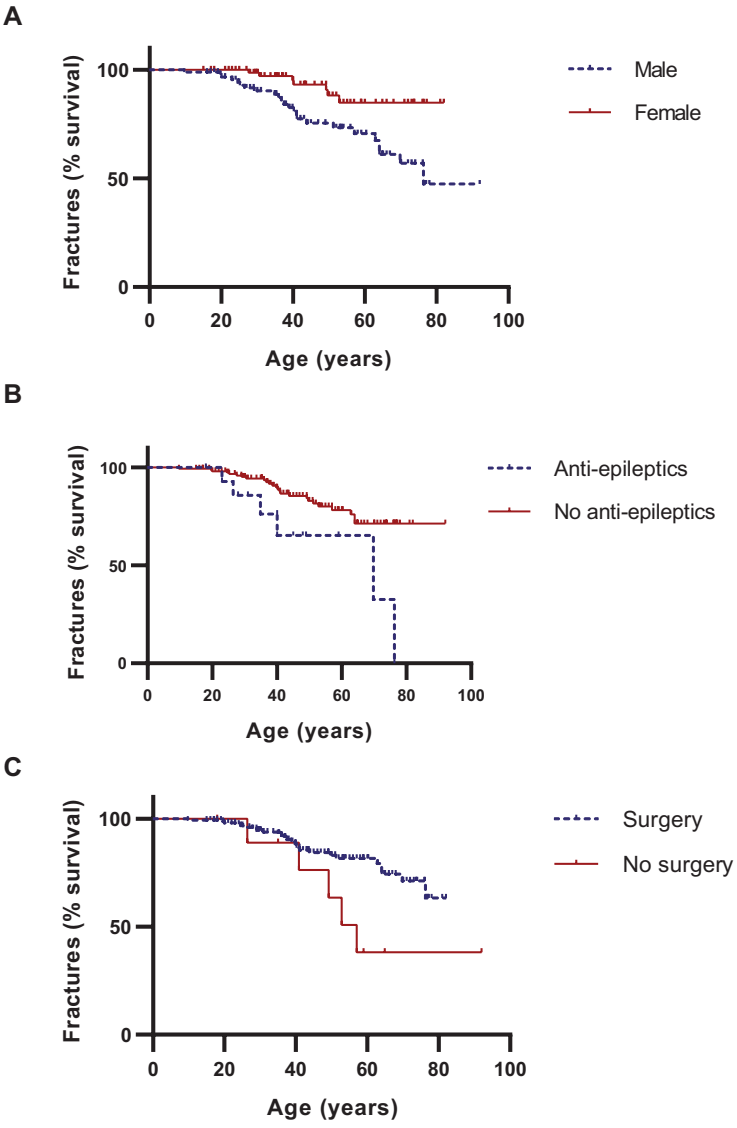
Sex steroid replacement therapy	0.7 (0.3-1.7)	0.41		
Diabetes insipidus	1.4 (0.6-3.4)	0.42		
Visual impairment	1.1 (0.4-2.9)	0.91		
BMD increasing medication	2.4 (0.7-8.3)	0.18	5.8 (1.1-29.7)	0.03
Scannertype: iDXA	1.6 (0.5-5.2)	0.44		
Scannertype: Prodigy	0.3 (0.0-1.8)	0.19		
Scannertype: DPXL	0.7 (0.1-4.0)	0.67		

	T-score ≤-2.5 or Z-score ≤-2			
	Univariable analysis		Multivariable analysis	
Variables	OR (95% CI)	*P*	OR (95% CI)	*P*
Age at DXA-scan	**1.03 (1.004-1.1)**	**0.03**	1.032 (1.003-1.1)	**0.03**
BMI	0.9 (0.9-1.0)	0.28		
Weight	1.0 (1.0-1.0)	0.49		
Diabetes mellitus	1.3 (0.3-5.2)	0.75		
Hydrocortisone dose	1.1 (1.0-1.2)	0.06		
Childhood onset disease	1.7 (0.5-4.4)	0.32		
Female sex	0.6 (0.2-1.6)	0.33		
Hydrocephalus	1.4 (0.6-3.4)	0.44		
Hypothalamic damage	0.4 (0.1-1.4)	0.16		
Medication for epilepsy	1.3 (0.2-7.4)	0.74		
Growth hormone deficiency**	0.2 (0.03-1.2)	0.07	0.2 (0.03-1.4)	0.10

GHRT	0.5 (0.1-1.6)	0.25		
Adrenal axis deficiency	1.3 (0.3-5.0)	0.74		
Gonadal axis deficiency	0.7 (0.2-2.9)	0.60		
TSH deficiency	NA*	NA*		
Sex steroid replacement therapy	0.7 (0.23-1.9)	0.44		
Diabetes insipidus	1.9 (0.64-5.8)	0.25		
Visual impairment	3.4 (0.7-15.8	0.13		
BMD increasing medication	2.4 (0.7-8.3)	0.18		
Scannertype: iDXA	0.7 (0.2-2.8)	0.64		
Scannertype: Prodigy	0.3 (0.0-3.2	0.33		
Scannertype: DPXL	0.9 (0.1-6.6)	0.93		

Abbreviations: BMD = bone mineral density; BMI = Body Mass Index; DXA = Dual-Energy X-ray Absorptiometry; GHRT = Growth Hormone Replacement Therapy; ROC = Receiver Operating Characteristic; TSH = Thyroid Stimulating Hormone. Univariable and multivariable model for low bone mineral density (BMD) (T- or Z-score at last DXA-scan ≤-1) and osteoporosis (T-score ≤-2.5 or Z-score ≤-2 at last DXA-scan). Height at last DXA-scan violates assumption of linearity of logit and is thus not included. No significant multivariable model for low BMD could be established. The best available model is shown. Several outliers were identified but not excluded from analysis. Omnibus Chi^2 $P = 0.09$, Nagelkerke $R^2=0.14$, Hosmer and Lemeshow $P = 0.50$. *NA: there were no cases of patients with osteoporosis and no TSH deficiency, bariatric surgery, surgery as tumor treatment, or Yttrium treatment. Multivariable logistic regression model osteoporosis: Omnibus Chi^2 $P = 0.02$, Nagelkerke $R^2=0.13$, Hosmer and Lemeshow $P = 0.11$. ROC-curve 0.65. **Small sample size for non-growth hormone deficiency: only 5 patients do not have a growth hormone deficiency and have DXA-scan results available.

Figure 1 Fracture free survival curves in craniopharyngioma survivors



Fracture free survival (Kaplan Meier) curves. Multivariable Cox regression model hazard ratios: (A) sex 0.4 (95%CI 0.2-0.83, *P* = 0.02), (B) for surgery 0.32 (0.12-0.84, *P* = 0.02) and (C) for anti-epileptics use 2.68 (1.08-6.69, *P* = 0.03) (-2LL Chi^2 <0.001, Omnibus Chi^2 0.001).

Discussion

Eighteen percent of the CP had suffered from at least one fracture during a median follow-up of 16 years. Fractures occurred more often in males than females and males were less often treated with medication intended to improve BMD, which could indicate that males were undertreated. DXA revealed overall mean BMD values in the normal range in our cohort and no differences in low BMD or T- or Z-scores between patients with and without fractures. However, the spread was wide: 46% had osteopenia and 24% had osteoporotic BMD T- or Z-scores.

In our study, osteoporosis was not a predictor of fracture risk. Obviously, there are more factors than only BMD that contribute to fracture risk. In the literature, prior and current exposure to glucocorticoids was associated with an increased fracture risk of substantial importance beyond explanations by BMD.[42] Using our logistic regression model for fractures, we did not find any role for pituitary hormonal deficiencies in the increased fracture rates. This is probably due to adequate replacement therapy, as illustrated by replacement rates of 94% for hypogonadotropic hypogonadism. As expected, epilepsy medication accelerated fracture risk, while previous treatment with surgery and female sex were protective. Anti-epileptic drugs can induce poor balance or clumsiness; certain antiepileptic drugs are associated with an increased rate of bone loss,[17] and some may cause hyponatremia, which correlates with fractures and low BMD.[43]

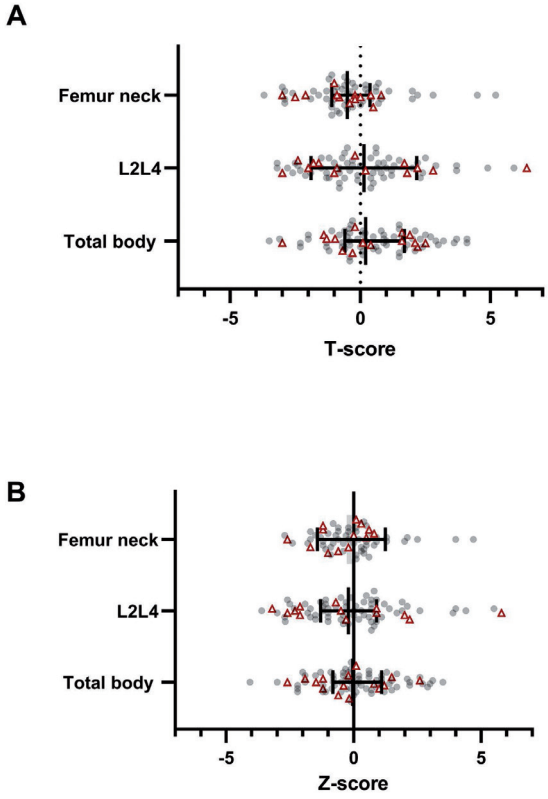
The observation that surgery as previous tumour treatment was identified as 'protective' factor, was surprising. Only a few patients were not treated with surgery; perhaps these are patients severely affect by comorbidities where surgery could not safely be performed, thereby having an increased risk for fractures as well. Patients with a fracture in their history had a higher mean age, were more often male, and had less often surgery. Female sex was observed to be a protective factor, which corresponds with the findings of Müller et al.[24] This might be related to the relatively young age of our study population, and to a higher BMI in women. In the premenopausal general population, higher

frequencies of fractures and lower osteoporosis preventive medication have been described in men, while in the postmenopausal general population, female sex is a risk factor for fractures.[44-46] A trend was shown for radiotherapy being protective for fractures. The general aim of the surgeons in treatment of CP have shifted from attempting gross total resection towards aiming at a more limited resection today, which may leave more tumour mass behind and cause the need to additional treatment with radiotherapy. Presumably, the lower risk for fractures observed in our study is therefore explained by a less damaging treatment strategy.

We found a high percentage of fractures in our study, since the cumulative fracture incidence in a European population was reported to be 7.8% in men and 9.5% in women.[19] In our previous study, no higher risk was found for fractures in our CP cohort.[10] This may partly be related to a lack of reliable Dutch reference data, which prompted Wijnen et al. to compare fracture incidence in Dutch patients with Swedish reference data. Fracture rate may differ between populations: it has been reported that Scandinavian countries have higher fracture rates than western Europe.[47] Olsson et al. previously showed an increased fracture risk.[1]

In a GHD population from the KIMS study (Pfizer International Metabolic Database), fracture rates were reported to be even higher compared to our study (27% in males and 29% in females); no significant differences of fracture prevalence between patients with or without (multiple) other hormonal pituitary deficiencies were found.[21] This suggests that GHD itself is an important factor in fracture risk, freestanding from other hormonal pituitary deficiencies. Given the high prevalence of GHD patients in our study, one might expect an even higher a frequency of fractures. Again, a demographic factor could play a role for a higher frequency of fractures, or a registration bias, or compensation of BMD by the protective factor of obesity in CP.[48] Obesity occurred in up to 75% of our patients[34] and a trend was shown for BMI as protective factor for fractures in our univariate logistic regression analysis. Gonadal axis hormonal deficiency does not seem to be a major contributor for a higher frequency of fractures, as demonstrated by our results; probably because 94% of premenopausal women and men received hormonal replacement therapy.

Figure 2 Scatterplot of T- and Z-scores of bone mineral density in survivors of craniopharyngioma with and without fractures



Scatterplot of T- (A) and Z-scores (B) of bone mineral density of the femur neck, L2-L4 and total body in survivors of craniopharyngioma. The error bars express median and quartiles. Every circle or triangle represents a measurement at last Dual-Energy X-ray Absorptiometry scan. The red triangles represent patients who had fractures in the past; the grey circles are patients who did not have fractures. A wide spread is shown in both patients with and without fractures in the past.

The mean BMD at last DXA-scan was normal in the total cohort, but low BMD occurred in 50% and osteoporosis in 24% (Table 7). The dispersion was rather large, and similar in patients with and without fractures as showed in Figure 1. BMD T- and Z-scores were similar between genders except for Z-score of the femur neck, which was lower in males (-0.4 ± 1.2 vs. 0.2 ± 1.4, $P = 0.08$). Sex or GHD were not a contributing factor in our logistic regression model for osteoporosis.

Final height in our patients was not impaired: the mean SDS score was -0.3 ± 1.2. It was also similar between AO CP and CO CP, and between genders. Apparently, GHRT had been sufficiently administered to these children. Indeed, 87% of patients with CO CP and GHD received GHRT. This is different from the findings of Kendall-Taylor et al., who did find for AO CP a significant difference in height (although it was on average only 3 cm or 0.6 SDS).[19] Perhaps the large size of their European cohort made it possible to find smaller differences, or there may be regional differences in treatment of CP, particularly the age at which GHRT is started.

Due to the retrospective cross-sectional design, there are some limitations of our study. Factors that may have contributed to a possible selection bias are the size and weight limitations of DXA-scanners,[49] DXA-scans were performed based on individual care plan decided by the insight of the clinicians. All types of DXA-scanners were included, while measurements by different DXA-scanners have shown intra-individual variability.[50] However, agreement between all DXA-devices is high.[51] By using age- and sex-matched SDS and applying generally used cut-offs to identify osteopenic or osteoporotic BMD values, we tried to limited the impact on our data. In our logistic regression model for osteoporosis, scanner type was not a contributing factor. Due to the retrospective design, relevant data on duration of GHRT, smoking, alcohol use, dietary intake, fractures of parents, physical activity and level of 25-hydroxyvitamin D were lacking. The percentage of fractures could be underestimated due to a reporting bias. Still, since studies on bone health in CP are so limitedly available, our study provides more insight and should create awareness for the importance of attention to bone health in CP.

This is the first study to date that investigated contributory factors for fractures in CP, and that describes bone health in a cohort existing of both CO and AO long-term survivors, with a large sample size. With this study, we hope to raise more awareness for the evaluation of bone health. The decision to either or not perform a DXA-scan is important in this respect, although DXA-scans alone do not evaluate the quality of the bone or fracture risk sufficiently. Fracture rate could also

be compromised by for example neurological defects. Nevertheless, clinicians should consider treatments that increase bone health, especially in males.

In conclusion, our cohort of CP showed a high frequency of fractures. Although men had a higher frequency of fractures, they received BMD increasing treatments less frequently. Pituitary hormone replacement therapy in our cohort overall seemed adequately managed, since pituitary hormonal deficiencies did not contribute significantly to fracture risk in our logistic regression model. Also of interest was our finding that osteoporotic BMD values did not predict the fracture risk. Our study suggests the importance of regularly evaluating fracture risk in every CP. We advise to take other risk factors than BMD into account, and suggest earlier evaluation of bone health increasing medication, especially in men.

References

1. Olsson DS, Andersson E, Bryngelsson IL, Nilsson AG, Johannsson G. Excess mortality and morbidity in patients with craniopharyngioma, especially in patients with childhood onset: a population-based study in Sweden. J Clin Endocrinol Metab. 2015;100(2):467–474.
2. Müller HL, Merchant TE, Warmuth-Metz M, Martinez-Barbera JP, Puget S. Craniopharyngioma. Nat Rev Dis Primers. 2019;5(1):75.
3. Wijnen M, van den Heuvel-Eibrink MM, Janssen JAMJL, et al. Very long-term sequelae of craniopharyngioma. Eur J Endocrinol. 2017;176(6):755–767.
4. Hyer SL, Rodin DA, Tobias JH, Leiper A, Nussey SS. Growth hormone deficiency during puberty reduces adult bone mineral density. Arch Dis Child. 1992;67(12):1472–1474.
5. Colao A, Di Somma C, Pivonello R, et al. Bone loss is correlated to the severity of growth hormone deficiency in adult patients with hypopituitarism. J Clin Endocrinol Metab. 1999;84(6):1919–1924.
6. Chemaitilly W, Li Z, Huang S, et al. Anterior hypopituitarism in adult survivors of childhood cancers treated with cranial radiotherapy: a report from the St Jude Lifetime Cohort study. J Clin Oncol. 2015;33(5):492–500.
7. Molitch ME, Clemmons DR, Malozowski S, Merriam GR, Vance ML; Endocrine Society. Evaluation and treatment of adult growth hormone deficiency: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab. 2011;96(6):1587–1609.
8. Barake M, Arabi A, Nakhoul N, et al. Effects of growth hormone therapy on bone density and fracture risk in age-related osteoporosis in the absence of growth hormone deficiency: a systematic review and meta-analysis. Endocrine. 2018;59(1):39–49.
9. Debono M, Ross RJ, Newell-Price J. Inadequacies of glucocorticoid replacement and improvements by physiological circadian therapy. Eur J Endocrinol. 2009;160(5):719–729.
10. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. Excess morbidity and mortality in patients with craniopharyngioma: a hospital-based retrospective cohort study. Eur J Endocrinol. 2018;178(1):93–102.
11. Holmer H, Popovic V, Ekman B, Follin C, Siversson AB, Erfurth EM. Hypothalamic involvement and insufficient sex steroid supplementation are associated with low bone mineral density in women with childhood onset craniopharyngioma. Eur J Endocrinol. 2011;165(1):25–31.

12. Roth C, Wilken B, Hanefeld F, Schr ter W, Leonhardt U. Hyperphagia in children with craniopharyngioma is associated with hyperleptinaemia and a failure in the downregulation of appetite. Eur J Endocrinol. 1998;138(1):89–91.

13. Brabant G, Horn R, Mayr B, von zur Mühlen A, Honegger J, Buchfelder M. Serum leptin levels following hypothalamic surgery. Horm Metab Res. 1996;28(12):728–731.

14. Ducy P, Amling M, Takeda S, et al. Leptin inhibits bone formation through a hypothalamic relay: a central control of bone mass. Cell. 2000;100(2):197–207.

15. Syrjälä P, Luukinen H, Pyhtinen J, Tolonen U. Neurological diseases and accidental falls of the aged. J Neurol. 2003;250(9): 1063–1069.

16. Myers AH, Young Y, Langlois JA. Prevention of falls in the elderly. Bone. 1996;18(1 Suppl):87S–101S.

17. Koppel BS, Harden CL, Nikolov BG, Labar DR. An analysis of lifetime fractures in women with epilepsy. Acta Neurol Scand. 2005;111(4):225–228.

18. Vidal M, Thibodaux RJ, Neira LFV, Messina OD. Osteoporosis: a clinical and pharmacological update. Clin Rheumatol. 2019;38(2):385–395.

19. Kendall-Taylor P, Jönsson PJ, Abs R, et al. The clinical, metabolic and endocrine features and the quality of life in adults with childhood-onset craniopharyngioma compared with adult-onset craniopharyngioma. Eur J Endocrinol. 2005;152(4):557–567.

20. Yuen KC, Koltowska-Häggström M, Cook DM, et al. Clinical characteristics and effects of GH replacement therapy in adults with childhood-onset craniopharyngioma compared with those in adults with other causes of childhood-onset hypothalamicpituitary dysfunction. Eur J Endocrinol. 2013;169(4):511–519.

21. Wüster C, Abs R, Bengtsson BA, et al.; KIMS Study Group and the KIMS International Board. Pharmacia & Upjohn International Metabolic Database. The influence of growth hormone deficiency, growth hormone replacement therapy, and other aspects of hypopituitarism on fracture rate and bone mineral density. J Bone Miner Res. 2001;16(2):398–405.

22. Zhao Y, Gan X, Luo P, et al. The risk of osteopenia in premenopausal women with various sellar tumors. Gynecol Endocrinol. 2012;28(12):945–948.

23. Okinaga H, Matsuno A, Okazaki R. High risk of osteopenia and bone derangement in postsurgical patients with craniopharyngiomas, pituitary adenomas and other parasellar lesions. Endocr J. 2005;52(6):751–756.

24. Müller HL, Schneider P, Bueb K, et al. Volumetric bone mineral density in patients with childhood craniopharyngioma. Exp Clin Endocrinol Diabetes. 2003;111(3):168–173.

25. DeVile CJ, Grant DB, Hayward RD, Stanhope R. Growth and endocrine sequelae of craniopharyngioma. Arch Dis Child. 1996;75(2):108–114.

26. Ono N, Kohga H, Zama A, Inoue HK, Tamura M. A comparison of children with suprasellar germ cell tumors and craniopharyngiomas: final height, weight, endocrine, and visual sequelae after treatment. Surg Neurol. 1996;46(4):370–377.

27. Jensen BL, Jensen KE, Kastrup KW, Pedersen SA, Wagner A. Final height and craniofacial development after surgical resection of craniopharyngioma. J Craniofac Genet Dev Biol. 1997;17(4):190–197.

28. Price DA, Wilton P, J nsson P, et al. Efficacy and safety of growth hormone treatment in children with prior craniopharyngioma: an analysis of the Pharmacia and Upjohn International Growth Database (KIGS) from 1988 to 1996. Horm Res. 1998;49(2):91–97.

29. de Vries L, Lazar L, Phillip M. Craniopharyngioma: presentation and endocrine sequelae in 36 children. J Pediatr Endocrinol Metab. 2003;16(5):703–710.

30. Müller HL, Emser A, Faldum A, et al. Longitudinal study on growth and body mass index before and after diagnosis of childhood craniopharyngioma. J Clin Endocrinol Metab. 2004;89(7):3298–3305.

31. Sorva R. Children with craniopharyngioma. Early growth failure and rapid postoperative weight gain. Acta Paediatr Scand. 1988;77(4):587–592.

32. Tosta-Hernandez PDC, Siviero-Miachon AA, da Silva NS, Cappellano A, Pinheiro MM, Spinola-Castro AM. Childhood craniopharyngioma: a 22-year challenging follow-up in a single center. Horm Metab Res. 2018;50(9):675–682.

33. Güemes Hidalgo M, Muñoz Calvo MT, Fuente Blanco L, Villalba Castaño C, Martos Moreno GA, Argente J. Endocrinological outcome in children and adolescents survivors of central nervous system tumours after a 5 year follow-up. An Pediatr (Barc). 2014;80(6):357–364.

34. van Santen SS, Olsson DS, Hammarstrand C, et al. Diagnosing metabolic syndrome in craniopharyngioma patients: body composition versus BMI. Eur J Endocrinol. 2019;181(2): 173–183.

35. van Santen S, Olsson D, van den Heuvel-Eibrink M, et al. Supplemental tables of "Bone health and final height in craniopharyngioma patients with a follow-up of 16 years" DANS 2020;1-6. https://doi.org/10.17026/dans-xwm-jxy2

36. Cummings SR, Bates D, Black DM. Clinical use of bone densitometry: scientific review. JAMA. 2002;288(15):1889–1897.

37. Centraal bureau voor Statistiek (CBS). ProMED-mail website. https://www.cbs.nl/. Accessed 23 August 2019.

38. Wikland KA, Luo ZC, Niklasson A, Karlberg J. Swedish population-based longitudinal reference values from birth to 18 years of age for height, weight and head circumference. Acta Paediatr. 2002;91(7):739–754.

39. Sjöberg A, Barrenäs ML, Brann E, et al. Body size and lifestyle in an urban population entering adulthood: the 'Grow up Gothenburg' study. Acta Paediatr. 2012;101(9):964–972.

40. Werner B, Bodin L. Growth from birth to age 19 for children in Sweden born in 1981: descriptive values. Acta Paediatr. 2006;95(5):600–613.

41. Kanis JA, Johansson H, Oden A, et al. A meta-analysis of prior corticosteroid use and fracture risk. J Bone Miner Res. 2004;19(6):893–899.

42. Upala S, Sanguankeo A. Association between hyponatremia, osteoporosis, and fracture: a systematic review and meta-analysis. J Clin Endocrinol Metab. 2016;101(4):1880–1886.

43. Lobo E, Marcos G, Santab rbara J, et al.; ZARADEMP Workgroup. Gender differences in the incidence of and risk factors for hip fracture: a 16-year longitudinal study in a southern European population. Maturitas. 2017;97:38–43.

44. Duyvendak M, Naunton M, Atthobari J, van den Berg PB, Brouwers JR. Corticosteroid-induced osteoporosis prevention: longitudinal practice patterns in The Netherlands 2001-2005. Osteoporos Int. 2007;18(10):1429–1433.

45. Lagro-Janssen T, Grosicar J. Morbidity figures from general practice: sex differences in traumatology. J Eval Clin Pract. 2010;16(4):673–677.

46. Kanis JA, McCloskey EV, Johansson H, Cooper C, Rizzoli R, Reginster JY; Scientific Advisory Board of the European Society for Clinical and Economic Aspects of Osteoporosis and Osteoarthritis (ESCEO) and the Committee of Scientific Advisors of the International Osteoporosis Foundation (IOF). European guidance for the diagnosis and management of osteoporosis in postmenopausal women. Osteoporos Int. 2013;24(1):23–57.

47. Reid IR. Fat and bone. Arch Biochem Biophys. 2010;503(1):20–27.

48. Seabolt LA, Welch EB, Silver HJ. Imaging methods for analyzing body composition in human obesity and cardiometabolic disease. Ann N Y Acad Sci. 2015;1353:41–59.

49. LaForgia J, Dollman J, Dale MJ, Withers RT, Hill AM. Validation of DXA body composition estimates in obese men and women. Obesity (Silver Spring). 2009;17(4):821–826.

50. Hull H, He Q, Thornton J, et al. iDXA, Prodigy, and DPXL dual-energy X-ray absorptiometry whole-body scans: a crosscalibration study. J Clin Densitom. 2009;12(1):95–102.

4

Body Composition and Bone Mineral Density in Craniopharyngioma Patients: A Longitudinal Study Over 10 Years

Selveta S. van Santen,[1,2,5] Daniel S. Olsson,[3,4]
Casper Hammarstrand,[3,4] Mark Wijnen,[1,2]
Marta Fiocco,[5,6,7] Marry M. van den Heuvel-Eibrink,[2,5]
Gudmundur Johannsson,[3] Joseph A. M. J. L. Janssen,[1]
Aart J. van der Lely,[1] Sebastian J. C. M. M. Neggers.[1,2,5]

[1]Department of Medicine, Endocrinology; Erasmus Medical Center, Rotterdam, The Netherlands;
[2]Department of Paediatric Oncology/Haematology, Erasmus MC – Sophia Children's Hospital, Rotterdam, The Netherlands;
[3]Department of Medicine, Endocrinology; Sahlgrenska University Hospital, Gothenburg, Sweden;
[4]Department of Internal Medicine and Clinical Nutrition, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden;
[5]Princess Máxima Center for Paediatric Oncology, Utrecht, The Netherlands;
[6]Mathematical Institute, Leiden University, Leiden, The Netherlands;
[7]Department of Biomedical Data Sciences, Medical Statistics Section, Leiden University Medical Center, Leiden, The Netherlands.

The Journal of Clinical Endocrinology & Metabolism

Abbreviations: AO, adulthood onset; BF%, body fat percentage; BMD, bone mineral density; BMI, body mass index; CO, childhood onset; CP, craniopharyngioma; DI, diabetes insipidus; DXA, dual energy X-ray absorptiometry; FMI, fat mass index; FFMI, fat free mass index; GHRT, growth hormone replacement therapy.

Abstract

Context: Patients with craniopharyngioma suffer from obesity and impaired bone health. Little is known about longitudinal changes in body composition and bone mineral density (BMD).

Objective: To describe body composition and BMD (change).

Design: Retrospective longitudinal study.

Setting: Two Dutch/Swedish referral centers.

Patients: Patients with craniopharyngioma (n = 112) with a dual-energy X-ray absorptiometry (DXA) scan available (2 DXA scans, n = 86; median Δtime 10.0 years; range 0.4-23.3) at age ≥ 18 years (58 [52%] male, 50 [45%] childhood onset).

Main outcome measures: Longitudinal changes of body composition and BMD, and associated factors of ΔZ-score (sex and age standardized).

Results: BMI (from 28.8 ± 4.9 to 31.2 ± 5.2 kg/m^2, $P < 0.001$), fat mass index (FMI) (from 10.5 ± 3.6 to 11.9 ± 3.8 kg/m^2, $P = 0.001$) and fat free mass index (FFMI) (from 18.3 ± 3.2 to 19.1 ± 3.2 kg/m^2, $P < 0.001$) were high at baseline and increased. Fat percentage and Z-scores of body composition did not increase, except for FFMI Z-scores (from 0.26 ± 1.62 to 1.06 ± 2.22, $P < 0.001$). Z-scores of total body, L2-L4, femur neck increased (mean difference 0.61 ± 1.12, $P < 0.001$; 0.74 ± 1.73, $P < 0.001$; 0.51 ± 1.85, $P = 0.02$). Linear regression models for ΔZ-score were positively associated with growth hormone replacement therapy (GHRT) [femur neck: beta 1.45 (95% CI 0.51–2.39)]; and negatively with radiotherapy [femur neck: beta -0.79 (-1.49--0.09)], glucocorticoid dose [total body: beta -0.06 (-0.09--0.02)] and medication to improve BMD [L2-L4: beta -1.06 (-1.84--0.28)].

Conclusions: Z-scores of BMI, fat percentage, and FMI remained stable in patients with craniopharyngioma over time, while Z-scores of FFMI and BMD increased. Higher glucocorticoid dose and radiotherapy were associated with BMD loss and GHRT with increase.

Keywords: craniopharyngioma, DXA, body composition, sarcopenia, bone mineral density, longitudinal study

Introduction

Patients with craniopharyngioma (CP) suffer from a tumor that may affect the pituitary, optic nerve and hypothalamus, causing damage and impaired function of these important structures.[1] The age at presentation of the disease has a bimodal distribution, with incidence peaks observed in children aged 5-14 [childhood onset (CO) patients with CP] and in adults aged 50-74 years [adulthood onset (AO) patients with CP].[2,3] Patients with CP have excess mortality,[1,3,4] especially due to obesity-related disorders (standardized mortality ratio 3.2- 19).[3-7] Neurological or endocrinological dysfunction may increase fracture risk[8] and obesity. Obesity was reported in up to 75% of the patients.[9] Several factors may contribute to obesity in patients with CP, such as endocrine aberrations related to hypopituitarism, visual and neurological sequelae with decreased sympathetic nerve activity, sleep disturbances and daytime somnolence.[10] The simplest way to investigate obesity is to calculate body mass index (BMI): it is non-invasive and inexpensive. Unfortunately, it does not well differentiate body fat from fat free mass.[11] Dual-Energy X-ray absorptiometry (or DXA-scan) is considered a reliable method to determining body composition in case of suspected discordance of BMI and adiposity,[9,12] and can be used to measure bone mineral density (BMD).[13] Only a couple of small studies have published cross-sectional data on DXA-scan derived body composition measures in patients with CP.[14-17] In a study including 11 patients with CP, it was found that resting energy rate was significantly reduced in obese subjects, even after controlling for fat free mass.[16] In another study with 185 CO patients with CP, obesity was found to be associated with higher BMI at diagnosis, higher maternal BMI, presence of ventriculoperitoneal shunts and hypothalamic tumor involvement.[18]

Studies in patients with CP investigating the longitudinal changes in BMI[18-21] or body composition[15] are scarce. Weight gain in patients with CP was observed mostly before[19] or in the first year after treatment/ diagnosis.[18,19,22,23] The only study investigating changes in body fat percentage (BF%) in 19 patients with CP receiving growth hormone replacement treatment (GHRT) reported no change in BF% after 60 months of follow-up.[15] To the best of our knowledge, no longitudinal

studies in patients with CP on sarcopenia or BMD change and influencing factors for BMD change have been performed to date.[1,24-28] Therefore, the aim of the present study was to describe the course of body composition and BMD in patients with CP, during follow-up, and to identify potential influencing factors for BMD changes.

Materials and Methods

This longitudinal retrospective study included patients who were treated for craniopharyngioma either at the Erasmus MC (Rotterdam, the Netherlands), or Sahlgrenska University Hospital (Gothenborg, Sweden). Only DXA-scans from patients over 18 years of age were included in the study. The research proposal was approved by the Ethical Review Board of the Erasmus MC and the Regional Ethical Review Board of the Gothenborg University; all patients gave their informed consent.

Data collection and patient identification methods have previously been described.[4, 8, 24] Data were collected from the first and most recent DXA scan regardless of scanner type. DXA scan data included pooled data of Lunar DPXL, Lunar iDXA, Lunar Prodigy. The study evaluated the Lunar DPXL and Lunar Prodigy data separately, since different scanner types might generate different results.[8] We did not do this for the Lunar iDXA because there was only a limited number of results from this scanner type. Gathered data existed of body composition measures (such as BF%, fat mass index [FMI] and fat free mass index [FFMI]), BMI, and BMD values and corresponding sex-specific T-scores and sex- and age-specific Z-scores as described in DXA scanner reports. The mean of the differences between first and last DXA scan was calculated. Furthermore, sex- and age-specific Z-scores were computed;[29-38] for body composition measures, Swedish references were used if Dutch references were unavailable.[9, 34]

All definitions were previously described.[8] Hypothalamic damage was defined as tumor- and/or treatment-related injury to the hypothalamus and/or third ventricle as visualized by neuroimaging.[9,24] In our analysis we evaluated patients who had radiotherapy and

hypothalamic damage together as 1 group, as we assumed that they had hypothalamic dysfunction. Medication to improve BMD was defined as current or past use of bisphosphonates, vitamin D, or calcium (gonadal axis replacement therapy or other hormonal replacement therapy were excluded for this definition). Patients were categorized as using high or low glucocorticoid dose, where patients with mean or higher glucocorticoid dose (17.7 mg of hydrocortisone equivalent dose) were considered to have a high dose. FFMI was calculated as lean mass and mineral bone mass (kg) per square height (meters); FMI as body fat mass (kg) per square height (meters); and BMI as total body weight per square height (meters).

Cut-offs of categories for BMD (osteopenia and osteoporosis)[39] and for high or low body composition measures are shown in Table 1 and Supplementary Table 1AB.[40] BMD categories may be specified through single or multiple sites (at either of femur neck, L2-L4, and/or total body). Patients were considered as obese based on BMI Z-scores >2.0.

Table 1 Definitions of cutoffs for body composition and BMD measures

	Male	Female
Body composition		
High BMI	≥ 30 kg/m^2 or Z-score ≥2	≥ 30 kg/m^2 or Z-score ≥2
High BF%	≥26% if aged 20-39 years old or ≥29% if aged 40-59 years old, or Z-score ≥2[38]	≥39% if aged 20-39 years old; ≥41% if aged 40-59 years old, or Z-score ≥2
High FMI	≥ 9 or Z-score ≥2[37]	≥ 13 or Z-score ≥2
Low FFMI	Z-score ≤-2	Z-score ≤-2
BMD		
Osteopenia	T-score -1 - -2.5 or Z-score -1 - -2	
Osteoporosis	T-score ≤-2.5 or Z-score ≤-2	

Abbreviations: BF% = body fat percentage; BMD = bone mineral density; BMI = body mass index; FMI = fat mass index; FFMI = fat free mass index.

Statistical analysis

Statistical analysis was performed using Version 25.0 of IBM SPSS Statistics for Windows, and Graphpad Version 8.01. Data are presented as mean and standard deviation (SD) unless stated otherwise. *P*-values of < 0.05 were considered significant. Comparisons of proportions were performed by using either the $\chi 2$ test, the Fisher's exact test or McNemar's test as appropriate. For normally distributed data, T-test was used for group comparisons, while nonparametric equivalents were used in case of violation of the normality assumption. Univariable and multivariable linear regression models were estimated to investigate determinants of changes in BMD Z-scores; baseline Z-scores and age at first DXA-scan were incorporated in each model. The model with the biggest R^2 and most significant variables was ultimately chosen. A diagnostic check was performed on model fitting, homoscedasticity, multicollinearity, influential cases and outliers.

Results

Baseline characteristics of subjects

Baseline characteristics of 112 included patients with CP are shown in Table 2. Most of the included patients originate from a previously reported cohort.[4, 24] Patients were previously treated with surgery (n = 108; in 64 patients a transsphenoidal approach and in 29 patients a transcranial approach), radiotherapy (n = 61), and yttrium (n = 13). The median number of surgical procedures was 1 (mean 1.3 ± 0.9, range 0-5). The median age at first presentation of CP was 25 years (range 0-73). Mean follow-up time was 19 ± 11 years (range 1-62). At last follow-up, 15 patients (13%) had died. Men received less often medication to improve BMD (4 [7%] vs. 12 [22%], *P* = 0.02) than women (as previously reported)[8]. Gonadal deficiencies were not different between men and women (95% vs. 85%, *P* = 0.15). Gonadal axis replacement therapy was more often administered to men than women (91% vs. 57% *P* < 0.0001) but not different if postmenopausal women (defined as >51 years) were excluded (91% vs. 94%, *P* = 1.00). Of 105 growth hormone deficient patients, 93 were using GHRT (89%). Patients

diagnosed before the year 2000 or from 2000 and onwards did not show any differences in body composition or BMD, except for total body BMD Z-score (−0.22 ± 1.48 vs. 0.51 ± 1.24, $P = 0.04$).[40]

Table 2 Baseline characteristics of craniopharyngioma patients

	All patients with CP (n=112)	Males	Females	*P*-value
Age at last follow-up (years)[a]	49 (16-82)	52 (16-78)	46 (18-82)	0.36
Age at presentation (years)[a]	25 (0-73)	27 (0-62)	23 (6-73)	0.93
(Female/ corresponding) gender (n [%])	54 (48%)	58 (100%)	55 (100%)	0.78
Childhood onset disease (n [%])	50 (45%)	24 (41%)	26 (48%)	0.47
Tumor location at last follow-up (n [%])				
Intrasellar	2 (2%)	1 (2%)	1 (2%)	1.0
Suprasellar	45 (40%)	22 (38%)	23 (43%)	0.50
Intra-/suprasellar	63 (56%)	35 (60%)	28 (52%)	0.49
CP treatment				
Surgery	108 (96%)	58 (100%)	50 (93%)	0.05
Radiation	62 (55%)	31 (53%)	31 (57%)	0.76
Pituitary deficiencies				
GH deficiency[c]	105 (94%) / 93 (83%)	53 (91%) / 48 (83%)	52 (96%) / 45 (83%)	0.44
TSH deficiency[c]	107 (96%) / 107 (96%)	58 (100%) / 58 (100%)	49 (91%) / 49 (91%)	**0.02**

Hypogonadism[bc]	102 (91%) / 84 (75%)	55 (95%) / 53 (91%)	47 (87%) / 31 (57%)	0.19
Corticotropic deficiency[c]	95 (85%) / 92 (84%)	53 (91%) / 50 (89%)	42 (78%) / 42 (78%)	0.045
ADH deficiency[c]	76 (68%) / 75 (67%)	40 (69%) / 40 (69%)	36 (67%) / 35 (65%)	0.80
Medical history				
Recurrence/ progression	40 (36%)	19 (33%)	21 (39%)	0.54
Hypothalamic damage	38 (36%)	21 (38%)	17 (35%)	0.77
Hydrocephalus ever	26 (23%)	11 (19%)	15 (28%)	0.29
Visual impairment	81 (76%)	42 (76%)	39 (75%)	0.99
Epilepsy	14 (13%)	5 (9%)	9 (17%)	0.20
Diabetes mellitus	13 (12%)	7 (12%)	6 (11%)	0.85
Fracture in history	21 (19%)	16 (28%)	5 (9%)	**0.01**
BMD medication	16 (14%)	4 (7%)	12 (22%)	**0.02**

The bold values indicate that they are statistically significant ($P < 0.05$). Tumor location and recurrence or progression were unknown in 2 and 1 patient(s), respectively. Abbreviations: ADH, antidiuretic hormone; BMD medication, bone mineral density increasing medication; CP, Craniopharyngioma; GH, growth hormone; N, number; TSH, thyroid stimulating hormone. [a]Median (range). [b]In females aged 50 years or younger and men, 83/90 subjects (92%) used gonadal hormone replacement therapy. [c]All/using replacement therapy.

Baseline values and longitudinal change in body composition

Median age at first DXA scan was 36 years (range 18-79) and age at last follow-up was 49 years (range 16-82). The median time since first tumor related treatment at first DXA scan was 9.9 years (range –9.2 to 47.0) and the median time between the first and last DXA scan was 10.0 years (range 0.41-23.3). The proportion of patients with obesity increased when defined by BMI Z-scores (from 28 [41%] to 38 [55%], $P = 0.02$) and FMI Z-scores (from 28 [48%] to 36 [61%], $P = 0.04$); other definitions of obesity did not change during follow-up (Table 3). Unstandardized values of BMI (from 28.8 ± 4.9 to 31.2 ± 5.2 kg/m^2, $P < 0.001$), FMI (from 10.5 ± 3.6 to 11.9 ± 3.8 kg/m^2, $P = 0.001$) and FFMI (from 18.3 ± 3.2 to 19.1 ± 3.2 kg/m^2, $P < 0.001$) were high at baseline and increased significantly (Table 4, Figure 1, and Supplementary Tables[40]). However, if standardized, only FFMI Z-scores increased (from 0.26 ± 1.62 to 1.06 ± 2.22, $P < 0.001$) (Figure 2). BF% (Z-scores) did not change (Table 4). There was no difference in change of body composition measures or body composition Z-scores between men and women over time (Figures 1 and 2). Patients treated for CP before 2000 had longer time difference from first to last DXA scan (12.4 ± 6.5 vs. 6.3 ± 4.2, $P < 0.001$) but no differences in BMI or body composition values at first DXA scan, last DXA scan or change over time (data not shown).

Subgroups: childhood onset vs. adulthood onset patients

The follow-up time since first presentation was significantly longer in CO patients than AO patients (22.7 ± 12.4 vs. 16.3 ± 9.0 years, $P = 0.03$). At baseline, CO patients had a higher proportion of subjects with normal BMI than AO patients (30% vs. 10%, $P = 0.02$) and had higher BF% (39.5 ± 13.2 vs. 34.2 ± 9.2, $P = 0.048$), but lower FFMI (16.6 ± 2.8 vs. 19.7 ± 3.0, $P < 0.001$) and FFMI Z-score values (–0.8 ± 1.4 vs. 0.7 ± 1.4, $P < 0.001$) than AO patients. During follow-up, CO patients increased more in BMI than AO patients (mean difference 2.8 ± 3.9 vs. 1.3 ± 2.9, $P = 0.008$) and increased in BF% Z-scores while AO patients decreased (mean difference 0.2 ± 0.9 vs. –0.3 ± 0.6, $P = 0.04$).

Subgroups: patients with vs. without radiotherapy/ hypothalamic damage

This group consisted of 34 patients who had radiotherapy only, 16 patients who had hypothalamic damage only, and of 22 patients who had both. At baseline, patients with radiotherapy or hypothalamic damage had a significant higher BMI (29.6 ± 5.1 vs. 27.2 ± 4.3, *P* = 0.02), higher BMI Z-scores (1.7 ± 1.9 vs. 0.7 ± 1.7, *P* = 0.004), and higher FMI Z-scores (1.6 ± 1.5 vs. 1.0 ± 1.2, *P* = 0.06). As a group, they had an increase in BF% Z-scores instead of a decrease in patients without radiotherapy or hypothalamic damage (mean change 0.1 ± 0.7 vs. −0.3 ± 0.8, *P* = 0.02) and increased more in FMI (mean change 1.3 ± 2.6 vs. 0.2 ± 2.1, *P* = 0.03). At last DXA scan, patients with radiotherapy or hypothalamic damage had not only a higher total body mass and fat-related body composition measures (BMI 32.3 ± 5.6 vs. 29.2 ± 3.5, *P* = 0.01; BMI Z-scores 1.8 ± 1.7 vs. 1.0 ± 1.2, *P* = 0.01; BF% Z-scores 1.8 ± 0.8 vs. 1.3 ± 1.1, *P* = 0.03; FMI Z-scores 2.5 ± 1.6 vs. 1.4 ± 1.3, *P* = 0.007), but also higher muscle mass-related measures (FFMI Z-scores 1.5 ± 2.6 vs. 0.3 ± 1.4, *P* = 0.03) than patients without radiotherapy or hypothalamic damage.

Table 3 Baseline DXA-scan and changes of BMD and body composition categories

	All CP		
	First DXA-scan	**Last DXA-scan**	***P***
Age (years)	37 ± 15	48 ± 16	**<0.001**
Osteoporosis (T/Z-score)	19/79 (24%)	20/79 (24%)	1.00
Osteoporosis (T-score)	13/75 (17%)	18/75 (24%)	0.38
Osteoporosis (Z-score)	19/77 (25%)	13/77 (17%)	0.24
Osteopenia (T/Z-score)	47/78 (60%)	42/79 (49%)	0.41
Osteopenia (T-score)	38/75 (51%)	27/75 (36%)	**0.04**
Osteopenia (Z-score)	39/77 (51%)	34/77 (44%)	0.46
BMI	28.8 ± 4.9	31.2 ± 5.1	**<0.001**
BMI Z-score	1.4 ± 1.9	1.5 ± 1.6	0.38
Low BMI	2/69 (3%)	1/69 (1%)	1.00

Normal BMI	12/69 (18%)	5/69 (7%)	**0.04**
Overweight	27/69 (40%)	25/69 (37%)	0.83
Obese	28/69 (41%)	38/69 (55%)	**0.02**
FMI			
Low FMI	0/59 (0%)	0/59 (0%)	NA
Normal FMI	10/59 (17%)	6/59 (10%)	0.29
Excess fat FMI	21/59 (36%)	17/59 (29%)	0.48
Obese FMI	28/59 (48%)	36/59 (61%)	**0.04**
FFMI			
Low FFMI	5/41 (12%)	2/41 (5%)	0.25
Normal FFMI	29/41 (71%)	29/41 (71%)	1.00
High FFMI	7/41 (17%)	10/41 (24%)	0.25
BF%			
Low BF%	0/48 (0%)	0/48 (0%)	NA
Normal BF%	16/48 (33%)	12/49 (25%)	0.34
High BF%	31/48 (65%)	36/48 (75%)	0.18

Baseline first DXA-scan and changes during follow-up (Δ). Data are presented as mean ± SD or n (%) as applicable. The bold values indicate that they are statistically significant ($P < 0.05$). For applied cutoffs, see Table 1 and Supplementary Table 1.[40] There are no significant differences between male and females at first or last DXA-scan. Abbreviations: BF% = Body Fat Percentage; BMI = Body Mass Index; DXA = Dual-Energy X-ray Absorptiometry Scan; FFMI = Fat Free Mass Index; FMI = Fat Mass Index; NA = Not Applicable (empty groups); *P* = *P*-value.

Subgroups: patients with vs. without certain pituitary hormone deficiencies

Patients with gonadal axis deficiency increased in BMI Z-scores (mean difference 0.2 ± 1.4 vs. −0.7 ± 1.0, $P = 0.03$) and increased more in higher FFMI at last DXA scan than patients without gonadal axis deficiency (mean change 1.4 ± 1.9 vs. −0.2 ± 1.2, $P = 0.009$). Patients with adrenocorticotropin deficiency increased during follow-up in BMI Z-scores (mean difference 0.2 ± 1.5 vs. −0.6 ± 1.0, $P = 0.03$) and FMI Z-scores (mean difference 0.1 ± 0.7 vs. −0.4 ± 1.5, $P = 0.03$). If patients

with diabetes insipidus (DI) were compared with patients without DI, they increased more in BMI (mean difference 2.6 ± 3.6 vs. 0.8 ± 2.6, *P* = 0.02), FMI Z-scores (mean difference 0.2 ± 0.7 vs. −0.2 ± 1.1, *P* = 0.006), and FFMI Z-scores from first to last DXA scan (mean difference 1.4 ± 1.4 vs. 0.1 ± 1.1, *P* = 0.005). Patients with high glucocorticoid dose had no significant differences from patients with low glucocorticoid dose except for higher unstandardized FFMI at first DXA scan (19.0 ± 3.2 vs. 16.9 ± 2.6, *P* = 0.003) and at last DXA scan (19.6 ± 3.3 vs. 18.1 ± 2.7, *P* = 0.04).

Table 4 DXA-results of any DXA-scanner type – Bone Mineral Density and Body Composition in all patients with craniopharyngioma

	First DXA-scan	Last DXA-scan	Δ	*P*-value
Age (years)	37.0 ± 14.9	48.0 ± 16.5	11.0 ± 6.5	**<0.001**
Bone				
Bone mineral density				
Total body	1.17 ± 0.13	1.19 ± 0.16	0.04 ± 0.10	**0.001**
L2-L4	1.13 ± 0.19	1.22 ± 0.26	0.11 ± 0.17	**<0.001**
Femur neck	0.96 ± 0.18	0.99 ± 0.22	0.04 ± 0.26	0.29
Femur	1.03 ± 0.21	1.04 ± 0.21	0.03 ± 0.32	0.70
T-score				
Total body	-0.10 ± 1.49	0.43 ± 1.68	0.66 ± 1.07	**<0.001**
L2-L4	-0.70 ± 1.57	0.14 ± 2.03	0.78 ± 1.63	**<0.001**
Femur neck	-0.64 ± 1.48	-0.40 ± 1.60	0.23 ± 1.95	0.44
Femur	-0.27 ± 1.61	-0.10 ± 1.52	0.26 ± 2.19	0.53
Z-score				
Total body	-0.49 ± 1.37	-0.05 ± 1.44	0.61 ± 1.12	**<0.001**
L2-L4	-0.77 ± 1.59	-0.03 ± 1.97	0.74 ± 1.73	**<0.001**
Femur neck	-0.62 ± 1.32	-0.09 ± 1.33	0.51 ± 1.85	**0.02**
Femur	-0.32 ± 1.44	-0.01 ± 1.35	0.29 ± 2.11	0.55
Body composition				
BMI	28.8 ± 4.9	31.2 ± 5.1	2.01 ± 3.36	**0.001**
BMI SDS	1.39 ± 1.92	1.52 ± 1.55	0.09 ± 1.41	0.38

FMI	10.5 ± 3.6	11.9 ± 3.8	0.97 ± 2.43	**0.001**
FMI SDS	1.37 ± 1.41	2.14 ± 1.46	0.10 ± 0.90	0.43
FFMI	18.3 ± 3.2	19.1 ± 3.2	1.13 ± 1.87	**<0.001**
FFMI SDS	0.26 ± 1.62	1.06 ± 2.22	0.91 ± 1.39	**<0.001**
BF%	36.4 ± 11.6	37.5 ± 8.4	0.28 ± 9.37	0.15
BF% SDS	1.41 ± 1.06	1.62 ± 0.96	-0.02 ± 0.73	0.61

Data are expressed as mean ± SD. The **bold** values indicate that they are statistically significant ($P < 0.05$). Δ = mean difference; BF% = Body Fat Percentage; BMD = Bone Mineral Density; BMI = Body Mass Index; DXA = Dual-Energy X-ray Absorptiometry; FFMI = Fat Free Mass Index; FMI = Fat Mass Index; N= number.

Longitudinal change in BMD

BMD, and corresponding T- and Z-scores were slightly below average at baseline, but increased for total body and L2-L4, and the Z-score for femur neck as well (mean difference Z-scores 0.61 ± 1.12, $P < 0.001$; 0.74 ± 1.73, $P < 0.001$; 0.51 ± 1.85, $P = 0.02$, respectively) (Table 4 and Figure 3). There was no significant difference in (change in) Z-scores between patients with or without radiotherapy, with or without hypothalamic damage, with or without tumor progression, or CO or AO patients (Figure 3). There was a decrease in osteopenia rates from first to last DXA scan in the entire group if it defined by low T-score (38 [51%] vs. 27 [36%], $P = 0.04$) but no difference in proportion of osteoporosis at first (19 [24%]) or last (20 [24%]) DXA scan ($P = 1.00$) (Table 3). Patients treated for CP before 2000 had lower total body Z-scores at first (–0.77 ± 1.30 vs. 0.11 ± 1.36, $P = 0.02$) and last (–0.22 ± 1.48 vs. 0.51 ± 1.24, $P = 0.04$) DXA scan than patients treated from 2000 and onwards. Other BMD-related values did not differ (data not shown).

Prodigy and DPXL

Using the Lunar Prodigy, almost all body composition results increased significantly, but not using DPXL.[40] The BMD results follow a similar pattern, except for femur neck: femur neck BMD, T-scores, and Z-scores significantly improved if measured using Lunar DPXL, in contrast to Lunar Prodigy.

Effect of prognostic factors for change in BMD Z-scores

Univariable and multivariable linear regression models were estimated to identify possible predictors of changes in total body Z-score, L2-L4 Z-score, and femur neck Z-score (Table 5).

In a univariable model, time from first to last DXA scan (beta 0.04 [0.003-0.08]), hydrocortisone equivalent dose (beta −0.06 [−0.10 to −0.02]) and medication to improve BMD (beta −0.95 [1.83 to −0.08]) were associated with change in total body Z-score. In the multivariable analysis, time from first to last DXA scan was not associated with change in total body Z-score (beta 0.03 [−0.005 to 0.07]) while hydrocortisone equivalent dose was (beta −0.06 [−0.09 to −0.02]). In a univariable model, medication to improve BMD and progression were not associated with change in L2-L4 Z-score (beta −1.30 [−2.78 to 0.18] and beta 0.86 [0.02-1.73], respectively). In multivariable analysis, medication to improve BMD was associated with change in L2-L4 Z-score (beta −1.06 [−1.84 to −0.28]), while tumor progression was not (beta 0.04 [−0.45 to 0.52]). In the univariable model for change in femur neck Z-score, GHRT (beta 1.45 [0.51-2.39]) and previous radiotherapy (beta −0.79 [−1.49 to −0.09]) were significant explanatory variables. In multivariable analysis, GHRT and radiotherapy were however not identified as significant prognostic factors (with corresponding estimated regression coefficients of beta −0.37 [−1.27 to 0.53]) and −0.05 [−0.67 to 0.57]).

Table 5 Linear regression model: univariable model and multivariable models of change in Z-scores of BMD

	Univariable linear regression model					
	Δ Total body Z-score		Δ L2-L4 Z-score		Δ Femur neck Z-score	
	Beta	95%CI	Beta	95%CI	Beta	95%CI
Time from first to last DXA	0.04	0.003 – 0.08	0.02	-0.04 – 0.09	-0.02	-0.08 – 0.04
Time since first treatment at last DXA	0.01	-0.01 – 0.03	-0.01	-0.04 – 0.03	0.001	-0.03 – 0.03
Female sex	-0.06	-0.57 – 0.44	0.05	-0.85 – 0.95	0.47	-0.26 – 1.19
BMI at last DXA	0.03	-0.03 – 0.08	-0.002	-0.09 – 0.08	0.05	-0.03 – 0.12
BMD medication	-0.95	-1.83 – -0.08	-1.30	-2.78 – 0.18	-0.67	-2.08 – 0.74
Adulthood onset	0.35	-0.38 – 1.08	0.93	-0.36 – 2.22	-0.30	-1.32 – 0.73
Hypothalamic damage	0.27	-0.31 – 0.85	-0.06	-1.06 – 0.95	-0.18	-1.04 – 0.68
Growth hormone replacement therapy	0.40	-0.48 – 1.27	1.03	-0.31 – 2.38	1.45	0.51 – 2.39
Gonadal replacement therapy	0.49	-0.13 – 1.10	0.64	-0.38 – 1.66	-0.17	-1.01 – 0.68
Hydrocortisone equivalent dose	-0.06	-0.10 – -0.02	-0.03	-0.10 – 0.05	0.05	-0.03 – 0.12
Epilepsy medication	0.78	-0.01 – 1.57	0.15	-1.24 – 1.53	-0.14	-1.24 – 0.96
Ever hydrocephalus	-0.16	-0.75 – 0.44	0.06	-1.01 – 1.12	0.18	-0.73 – 1.09
Progression	0.31	-0.21 – 0.83	0.86	-0.02 – 1.73	-0.16	-0.89 – 0.59
Surgery	-0.29	-2.34 – 1.76	0.71	-2.77 – 4.18	1.11	-1.58 – 3.79
Radiotherapy	-0.11	-0.61 – 0.39	0.14	-0.72 – 1.01	-0.79	-1.49 – -0.09
Yttrium	-0.32	-1.19 – 0.55	-0.05	-1.85 – 1.76	-0.22	-1.62 – 1.18

	Multivariable linear regression model					
	Δ Total body Z-score R^2=0.23		Δ L2-L4 Z-score R^2=0.15		Δ Femur neck Z-score R^2=0.65	
	Beta	95%CI	Beta	95%CI	Beta	95%CI
Time from first to last DXA	0.03	-0.005 – 0.07				
Time since first treatment at last DXA						
Female sex						
BMI at last DXA						
BMD medication			-1.06	-1.84 – -0.28		
Adulthood onset						
Hypothalamic damage						
Growth hormone replacement therapy					-0.37	-1.27 – 0.53
Gonadal replacement therapy						
Hydrocortisone equivalent dose	-0.06	-0.09 – -0.02				
Epilepsy medication						
Ever hydrocephalus						
Progression			0.04	-0.45 – 0.52		
Surgery						
Radiotherapy					-0.05	-0.67 – 0.57
Yttrium						

All models are corrected for baseline Z-score and age at first DXA-scan. Hydrocortisone dose is a hydrocortisone equivalent dose (mg). Abbreviations: BMD medication = medication to improve bone mineral density; CI = Confidence Interval; DXA = Dual-Energy X-ray Absorptiometry scan; BMI = Body Mass Index; *P* = *P*-value.

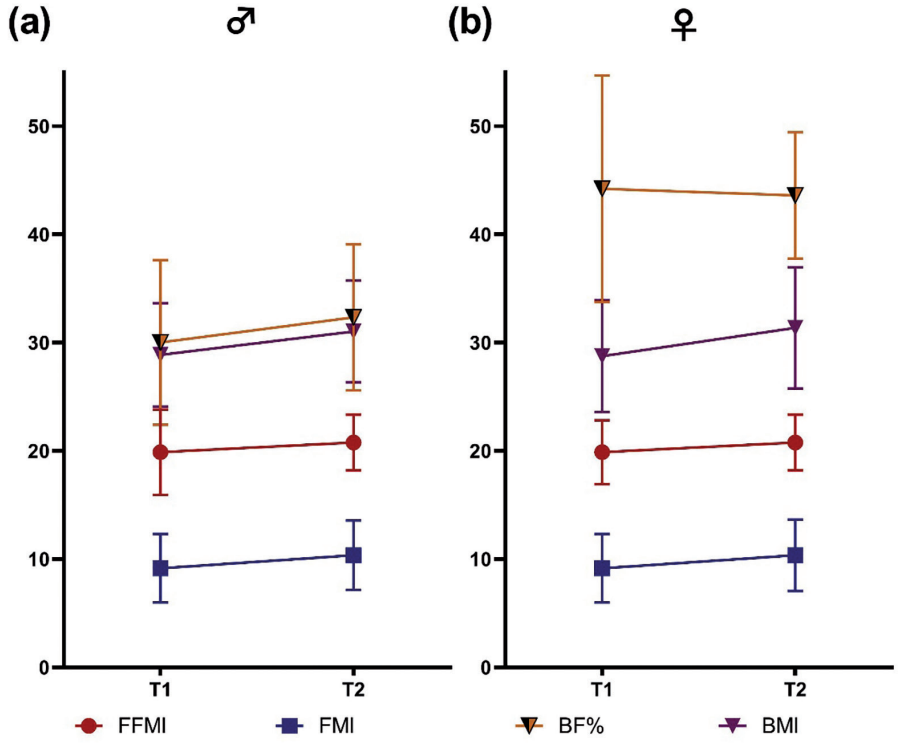
Discussion

This is the first study assessing longitudinally changes in 86 patients with CP, comparing anthropometric and BMD data, with a follow-up of 10 years. In addition, we investigated prognostic factors for BMD change.

Body composition improved in patients with CP during follow-up: FFMI increased and was relatively high, implying an improvement in muscle mass and bone mass, and suggesting that there was hardly any sign of sarcopenic obesity. This increase in age-standardized FFMI was peculiar, since in the general population FFMI usually decreases with advancing age.[41, 42] One may suggest that this increase in muscle and bone mass might be related to physical functioning. However, it is unclear whether gain in muscle mass was a reflection of improved physical performance or sufficient hormonal replacement therapy. In the elderly general population, impaired physical activity performance was associated with FMI rather than with FFMI,[43] and in the male general population, FFMI had a U-shaped relationship with mortality.[44] It is unknown whether the increase in FFMI in our study population affected the mortality in any way. We observed an increase in unstandardized values of body mass and fat-related measures, but not in the age- and sex-specific scores, suggesting that patients did gain fat and total weight with aging, similarly as the general population does. Loss of fat mass is a desired goal of treatment, as obesity and the metabolic syndrome occur very frequently in patients with CP, making improvement of their cardiovascular risk profile an enormous challenge.[9] Both physical activity level and basal metabolic rate are usually low in patients with CP, regardless of fat free mass.[16, 23] Patients may already gain weight in the first year after onset and even before diagnosis.[19] This underlines the difficulty of developing an effective strategy in patients with CP to prevent the development of obesity. Thus, anti-obesity treatment should ideally be started as soon as possible and preferably directly after confirmation of the CP diagnosis.

Interestingly, bone density of the total body, femur neck and L2-L4 improved during follow-up. Independent risk factors for a decrease in bone density scores were glucocorticoids, previous radiotherapy, and,

Figure 1 Body composition measures men and women with craniopharyngioma



Unstandardized mean and SD of fat free mass index (FFMI; red circles), fat mass index (FMI; blue squares), body fat percentage (BF%; black/orange triangles) and body mass index (BMI; purple triangles) are shown at the first Dual-Energy X-ray Absorptiometry (DXA-) scan (T1) and last DXA-scan (T2) in male (A) and female (B) patients with craniopharyngioma. As expected, men had significantly lower FMI values than women at T1 and T2 ($P \leq 0.001$), lower BF% ($P < 0.001$), and higher FFMI ($P < 0.001$); there was no difference in BMI or in changes from T1 to T2. There was a significant increase from T1 to T2 in all patients in BMI ($P < 0.001$), FMI ($P < 0.001$) and FFMI ($P < 0.001$), but not BF% ($P = 0.15$).

remarkably, medication given to improve BMD. It is well known that excessive glucocorticoids can reduce bone mass.[13] Cranial radiotherapy is not only an important risk for pituitary deficiencies and hypothalamic dysfunction, but also for obesity.[45] The authors hypothesized that the improvement in BMD in our cohort was explained by the high obesity rates in patients with CP, as obesity can induce an increase in bone mass and is generally considered to be a protective factor for osteoporosis.[43]

Factors involved in this increase of bone mass in obesity are augmented leptin levels and an increased peripheral conversions of androgens to estrogens.[21] However, in our study, BMI was not associated with bone density change. This is in line with new findings on obesity, if implicated with low-grade inflammation, does not necessarily concur with benefits on bone mass.[43] Obesity may cause a state of leptin resistance.[43] Central leptin resistance or insensitivity due to hypothalamic damage could mediate negative effects of obesity on bone metabolism.[43]

The unexpected finding that medication administered to improve BMD was associated with a decrease in BMD in our study might be due to bias by indication: this medication was likely to be given to patients who already had developed a poor bone health. Administration of GHRT was found to be associated with an increase in bone density. This is consistent with previous literature which describes that discontinuation of GHRT in CO patients before establishing peak bone mass in the first 3 decades of life may be associated with a high prevalence of pathological bone densities.[21]

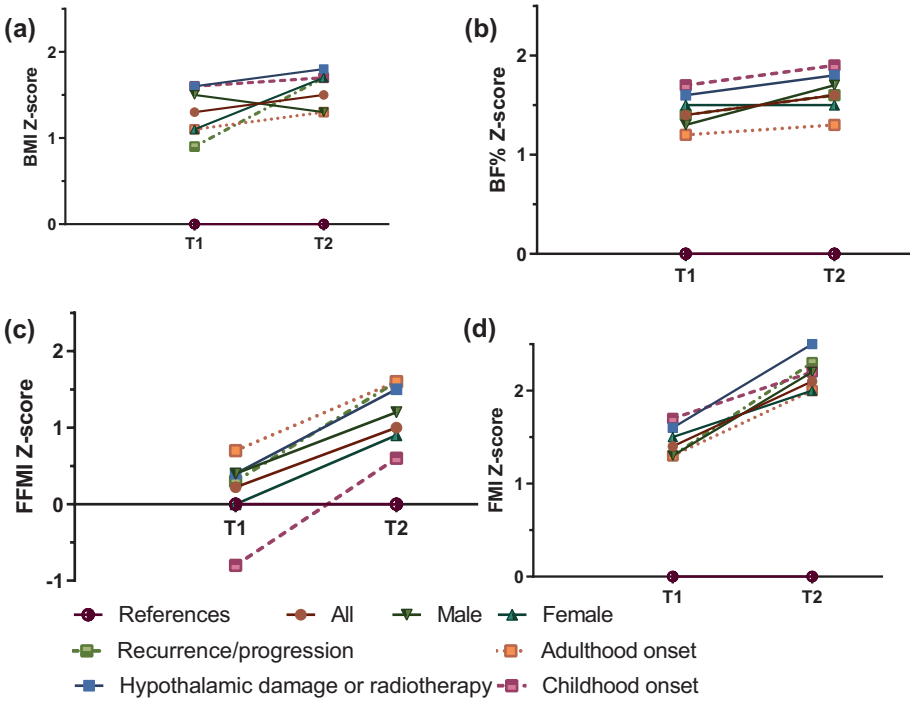
CO patients and patients with previous radiotherapy or hypothalamic damage seemed to be at higher risk for a less favorable body composition than their counter-subgroup. CO patients showed more increase in body mass, fat mass, and fat percentage, and less muscle mass at baseline than AO patients. This may have contributed to their previously found excess risk of diabetes mellitus type 2, cerebral infarction and total mortality,[4] and of morbid obesity, and be related to the higher risk of hypothalamic involvement and panhypopituitarism of CO patients.[24] In addition, the increase in FFMI Z-scores may be, at least partly, related to GHRT.[28, 46, 47] Treatment of CP has changed from gross total resection to more conservative surgery,[24] which could affect body composition and BMD. However, patients treated before and from 2000 onwards almost did not differ, except for total body Z-score, follow-up time, and cohort size.

There are some limitations in this research due to the retrospective design. As discussed before,[8,9] differences in observation can be perceived from different types of DXA-scanners; in 15 out of 198 scan-

results, the scanner type was unknown due to a time period where both DPXL and Prodigy were used. We coped with this aspect by also analyzing data separately per DXA-scanner for the most often used scanner types, and using Z-scores. This study is unique in reporting factors on BMD change in patients with CP, and it is one among few studies reporting longitudinal change in body composition, and represents a large cohort with an extensive follow-up.

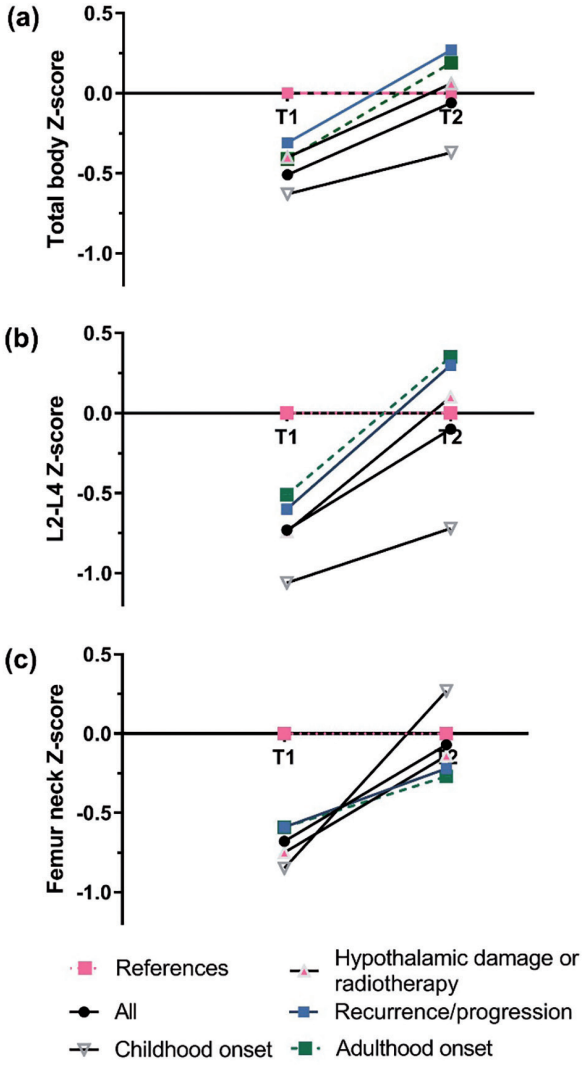
Summarizing, patients with CP remained as stable in body composition as the general population over time, and even showed an increase in bone and muscle mass. Higher glucocorticoid dose and central nervous system irradiation negatively influenced this increase of BMD, while GHRT shows the opposite and it is associated with an increase in bone density. Even though these results are beneficial at glance, there still remains considerable need for improvement, as cardiovascular risk and fracture rates remain unacceptably high in patients with CP.[4,8] In addition, subgroups of CP patients like those with CO disease and hypothalamic damage remain at high risk for adverse anthropometric effects. Future studies among patients with CP should therefore focus on development of methods for early prevention of obesity, interventions to influence body composition and improvement of bone quality in affected individuals, already from the diagnostic phase.

Figure 2 BMI Z-scores at first and last DXA-scan



Sex- and age standardized Z-scores are presented of (A) body mass index (BMI), (B) body fat percentage (BF%), (C) fat free mass index (FFMI) and (D) fat mass index, at first (T1) and last (T2) Dual-Energy X-ray Absorptiometry (DXA-) scan. Data are given as mean ± standard deviation. Male and female patients did not differ in body composition Z-scores. Patients with recurrence or progression of disease increased in BMI Z-scores while patients without recurrence or progression decreased (mean change 0.7 ± 1.3 vs. -0.3 ± 1.4, $P = 0.04$). Patients with hypothalamic damage or radiotherapy had higher BMI Z-scores at T1 (1.7 ± 1.9 vs. 0.7 ± 1.7, $P = 0.004$), T2 (1.8 ± 1.7 vs. 1.0 ± 1.2, $P = 0.01$), higher BF% Z-scores at T1 (1.6 ± 1.0 vs. 1.1 ± 1.2, $P = 0.03$), T2 (1.8 ± 0.8 vs. 1.3 ± 1.1, $P = 0.03$) and differed in change of BF% Z-scores from T1 to T2 (0.1 ± 0.7 vs. -0.3 ± 0.8, $P = 0.02$). They also had a higher FMI Z-scores at T1 (1.6 ± 1.5 vs. 1.0 ± 1.2, $P = 0.06$) and T2 (2.5 ± 1.4 vs. 1.4 ± 1.3, $P = 0.007$), and FFMI Z-scores at T2 (1.5 ± 2.6 vs. 0.3 ± 1.4, $P = 0.03$). Childhood onset patients had higher BF% Z-scores at T2 (1.9 ± 0.8 vs. 1.3 ± 1.0, $P = 0.02$) than adulthood onset patients and increased instead of decreased in BF% Z-scores from T1 to T2 (mean difference 0.2 ± 0.8 vs. -0.3 ± 0.6, $P = 0.04$). They also had lower FFMI Z-scores at T1 (-0.8 ± 1.4 vs. 0.7 ± 1.4, $P < 0.001$) and borderline lower FFMI Zscores at T2 (0.6 ± 2.4 vs. 1.6 ± 2.0, $P = 0.07$).

Figure 3 Total body, L2-L4 and femur neck Z-scores of the first and last DXA-scan



Z-scores at first (T1) and last (T2) Dual-Energy X-ray Absorptiometry (DXA-) scan in all craniopharyngioma patients of (A) total body, (B) L2-L4 and (C) femur neck. Data is given as mean ± standard deviation. Z-scores improved significantly in all patients from T1 to T2 of total body (from -0.49 ± 1.37 to -0.05 ± 1.44, $P < 0.001$), L2-L4 (from -0.77 ± 1.59 to -0.03 ± 1.97, $P < 0.001$) and femur neck (from -0.62 ± 1.32 to -0.09 ± 1.33, $P = 0.02$). There was no significant difference in Z-scores between patients with or without radiotherapy/recurrence or progression/hypothalamic damage or patients with childhood- or adulthood onset disease, at T1, T2 or change from T1 to T2.

References

1. Müller HL. Craniopharyngioma. Handb Clin Neurol. 2014;124:235-253.
2. Bunin GR, Surawicz TS, Witman PA, Preston-Martin S, Davis F, Bruner JM. The descriptive epidemiology of craniopharyngioma. J Neurosurg. 1998;89(4):547-551.
3. Olsson DS, Andersson E, Bryngelsson IL, Nilsson AG, Johannsson G. Excess mortality and morbidity in patients with craniopharyngioma, especially in patients with childhood onset: a population-based study in Sweden. J Clin Endocrinol Metab. 2015;100(2):467-474.
4. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. Excess morbidity and mortality in patients with craniopharyngioma: a hospital-based retrospective cohort study. Eur J Endocrinol. 2018;178(1):93-102.
5. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. The metabolic syndrome and its components in 178 patients treated for craniopharyngioma after 16 years of follow-up. Eur J Endocrinol. 2018;178(1):11-22.
6. Bülow B, Hagmar L, Mikoczy Z, Nordström CH, Erfurth EM. Increased cerebrovascular mortality in patients with hypopituitarism. Clin Endocrinol (Oxf). 1997;46(1):75-81.
7. Tomlinson JW, Holden N, Hills RK, et al. Association between premature mortality and hypopituitarism. West Midlands Prospective Hypopituitary Study Group. Lancet. 2001;357(9254):425-431.
8. van Santen SS, Olsson DS, van den Heuvel-Eibrink MM, et al. Fractures, bone mineral density, and final height in craniopharyngioma patients with a follow-up of 16 years. J Clin Endocrinol Metab. 2020;105(4):e1397-e1407.
9. van Santen SS, Olsson DS, Hammarstrand C, et al. Diagnosing metabolic syndrome in craniopharyngioma patients: body composition versus BMI. Eur J Endocrinol. 2019;181(2):173-183.
10. Lamas Oliveira C. Metabolic consequences of craniopharyingioma and their management. Endocrinol Nutr. 2013;60(9):529-534.
11. Chung S. Body mass index and body composition scaling to height in children and adolescent. Ann Pediatr Endocrinol Metab. 2015;20(3):125-129.
12. Nederlands Centrum Jeugdgezondheid, Utrecht. Richtlijn overgewicht. Vol 20182012:Guidelines for obesity for children in the Dutch population. https://www.ncj.nl/richtlijnen/alle-richtlijnen/richtlijn/overgewicht
13. Mazziotti G, Frara S, Giustina A. Pituitary diseases and bone. Endocr Rev. 2018;39(4):440-488.

14. Srinivasan S, Ogle GD, Garnett SP, Briody JN, Lee JW, Cowell CT. Features of the metabolic syndrome after childhood craniopharyngioma. J Clin Endocrinol Metab. 2004;89(1):81-86.

15. Profka E, Giavoli C, Bergamaschi S, et al. Analysis of shortand long-term metabolic effects of growth hormone replacement therapy in adult patients with craniopharyngioma and non-functioning pituitary adenoma. J Endocrinol Invest. 2015;38(4):413-420.

16. Kim RJ, Shah R, Tershakovec AM, et al. Energy expenditure in obesity associated with craniopharyngioma. Childs Nerv Syst. 2010;26(7):913-917.

17. Nagasaki K, Tsumanuma I, Yoneoka Y, et al. Metabolic effects of growth hormone replacement in two pediatric patients with growth without growth hormone. Endocr J. 2010;57(9):771-775.

18. Müller HL, Bueb K, Bartels U, et al. Obesity after childhood craniopharyngioma–German multicenter study on pre-operative risk factors and quality of life. Klin Padiatr. 2001;213(4):244-249.

19. Müller HL, Emser A, Faldum A, et al. Longitudinal study on growth and body mass index before and after diagnosis of childhood craniopharyngioma. J Clin Endocrinol Metab. 2004;89(7):3298-3305.

20. Khan MJ, Humayun KN, Donaldson M, Ahmed SF, Shaikh MG. Longitudinal changes in body mass index in children with craniopharyngioma. Horm Res Paediatr. 2014;82(6): 372-379.

21. Andereggen L, Hess B, Andres R, et al. A ten-year follow-up study of treatment outcome of craniopharyngiomas. Swiss Med Wkly. 2018;148:w14521.

22. Ahmet A, Blaser S, Stephens D, Guger S, Rutkas JT, Hamilton J. Weight gain in craniopharyngioma–a model for hypothalamic obesity. J Pediatr Endocrinol Metab. 2006;19(2): 121-127.

23. Holmer H, Pozarek G, Wirf lt E, et al. Reduced energy expenditure and impaired feeding-related signals but not high energy intake reinforces hypothalamic obesity in adults with childhood onset craniopharyngioma. J Clin Endocrinol Metab. 2010;95(12):5395-5402.

24. Wijnen M, van den Heuvel-Eibrink MM, Janssen JAMJL, et al. Very long-term sequelae of craniopharyngioma. Eur J Endocrinol. 2017;176(6):755-767.

25. Müller HL, Faldum A, Etavard-Gorris N, et al. Functional capacity, obesity and hypothalamic involvement: cross-sectional study on 212 patients with childhood craniopharyngioma. Klin Padiatr. 2003;215(6):310-314.

26. Müller HL, Gebhardt U, Etavard-Gorris N, Kolb R, Warmuth-Metz M, S rensen N. Current strategies in diagnostics and endocrine treatment of patients with childhood craniopharyngioma during follow-up–recommendations in KRANIOPHARYNGEOM 2000. Onkologie. 2005;28(3):150-156.

27. Müller HL, Gebhardt U, Teske C, et al.; Study Committee of KRANIOPHARYNGEOM 2000. Post-operative hypothalamic lesions and obesity in childhood craniopharyngioma: results of the multinational prospective trial KRANIOPHARYNGEOM 2000 after 3-year follow-up. Eur J Endocrinol. 2011;165(1): 17-24.

28. Kendall-Taylor P, Jönsson PJ, Abs R, et al. The clinical, metabolic and endocrine features and the quality of life in adults with childhood-onset craniopharyngioma compared with adult-onset craniopharyngioma. Eur J Endocrinol. 2005;152(4): 557-567.

29. Blokstra A, Vissink P, Venmans L, et al. Measuring the Netherlands. A monitoring study of risk factors in the general population, 2009– 2010. 2012. Contract No.: RIVM Rapport 260152001/260152011. https://www.rivm.nl/publicaties/nederland-maat-genomen-2009-2010-monitoring-van-risicofactoren-in-al-gemene-bevolking

30. Alwis G, Rosengren B, Stenevi-Lundgren S, Düppe H, Sernbo I, Karlsson MK. Normative dual energy X-ray absorptiometry data in Swedish children and adolescents. Acta Paediatr. 2010;99(7):1091-1099.

31. Berg C, Rosengren A, Aires N, et al. Trends in overweight and obesity from 1985 to 2002 in G teborg, West Sweden. Int J Obes (Lond). 2005;29(8):916-924.

32. Fredriks AM, van Buuren S, Wit JM, Verloove-Vanhorick SP. Body index measurements in 1996-7 compared with 1980. Arch Dis Child. 2000;82(2):107-112.

33. Gavriilidou NN, Pihlsgård M, Elmst hl S. Anthropometric reference data for elderly Swedes and its disease-related pattern. Eur J Clin Nutr. 2015;69(9):1066-1075.

34. Larsson I, Lissner L, Samuelson G, et al. Body composition through adult life: Swedish reference data on body composition. Eur J Clin Nutr. 2015;69(7):837-842.

35. van der Sluis IM, de Ridder MA, Boot AM, Krenning EP, de Muinck Keizer-Schrama SM. Reference data for bone density and body composition measured with dual energy x ray absorptiometry in white children and young adults. Arch Dis Child. 2002;87(4):341-7; discussion 341.

36. Gültekin T, Ozer BK, Akin G, Bektaş Y, Sağir M, Güle E. Prevalence of overweight and obesity in Turkish adults. Anthropol Anz. 2009;67(2):205-212.

37. Kelly TL, Wilson KE, Heymsfield SB. Dual energy x-ray absorptiometry body composi-tion reference values from NHANES. PLOS One. 2009;4(9):e7038.

38. Perreault L. Determining body composition in adults. In: Pi-Sunyer F, Sullivan D, eds. Vol 2018. Waltham, MA: UpToDate; 2017.

39. Cummings SR, Bates D, Black DM. Clinical use of bone densitometry: scientific review. JAMA. 2002;288(15):1889-1897.

40. van Santen SS, Olsson DS, Hammarstrand C, et al. Body composition and bone mineral density in craniopharyngioma patients: a longitudinal study over 10 years: supplementary tables. Figshare; 2020. doi:10.6084/m9.figshare.12562217.v2. Posted August 21, 2020. https://figshare.com/articles/journal_contribution/20200623_longitudinal_article_supplementary_tables_docx/12562217

41. Ramírez Torres M, Ruiz Valenzuela RE, Esparza-Romero J, López Teros MT, Alemán-Mateo H. The fat mass index, not the fat-free mass index, is associated with impaired physical performance in older adult subjects: Evidence from a cross-sectional study. Clin Nutr. 2019;38(2):877-882.

42. Hull HR, Thornton J, Wang J, et al. Fat-free mass index: changes and race/ethnic differences in adulthood. Int J Obes (Lond). 2011;35(1):121-127.

43. Upadhyay J, Farr OM, Mantzoros CS. The role of leptin in regulating bone metabolism. Metabolism. 2015;64(1): 105-113.

44. Lee DH, Keum N, Hu FB, et al. Predicted lean body mass, fat mass, and all cause and cause specific mortality in men: prospective US cohort study. BMJ. 2018;362:k2575. https://pubmed.ncbi.nlm.nih.gov/29970408/

45. Lustig RH. Hypothalamic obesity after craniopharyngioma: mechanisms, diagnosis, and treatment. Front Endocrinol (Lausanne). 2011;2:60. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3356006/

46. Gasco V, Caputo M, Lanfranco F, Ghigo E, Grottoli S. Management of GH treatment in adult GH deficiency. Best Pract Res Clin Endocrinol Metab. 2017;31(1):13-24.

47. Giagulli VA, Castellana M, Perrone R, Guastamacchia E, Iacoviello M, Triggiani V. GH supplementation effects on cardiovascular risk in GH deficient adult patients: a systematic review and meta-analysis. Endocr Metab Immune Disord Drug Targets. 2017;17(4):285-296.

Supplementary Tables

Supplementary Table 1A BMI and FFMI cutoffs

	Low	Normal	High/excess fat	Obese
BMI	<20 kg/m^2	20-24,99 kg/m^2	25-29,99 kg/m^2 (overweight)	≥ 30 kg/m^2 or ≥ 2 SDS
FFMI	≤ -2 SDS	-2 – 2 SDS	≥ 2 SDS	
FMI (M / F)	<3 / <5	3 – 6 / 5 – 9	>6 – 9 / >9 – 13	>9 / >13

Abbreviations: BMI = body mass index; FFMI = fat free mass index; FMI = fat mass index; M / F = Male / Female references; SDS = standardized deviation score. High BMI can be interpreted as an overweight individual. FMI is expressed as fat mass per squared height (kg/m^2).

Supplementary Table 1B BF% cutoffs

	Age 20-39 years		Age 40-59 years	
Sex	**Men**	**Women**	**Men**	**Women**
BF% category				
Low	≤8% or SDS-score ≤-2	≤21% or SDS-score ≤-2	≤11% or SDS-score ≤-2	≤23% or SDS-score ≤-2
Normal	8-26%	21-39%	11-29%	23-41%
High	≥26% or SDS ≥2	≥39% or SDS ≥2	≥29% or SDS ≥2	≥41% or SDS ≥2

Cutoff-values for low, normal and BF%.[1] Abbreviations: BF% = body fat percentage; SDS = standardized deviation scores.

Supplementary Table 2 Patients with craniopharyngioma diagnosed before and after 2000

	Diagnosis before 2000 (n=78)	Diagnosis after 2000 (n=32)	*P*-value
Female sex	38 (49%)	14 (44%)	0.64
Median age at first treatment (range)	20.0 (4 – 62)	9 (6 – 74)	0.08
Childhood onset disease	40 (51%)	10 (31%)	0.06
Surgery	77 (99%)	31 (97%)	1.00
Radiotherapy	43 (55%)	19 (59%)	0.68
Yttrium brachytherapy	11 (14%)	2 (7%)	0.31
Visual deficiencies	56 (77%)	25 (78%)	0.87
Recurrence/progression	28 (36%)	12 (38%)	0.91
Hypothalamic damage	22 (29%)	14 (48%)	0.08
Hydrocephalus	16 (21%)	10 (31%)	0.20
GH deficiency	73 (94%)	31 (97%)	0.49
TSH deficiency	75 (96%)	32 (100%)	0.63
Hypogonadism	71 (91%)	30 (94%)	0.93
Corticotropic deficiency	68 (87%)	27 (84%)	0.93
ADH deficiency	53 (68%)	23 (72%)	0.69
Fracture in history	14 (18%)	7 (22%)	0.63
BMD medication	14 (18%)	2 (6%)	0.20
Median time (years) since first presentation at first DXA	9.8 (0.8 – 57)	3.3 (-0.9 – 12.73)	**0.004**
Median age (years) at first DXA-scan (range)	35 (18-77)	32 (18 – 79)	0.57
Time difference from first to last DXA-scan (years)	12.4 ± 6.5	6.3 ± 4.2	**<0.001**
BMI at first DXA-scan	28.4 ± 4.7	30.0 ± 5.3	0.11
BMI at last DXA-scan	30.9 ± 4.6	32.2 ± 6.5	0.41
Change in BMI	2.3 ± 3.3	1.2 ± 3.4	0.20
BMI Z-score at first DXA-scan	1.2 ± 1.8	1.9 ± 2.1	0.05
BMI Z-score at last DXA-scan	1.4 ± 1.4	2.0 ± 1.89	0.10
Change in BMI Z-score	0.1 ± 1.4	0.0 ± 1.5	0.43

FMI at first DXA-scan	10.3 ± 3.3	11.1 ± 4.2	0.25
FMI at last DXA-scan	11.8 ± 3.7	12.4 ± 4.0	0.57
Change in FMI	1.2 ± 2.4	0.3 ± 2.4	0.24
FMI Z-score at first DXA-scan	1.3 ± 1.3	1.6 ± 1.6	0.21
FMI Z-score at last DXA-scan	2.1 ± 1.4	2.3 ± 1.6	0.60
Change in FMI Z-score	0.1 ± 1.0	0.2 ± 0.8	0.61
FFMI at first DXA-scan	18.1 ± 3.2	18.7 ± 3.2	0.46
FFMI at last DXA-scan	19.0 ± 3.0	19.5 ± 3.6	0.74
FFMI change	1.2 ± 1.9	0.9 ± 1.9	0.33
FFMI Z-score at first DXA-scan	0.1 ± 1.6	0.6 ± 1.7	0.37
FFMI Z-score at last DXA-scan	0.9 ± 2.2	1.4 ± 2.4	0.43
FFMI Z-score change	1.0 ± 1.4	0.6 ± 1.4	0.33
BF% at first DXA-scan	36.5 ± 12.0	36.3 ± 10.7	0.63
BF% at last DXA-scan	37.2 ± 8.5	38.5 ± 8.4	0.63
BF% change	0.2 ± 10.3	0.2 ± 5.6	0.47
BF% Z-score at first DXA-scan	1.3 ± 1.1	1.7 ± 0.7	0.06
BF% Z-score at last DXA-scan	1.6 ± 1.0	1.7 ± 0.8	0.71
BF% Z-score change	0.0 ± 0.8	-0.12 ± 0.57	0.62
Total body BMD at first DXA-scan	1.15 ± 0.12	1.21 ± 0.15	0.06
Total body BMD at last DXA-scan	1.18 ± 0.16	1.23 ± 0.14	0.31
Total body BMD change	0.05 ± 0.11	0.01 ± 0.08	0.23
Total body BMD T-score at first DXA-scan	-0.31 ± 1.45	0.50 ± 1.50	0.07
Total body BMD T-score at last DXA-scan	0.28 ± 1.71	1.00 ± 1.51	0.11
Total body BMD T-score change	0.76 ± 1.12	0.43 ± 0.99	0.38
Total body BMD Z-score at first DXA-scan	-0.77 ± 1.30	0.11 ± 1.36	**0.02**
Total body BMD Z-score at last DXA-scan	-0.22 ± 1.48	0.51 ± 1.24	**0.04**
Total body BMD Z-score change	0.71 ± 1.05	0.49 ± 0.91	0.94

L2-L4 BMD at first DXA-scan	1.13 ± 0.18	1.14 ± 0.22	0.73
L2-L4 BMD at last DXA-scan	1.21 ± 0.27	1.27 ± 0.21	0.19
L2-L4 BMD change	0.11 ± 0.17	0.13 ± 0.15	0.63
L2-L4 BMD T-score at first DXA-scan	-0.64 ±1.61	-0.85 ± 1.53	0.53
L2-L4 BMD T-score at last DXA-scan	-0.05 ± 2.21	0.43 ± 1.69	0.21
L2-L4 BMD T-score change	0.75 ± 1.70	1.04 ± 1.23	0.48
L2-L4 BMD Z-score at first DXA-scan	-0.65 ± 1.56	-0.91 ± 1.52	0.67
L2-L4 BMD Z-score at last DXA-scan	-0.19 ± 2.16	0.20 ± 1.58	0.23
L2-L4 BMD Z-score change	0.50 ± 1.91	1.21 ± 1.09	0.11
Femur neck BMD at first DXA-scan	0.95 ± 0.18	0.99 ± 0.20	0.21
Femur neck BMD at last DXA-scan	0.98 ± 0.23	1.03 ± 0.19	0.26
Femur neck BMD change	0.06 ± 0.27	0.00 ± 0.29	0.58
Femur neck BMD T-score at first DXA-scan	-0.69 ± 1.48	-0.46 ± 1.56	0.50
Femur neck BMD T-score at last DXA-scan	-0.44 ± 1.72	-0.26 ± 1.10	0.37
Femur neck BMD T-score change	0.42 ± 1.97	-0.25 ± 1.82	0.52
Femur neck BMD Z-score at first DXA-scan	-0.67 ±1.31	-0.47 ± 1.40	0.71
Femur neck BMD Z-score at last DXA-scan	-0.10 ± 1.41	-0.06 ± 1.02	0.81
Femur neck BMD Z-score change	0.69 ± 1.83	-0.06 ± 1.88	0.34

Data are given as number and percentage, or mean + SD unless stated otherwise. BF% = body fat percentage; BMD = bone mineral density; BMI = body mass index; DXA-scan = Dual-Energy X-ray Absorptiometry Scan; FFMI = fat free mass index; FMI = fat mass index.

Supplementary Table 3A DXA-results of any DXA-scanner type – Bone mineral Density and Body Composition in male craniopharyngioma patients

	First DXA-scan	Last DXA-scan	Δ	*P*-value
Age (years)	38.2 ± 14.6	49.5 ± 18.2	10.8 ± 6.7	**<0.001**
Bone				
Bone mineral density				
Total body	1.21 ± 0.14	1.24 ± 0.15	0.05 ± 0.09	**0.002**
L2-L4	1.16 ± 0.20	1.31 ± 0.27	0.14 ± 0.20	**<0.001**
Femur neck	0.97 ± 0.21	0.96 ± 0.20	-0.03 ± 0.29	0.41
Femur	1.05 ± 0.23	1.02 ± 0.20	-0.05 ± 0.32	0.28
T-score				
Total body	-0.06 ± 1.74	0.48 ± 1.74	0.72 ± 1.03	**<0.001**
L2-L4	0.60 ± 1.61	0.55 ± 2.21	0.91 ± 2.00	**0.003**
Femur neck	-0.80 ± 1.71	-0.68 ± 1.49	0.02 ± 2.20	0.71
Femur	-0.30 ± 1.80	-0.28 ± 1.46	0.02 ± 2.42	0.91
Z-score				
Total body	-0.54 ± 1.54	-0.02 ± 1.59	0.56 ± 1.30	**<0.001**
L2-L4	-0.52 ± 1.65	0.33 ± 2.23	0.60 ± 2.32	**0.03**
Femur neck	-0.69 ± 1.51	-0.31 ± 1.21	0.25 ± 2.09	0.48
Femur	-0.33 ± 1.66	-0.19 ± 1.31	0.03 ± 2.25	0.81
Body composition				
BMI	28.9 ± 4.7	31.0 ± 4.6	1.58 ± 2.50	**0.001**
BMI SDS	1.63 ± 2.28	1.36 ± 1.51	-0.26 ± 1.38	0.73
FMI	9.1 ± 3.2	10.4 ± 3.2	0.83 ± 2.33	**0.03**
FMI SDS	1.25 ± 1.50	2.24 ± 1.44	0.25 ± 0.92	0.25
FFMI	19.9 ± 2.9	20.8 ± 2.5	0.99 ± 1.93	**0.002**
FFMI SDS	0.45 ± 1.53	1.20 ± 1.79	0.81 ± 1.22	**0.009**
BF%	29.9 ± 7.8	32.4 ± 6.7	1.31 ± 6.42	0.29
BF% SDS	1.34 ± 1.15	1.72 ± 0.90	0.06 ± 0.89	0.91

Δ = mean difference; BF% = Body Fat Percentage; BMD = Bone Mineral Density; BMI = Body Mass Index; DXA = Dual-Energy X-ray Absorptiometry; FFMI = Fat Free Mass Index; FMI = Fat Mass Index; SDS = Standardized Deviation Scores. Data are expressed as mean ± SD.

Supplementary Table 3B DXA-results of any DXA-scanner type – Bone mineral Density and Body Composition in female craniopharyngioma patients

	First DXA-scan	Last DXA-scan	Δ	*P*-value
Age (years)	35.3 ± 15.2	46.4 ± 14.6	11.3 ± 6.4	**<0.001**
Bone				
Bone mineral density				
Total body	1.12 ± 0.09	1.13 ± 0.15	0.02 ± 0.11	0.24
L2-L4	1.10 ± 0.18	1.15 ± 0.22	0.08 ± 0.13	**0.003**
Femur neck	0.94 ± 0.14	1.02 ± 0.24	0.11 ± 0.23	**0.02**
Femur	0.98 ± 0.17	1.05 ± 0.23	0.16 ± 0.27	0.07
T-score				
Total body	-0.15 ± 1.16	0.38 ± 1.63	0.59 ± 1.13	**0.01**
L2-L4	-0.83 ± 1.52	-0.37 ± 1.89	0.65 ± 1.11	**0.01**
Femur neck	-0.43 ± 1.12	-0.16 ± 1.67	0.46 ± 1.63	0.15
Femur	-0.22 ± 1.26	0.06 ± 1.58	0.63 ± 1.84	0.21
Z-score				
Total body	-0.44 ± 1.13	-0.07 ± 1.26	0.66 ± 0.89	**0.001**
L2-L4	-0.95 ± 1.38	-0.47 ± 1.77	0.76 ± 0.97	**<0.001**
Femur neck	-0.52 ± 1.06	0.10 ± 1.41	0.80 ± 1.55	**0.007**
Femur	-0.29 ± 0.95	0.15 ± 1.38	0.74 ± 1.84	0.17
Body composition				
BMI	28.8 ± 5.2	31.4 ± 5.6	2.43 ± 4.03	**0.001**
BMI SDS	1.12 ± 1.39	1.67 ± 1.60	0.44 ± 1.38	0.11
FMI	12.3 ± 3.3	13.7 ± 3.6	1.14 ± 2.59	**0.01**
FMI SDS	1.52 ± 1.30	2.05 ± 1.50	-0.03 ± 0.87	0.96
FFMI	16.0 ± 1.8	17.2 ± 2.7	1.31 ± 1.81	**0.001**
FFMI SDS	0.05 ± 1.72	0.92 ± 2.58	1.00 ± 1.56	**0.006**
BF%	44.2 ± 10.5	43.6 ± 5.8	-0.97 ± 12.04	0.20
BF% SDS	1.50 ± 1.95	1.52 ± 1.02	-0.11 ± 0.53	0.37

Δ = mean difference; BF% = Body Fat Percentage; BMD = Bone Mineral Density; BMI = Body Mass Index; DXA = Dual-Energy X-ray Absorptiometry; FFMI = Fat Free Mass Index; FMI = Fat Mass Index; SDS = Standardized Deviation Scores. Data are expressed as mean ± SD.

Supplementary Table 4 DXA-results of the Prodigy – Body composition and bone mineral density in all craniopharyngioma patients

	First DXA-scan	Last DXA-scan	Δ	*P*-value
Body composition				
BMI	29.3 ± 5.5	31.2 ± 5.3	1.7 ± 3.1	**<0.001**
BMI SDS	1.9 ± 4.0	1.4 ± 1.6	0.2 ± 1.4	0.41
FMI	11.3 ± 3.6	12.3 ± 3.7	1.2 ± 2.2	**0.001**
FMI SDS	1.8 ± 1.4	2.2 ± 1.4	0.3 ± 0.8	**0.04**
FFMI	17.8 ± 2.8	18.8 ± 3.4	1.3 ± 1.7	**<0.001**
FFMI SDS	0.1 ± 1.7	1.3 ± 2.6	1.1 ± 1.3	**<0.001**
BF%	37.9 ± 8.1	39.7 ± 8.3	1.9 ± 4.8	**0.005**
BF% SDS	1.8 ± 1.2	1.9 ± 0.9	0.3 ± 0.7	**0.03**
Bone mineral density				
Total body	1.17 ± 0.14	1.21 ± 0.15	0.04 ± 0.08	**0.002**
L2-L4	1.16 ± 0.20	1.26 ± 0.25	0.08 ± 0.17	**0.004**
Femur neck	0.99 ± 0.18	1.01 ± 0.22	0.02 ± 0.24	0.79
Femur	1.03 ± 0.18	1.06 ± 0.21	0.03 ± 0.26	0.63
T-score				
Total body	0.00 ± 1.41	0.71 ± 1.63	0.56 ± 0.95	**<0.001**
L2-L4	-0.42 ± 1.65	0.27 ± 2.04	0.51 ± 1.65	0.02
Femur neck	-0.39 ± 1.45	-0.34 ± 1.64	0.05 ± 1.78	0.71
Femur	-0.21 ± 1.38	-0.02 ± 1.54	0.23 ± 1.85	0.72
Z-score				
Total body	-0.40 ± 1.30	0.13 ± 1.47	0.54 ± 0.85	**<0.001**
L2-L4	-0.48 ± 1.68	-0.03 ± 2.05	0.34 ± 1.78	0.08
Femur neck	-0.30 ± 1.27	0.00 ± 1.39	0.25 ± 1.74	0.26
Femur	-0.21 ± 1.30	0.11 ± 1.36	0.32 ± 1.80	0.32

Δ = mean difference; BF% = Body Fat Percentage; BMD = Bone Mineral Density; BMI = Body Mass Index; DXA = Dual-Energy X-ray Absorptiometry; FFMI = Fat Free Mass Index; FMI = Fat Mass Index; SDS = Standardized Deviation Scores.
Data are expressed as mean ± SD.

Supplementary Table 5 DXA-results of the DPXL – Body composition and bone mineral density in all craniopharyngioma patients

	First DXA-scan	Last DXA-scan	Δ	*P*-value
Body composition				
BMI	28.4 ± 5.1	28.7 ± 4.1	1.0 ± 2.5	0.06
BMI SDS	1.0 ± 1.3	1.0 ± 1.2	0.1 ± 1.1	0.22
FMI	10.7 ± 4.1	9.8 ± 3.6	-1.0 ± 3.9	0.21
FMI SDS	1.2 ± 1.3	1.2 ± 1.3	-0.2 ± 0.9	0.19
FFMI	18.0 ± 3.3	17.4 ± 4.4	-0.4 ± 3.4	0.61
FFMI SDS	0.3 ± 1.9	-0.1 ± 1.7	0.1 ± 0.8	0.77
BF%	35.5 ± 10.0	33.3 ± 9.9	-2.1 ± 5.0	**0.04**
BF% SDS	1.3 ± 1.3	1.2 ± 1.3	-0.4 ± 0.6	**0.001**
Bone mineral density				
Total body	1.14 ± 0.11	1.16 ± 0.11	0.03 ± 0.07	**0.005**
L2-L4	1.13 ± 0.16	1.23 ± 0.21	0.10 ± 0.09	**<0.001**
Femur neck	0.90 ± 0.13	0.97 ± 0.14	0.06 ± 0.08	**0.005**
T-score				
Total body	-0.29 ± 1.38	-0.14 ± 1.19	0.27 ± 0.87	0.07
L2-L4	-0.68 ± 1.39	0.14 ± 1.74	0.85 ± 0.77	**<0.001**
Femur neck	-1.04 ± 1.16	-0.46 ± 1.20	0.51 ± 0.96	**0.01**
Z-score				
Total body	-0.76 ± 1.16	-0.44 ± 1.26	0.45 ± 0.56	**0.001**
L2-L4	-0.72 ± 1.25	-0.12 ± 1.58	0.60 ± 0.88	**0.006**
Femur neck	-0.97 ± 1.08	-0.36 ± 0.98	0.60 ± 0.84	**0.002**

Δ = mean difference; BF% = Body Fat Percentage; BMD = Bone Mineral Density; BMI = Body Mass Index; DXA = Dual-Energy X-ray Absorptiometry; FFMI = Fat Free Mass Index; FMI = Fat Mass Index; SDS = Standardized Deviation Scores. Data are expressed as mean ± SD.

References

1. Perreault L. Determining body composition in adults. 2017; https://www-uptodate-com.eur.idm.oclc.org/contents/determining-bodycomposition-inadults?search=body%20composition%20adults&source=search_result&selectedTitle=1~116&usage_type=default&display_rank=1#H3702267332. Accessed July 4th, 2018, 2018.
2. Kelly TL, Wilson KE, Heymsfield SB. Dual Energy X-Ray Absorptiometry Body Composition Reference Values from NHANES. PLoS ONE. 2009;4(9):e7038.

5

Diagnosing Metabolic Syndrome in Craniopharyngioma Patients: Body Composition Versus BMI

Selveta S. van Santen,[1,2] Daniel S. Olsson,[3,4]
Casper Hammarstrand,[3,4] Mark Wijnen,[1]
Marry M. van den Heuvel-Eibrink,[1,2] Aart J. van der Lely,[1]
Gudmundur Johannsson,[3,4] Joseph A. M. J. L. Janssen,[1]
Sebastian J. C. M. M. Neggers.[1,2]

[1] Department of Internal Medicine, Endocrinology, Erasmus Medical Center, Rotterdam, The Netherlands;
[2]Princess Máxima Center for Pediatric Oncology, Utrecht, The Netherlands;
[3]Department of Internal Medicine, Endocrinology, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Göteborg, Sweden;
[4]Department of Internal Medicine and Clinical Nutrition, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Göteborg, Sweden.

European Journal of Endocrinology 2019

Abstract

Objective: Craniopharyngioma patients often have poor metabolic profiles due to hypothalamic–pituitary damage. Previously, using BMI as obesity marker, the occurrence of the metabolic syndrome in these patients was estimated at 46%. Our aim was to determine if dual-energy X-ray absorptiometry (DXA) scan in evaluation of obesity and metabolic syndrome would be superior.

Design: Retrospective study of craniopharyngioma patients for whom DXA scan results were available.

Methods: BMI, fat percentage and fat mass index were used to evaluate obesity and as components for obesity in metabolic syndrome.

Results: Ninety-five craniopharyngioma patients were included (51% female, 49% childhood-onset disease). Metabolic syndrome occurred in 34-53 (45-51%) subjects (depending on the definition of obesity, although all definitions occurred in higher frequency than in the general population). Metabolic syndrome frequency was higher if obesity was defined by fat percentage (52% vs. 42%) or fat mass index (51% vs. 43%) compared to BMI. Misclassification appeared in 9% (fat percentage vs. BMI) and 7% (fat mass index vs. BMI) for metabolic syndrome and 29% and 13% for obesity itself, respectively. For metabolic syndrome, almost perfect agreement was found for BMI compared with fat percentage or fat mass index. For obesity, agreement was fair to moderate (BMI vs. fat percentage).

Conclusion: Using BMI to evaluate obesity underestimates the true prevalence of metabolic syndrome in patients with craniopharyngioma. Furthermore, fat percentage contributes to a better evaluation of obesity than BMI. The contribution of DXA scan might be limited for identification of the metabolic syndrome.

Keywords: body composition, craniopharyngioma, metabolic syndrome, DXA-scan.

Introduction

Craniopharyngiomas (CPs) are tumors in the (supra)sellar region that occur both in children and adults, with peak incidences between ages 5–9 and 40–44 years.[1, 2] These tumors are rare with an the estimated incidence rate of 1.7 per million person years[1, 2] and relatively high 10-year survival rates at between 77 and 93%.[2, 3] Still, the burden to patients, their families[2, 4] and to the community is substantial, particularly because of the increased risk of long-term morbidity[2, 5, 6] and mortality compared to the general population.[2, 3, 5, 7] The standardized mortality ratio from cardio- and cerebrovascular disease has been reported to be between 3.2 and 19.4.[2, 6, 8]

Although histologically benign, CP can physically disturb important neurovascular structures,[9] particularly the hypothalamus, pituitary and optic nerves.[3, 9] The hypothalamus plays a major role in integrating hormonal, metabolic and neural signals for energy homeostasis.[7] Failure of the hypothalamo–pituitary axis is strongly linked with the development of obesity, metabolic syndrome[3, 7, 10, 11, 12] and subsequent cardiovascular disease.[8, 13] Visual impairment (i.e. due to tumor growth near the optic nerve) occurs in approximately 75% of the patients[3] and is a barrier to engage in physical activity.

Up to two-thirds of the patients are found to be overweight or obese,[3, 14] and in a study of 224 Dutch and Swedish patients, 9% has been diagnosed with type II diabetes mellitus (T2D).[5] Obesity and T2D are both associated with cardiovascular disease and cerebral infarction;[15, 16] the latter has been shown to be an important cause of mortality.[5] Severe weight gain is especially seen in the first year after the operation.[17] Both levels of physical activity and basal metabolic rate are low in patients with CP.[7, 18]

Obesity is an important contributor to the metabolic syndrome (MetS) in CP survivors.[6] Many definitions of MetS have been formulated, but all include obesity, dyslipidemia, hypertension and altered glucose metabolism as components.[6] MetS and T2D are two important risk factors for cardiovascular disease, giving a two-fold and five-fold

elevated risk respectively.[19] MetS is associated with an increase in all-risk mortality.[20] The prevalence of MetS has been estimated to be 46% in a previous study of CP survivors.[6] This is very high compared to the general population (15% in the Swedish and the 26% in Dutch populations of that age).[6] Obesity rates in the previous study were estimated based on BMI. BMI is the most accessible method for estimating obesity in a clinical setting, but does not take into account an altered body composition. Dual-Energy X-ray absorptiometry (DXA scan) is considered the gold standard for determining body composition where there is suspected discordance between BMI and adiposity.[21] Furthermore, compared with measurement of body fat percentage (BF%) by DXA scan, BMI has been found to underestimate obesity in adult, non-CP childhood cancer survivors.[22] Therefore, considering the relatively substantial contribution of adiposity to MetS in CP survivors, we hypothesized that the occurrence of MetS defined using BMI is underestimated in our cohort of CP patients due to their altered body composition.

Our aim was to investigate the value of DXA scan in cardiovascular risk assessment in CP patients. Our main objective was to compare the relative effectiveness of BMI and DXA scan-derived body composition measures such as BF% and fat mass index (FMI) in the evaluation of MetS. Our secondary objective was also to determine influencing factors of obesity and MetS in CP survivors. Our approach was to determine the agreement between methods and to perform logistic regression models.

Subjects and methods

Patients and study design

The study had a cross-sectional and retrospective design. Patients treated for CP at the Erasmus University Medical Center (Erasmus MC, Rotterdam, the Netherlands) and Sahlgrenska University Hospital (Gothenburg, Sweden) were included as described before.[6] Only patients with at least one DXA scan available were included in the study (included, n = 95; excluded, n = 61, description shown in Supplementary Table 1, see section on supplementary data given at the end of this

article). The decision to perform a DXA scan was made by the physicians in the context of general care. Although it was not possible to retrieve much information on why the decision was made, it may be assumed this was done to evaluate bone mineral density (BMD). If BMD was not of interest, bio-electrical impedance analysis measurements were an option in the Erasmus MC, which are less costly, easier to plan and do not involve any radiation. The most frequently used DXA scanners were Lunar DPXL, Lunar Prodigy and Lunar iDXA (see Supplementary Table 2). The study was approved by the Ethical Review Board of Erasmus MC and the Regional Ethical Review Board, Gothenburg University. All patients gave their informed consent.

Table 1 Definition of the metabolic syndrome: adjustments to the Joint Interim Statement[6]

At least three of the following five criteria:
Obesity defined by high body mass index (BMI), or defined by high body fat percentage (BF%), or defined by high fat mass index (FMI)
Fasting glucose ≥5.6 mmol/L (or corresponding HbA1c of ≥5.2% or 33.3 mmol/mol) or drug treatment for increased glucose
Triglycerides ≥1.7 mmol/L or drug treatment for elevated triglycerides
HDL cholesterol <1.0 mmol/L in males and <1.3 mmol/L in females or drug treatment for reduced HDL cholesterol
Blood pressure ≥130/85 mmHg or drug treatment for hypertension

HDL= High-density lipoprotein.
BMI was calculated by dividing the weight (kg) per squared height (meters). FMI was calculated by total fat mass (kg) per square height (meters).
Obesity (high BMI): defined by BMI >30, or by ≥2 SDS-score.
Obesity (high BF%): defined by high BF% with limits defined according to sex and age (for females as a BF% ≥39% if aged 20-39 years old or ≥41% between 40-59 years old, and for males ≥26% if 20-39 years old or ≥29% in the age category of 40-59), or by SDS-score ≥2).
Obesity (high FMI): defined by high FMI with limits defined according to sex and age (for adult females ≥ 13; for males ≥ 9).

Data collection

The methods and definitions for data collection were described previously.[6] Hypothalamic damage is defined as tumor- and/or treatment-related injury to the hypothalamus and/or third ventricle as visualized by neuroimaging.[6] If patients were treated for ACTH deficiency with cortisone acetate, the dose was converted to a hydrocortisone equivalent dose (described previously[23]). For the current study, the presence of MetS criteria was evaluated as close as possible (±3 years) to the last DXA scan. The results of DXA scan, including BMI, FMI and BF% were gathered. If patients had more than one scan, data from the first and last DXA scan were gathered.

The criteria used in the definition for MetS were according to the joint interim statement as described previously[6] with modification of the criterion of obesity and matching HbA1c values for high glucose (Table 1). MetS includes components of obesity, low high-density lipoprotein (HDL) cholesterol and high triglycerides, fasting glucose/HbA1c and blood pressure or pharmaceutical treatment (see below and Table 1). Three different definitions of obesity were used as criterion for MetS: based on high BMI, high BF% and high FMI. High BMI was defined as >30 kg/m^2 or SDS >2.[24, 25] High BF% was defined through total body fat percentage as measured by DXA; for females as a BF% ≥39% if aged 20–39 years old or ≥41% if aged 40–59 years old. For males, this was ≥26% if aged 20–39 years old or ≥29% if aged 40–59 years old, or by SDS-score ≥2.[21] High FMI (kg fat mass/m^2, measured by DXA scan) was defined for adult females as ≥13, for adult males ≥9[26] (Table 1). The MetS definitions are referred to as MetS (BMI), MetS (BF%) and MetS (FMI), since the obesity component is the only component varying. The cutoff value for BMI of >30 kg/m^2 was chosen, because this conforms with our previous research[6] and is an alternative for waist circumference according to the International Diabetes Federation (IDF) consensus[27] and World Health Organization (WHO) definition.[28]

Standardized deviation scores (SDS) of BMI, FMI and BF% were calculated with the corresponding reference groups.[29-40] For BF% in adults and FMI, data from a Swedish reference group was used.[29] If

the patient was of a different ethnic background from the majority, a reference group from the country of origin was used instead, if available. DXA data from any type of scanner were accepted. When the age of our subjects at the moment of DXA scan conformed to the reference population (30–70 for the Dutch and 45–69 years for the Swedish population), a comparison of occurrence of MetS at last DXA scan was made and a weighted mean was calculated based on the sex and country of origin.[30, 41] The same was done for the components of MetS; a comparison with the general population was made with two definitions for low HDL cholesterol and hypertension since the study (the NL de Maat study) used to compare the Dutch figures had different cutoff values and did not regard medication.[30]

Statistical analysis

All statistical analyses were performed using IBM SPSS Statistics for Windows, version 24.0 and Graphpad version 7.04. Unless stated otherwise, data are presented as mean and standard deviation (s.d.). Statistical significance was accepted at a $P \leq 0.05$. For normally distributed data, an unpaired T-test was used for group comparisons; Mann–Whitney U tests were used for non-normally distributed data. Comparisons of proportions were conducted using the Fisher's exact test or the $\chi 2$ test as indicated. McNemar's test was performed in the case of two related dichotomous variables. The Kruskal–Wallis test was used to compare multiple groups and non-normally distributed outcomes. To identify risk factors for MetS and its components, a univariable logistic regression model was performed first; only significant predictors ($P < 0.05$) were entered in the multiple logistic regression analysis thereafter. Refinement was performed by adding variables with P values < 0.20 if the number of events allowed it (with rule of thumb ~10 events per variable). The most significant contributor was left in the model if more candidates were present than number of variables to fill; when equal options were available, the highest Nagelkerke R^2 and best discrimination was chosen. The dependent variables were studied with forced entry. The Student's T-test was used to compare normally distributed variables, and the Wilcoxon rank-sum test was

used to compare non-normally distributed variables. The one sample binomial T-test was used to compare our data with the reference population Bonferroni adjustments were used if applicable to correct for multiple testing. Cohen's kappa and the Altman–Bland method (with the difference of the mean of two methods) were used to check for agreement between methods of measuring MetS and obesity to compare the definitions.

Results

Baseline characteristics

The baseline characteristics of the 95 CP patients included are shown in Table 2. Childhood onset (CO) of the disease was observed in 46 patients (49%) and 51% of all patients were female. The median follow-up time was 19 years (range 1–62). The median age at last follow-up was 45 years (range 16–78), mean age at first DXA scan was 35 years (range 11–79) and the mean time from first to last DXA scan was 10.4 years (range 0.1–23.3). The mean time from first treatment to first DXA scan was 8.9 years (s.d. 9.8, range −8.7 to 47.0). Patient characteristics did not differ significantly between males and females, except for pituitary deficiencies which were more prevalent in males (Table 2). In males, there was a trend for higher frequency of TSH deficiency (47 (100%) vs. 43 (90%), $P = 0.06$) and for higher ACTH deficiency (43 (92%) vs. 36 (77%), $P = 0.05$). Sex hormone replacement therapy was used by 74 patients with hypogonadotropic hypogonadism (79% of males and premenopausal females). Use of sex hormone replacement therapy was significantly higher in males than females (94 vs. 65%, $P = 0.001$), also after postmenopausal women were excluded (96 vs. 78%, $P = 0.012$). Patients with CO craniopharyngioma (CO CP) had a longer follow-up (median 22 vs. 16 years, $P < 0.05$), higher frequency of radiotherapy in the past (65 vs. 42%, $P = 0.02$) and greater occurrence of epilepsy (22 vs. 6%, $P = 0.03$) compared with patients with adulthood-onset (AO) CP. A comparison of the included and excluded patients from the original cohort[6] is presented in Supplementary Table 1. Of 95 patients, 93 had two or more DXA scans available; at first and last

DXA scan, respectively 66 and 74 patients had enough data available to evaluate MetS (BMI), 66 and 69 to evaluate MetS (BF%), and 61 and 67 patients to evaluate MetS (FMI).

Table 2 Characteristics of our cohort of 95 Dutch and Swedish craniopharyngioma patients

	All cranio-pharyngiomas (n=95)	Females (n=48)	Males (n=47)
Baseline characteristics			
Age at presentation (years)[a]	22 (3-62)	22 (4-62)	16 (3-62)
Childhood-onset (n [%])	47 (50)	24 (50)	23 (49)
Adult-onset (n [%])	48 (51)	24 (50)	24 (51)
Follow-up since presentation (years)[a]	19 (1-62)	20 (3-62)	17 (1-39)
Age at last follow-up (years)[a]	45 (16-78)	45 (17-77)	53 (16-78)
Tumor characteristics			
Location (n [%]) at presentation			
Intrasellar	2 (2)	1 (2)	1 (2)
Suprasellar	47 (50)	25 (52)	22 (47)
Intra-/suprasellar	39 (41)	21 (44)	18 (38)
No information available	7 (7)	1 (2)	6 (13)
Hydrocephalus (n [%])	20 (21)	12 (26)	8 (19)
Hypothalamic damage (n [%])	31 (34)	16 (34)	15 (34)

Craniopharyngioma treatment			
Neurosurgery (n [%])	93 (98)	46 (96)	47 (100)
Radiotherapy (n [%])	51 (54)	25 (52)	26 (55)
^{90}Yttrium brachytherapy (n [%])	9 (10)	7 (15)	2 (4)
Recurrence (n [%])	33 (35)	18 (38)	14 (31)
Long-term health outcome			
Pituitary hormone deficiencies (n [%])			
GH	89 (95)[b]	45 (96)	44 (94)
FSH/LH	85 (90)[c]	41 (85)	44 (94)
TSH	90 (95)[d]	43 (90)	47 (100)*
ACTH	79 (84)[d]	36 (77)	43 (92)**
AVP	65 (69)[d]	34 (71)	31 (67)
Visual impairment (n [%])	72 (78)[e]	37 (80)	35 (76)
Treatment for epilepsy (n [%])	6 (6)	3 (6)	4 (9)
Treatment for psychiatric illness (n [%])	12 (13)	6 (13)	6 (13)

[a]Median (range). [b]Replacement therapy was used by 80 patients with growth hormone deficiency (91%). [c]Replacement therapy was used by 74 patients with hypogonadotropic hypogonadism (79 % of males and premenopausal females); it was significantly higher in males than females (94% vs. 65%, $P = 0.001$), also after exclusion of postmenopausal women (96% vs. 78%, $P = 0.012$). [d]All patients were substituted with replacement therapy except for one patient, who was diagnosed with ACTH deficiency by metyraponetest but did not receive treatment later. [e]Missing data for 3 subjects; 76% of the entire cohort. ACTH = Adrenocorticotropic hormone; AVP = arginine vasopressin or antidiuretic hormone; FSH/LH = Follicle stimulating hormone/luteinizing hormone; GH = Growth hormone; n = Number; TSH = Thyroid stimulating hormone. *$P = 0.06$. **$P = 0.05$. Mean hydrocortisone dose or equivalent was 20.9 ± 6.2 mg, range 5-50 (data available for n=76).

The MetS and its components

Information on MetS (components) is presented in Tables 3 and 4. In 64 cases (78%), MetS components were evaluated within ±1 year of the last DXA scan. In Supplementary Table 3, a comparison is made for patients where evaluation of MetS criteria was performed within 1 year or more than 1 year before or after DEXA scan. Patients evaluated within 1 year of DXA scan had a lower rate of obesity based on BF% at last DXA scan (46 vs. 80%, $P = 0.03$).

At last DXA scan, MetS defined by BMI, BF% and FMI occurred in 33 (45%), 35 (51%), and 34 (51%) subjects, respectively (Table 3). The frequency of MetS was comparable to our previous study of CP patients in which DXA scans were not used (*P* values for BMI, BF% and FMI respectively $P = 0.45$, $P = 0.25$, and $P = 0.26$)[6], and were at all rates higher than those in the general population ($P < 0.001$). At last DXA scan, MetS is estimated to be (borderline) higher if defined by BF% or FMI vs. BMI ($P = 0.03$, $P = 0.06$) in the current cohort. Subjects had an increased occurrence of obesity compared to the general population (range: 53–67% vs. 30–31% depending on the definition, $P < 0.05$) (Table 3). The occurrence of hypertension and low HDL cholesterol was higher than the general population, with frequencies of 40 ($P < 0.001$) and 62% ($P < 0.001$), respectively. Here MetS criteria took into account medication use. MetS was significantly more prevalent in obese compared to non-obese subjects if obesity was defined with BMI (62 vs. 21%, $P = 0.001$), BF% (55 vs. 15%, $P = 0.01$) and FMI (63 vs. 25%, $P < 0.01$). Visual deficits were also more prevalent in obese compared with non-obese patients with CP (if defined by increased BF% at first DXA scan 85 vs. 59%, $P = 0.01$; if defined by increased FMI at first DXA scan 90 vs. 79%, $P < 0.05$; if defined by high FMI at last DXA scan 90 vs. 69%, $P = 0.03$). Unexpectedly, there was no significant difference in hypothalamic damage between obese and non-obese subjects, but a trend was shown for some definitions (based on BMI at first DXA scan 41 vs. 25%, $P = 0.14$; at last DXA scan 44 vs. 25%, $P = 0.07$, based on BF% at first DXA scan 40 vs. 29%, $P = 0.31$; at last DXA scan 46 vs. 32%, $P = 0.23$, based on FMI at first DXA scan 43 vs. 24%, $P = 0.06$; at last DXA scan 46 vs. 27%, $P = 0.09$).

CO CP patients had a higher occurrence of obesity defined by BF% at last DXA scan (87 vs. 59%, $P = 0.03$) and a lower occurrence of the MetS component hypertension (31 vs. 60%, $P < 0.01$) than AO CP. The presence or absence of growth hormone replacement treatment in patients had no effect on MetS components.

Longitudinal change of metabolic parameters

The following numbers include only patients that have MetS (components) evaluated at both first and last DXA scan to evaluate possible longitudinal changes. There was no significant change in frequency of MetS of any definition during follow-up for MetS based on BMI from 31 (49%) to 25 (40%), $P = 0.31$; for MetS based on BF% from 31 (53%) to 27 (47%), $P = 0.52$; for MetS based on FMI from 32 (56%) to 27 (47%), $P = 0.41$). Obesity frequencies defined by BMI showed a trend to increase between first and last visit (34 (44%) vs. 43 (56%), $P = 0.06$), but obesity rates defined by BF% and FMI did not change (respectively from 42 (66%) to 44 (69%), $P = 0.82$; from 27 (47%) to 32 (56%), $P = 0.27$. Importantly, the number of patients with hypertension increased significantly (19 (26%) vs. 34 (46%), $P = 0.008$). However, the MetS component low HDL cholesterol did not change significantly (from 38 (54%) to 27 (39%), $P = 0.09$). Neither was there a change in the components of high glucose (from 42 (62%) at first to 44 (65%) at last DXA scan, $P = 0.84$) or low HDL cholesterol (from 38 (54%) to 27 (39%), $P = 0.09$). The component high triglycerides decreased from 35 (51%) to 19 (28%) ($P = 0.01$).

Table 3 Metabolic syndrome and its components in patients with craniopharyngioma

	All CP	Ref. CP	Norm. data	*P*
MetS				
BMI	33 (45%)	20 (57%)	19%	**<0.001**
BF%	35 (51%)	20 (67%)	19%	**<0.001**
FMI	34 (51%)	19 (63%)	17%	**<0.001**

Metabolic components				
Obesity (FD)				
BMI	35 (43%)	16 (53%)	31%	**<0.05**
BF%	53 (64%)	21 (66%)	31%	**<0.05**
FMI	30 (47%)	17 (57%)	30%	**<0.05**
Obesity (LD)				
BMI	49 (57%)	29 (58%)	31%	**<0.05**
BF%	48 (75%)	18 (62%)	31%	**<0.05**
FMI	41 (59%)	22 (67%)	31%	**<0.05**
High glucose	52 (67%)	NA	NA	NA
High triglycerides	22 (29%)	NA	NA	NA
Low HDL	32 (41%)	4 (7%)*	7%	0.50
High blood pressure	37 (46%)	12 (22%)**	32%	0.07

	Dutch (n=35) vs. Sweden (n=60)	***P***	**♀ (n=48) vs. ♂ (n=47)**	***P***
MetS				
BMI	73% vs. 37%	**0.01**	43% vs. 46%	0.82
BF%	65% vs. 46%	0.18	54% vs. 46%	0.91
FMI	78% vs. 47%	0.15	52% vs. 50%	0.90
Metabolic components				
Obesity (FD)				
BMI	38% vs. 45%	0.58	40% vs. 46%	0.57
BF%	44% vs. 72%	**0.01**	59% vs. 69%	0.32
FMI	33% vs. 49%	0.48	40% vs. 53%	0.30
Obesity (LD)				
BMI	63% vs. 54%	0.45	52% vs. 61%	0.40
BF%	63% vs. 80%	0.21	69% vs. 81%	0.25
FMI	50% vs. 60%	0.54	55% vs. 62%	0.57
High glucose	75% vs. 64%	0.36	53% vs. 80%	**0.01**
High triglycerides	63% vs. 20%	**0.004**	42% vs. 18%	**0.02**

Low HDL	58% vs. 36%	0.09	51% vs. 31%	0.07
High blood pressure	38% vs. 48%	0.42	41% vs. 50%	0.42

Abbreviations: BF% = body fat percentage; BMI = body mass index; CP = craniopharyngioma; FD = first DXA-scan (Dual-Energy X-ray Absorptiometry scan); FMI = fat mass index; HDL = High-density lipoprotein cholesterol; LD = Last DXA-scan; Norm. = Normative; MetS = metabolic syndrome; n = number; NA = Not Available; *P* = *P*-value; Ref. CP = referenced CP (all CP in the same age group as reference population); ♀ = Female; ♂ = Male. Baseline differences between Dutch and Swedish patients were age at presentation (median 16 vs. 30 years; *P* = 0.021), growth hormone replacement therapy (74% vs. 92%, *P* = 0.018), hypothalamic damage (59% vs. 22%; *P* = 0.01). MetS (component) definitions: see Table 1. MetS (components) are components evaluated at LD, except obesity (FD and LD). *For all adult patients HDL ≤ 0.9 mmol/l regardless sex/ medication to compare with reference data. If defined as in Table 1, low HDL occurs in 15 patients (40%) (higher than the normative data (*P*<0.001)). **Defined as blood pressure >140/90 mmHg, regardless use of medication. If defined as in Table 1, it occurs in 24 patients (62%) (higher than normative data (*P*<0.001)). Respectively 74, 69 and 67 patients had enough data available to evaluate MetS (BMI), MetS (BF%), MetS (FMI). The data set was not complete in all patients.

Agreement between anthropometric measurements and DXA scan for MetS and obesity

A Cohen's kappa measurement of agreement was performed with MetS and obesity (Supplementary Table 4). Almost perfect agreements were found between MetS evaluated by BMI and either MetS evaluated by BF% (Kappa 0.82, *P* < 0.001) or MetS evaluated by FMI (Kappa 0.85, *P* < 0.001). MetS was evaluated incorrectly by BMI compared to BF% in six patients (9%) and FMI in five patients (7%) (Table 5). At last DXA scan, the estimated occurrence of MetS was lower when evaluated by BMI as compared to BF% and FMI (respectively 42 vs. 52% (*P* = 0.03), 43 vs. 51% (*P* = 0.06)). This was not the case for the first DXA scan (respectively 49 vs. 54%, *P* = 0.38; 48 vs. 53%, *P* = 0.25), suggesting the difference in estimation of MetS becomes less accurate as patients become older. In Figure 1, BMI was set out vs. BF% and FMI to illustrate discordances in the evaluation of obesity. The occurrence of obesity was lower if assessed by BMI when compared with obesity assessed by BF% (43 vs. 65%, *P* < 0.001 at first DXA scan, 58 vs. 74%, *P* = 0.03 at last DXA scan) but not when compared to FMI (41% vs. 47%, *P* = 0.22

at first DXA scan; 57 vs. 58%, $P = 1.00$ at last DXA scan). At last DXA scan, obesity estimations are discordant if BMI is compared to BF% (18 patients (29%)) and FMI (9 patients (13%)) at last DXA scan (see Table 6 for concordance of obesity measures). A Bland–Altman plot and analysis[42] was performed for BMI SDS with BF% and FMI SDS (Figure 2 and Supplementary Table 5). An upward trend is seen in the difference of the average for BMI/BF% SDS at first and last DXA scan (Figure 2). Differences outside the 95% agreement interval are observed, especially at the lower and higher averages, suggesting discrepancies occur mostly in the upper and lower end of the ranges. This effect was not observed for FMI.

Table 4 Occurrence of the criteria for metabolic syndrome in craniopharyngioma patients at last DXA-scan

MetS	MetS (BMI) (n, (%))	MetS (BF%) (n, (%))	MetS (FMI) (n, (%))
No criterion	6 (8%)	4 (6%)	5 (7%)
One criterion	13 (18%)	9 (13%)	11 (16%)
Two criteria	19 (26%)	16 (23%)	16 (24%)
Three criteria	15 (20%)	15 (22%)	18 (27%)
Four criteria	10 (14%)	5 (7%)	5 (8%)
Five criteria	4 (5%)	1 (1%)	4 (6%)
NA	7 (9%)	19 (28%)	8 (12%)

MetS = Metabolic Syndrome; MetS (BMI) = component of obesity defined with high body mass index (BMI >30) or increased standardized deviation score (SDS); MetS (BF%) = component of obesity defined with high fat percentage with limits defined according to sex and age, or by high SDS; MetS (FMI) = component of obesity defined with increased FMI with limits defined according to sex and age or SDS; n = number; NA = not available (not all criteria are known). The data set was not complete in all patients.

Table 5 Concordance in metabolic syndrome and obesity of BMI, BF% and FMI

MetS			**First DXA-scan**		**Last DXA-scan**	
			BMI		BMI	
			Non-MetS (n, %)	MetS (n, %)	Non-MetS (n, %)	MetS (n, %)
	BF%	Non-MetS (n, %)	28 (44%)	**1 (2%)**	32 (49%)	**0 (0%)**
		MetS (n, %)	**4 (6%)**	30 (49%)	**6 (9%)**	28 (42%)
	FMI	Non-MetS (n, %)	29 (48%)	**0 (0%)**	33 (49%)	**0 (0%)**
		MetS (n, %)	**3 (5%)**	29 (48%)	**5 (7%)**	29 (43%)
Obesity			**Non-obese (n, %)**	**Obese (n, %)**	**Non-obese (n, %)**	**Obese (n, %)**
	BF%	Non-obese (n, %)	26 (34%)	**1 (1%)**	12 (19%)	**4 (7%)**
		Obese (n, %)	**18 (23%)**	32 (42%)	**14 (23%)**	32 (52%)
	FMI	Non-obese (n, %)	33 (52%)	**1 (2%)**	25 (36%)	**4 (6%)**
		Obese (n, %)	**5 (8%)**	25 (39%)	**5 (7%)**	35 (51%)

Concordance of metabolic syndrome of for definitions of the criterion obesity and obesity itself with body mass index (BMI), body fat percentage (BF%) and fat mass index (FMI) at first and last DXA-scan in craniopharyngioma patients. N = number. Non-MetS represent subjects without presence of metabolic syndrome, MetS with presence of metabolic syndrome. The bold numbers and percentages are discordances of the different definitions. Metabolic syndrome is underestimated in 9% for BF% and 8% for FMI compared to BMI. At last DXA-scan, obesity is underestimated in 23% of the cases if defined by BF% in comparison to BMI; for FMI this percentage is much lower. The numbers may not add up to 100% due to rounding issues. The data set was not complete in all patients.

Determinants for occurrence of obesity and MetS and its components in patients with craniopharyngioma

Results of the univariable and multivariable logistic regression analyses on explanatory factors for MetS and its components at last DXA scan are shown in Supplementary Table 6. Explanatory factors in univariable models for MetS (BMI, BF% and FMI) were age at last DXA scan, recurrence or progression, hydrocephalus and visual impairment. These factors remained significant in multivariable analysis in one of the models with the exception of visual impairment. The explanation of the models was limited (Nagelkerke's R^2 0.15–0.29). The dose of glucocorticoid replacement was not a significant contributor (data not shown).

Figure 1 BMI versus body fat percentage and FMI in obesity

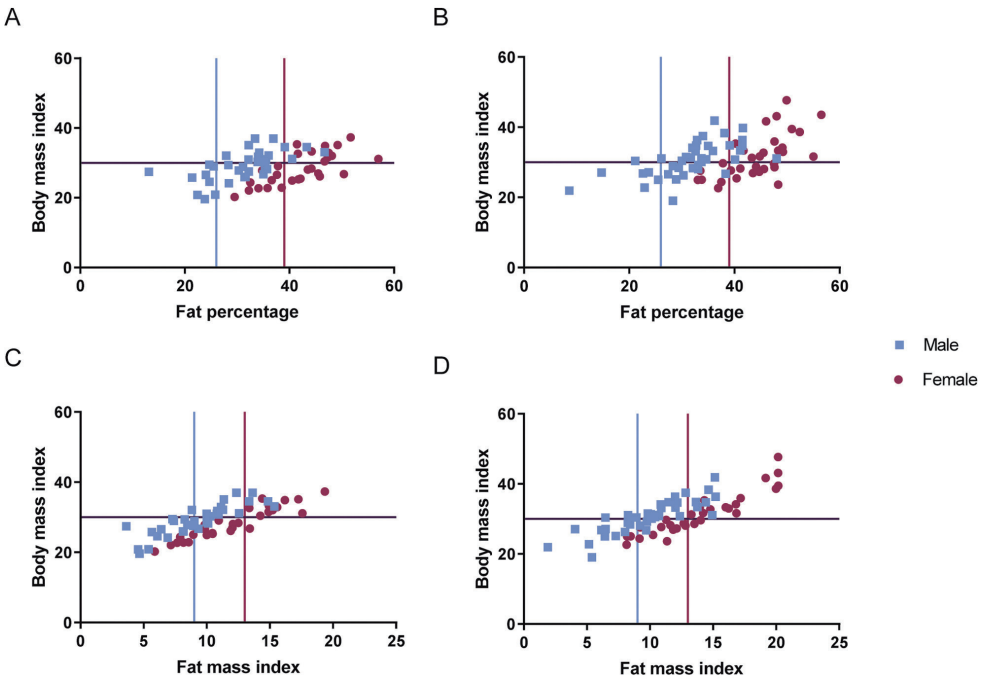


Figure 1: Body mass index (BMI) vs. body fat percentage (BF%) and fat mass index (FMI) at first (AC) and last (BD) DXA-scan in craniopharyngioma patients. Obesity based on BF% or FMI are bordered by the blue vertical line for males and by the red vertical line for females on the X-axis. The purple horizontal line borders obesity based on BMI. Female and male patients in the right lower quadrant are considered obese by fat percentage/ FMI, but not by BMI.

Type II diabetes mellitus and cardio- and cerebrovascular morbidity

Eleven patients with CP (12%) had diabetes mellitus of which one had type I diabetes mellitus. The frequency of T2D was higher in patients with CP compared with normative data (14% in subjects with the same age category as the normative data, vs. 6% in normative data; $P < 0.05$), but there was no difference in incidence between obese and non-obese CP patients. Myocardial infarctions occurred in four patients with CP (4%) with a median onset at 58 years of age (range 39–75). Nine subjects (10%) had experienced cerebrovascular accidents with a median age of onset of 41 years (range 16–66) and with a median of 8 years since first presentation of CP (range 0–25).

Discussion

This is the first study to assess the value of body composition measured by DXA scan in evaluating MetS in CP survivors. A discordancy in evaluation of MetS occurs in 9% of the cases if BMI is used as opposed to BF%. Moreover, the occurrence of obesity was estimated 22% higher if measured by BF% compared to BMI. Since obesity itself is a predictor of cardiovascular disease,[43] this suggests that DXA scan can be of great value in certain cases. In the Rotterdam/Gothenborg cohort, a standardized incidence ratio of 4.9 for cerebrovascular accidents was observed.[5] The risk of T2D was approximately doubled in CP patients compared to the general population. The long-term care of patients with CP should include cardiovascular risk management. Figure 2 suggests that discrepancy is most evident in the extremes, the very thin and very fat CP patients. Despite these results, the agreement between methods for MetS is good, as evaluated by Cohen's kappa. In clinical practice, confirmation by DXA scan will not be of added value if obesity is obviously apparent from BMI. Although discordance is not as impressive as found in childhood cancer patients,[22] obesity evaluation is discordant in one out of five patients and MetS evaluation is discordant in one out of ten if BMI is used instead of BF%. Therefore, our recommendation is to decide on a case-by-case basis if a DXA scan should be performed.

The risk of a decreased BMD should also be taken into account when deciding if a DXA scan should be included in the patients' management.

There was a striking sex-dependent difference in MetS components: while men had an increased risk of having the high glucose component, women were more likely to have high triglycerides (Table 3). This disparity may be explained by the higher risk for men to develop T2D,[44] and estrogen replacement therapy raising triglycerides in women.[45] Still, this effect should be limited: premenopausal females seem to be undertreated with gonadal replacement therapy, as they receive significantly less frequently replacement treatment compared with men. Being female has been shown to positively correlate with MetS,[46] but in our cohort, the MetS rate was equal between genders. Female sex was a protective factor for high glucose and a risk factor for high triglycerides and low HDL.

To determine factors that influence MetS and obesity, a logistic regression model was established. Age is a main predictor for all definitions of MetS (based on BMI, BF% and FMI; Supplementary Table as the prevalence of MetS increases with age.[47] Another important predictor for MetS (BF%) is visual impairment, which is in line with the findings of the previous study of Wijnen et al.,[6] who studied a larger cohort. To our knowledge, the only other study investigating MetS (components) in CP patients, is Tosta et al.,[46] who found that BMI SDS at diagnosis positively influenced the presence of the MetS posttreatment. Radiation was expected to be a factor involved, as this is a risk factor in developing the MetS and obesity in cancer survivors,[48] but it was not found to be so in our study. Although hypothalamic damage was expected to be a significant contributor, it was only borderline significant for two definitions of MetS (MetS (BMI) and MetS (BF%); Supplementary Table 6). Our cohort may have been underpowered to define hypothalamic damage and radiation therapy as a significant predictors. Although the hypothalamus plays a major role in weight maintenance, it has been shown that patients develop morbid obesity under hypothalamus-sparing operative strategies,[49] suggesting that obesity is multifactorial. Hydrocephalus[50] is associated with obesity, so

finding this as a significant contributor for MetS was not unexpected.

Predictors of poor outcomes are visual deterioration,[51] hydrocephalus and multiple operations due to tumor recurrence.[5,52] Recurrence or progression was a key predictor in all definitions. All predictors could be a sign of advanced brain disease and may be interrelated. On the other hand, no interaction was found while establishing the model. Advanced brain disease is likely to influence aspects of lifestyle such as physical activity.

Figure 2 Bland Altman Plot for BMI SDS and BF%/FMI SDS at first and last DXA-scan

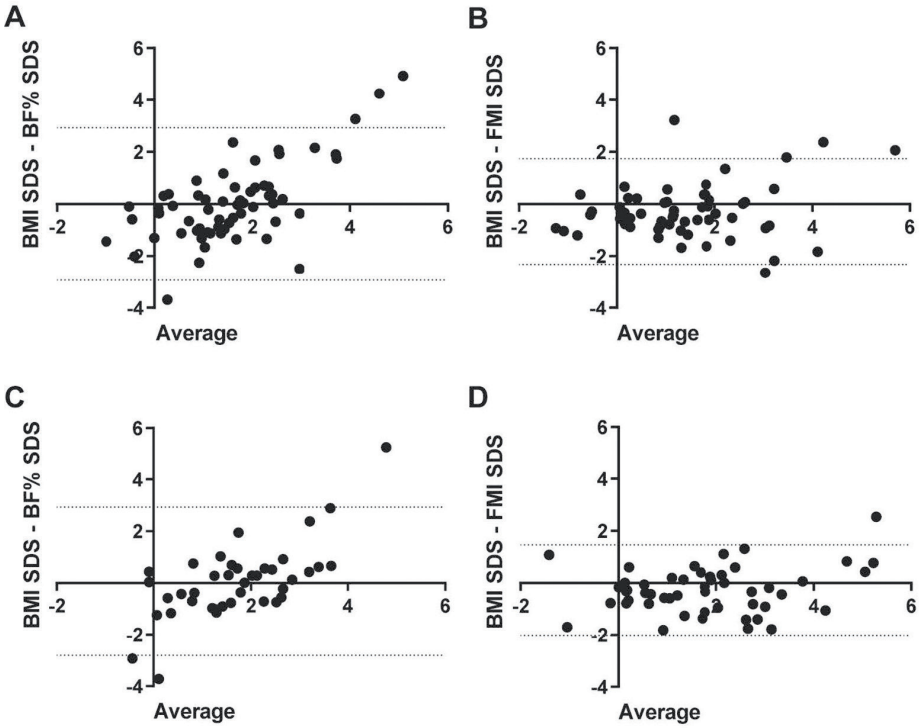


Figure 2: Bland Altman Plots. Plot A and B present data on the first DXA-scan; plot C and D of the last DXA-scan. Respectively, on the X-axis the average, and on the Y-axis the difference is expressed of body mass index standardized deviation scores (SDS) and body fat percentage SDS (AC) or fat mass index SDS (BD). An upward trend is seen in the difference of the average for BF% at first and last DXA-scan (AC). The difference in SDS of BMI and BF% seems larger at the end of the ranges, since the differences are observed outside the 95% agreement interval. These suggested discrepancies appear mostly in the upper and lower end of the ranges. For the difference of the average for FMI, the upward trend is not obviously shown.

There are some limitations in this study. Several factors contribute to a possible selection bias: there are size and weight limitations of DXA scanners,[53] the study was performed in referral centers, and the decision to perform a DXA scan was made by clinical judgment of the physicians, which is why only a subset of our previously studied cohort was assessed.[6] MetS criteria were evaluated within a window spanning 3 years either side of the last DXA scan. This period was chosen since earlier studies have shown that BMI change is central to other correlated features in the MetS[54] and BMI was stable for 3 years in a group of childhood cancer survivors.[22] Also, in a group of over 5000 French subjects, MetS was found to be stable over a period of 3 years.[55] Furthermore, any DXA scanner was accepted, even though it is known that the DXA scan has a considerable intraindividual variability[56] and results may differ between machines; it would have been ideal to cross-calibrate all scanners.[57] To cope with this, we used age- and sexmatched s.d. scores. The DXA scan is still considered to be the method of preference when BMI and adiposity are considered discordant.[21, 25] For evaluation of obesity by BMI, a cutoff of ≥30 kg/m^2 for reasons of conformity with our earlier study[6] and by the WHO guidelines,[58] as well as to avoid overdiagnosis. However, we realize that cutoffs used for the waist circumference according to the International Diabetes Federation (IDF) values were closer to matching a BMI of 25 kg/m^2 in males. The IDF definition notes that, if BMI >30 kg/m^2, central obesity can be assumed and waist circumference does not need to be measured.[27] This remains however a topic of discussion. Since we were unable to retrieve information on waist circumference and to avoid overdiagnosing as well as to be able to make comparisons with our previous research, we decided to use a cutoff level of >30 kg/m^2 for BMI.

Since the availability of literature is extremely limited on the topic, the data we present here are still highly important for clinicians in considering the use of DXA scans in long-term follow-up, despite its limitations. It is the only study in patients with CP that compares body composition measures vs. BMI for evaluation of obesity and the MetS as well as contains longitudinal follow-up.

In conclusion, patients with CP are at high risk of obesity and MetS. Obesity and MetS are underestimated if measured by BMI. Moreover, age, visual impairment and recurrence or progression are predictors of MetS or its components. This study is single in its existence providing evidence that DXA scan measured BF% is a better alternative for evaluation of obesity and MetS in certain patients with CP, especially those in the upper and lower range of BMI or BF%. The cost-effectiveness of performing DXA-scans remains to be determined before general clinical use in CP survivors can be recommended.

References

1. Nielsen EH, Feldt-Rasmussen U, Poulsgaard L, Kristensen LO, Astrup J, Jorgensen JO, Bjerre P, Andersen M, Andersen C, Jorgensen J et al. Incidence of craniopharyngioma in Denmark (n = 189) and estimated world incidence of craniopharyngioma in children and adults. Journal of Neuro-Oncology 2011; 104(3): 755-63. (https://doi.org/10.1007/s11060-011-0540-6)

2. Olsson DS, Andersson E, Bryngelsson IL, Nilsson AG & Johannsson G. Excess mortality and morbidity in patients with craniopharyngioma, especially in patients with childhood onset: a population-based study in Sweden. Journal of Clinical Endocrinology and Metabolism 2015; 100: 467–474. (https://doi.org/10.1210/jc.2014-3525)

3. Wijnen M, van den Heuvel-Eibrink MM, Janssen JAMJL, Catsman-Berrevoets CE, Michiels EMC, van Veelen-Vincent MC, Dallenga AHG, van den Berge JH, van Rij CM, van der Lely AJ et al. Very long-term sequelae of craniopharyngioma. European Journal of Endocrinology 2017; 176: 755–767. (https://doi.org/10.1530/EJE-17-0044)

4. Jackson AC, Tsantefski M, Goodman H, Johnson B & Rosenfeld J. The psychosocial impacts on families of low-incidence, complex conditions in children: the case of craniopharyngioma. Social Work in Health Care 2003; 38: 81–107.

5. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, Hammarstrand C, Janssen JAMJL, van der Lely AJ, Johannsson G & Neggers SJCMM. Excess morbidity and mortality in patients with craniopharyngioma: a hospital-based retrospective cohort study. European Journal of Endocrinology 2018; 178: 93–102. (https://doi. org/10.1530/EJE-17-0707)

6. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, Hammarstrand C, Janssen JAMJL, Van der Lely AJ, Johannsson G & Neggers SJCMM. The metabolic syndrome and its components in 178 patients treated for craniopharyngioma after 16 years of follow-up. European Journal of Endocrinology 2018; 178: 11–22. (https://doi.org/10.1530/EJE-17-0387)

7. Holmer H, Pozarek G, Wirfalt E, Popovic V, Ekman B, Bjork J & Erfurth EM. Reduced energy expenditure and impaired feedingrelated signals but not high energy intake reinforces hypothalamic obesity in adults with childhood onset craniopharyngioma. Journal of Clinical Endocrinology and Metabolism 2010; 95: 5395–5402. (https://doi.org/10.1210/jc.2010-0993)

8. Bulow B, Attewell R, Hagmar L, Malmstrom P, Nordstrom CH & Erfurth EM. Postoperative prognosis in craniopharyngioma with respect to cardiovascular mortality, survival, and tumor recurrence. Journal of Clinical Endocrinology and Metabolism 1998; 83: 3897–3904. (https://doi.org/10.1210/jcem.83.11.5240)

9. Muller HL. Craniopharyngioma. Handbook of Clinical Neurology 2014; 124: 235–253. (https://doi.org/10.1016/B978-0-444-59602-4.00016-2)

10. Berthon BS, MacDonald-Wicks LK & Wood LG. A systematic review of the effect of oral glucocorticoids on energy intake, appetite, and body weight in humans. Nutrition Research 2014; 34: 179–190. (https://doi.org/10.1016/j.nutres.2013.12.006)

11. Erfurth EM. Endocrine aspects and sequel in patients with craniopharyngioma. Journal of Pediatric Endocrinology and Metabolism 2015; 28: 19–26. (https://doi.org/10.1515/jpem-2014-0419)

12. Morley JE. Hormones and sarcopenia. Current Pharmaceutical Design 2017; 23: 4484–4492. (https://doi.org/10.2174/1381612823666161123150032)

13. Bulow B, Hagmar L, Mikoczy Z, Nordstrom CH & Erfurth EM. Increased cerebrovascular mortality in patients with hypopituitarism. Clinical Endocrinology 1997; 46: 75–81. (https://doi.org/10.1046/j.1365-2265.1997.d01-1749.x)

14. Wang KW, Fleming A, Johnston DL, Zelcer SM, Rassekh SR, Ladhani S, Socha A, Shinuda J, Jaber S, Burrow S et al. Overweight, obesity and adiposity in survivors of childhood brain tumours: a systematic review and meta-analysis. Clinical Obesity 2018; 8: 55–67. (https://doi.org/10.1111/cob.12224)

15. Yuen KCJ, Mattsson AF, Burman P, Erfurth EM, Camacho-Hubner C, Fox JL, Verhelst J, Geffner ME & Abs R. Relative risks of contributing factors to morbidity and mortality in adults With craniopharyngioma on growth hormone replacement. Journal of Clinical Endocrinology and Metabolism 2018; 103: 768–777. (https://doi.org/10.1210/jc.2017-01542)

16. Hatzitolios AI, Didangelos TP, Zantidis AT, Tziomalos K, Giannakoulas GA & Karamitsos DT. Diabetes mellitus and cerebrovascular disease: which are the actual data? Journal of Diabetes and Its Complications 2009; 23: 283–296. (https://doi.org/10.1016/j. jdiacomp.2008.01.004)

17. Muller HL, Bueb K, Bartels U, Roth C, Harz K, Graf N, Korinthenberg R, Bettendorf M, Kuhl J, Gutjahr P et al. Obesity after childhood craniopharyngioma – German multicenter study on preoperative risk factors and quality of life. Klinische Padiatrie 2001; 213: 244–249. (https://doi.org/10.1055/s-2001-16855)

18. Kim RJ, Shah R, Tershakovec AM, Zemel BS, Sutton LN, Grimberg A & Moshang T. Energy expenditure in obesity associated with craniopharyngioma. Child's Nervous System 2010; 26: 913–917. (https://doi.org/10.1007/s00381-009-1078-1)

19. Cornier MA, Dabelea D, Hernandez TL, Lindstrom RC, Steig AJ, Stob NR, Van Pelt RE, Wang H & Eckel RH. The metabolic syndrome. Endocrine Reviews 2008; 29: 777–822. (https://doi.org/10.1210/er.2008-0024)

20. Ju SY, Lee JY & Kim DH. Association of metabolic syndrome and its components with all-cause and cardiovascular mortality in the elderly: A meta-analysis of prospective cohort studies. Medicine 2017; 96: e8491. (https://doi.org/10.1097/MD.0000000000008491)

21. Perreault L. Determining Body Composition in Adults. Eds F Pi-Sunyer & D Sullivan. Waltham, MA: UpToDate, 2017.

22. Blijdorp K, van den Heuvel-Eibrink MM, Pieters R, Boot AM, Delhanty PJ, van der Lely AJ & Neggers SJ. Obesity is underestimated using body mass index and waist-hip ratio in long-term adult survivors of childhood cancer. PLoS ONE 2012; 7 e43269. (https://doi.org/10.1371/journal.pone.0043269)

23. Webb R & Singer M. Oxford Handbook of Critical Care. Oxford university Press:Oxford and New York, 2005.

24. Perreault L. Obesity in Adults: Prevalence, Screening, and Evaluation. Eds F Pi-Sunyer, D Sullivan. Waltham, MA, 2018.

25. Kist-van Holthe J, Beltman M, Bulk-Bunschoten A, L'Hoir M, Kuijpers T, Pijpers F, Renders C, Timmermans-Leenders E, Blok D, van de Veer M & HiraSing R Nederlands Centrum Jeugdgezondheid/Dutch Center of Health of the Youth https://www.ncj.nl/richtlijnen/alle-richtlijnen/richtlijn/overgewicht Publication Published: May 2012

26. Kelly TL, Wilson KE & Heymsfield SB. Dual energy X-ray absorptiometry body composition reference values from NHANES. PLoS ONE 2009; 4 e7038. (https://doi.org/10.1371/journal. pone.0007038)

27. Alberti G, Zimmet P & Shaw J. The IDF Consensus Worldwide Definition of the Metabolic Syndrome. International Diabetes Federation, 2006.

28. Definition, Diagnosis and classification of diabetes mellitus and its complications: report of a WHO consultation. Geneva, Switzerland: World Health Organisation, 1999. (available at: https://apps.who.int/iris/bitstream/handle/10665/66040/WHO_NCD_NCS_99.2.pdf).

29. Larsson I, Lissner L, Samuelson G, Fors H, Lantz H, Naslund I, Carlsson LM, Sjostrom L & Bosaeus I. Body composition through adult life: Swedish reference data on body composition. European Journal of Clinical Nutrition 2015; 69: 837–842. (https://doi.org/10.1038/ejcn.2014.268)

30. Blokstra A, Vissink P, Venmans L, Holleman P, van der Schouw Y, Smit H & Verschuren W. Measuring the Netherlands. A monitoring study of risk factors in the general population, 2009–2010. 2011; Contract No.: RIVM Rapport 260152001/260152011. (https://www.rivm.nl/bibliotheek/rapporten/260152001.pdf)

31. Berg C, Rosengren A, Aires N, Lappas G, Toren K, Thelle D & Lissner L. Trends in overweight and obesity from 1985 to 2002 in Goteborg, West Sweden. International Journal of Obesity 2005; 29: 916–924. (https://doi.org/10.1038/sj.ijo.0802964)

32. Karlberg J, Luo ZC & Albertsson-Wikland K. Body mass index reference values (mean and SD) for Swedish children. Acta Paediatrica 2001; 90: 1427–1434. (https://doi.org/10.1111/j.1651-2227.2001.tb01609.x)

33. Fredriks AM, van Buuren S, Wit JM & Verloove-Vanhorick SP. Body index measurements in 1996–7 compared with 1980. Archives of Disease in Childhood 2000; 82: 107–112. (https://doi.org/10.1136/adc.82.2.107)

34. Alwis G, Rosengren B, Stenevi-Lundgren S, Duppe H, Sernbo I & Karlsson MK. Normative dual energy X-ray absorptiometry data in Swedish children and adolescents. Acta Paediatrica 2010; 99: 1091–1099. (https://doi.org/10.1111/j.1651-2227.2010.01713.x)

35. Gultekin T, Ozer BK, Akin G, Bektas Y, Sagir M & Gulec E. Prevalence of overweight and obesity in Turkish adults. Anthropologischer Anzeiger 2009; 67: 205–212. (https://doi.org/10.1127/0003-5548/2009/0022)

36. van der Sluis IM, de Ridder MA, Boot AM, Krenning EP & de Muinck Keizer-Schrama SM. Reference data for bone density and body composition measured with dual energy x ray absorptiometry in white children and young adults. Archives of Disease in Childhood 2002; 87: 341–347; discussion 341–347. (https://doi.org/10.1136/adc.87.4.341)

37. Coin A, Sergi G, Minicuci N, Giannini S, Barbiero E, Manzato E, Pedrazzoni M, Minisola S, Rossini M, Del Puente A et al. Fat-free mass and fat mass reference values by dual-energy X-ray absorptiometry (DEXA) in a 20–80 year-old Italian population. Clinical Nutrition 2008; 27: 87–94. (https://doi.org/10.1016/j.clnu.2007.10.008)

38. White EM, Wilson AC, Greene SA, McCowan C, Thomas GE, Cairns AY & Ricketts IW. Body mass index centile charts to assess fatness of British children. Archives of Disease in Childhood 1995; 72: 38–41. (https://doi.org/10.1136/adc.72.1.38)

39. Larsson I, Berteus Forslund H, Lindroos AK, Lissner L, Naslund I, Peltonen M & Sjostrom L. Body composition in the SOS (Swedish Obese Subjects) reference study. International Journal of Obesity and Related Metabolic Disorders 2004; 28: 1317–1324. (https://doi.org/10.1038/sj.ijo.0802732)

40. Gavriilidou NN, Pihlsgard M & Elmstahl S. Anthropometric reference data for elderly Swedes and its disease-related pattern. European Journal of Clinical Nutrition 2015; 69: 1066–1075. (https://doi.org/10.1038/ejcn.2015.73)

41. Hollman G & Kristenson M. The prevalence of the metabolic syndrome and its risk factors in a middle-aged Swedish population--mainly a function of overweight? European Journal of Cardiovascular Nursing 2008; 7: 21–26. (https://doi.org/10.1016/j. ejcnurse.2007.05.003)

42. Bland JM & Altman DG. Statistical methods for assessing agreement between two methods of clinical measurement. Lancet 1986; 1307–310. (https://doi.org/10.1016/j.ijnurstu.2009.10.001)

43. Eckel N, Li Y, Kuxhaus O, Stefan N, Hu FB & Schulze MB. Transition from metabolic healthy to unhealthy phenotypes and association with cardiovascular disease risk across BMI categories in 90 257 women (the Nurses' Health Study): 30 year follow-up from a prospective cohort study. Lancet Diabetes and Endocrinology 2018; 6: 714–724. (https://doi.org/10.1016/S2213-8587(18)30137-2)

44. RIVM. Diabetes meliltus: cijfers en context 2019. https://www. volksgezondheidenzorg.info/onderwerp/diabetes-mellitus/cijferscontext/huidige-situatie Accessed on April 15th, 2019

45. Walsh BW, Schiff I, Rosner B, Greenberg L, Ravnikar V & Sacks FM. Effects of postmenopausal estrogen replacement on the concentrations and metabolism of plasma lipoproteins. New England Journal of Medicine 1991; 325: 1196–1204. (https://doi.org/10.1056/NEJM199110243251702)

46. Tosta-Hernandez PDC, Siviero-Miachon AA, da Silva NS, Cappellano A, Pinheiro MM & Spinola-Castro AM. Childhood craniopharyngioma: A 22-year challenging follow-up in a Single Center. Hormone and Metabolic Research 2018; 50: 675–682. (https://doi.org/10.1055/a-0641-5956)

47. Ford ES, Giles WH & Dietz WH. Prevalence of the metabolic syndrome among US adults: findings from the third National Health and Nutrition Examination Survey. JAMA 2002; 287: 356–359. (https://doi.org/10.1001/jama.287.3.356)

48. Friedman DN, Tonorezos ES & Cohen P. Diabetes and metabolic syndrome in survivors of childhood cancer. Hormone Research in Paediatrics 2019; 1–10. (https://doi.org/10.1159/000495698)

49. Muller HL. Hypothalamic involvement in craniopharyngioma- Implications for surgical, radiooncological, and molecularly targeted treatment strategies. Pediatric Blood and Cancer 2018; 65: e26936. (https://doi.org/10.1002/pbc.26936)

50. Ahmet A, Blaser S, Stephens D, Guger S, Rutkas JT & Hamilton J. Weight gain in craniopharyngioma – a model for hypothalamic obesity. Journal of Pediatric Endocrinology and Metabolism 2006; 19: 121–127. (https://doi.org/10.1515/JPEM.2006.19.2.121)

51. Duff J, Meyer FB, Ilstrup DM, Laws ER, Jr, Schleck CD & Scheithauer BW. Long-term outcomes for surgically resected craniopharyngiomas. Neurosurgery 2000; 46: 291–302; discussion 302–295. (https://doi.org/10.1097/00006123-200002000-00007)

52. Poretti A, Grotzer MA, Ribi K, Schonle E & Boltshauser E. Outcome of craniopharyngioma in children: long-term complications and quality of life. Developmental Medicine and Child Neurology 2004; 46: 220–229.

53. Seabolt LA, Welch EB & Silver HJ. Imaging methods for analyzing body composition in human obesity and cardiometabolic disease. Annals of the New York Academy of Sciences 2015; 1353: 41–59. (https://doi.org/10.1111/nyas.12842)

54. Maison P, Byrne CD, Hales CN, Day NE & Wareham NJ. Do different dimensions of the metabolic syndrome change together over time? Evidence supporting obesity as the central feature. Diabetes Care 2001; 24: 1758–1763. (https://doi.org/10.2337/diacare.24.10.1758)

55. Balkau B, Vernay M, Mhamdi L, Novak M, Arondel D, Vol S, Tichet J & Eschwege E. The incidence and persistence of the NCEP (National Cholesterol Education Program) metabolic syndrome. The French D.E.S.I.R. Study. Diabetes and Metabolism 2003; 29: 526–532. (https://doi.org/10.1016/S1262-3636(07)70067-8)

56. LaForgia J, Dollman J, Dale MJ, Withers RT & Hill AM. Validation of DXA body composition estimates in obese men and women. Obesity 2009; 17: 821–826. (https://doi.org/10.1038/oby.2008.595)

57. Hull H, He Q, Thornton J, Javed F, Allen L, Wang J, Pierson RN & Gallagher D. iDXA, prodigy, and DPXL dual-energy X-ray absorptiometry whole-body scans: a cross-calibration study. Journal of Clinical Densitometry 2009; 12: 95–102. (https://doi.org/10.1016/j.jocd.2008.09.004)

58. Alberti KG, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, Fruchart JC, James WP, Loria CM, Smith SC, Jr et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation 2009; 120: 1640–1645. (https://doi.org/10.1161/CIRCULATIONAHA.109.192644)

Supplementary Tables

Supplementary Table 1 Comparison of craniopharyngioma patients with and without DXA-scan

		Patients with DXA scan (n=95)	Patients without DXA-scan (n=61)	*P*-value
Baseline characteristics	Age at presentation (years)[a]	22 (3-62)	22 (0-79)	0.43
	Childhood-onset (n [%])	46 (49)	28 (46)	0.66
	Follow-up since presentation (years [range])[a]	19 (1-62)	14 (3-33)	0.09
	Age at last follow-up (years [range])[a]	45 (16-78)	43 (18-92)	0.65
	Female sex (n [%])	48 (51)	26 (43)	0.38
	Swedish cohort (n [%])	69 (63)	10 (16)	**<0.001**
Tumor characteristics	Location (n [%]) at last imaging			
	Intrasellar	1 (1)	3 (5)	0.34
	Suprasellar	43 (51)	15 (26)	**0.005**
	Intra-/suprasellar	41 (48)	39 (68)	**0.02**
	Hydrocephalus (n [%])	20 (21)	18 (30)	0.28
	Hypothalamic damage (n [%])	31 (34)	25 (46)	0.17
Craniopharyn-gioma treatment	Neurosurgery (n [%])	93 (98)	55 (90)	0.06
	Radiotherapy (n [%])	51 (54)	28 (46)	0.34
	^{90}Yttrium brachytherapy (n [%])	9 (10)	13 (21)	**0.05**
	Recurrence (n [%])	33 (35)	26 (43)	0.31
Long-term health outcome	Pituitary hormone deficiencies (n [%])			
	GH	89 (95)	48 (80)	**0.005**
	FSH/LH	85 (90)	49 (82)	0.17
	TSH	90 (95)	56 (92)	0.47
	ACTH	79 (84)	49 (80)	0.55
	ADH	65 (69)	34 (56)	0.09
	Visual impairment (n [%])	72 (78)	42 (78)	0.95

	Treatment for epilepsy (n [%])	6 (6)	6 (10)	0.64
	Treatment for psychiatric illness (n [%])	12 (13)	7 (12)	0.84
	Bariatric surgery (n [%])	4 (4)	1 (2)	0.65
	OSAS (n [%])	21 (23)	6 (10)	0.04
	Diabetes mellitus (n [%])	11 (12)	14 (23)	0.08
	Cerebrovascular disease (n [%])	9 (10)	7 (12)	0.74
	Myocardial infarction (n [%])	4 (4)	5 (8)	0.31
	BMI (mean [range])	31.4 (19-47.7)	31.7 (16.9-59.5)	0.87
	MetS component obesity (based on BMI) (n [%])	54 (57)	30 (53)	0.613
	MetS component insulin resistance (n [%])	56 (74)	20 (26)	**0.006**
	MetS component low HDL cholesterol (n [%])	41 (46)	25 (38)	0.736
	MetS component high blood pressure (n [%])	48 (51)	31 (51)	0.976
	MetS component high triglycerides (n [%])	31 (34)	30 (54)	**0.017**
	Presence of MetS (n [%])	43 (47)	27 (44)	0.763
	Dead (n [%])	13 (14)	12 (20)	0.320

Comparison of 156 craniopharyngioma patients with (n=95) and without (n=61) DXA-scan. Data are given as mean and standard deviation (SD) unless stated otherwise [a]Median (range). ACTH = Adrenocorticotropic hormone; ADH = Antidiuretic hormone; FSH/LH = Follicle stimulating hormone/luteinizing hormone; GH = Growth hormone; n = Number; OSAS = obstructive sleep apnea syndrome; TSH = Thyroid stimulating hormone. Patients were only included for the measurement of BMI if they reached the age of 18 years at the moment of measurement; the measurement was at last DXA-scan or, if unavailable, at last follow-up. A lower occurrence of intra- and suprasellar location, Yttrium brachytherapy and MetS component of high triglycerides, and higher percentage of suprasellar location, GHD, obstructive sleep apnea syndrome and proportion of inclusion of Swedes appeared in the current cohort.

Supplementary Table 2 Included Dual X-ray Absorptiometry scanners

	First DXA-scan (n=95)	Last DXA-scan (n=93)
Unknown/other	1 (1%)	8 (9%)
Lunar iDXA Gothenborg University Hospital	0 (0%)	19 (20%)
Lunar DPXL total	58 (61%)	14 (15%)
DPXL Erasmus MC	32 (34%)	12 (13%)
DPXL Gothenborg University Hospital	26 (27%)	2 (2%)
Lunar Prodigy total	36 (38%)	52 (56%)
Prodigy (1) Erasmus MC	0 (0%)	9 (10%)
Prodigy (2) Erasmus MC	2 (2%)	4 (4%)
Prodigy Gothenborg University Hospital	34 (36%)	39 (42%)

DXA-scanners included in the research. Data is given as N (%). Prodigy (1) is used at the dept. of Nuclear Medicine in the Erasmus MC; Prodigy (2) at the dept. of Internal Medicine. Total includes all scanners of the same type in both centers.

Supplementary Table 3 Metabolic syndrome and its components in patients with craniopharyngioma

	CP <1yr	CP>1yr	*P*-value
MetS			
BMI	27 (44%)	6 (50%)	0.68
BF%	29 (50%)	6 (54%)	0.78
FMI	30 (52%)	4 (44%)	0.68
Metabolic components*			
Obesity (FD)			
BMI	26 (43%)	5 (39%)	0.68
BF%	42 (71%)	5 (39%)	0.05
FMI	27 (49%)	2 (29%)	0.43
Obesity (LD)			
BMI	34 (56%)	8 (50%)	0.68
BF%	37 (80%	6 (46%)	**0.03**
FMI	34 (61%)	5 (50%)	0.73
High glucose	39 (63%)	13 (81%)	0.17
High triglycerides	18 (29%)	4 (31%)	1.00
Low HDL	25 (39%)	7 (50%)	0.45
High blood pressure	31 (48%)	5 (31%)	0.21

Data are presented as n (%). CP <1yr or CP >1yr means that the patients have MetS components evaluated in within one year, or more than one year before or after DEXA-scan. Abbreviations: BF% = body fat percentage; BMI = body mass index; CP = craniopharyngioma; FD = first DXA-scan (Dual-Energy X-ray Absorptiometry scan); FMI = fat mass index; HDL = High-density lipoprotein cholesterol; LD = Last DXA-scan; Norm. = Normative; MetS = metabolic syndrome; n = number; NA = Not Available; *P* = *P*-value; ♀ = Female; ♂ = Male. MetS (components) are components evaluated at LD, except obesity (FD and LD). *MetS components are as defined in Table 1.

Supplementary Table 4 Bland-Altman bias of BMI SDS and BF%/FMI SDS scores at first and last DXA-scan

	Metabolic syndrome at last DXA scan	**Obesity**	
		First DXA-scan	Last DXA-scan
	Obesity defined with BMI	Defined with BMI	Defined with BMI
	Cut-off point (kappa; SE, (P))	Cut-off point (kappa; SE, (P))	Cut-off point (kappa; SE, (P))
Obesity (BF%)	0.82; 0.07 (<0.001)	0.53; 0.09 (<0.001)	0.37; 0.12 (0.002)
Obesity (FMI)	0.85; 0.06 (<0.001)	0.81; 0.07 (<0.001)	0.73; 0.08 (<0.001)

Agreement of anthropomorphic measurements and DXA-scan for evaluation of obesity and metabolic syndrome. For metabolic syndrome, the component of obesity is either defined with high body mass index (BMI), high body fat percentage (BF%), or high fat mass index (FMI). For obesity: defined on high body mass index (BMI >30) or increased standardized deviation score (SDS); high body fat percentage with limits defined according to sex and age, or by increased SDS; of with high fat mass index with limits defined according to sex and age or SDS; n=number. A Cohen's kappa measurement of agreement was set out with metabolic syndrome and obesity. For metabolic syndrome, an almost perfect agreement is found. For obesity, agreement is only fair to moderate.

Supplementary Table 5 Agreement of anthropomorphic measurements and DXA-scan for evaluation of obesity and metabolic syndrome

			BF% SDS	FMI SDS
First DXA-scan	Body mass index SDS	Bland- Altman bias	<0,001	-0,290
		SD of bias	1,496	-0,290
		LLA	-2,932	-2,319
		ULA	2,933	1,738
Last DXA-scan	Body mass index SDS	Bland- Altman bias	0,063	-0,273
		SD of bias	1,462	0,887
		LLA	-2,802	-2,012
		ULA	2,929	1,466

Bland-Altman bias of BMI SDS and BF%/FMI SDS scores in craniopharyngioma patients. BF% = body fat percentage; FMI = fat mass index; SD = standard deviation; SDS = standardized deviation scores; LLA = lower limit of 95% agreement interval; ULA = upper limit of 95% agreement interval. A Bland-Altman bias was performed of the mean difference of body mass index SDS and body fat percentage SDS/fat mass index SDS. The bias is only small. This table is complementary to Figure 1. Almost perfect agreement is found. For obesity, agreement is only fair to moderate.

Supplementary Table 6 Risk factors for the metabolic syndrome in patients with craniopharyngioma

	Univariable analysis					
	MetS (BMI)		MetS (BF%)		MetS (FMI)	
Variables	OR (95% CI)	*P*	OR (95% CI)	*P*	OR (95% CI)	*P*
Age at DXA-scan	**1.03 (1.00-1.06)**	**0.03**	**1.04 (1.00-1.07)**	**0.03**	**1.04 (1.01-1.07)**	**0.02**
Age at presentation	1.03 (0.99-1.06)	0.15	1.02 (0.98-1.05)	0.37	1.03 (0.99-1.07)	0.10
Female sex	0.90 (0.36-2.24)	0.82	1.06 (0.41-2.72)	0.91	1.07 (0.41-2.79)	0.90
Recurrence or progression	**4.03 (1.46-11.11)**	**<0.01**	**2.89 (1.02-8.17)**	**<0.05**	2.22 (0.80-6.21)	0.13
Hydrocephalus	**3.88 (1.24-12.22)**	**0.02**	2.60 (0.85-7.90)	0.094	2.61 (0.85-7.99)	0.09
Hypothalamic damage	2.38 (0.92-6.21)	0.08	2.10 (0.79-5.56)	0.14	1.63 (0.64-4.19)	0.31
Medication for epilepsy	4.44 (0.49-40.98)	0.18	5.86 (0.64-53.09)	0.12	0.59 (0.65-53.47)	0.12
Medication for psychiatric disease	1.24 (0.32-4.83)	0.75	2.29 (0.52-9.99)	0.27	1.25 (0.31-5.13)	0.76
Growth hormone deficiency	2.52 (0.25-25.49)	0.43	1.03 (0.06-17.16)	0.98	1.03 (0.14-7.80)	0.98
GHRT	1.71 (0.39-7.45)	0.47	1.03 (0.24-4.51)	0.97	1.03 (0.24-4.53)	0.96
Adrenal axis deficiency	2.34 (0.66-8.29)	0.19	1.25 (0.37-4.20)	0.72	0.69 (0.20-2.437)	0.56
Gonadal axis deficiency	3.76 (0.74-19.09)	0.11	1.34 (0.33-5.50)	0.69	1.67 (0.42-6.55)	0.46
Sex steroid replacement therapy	2.31 (0.56-9.63)	0.25	1.04 (0.27-4.01)	0.96	1.2 (0.33-4.43)	0.78
Diabetes insipidus	2.14 (0.75-6.12)	0.15	2.09 (0.73-5.60)	0.17	1.21 (0.42-3.50)	0.73
Radiotherapy	1.43 (0.57-3.59)	0.45	1.34 (0.52-3.44)	0.55	1.19 (0.45-3.13)	0.72
^{90}Yttrium brachytherapy	5.59 (0.59-51.97)	0.14	2.07 (0.35-12.09)	0.42	4.27 (0.45-40.37)	0.21
Visual impairment	2.80 (0.89-8.86)	0.08	**3.31 (1.01-10.84)**	**<0.05**	2.10 (0.62-7.13)	0.23

	Multivariable analysis[a]					
	MetS (BMI)		MetS (BF%)		MetS (FMI)	
Variables	OR (95% CI)	*P*	OR (95% CI)	*P*	OR (95% CI)	*P*
Age at DXA-scan	1.04 (1.00-1.08)	0.03	1.04 (1.00-1.07)	0.04	1.05 (1.01-1.08)	0.01
Age at presentation						
Female sex						
Recurrence or progression	3.17 (1.06-9.47)	0.04	2.04 (0.67-6.25)	0.21		
Hydrocephalus	5.64 (1.40-22.80)	0.02			4.07 (1.02-16.19)	<0.05
Hypothalamic damage						
Medication for epilepsy						
Medication for psychiatric disease						
Growth hormone deficiency						
GHRT						
Adrenal axis deficiency						
Gonadal axis deficiency						
Sex steroid replacement therapy						
Diabetes insipidus						
Radiotherapy						
^{90}Yttrium brachytherapy						
Visual impairment			2.64 (0.74-9.48)	0.14		

Univariable and multivariable logistic regression analysis. Abbreviations: CI = Confidence interval; GHRT = Growth hormone replacement therapy; MetS = Metabolic Syndrome; NA = not applicable; OR = Odds Ratio; O $\chi 2$ = Omnibus $\chi 2$; R^2 = Nagelkerke's R^2; SE = standard error. Hypothalamic damage at diagnosis. MetS is defined in Table 1. Age is

given in years. Hosmer Lemeshow test was non-significant in all models.

MetS (BMI): R^2 =0.29; O $\chi 2$ <0.001. MetS (BF%): R^2 =0.20; O $\chi 2$ =0.01. MetS (FMI): R^2 =0.18; O $\chi 2$ =0.008.

Explanatory (borderline) significant factors MetS components:

High glucose (univariable): female sex (OR 0.3, 95% CI 0.1-0.8, *P* = 0.01).

High blood pressure (univariable): age at DXA-scan (OR 1.1, 95% CI 1.0-1.1, *P* < 0.001), age of onset (OR 1.1, 95% CI 1.0-1.1, *P* = 0.001).

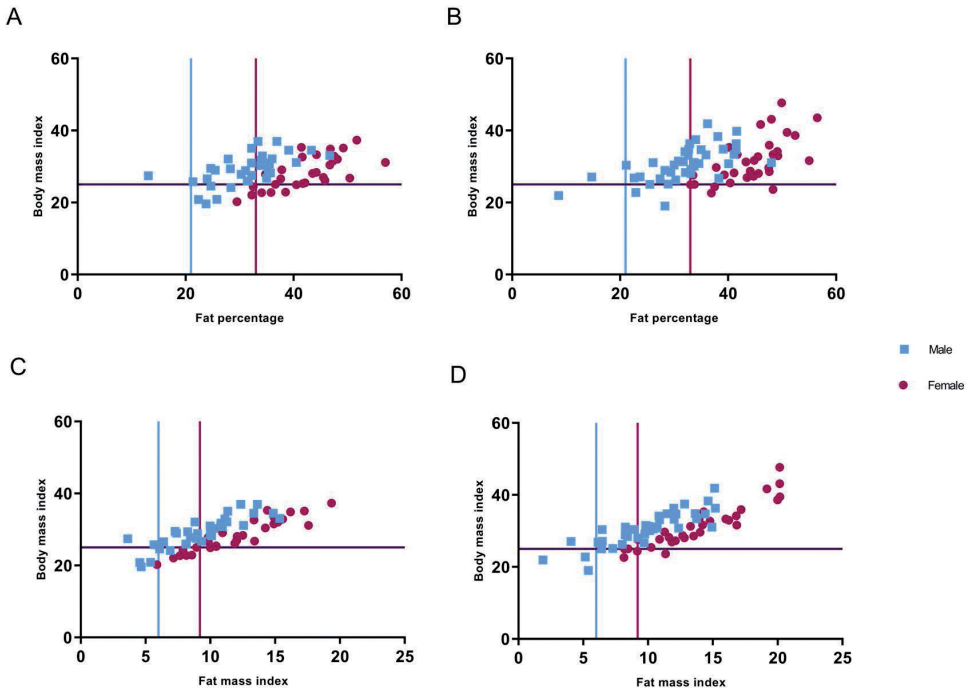
Low HDL (uni-/multivariable): female sex (OR 2.4, 95% CI 0.9-6.0, *P* = 0.07/ OR 3.9, 95% CI 1.1-13.1, *P* = 0.03), visual deficiency (OR 3.4, 95% CI 1.0-11.4, *P* = 0.05/ OR 5.0, 95% CI 0.9-29.1, *P* = 0.08), recurrence or progression (OR 5.8, 95% CI 2.1-15.8, *P* = 0.001/ OR 5.9, 95% CI 1.7-19.8, *P* = 0.004) and hypothalamic damage (OR 3.0, 95% CI 1.1-8.1, *P* = 0.03/ OR 3.9, 95% CI 1.0-16.1, *P* = 0.06). R^2 0.42, O $\chi 2$ (*P* < 0.001).

High triglycerides (uni-/multivariable): female sex (OR 3.3, 95% CI 1.1-9.4, *P* = 0.03/ OR 3.9, 95% CI 1.2-12.1, *P* = 0.02), recurrence or progression (OR 3.7, 95% CI 1.2-10.7, *P* = 0.02/ OR 3.8, 95% CI 1.2-11.4, *P* = 0.02), Yttrium brachytherapy (OR 15.3, 95% CI 1.7-140.2, *P* = 0.02/ NA), and borderline significant visual deficiency (OR 4.5, 95% CI 0.9-21.5, *P* = 0.06/ NA). R^2 0.21, O $\chi 2$ *P* = 0.003.

Obesity (BMI) (univariable): gonadal axis deficiency (OR 4.2, 95% CI 1.0-17.2, *P* = 0.04).

Obesity (FMI) (univariable): visual deficiency (OR 4.0 95% CI 1.1-14.8, *P* = 0.04).

Supplementary Figure 1 BMI versus body fat percentage and FMI in overweight subjects



Body mass index (BMI) vs. body fat percentage (BF%) and fat mass index (FMI) at first (AC) and last (BD) DXA-scan in craniopharyngioma patients. Overweight based on BF% or FMI are bordered by the blue vertical line for males and by the red vertical line for females on the X-axis. The purple horizontal line borders overweight based on BMI. Female and male patients in the right lower quadrant are considered overweight by fat percentage/ FMI, but not by BMI.[26,59]

6

Can Biomarkers Be Used To Improve Diagnosis and Prediction of Metabolic Syndrome in Childhood Cancer Survivors? A Systematic Review

Vincent G. Pluimakers,[*1] and Selveta S. van Santen,[*1,2]
Marta Fiocco,[1,3,4] Marie-Christine E. Bakker,[1,5]
Aart J. v.d. Lely,[2] Marry M. van den Heuvel-Eibrink,[**1]
and Sebastian J. C. M. M. Neggers.[**1,2]

[1]Princess Máxima Center for Pediatric Oncology, Utrecht, The Netherlands;
[2]Dept. of Medicine, Endocrinology, Erasmus Medical Center, Rotterdam, The Netherlands;
[3]Medical Statistics, dept. of Biomedical Data Science, Leiden UMC,
Leiden, The Netherlands;
[4]Mathematical Institute Leiden University, Leiden, The Netherlands;
[5]Department of Medicine, University Medical Center Utrecht, The Netherlands.
[*]Both authors contributed equally to the content of this manuscript.
[**]Both authors contributed equally to the content of this manuscript.

Obesity Reviews 2021

Running title: Novel metabolic syndrome biomarkers in CCS

Abbreviations: CVD = cardiovascular disease; T2DM = type 2 diabetes mellitus; MetS = metabolic syndrome; CCS = childhood cancer survivors; DXA = Dual-energy X-ray Absorptiometry; hsCRP = high sensitivity C-reactive protein; TNF-alpha = Tumor Necrosis Factor alpha; IL-1 = interleukin 1; IL-6 = interleukin 6; apoB = apolipoprotein B; Lp(a) = lipoprotein(a); AUC = area under the curve; ROC = receiver operating characteristic; OR = odds ratio; HR = hazard ratio; GRADE = Grading of Recommendations Assessment Development and Evaluation; BMI = body mass index; ALL = acute lymphoblastic leukemia; apoA1 = apolipoprotein A1; LDL = low density lipoproteins; HOMA-IR = Homeostatic Model Assessment of Insulin Resistance; HDL = high density lipoproteins.

Abstract

Background: Childhood cancer survivors (CCS) are at increased risk to develop metabolic syndrome (MetS), diabetes and cardiovascular disease. Common criteria underestimate adiposity and possibly underdiagnose MetS, particularly after abdominal radiotherapy.

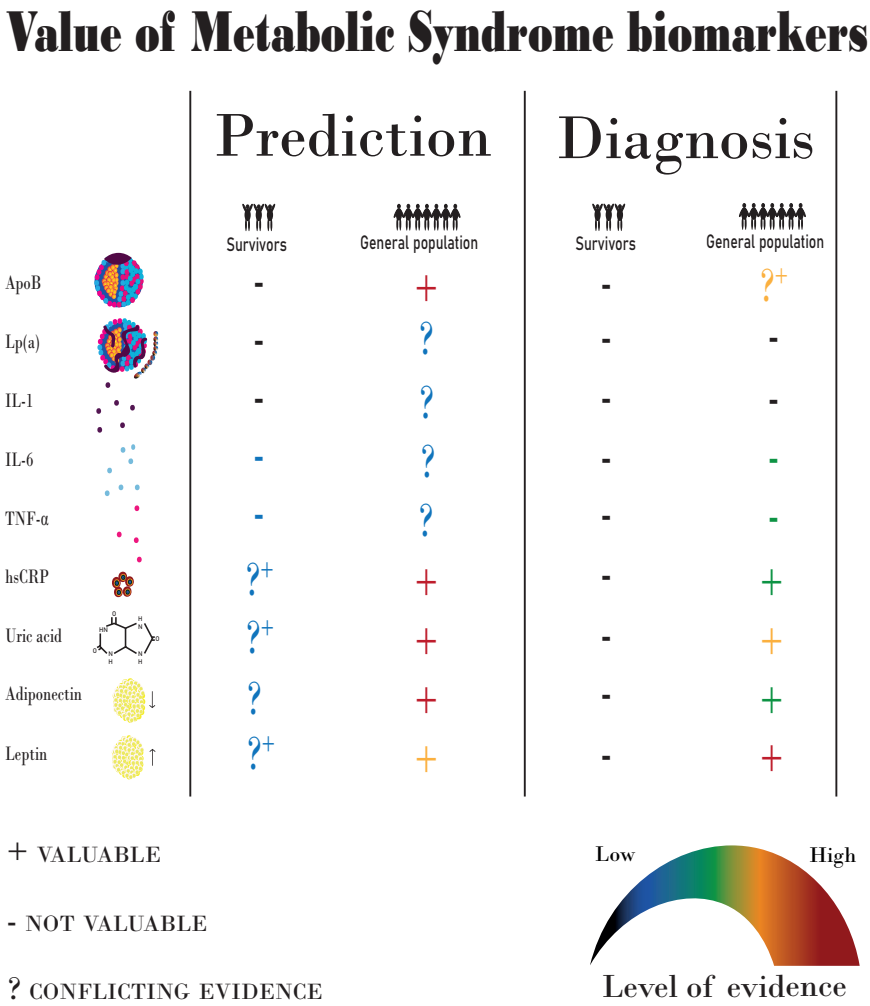
Design: A systematic literature review and meta-analysis on the diagnostic and predictive value of nine newer MetS related biomarkers (adiponectin, leptin, uric acid, hsCRP, TNF-alpha, IL-1, IL-6, apolipoprotein B (apoB), and lipoprotein(a) [lp(a)]) in survivors and adult non-cancer survivors was performed by searching PubMed and Embase. Evidence was summarized with GRADE after risk of bias evaluation (QUADAS-2/QUIPS). Eligible studies on promising biomarkers were pooled.

Results: We identified 175 general population, and 5 CCS studies. In the general population, valuable predictive biomarkers are uric acid, adiponectin, hsCRP and apoB (high level of evidence) and leptin (moderate level of evidence). Valuable diagnostic biomarkers are hsCRP, adiponectin, uric acid and leptin (low, low, moderate and high level of evidence, respectively). Meta-analysis showed OR for hyperuricemia of 2.94 (age-/sex-adjusted), OR per unit uric acid increase of 1.086 (unadjusted), and AUC for hsCRP of 0.71 (unadjusted).

Conclusions: Uric acid, adiponectin, hsCRP, leptin, and apoB can be alternative biomarkers in the screening setting for MetS in survivors, to enhance early identification of those at high risk of subsequent complications.

Keywords: childhood cancer survivors, the metabolic syndrome, biomarker, systematic review

Graphical abstract



Graphical abstract of systematic review for novel biomarkers for diagnosis and prediction of the metabolic syndrome in childhood cancer survivors and a young general population. Conclusions per biomarker are categorized as valuable, not valuable or conflicting evidence, and the level of evidence is expressed per color from low to high (black-blue-green-yellow-red; see figure on the lower right).

Introduction

Childhood cancer 5-year survival rates have increased from 5-30% in early seventies to more than 80% in the present time.[1, 2] Deployed therapies, such as chemotherapy, radiotherapy, and stem cell transplantation, better stratification and enhanced supportive care regimens, are responsible for increase in survival rates. However, intensification of treatment is also associated with long-term excess mortality and morbidities in survivors.[3] Survivors have a high level of frailty, suggesting their biological age progresses faster than their actual age. Consequently, survivors with an actual mean age of 33 have a biological age of 65 if they are compared to the general population.[4-9] At the age of 45-50 years, the prevalence of any chronic health condition is very high, from 95% up to 99%.[3, 10] One of these severe conditions is represented by cardiovascular disease (CVD), which is an important cause of premature death beyond 5 years cancer survival; the standardized mortality risk for CVD ranges from 1.9 to 12.7.[11-25]

This high risk of cardiovascular death is not only due to treatment effects, such as anthracycline exposure and cardiac irradiation;[26] survivors are also at high risk of type II diabetes mellitus (T2DM) and the metabolic syndrome (MetS).[11] These diseases are independent predictors of CVD and associated with factors such as adiposity, dyslipidemia, insulin resistance, and hypertension. These factors cluster together and form the 'deadly quartet', a MetS concept developed by Reaven in 1988.[27] The MetS had many definitions ever since.[11, 27-37] Patients with MetS carry a doubled risk of dying from cardio- and cerebrovascular disease.[11, 38] In addition, patients with the MetS are five times more likely to develop T2DM, which subsequently triples the risk of CVD.[11, 39-41]

As survivors develop cardiovascular complications at a relatively young age, there is a need for early diagnosis of MetS, to possibly prevent T2DM and CVD, and to improve long-term survival.[11] The occurrence of MetS may be underestimated especially in abdominally irradiated childhood cancer survivors (CCS), who have an unreliable waist circumference, while their MetS risk is even higher.[11, 42-44] Body mass index (BMI) and bioimpedance are alternative methods for

body composition measurement, but do not specifically measure abdominal fat, rely on hydration status and often underestimate body fat.[42, 45-47] Obviously, another alternative option to evaluate adiposity is measuring fat percentage by Dual-energy X-ray Absorptiometry (DXA) scan, which is the gold standard in case of suspected discordance of anthropomorphic measurements and adiposity.[42, 48, 49] However, performing DXA scans in all survivors on a routine basis is time-consuming and costly.[11] Additionally, there is currently no consensus for the threshold of fat percentage for diagnosing obesity.[50] Newer serum biomarkers may serve as another alternative for accurate early diagnosis or prediction of (disguised) MetS in CCS. Adult cardiologists currently apply multiple biomarkers that have been shown to improve risk estimation for CVD.[51]

Therefore, our primary objectives were to evaluate the value of the use of these newer serum biomarkers as (1) diagnostic marker, and as (2) additional independent predictor for the occurrence of MetS later in life, in survivors of childhood cancer specifically, as well as in a relatively young general, non-cancer population (studies with >75% of participants below 65 years). By including this selection of general population studies as well, we aimed to cover all available literature applicable and generalizable to young-adult survivors. To accomplish this, we performed a systematic literature search on adipokines adiponectin and leptin, uric acid, the inflammatory markers high sensitivity C-reactive protein (hsCRP), Tumor Necrosis Factor alpha (TNF-alpha), interleukin 1 (IL-1), and interleukin 6 (IL-6), and the lipid markers apolipoprotein B (apoB) and lipoprotein(a) [Lp(a)], and performed a meta-analysis of these outcomes for relevant recurrently published biomarkers. As secondary purpose, we screened for other new biomarkers that are not enlisted above, in order to reveal additional, potentially useful biomarkers.

Methods

The Systematic Search

A systematic literature review was performed in PubMed and Embase, to gather all published literature published between the first of October 2009 and September 3, 2020. Details of the search terms are available in Supplementary Table 1; in general, the search terms were related to adults/general population, as well as to (childhood) cancer survivors, and combined with all enlisted 9 separate biomarkers (adiponectin, leptin, uric acid, hsCRP, TNF-alpha, IL-1, IL-6, apoB and lp(a)) and the MetS. The AMSTAR checklist for systematic reviews was followed.[52] All titles and abstracts were screened by two independent reviewers (VP and SSvS), who were blinded to each other's judgement. Studies were included if they had the MetS as outcome, and one or more newer biomarker(s) as independent variable included in the model in predictive studies, or as discriminative variable in diagnostic studies. For studies performed in CCS, no limits were set for sample size or age. General population studies were eligible if the sample size was roughly 250 or larger and if 75% or more of this population was below 65 years of age, as they have comparable levels of frailty to a young adult survivor population.[5, 7, 8] We excluded studies with older adults since they are expected to have higher levels of frailty, comorbidities and aging factors, which may be confounders in the correlation between the newer biomarker and the metabolic syndrome. Multivariable analysis was mandatory for article inclusion of studies that investigated the prediction of MetS.

Studies were excluded if all included patients had an elevated biomarker; if all or none of the subjects had the MetS; if it was a selected cohort with pre-existing comorbidities (i.e. familial hypercholesterolemia, psoriasis, schizophrenia, polycystic ovary syndrome, obesity, hypertension); if all patients suffered from MetS or endpoint(s) such as T2DM, cardio- or cerebrovascular disease, or non-alcoholic fatty liver disease; if the article was a review, case study, expert opinion or conference abstract; if the article was written in a language other than English or Dutch, or if the full text was unavailable

(see Appendix A for an overview of selection criteria). Studies were only included if the outcome was presence or absence of MetS; those with separate MetS components or MetS risk score as outcome were out of the scope of this review. After all articles were screened based on title and abstract, the judgements were unblinded. Discrepancies were discussed and resolved by the two reviewers (VP and SSvS) and where necessary, two senior experts were consulted (MMvdHE and SJCMMN). A cross-reference check was performed with Scopus, to screen all forward and backward citations of included studies. The articles found by the cross-reference check were screened likewise. A flow diagram with the number of in- and excluded articles and reasons for exclusion illustrates this process (Figure 1).

Risk of bias assessment

The QUIPS tool was applied for critical appraisal of predictor studies[53, 54] (Supplementary Table 2) and QUADAS-2 tool for diagnostic studies (Supplementary Table 3). Definitions for low risk of bias judgement are shown in Appendix A. In case of doubt, the study was discussed with both reviewers and senior experts (VP, SSvS, MMvdHE, SJCMMN).

Data extraction enlisted novel biomarkers

Data of all included articles were extracted and summarized; the summaries of the enlisted newer biomarkers (adiponectin, leptin, uric acid, hsCRP, TNF-alpha, IL-1, IL-6, apoB and lp(a)) are depicted in Supplementary Table 4A-V. Data of interest are details regarding the size of the population and its type (survivors and their previous diagnosis, or general population), the study design (cross-sectional or longitudinal and retrospective or prospective), the biomarker (which and how it was measured), the exact outcome (MetS definition) and statistical analysis of choice. For studies investigating the diagnostic value of the biomarker for MetS, outcomes of interest were area under the curve (AUC) of receiver operating characteristic (ROC) curves, sensitivity and specificity. For the studies evaluating the predictive value of the biomarker of later development of the MetS, odds ratios (OR's) or beta-coefficients of multivariable logistic regression models, or hazard ratios (HR's) from multivariable Cox Proportional Hazards analysis were extracted from the publications.

Table 1 Commonly used metabolic syndrome definitions in selected studies

	NCEP ATP III	IDF 2006	Joint interim statement/ Harmonized definition	Japanese Obesity Society
Required for MetS diagnosis	3 or more criteria	Obesity plus 2 or more criteria	3 or more criteria	Obesity plus 2 or more criteria
Obesity	Waist circumference >102cm (men) or >88cm (women)	Waist circumference >90cm (men) or >80cm (women)	Waist circumference with ethnic-specific thresholds	Waist circumference >85cm (men) or >90cm (women)
Insulin resistance	Fasting plasma glucose ≥5.6 mmol/L or treatment	Fasting plasma glucose ≥5.6 mmol/L or treatment	Fasting plasma glucose ≥5.6 mmol/L or treatment	Fasting plasma glucose ≥6.1 mmol/L or treatment
Dyslipidemia	Triglycerides ≥1.7 mmol/L or treatment	Triglycerides ≥1.7 mmol/L or treatment	Triglycerides ≥1.7mmol/L or treatment	Triglycerides ≥1.7mmol/L, or HDL cholesterol <1 mmol/L (men) or <1.3 mmol/L (women), or treatment
	HDL cholesterol <1mmol/L (men) or <1.3 mmol/L (women) or treatment	HDL cholesterol <1mmol/L (men) or <1.3mmol/L (women) or treatment	HDL cholesterol <1mmol/L (men) or <1.3mmol/L (women) or treatment	
Hypertension	≥130/85mmHg or treatment	≥130/85mmHg or treatment	≥130/85mmHg or treatment	≥130/85mmHg or treatment

	Chinese Diabetes Society	Children and adolescents	Modified with BMI instead of waist circumference
Required for MetS diagnosis	3 or more criteria	3 or more criteria	
Obesity	Body mass index ≥25kg/m^2	Waist circumference ≥90th percentile	Body mass index ≥30 (Caucasians) or ≥25 (Asians) kg/m^2

Insulin resistance	Fasting plasma glucose ≥6.1 mmol/L or treatment	Fasting plasma glucose ≥5.6 mmol/L or treatment	
Dyslipidemia	Triglycerides ≥1.7 mmol/L, or HDL cholesterol <0.9 mmol/L (men) or <1.0mmol/L (women), or treatment	Triglycerides ≥1.7mmol/L or treatment	
		HDL cholesterol <1mmol/L or treatment	
Hypertension	≥140/90mmHg or treatment	≥130/85mmHg or treatment	

Table 2 Summary of outcomes

Summary of outcomes in diagnostic studies				
Biomarker	**Total number of studies and participants**	**Outcome**	**Number of studies**	**Range**
Leptin, in general population	6 studies, 8,209 participants[68,102,106,54,236,237]	AUC	5[68,102,106,236,237]	0.68-0.93
		Sensitivity	3[102,154,237]	48.0-92.6%
		Specificity	3[102,154,237]	56.3-72.0%
Uric acid, in general population	9 studies, 73,190 participants[66,101,150,237-242]	AUC	7[66,101,150,237,239,240,242]	0.56-0.85
		Sensitivity	3[237-239]	38.0-76.0%
		Specificity	3[237-239]	56.0-85.0%
Adiponectin, in general population	12 studies, 21,888 participants[63,65,67-69,102,106,140,143,243-245]	AUC	12[63,65,67-69,102,106,140,143,243-245]	0.55-0.92
		Sensitivity	2[102,243]	64.7-69.3%
		Specificity	2[102,243]	56.0-66.0%
hsCRP, in general population	7 studies, 18,211 participants[64,91,100,101,208,246,247]	AUC	6[64,91,100,101,208,247]	0.55-0.74
		Sensitivity	3[208,246,247]	51.0-69.0%
		Specificity	3[208,246,247]	56.6-72.0%
ApoB, in general population	1 study, 8,120 participants[111]	AUC	1[111]	0.68
TNF-alpha, in general population	1 study, 976 participants[64]	AUC	1[64]	0.54
IL-6, in general population	1 study, 976 participants[64]	AUC	1[64]	0.56
IL-1 and lp(a), in general population	No studies	n.a.	n.a.	n.a.

All biomarkers, in survivors	No studies	n.a.	n.a.	n.a.

Summary of outcomes in prognostic studies				
Biomarker	**Total number of studies and participants**	**Outcome**	**Number of studies**	**Range**
Uric acid, in general population	78 studies, 447,559 participants[71,72,75,77,82,88,90,92-96,98,99,101,150,207,210,238,239,241,242,248-303]	OR dichotomous	21[77,82,93-96,98,150,248,254,257,258,264,266,267,274,280,287,296,297,304]	1.00-5.17
		OR per unit	19[90,92,98,99,101,238,249,250,255,260,262,263,265,283,285,288,289,298]	1.001-2.14
		OR per unit logtransformed	2[239,302]	1.16, 2.08
		OR highest quantile	24[72,95,251,253,256-259,261,268,271,272,275,277-279,281,282,290-292,296,301,303]	1.00-8.04
		HR dichotomous	5[207,239,242,270,300]	1.06-2.99
		HR per unit	4[241,285,286,294]	1.10-2.35
		HR per SD	3[207,210,295]	0.86-1.36
		HR highest quantile	8[207,241,252,269,286,293-295]	0.74-3.47
		HR per unit longitudinal increase	2[207,285]	1.05, 1.31
		RR per unit logtransformed	1[276]	7.25 for men, 13.26 for women
		RR per SD	1[71]	1.10
		RR per 1.4mg/dl	1[88]	1.54 for men, 1.82 for women
		RR highest quantile	2[71,299]	1.69, 1.76
		PR	2[75,284]	1.47, 2.10
		IRR	1[75]	1.73

Uric acid, in survivors	2 studies, 390 survivors[58,59]	MetS prevalence in uric acid Q4 vs. Q1-3	1[59]	28.5% vs. 12.5% (P = 0.0044)
		MetS component(s) prevalence	1[58]	60% vs. 24% (P =0.04)
		high vs low uric acid		
Adipo-nectin, in general population	38 studies, 56,656 participants[65,67,69,70,73,74,76,78,81,102,103,105,140-148,159,245,305-319]	OR dichotomous (low adiponectin)	2[78,103]	0.90, 2.68
		OR per unit	9[67,73,76,141,144,148,159,309,314]	0.66-1.08
		OR per 5 units	1[69]	0.82 for men, 0.90 for women
		OR per unit logtransformed	4[74,146,245,306]	0.10-0.67
		OR per SD	2[315,317]	0.50-0.91
		OR per unit logtransformed Z-score	1[102]	0.76 for boys, 0.69 for girls
		OR highest quantile	13[69,105,140,142,143,145,147,305,308,315,317-319]	0.10-0.67
		OR lowest quantile	6[65,81,307,312,313,316]	1.82-18.6
		HR high baseline and increase during follow-up vs low baseline and decrease	1[311]	0.33
		HR decreased at follow-up	1[81]	4.37
		Time ratio of developing MetS Q1 vs Q4	1[310]	0.15 (=85% shorter time to develop MetS)

		Baseline ratio (value in	1[70]	1.27
		MetS subjects divided by value in non-MetS, adjusted for covariates)		
Adipo-nectin, in survivors	3 studies, 139 survivors[58,60,62]	OR highest quantile at baseline and follow-up	1[62]	0.5 (n.s.) for baseline, 0.9 (n.s.) for follow-up
		HR dichotomous (low adiponectin)	1[58]	6.7
		P-value of Kruskal-Wallis test median adiponectin in 0, 1, 2-4 MetS components	1[60]	n.s.
hsCRP, in general population	32 studies, 119,138 participants[70,74, 83-85,88,90,91,147,155,193, 195-199,208,246,249,265, 269,320-330]	OR dichotomous	2[155,196]	1.20, 2.74
		OR per unit	7[90,91,198,199,249, 328,330]	1.007-2.97
		OR per unit logtransformed	4[74,246,265,324]	1.15-3.2
		OR per SD	1[85]	1.21
		OR per unit logtransformed	2[208,325]	0.96, 1.07
		OR highest quantile	11[84,147,193,197,321-323,325-327,329]	1.07-7.11
		OR highest of three groups (<1.0, 1.0-3.0 and >3 µg/ml)	3[195,320,324]	1.65-18.86
		HR per unit logtransformed	1[269]	1.15
		RR per threefold increase	1[88]	1.13
		Baseline ratio	1[70]	0.80 (n.s.)
		P-value of likelihood test in multivariable model	1[83]	n.s.

hsCRP, in survivors	1 study, 87 survivors and 87 controls[61]	OR dichotomous	1[61]	7.26
ApoB, in general population	10 studies, 66,924 participants[74,79,82,86,87,108,109,331-333]	OR dichotomous	1[82]	2.55
		OR per unit	1[74]	2.99
		OR per 30mg/dl	1[87]	1.76 for men, 2.10 for women
		OR per SD	1[331]	1.56
		OR highest quantile	6[79,108,109,331-333]	0.96-6.03
		OR highest of three groups (<90, 90-119 and ≥120 mg/dl)	1[86]	2.69 for men, 1.69 for women
		RR per SD	1[331]	1.17 (n.s.)
		RR highest quantile	1[331]	1.79
ApoB, in survivors	No studies	n.a.	n.a.	n.a.
Leptin, in general population	17 studies, 28,797 participants[68,73,74,102,103,147-149,236,306,314,315,319,334-337]	OR dichotomous	1[103]	2.39
		OR per unit	4[73,148,314,336]	0.96-1.91
		OR per 10ng/ml	1[149]	1.06 (adjusted for WC), 1.22 (adjusted for BMI)
		OR per unit logtransformed	2[74,306]	1.47, 2.76
		OR per SD	3[68,315,335]	1.01-1.31
		OR per unit logtransformed Z-score	1[102]	1.81 for boys, 1.32 for girls
		OR highest quantile	6[147,236,315,319,334,337]	1.16-3.02

Leptin, in survivors	3 studies, 1 39 survivors [58,60,62]	OR highest quantile at baseline and follow-up	1[62]	4.8 for baseline, 5.7 for follow-up
		MetS component(s) prevalence high vs low leptin	1[58]	54% vs. 17% (P = 0.03)
		P-value of Kruskal-Wallis test median adiponectin in 0, 1, 2-4 MetS components	1[60]	n.s.
IL-6, in general population	5 studies, 3,370 participants[67,80,143,199,200]	OR per unit	2[67,199]	0.98-1.47
		OR highest quantile	2[143,200]	0.98 (n.s.), 4.10
		P-value in multivariable model	1[80]	n.s.
IL-6, in survivors	1 study, 87 survivors and 87 controls[61]	OR dichotomous	1[61]	1.53 (n.s.)
Lp(a), in general population	5 studies, 15,162 participants[89,320,338-340]	OR dichotomous	1[320]	8.27
		OR highest of three groups (<18.40, 18.40-33.84 and ≥33.85 µg/ml)	1[338]	0.82 (n.s.)
		OR per unit	1[339]	1.0 (n.s.)
		OR highest quantile	1[340]	0.45
		HR highest quantile	1[89]	1.01 (n.s.)
Lp(a), in survivors	No studies	n.a.	n.a.	n.a.

IL-1, in general population	4 studies, 1,594 participants[70,199,200,341]	OR per unit	2[199,341]	2.28 (IL-1 alpha), 1.009, 2.01 (IL-1 beta)
		OR highest quartile	1[200]	0.98 (n.s.)
		Baseline ratio	1[70]	1.17 (suggests effect in other direction)
TNF-alpha, in general population	3 studies, 1,458 participants[80,199,200]	OR per unit	1[199]	1.45
		OR highest quartile	1[200]	0.78 (n.s.)
		P-value in multivariable model	1[80]	n.s.
TNF-alpha, in survivors	1 study, 87 survivors and 87 controls[61]	OR dichotomous	1[61]	0.52 (n.s.)

Summary of evidence

The Grading of Recommendations Assessment Development and Evaluation (GRADE) tool was applied to summarize the quality of the evidence for each biomarker, per clinical research question (diagnosing or predicting MetS) and per population (general population and CCS).[55] The level of evidence was classified as insufficient, very low, low, moderate and high (Supplementary Table 4).[55] The applied thresholds for biomarkers are shown in Supplementary Table 5. An overview was made for studies assessing the same independent variables and outcome (Supplementary Table 6).

Data extraction non-enlisted biomarkers

As secondary objective we screened all articles for other biomarkers than the above enlisted nine biomarkers of our main interest (non-enlisted biomarkers). Details are discussed in Part 2 of the Appendix. These non-enlisted biomarkers were evaluated for presence of an effect if there were 4 or more publications with this biomarker in our search. As we did not search for these biomarkers systematically, evidence quality was not assessed with GRADE.

Meta-analysis

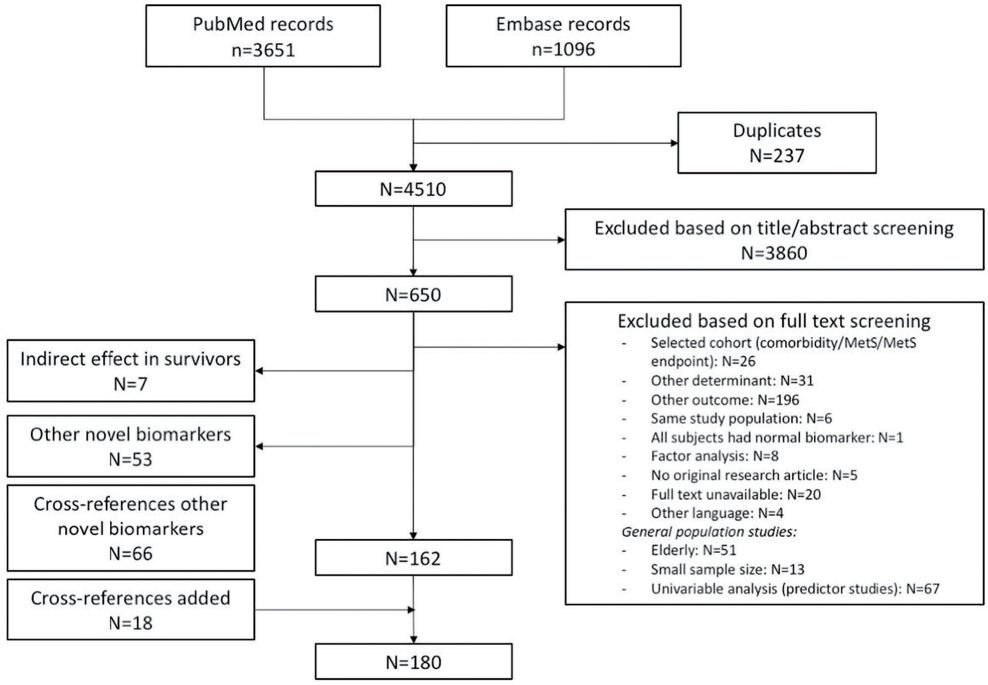
A meta-analysis was performed of relevant enlisted biomarkers with at least three publications on the same outcome measures and, if applicable, adjusted for the same covariates. Dichotomous outcomes were considered as comparable if the applied threshold differed less than the intra- and inter-assay variability for the biomarker as reported in literature. A random effects model with inverse variance weighting was used to estimate a pooled overall outcome measure. Overall heterogeneity (I-squared) and between-study variance (tau-squared) were calculated.[56] Meta-analysis was performed with the package *meta* in R.[57]

Results

Study selection

As shown in the flow chart (Figure 1), the literature search in PubMed and Embase yielded a total of 4,510 unique records. After title and abstract screening, 650 full-text articles were reviewed, after which 162 relevant studies remained. Backward and forward citation searching identified 18 additional studies. Hence, a total of 180 studies were identified that reported on the diagnostic and/or predictive value of one or more of the enlisted nine biomarkers of interest. Only five studies among the 180 were performed among a population of CCS.[58-62] All other studies were performed in the general population.

Figure 1 Flow chart of in- and excluded articles from the systematic literature search



Among 180 studies which included data regarding the 9 enlisted biomarkers, 60 also reported the value of other, non-enlisted newer biomarkers. Furthermore, we identified 119 other studies that only investigated non-enlisted newer biomarkers (other than the nine of our main interest), yielding a total of 179 studies for our secondary objective.

A detailed description of the critical appraisal of each of the 180 included studies for the nine predefined biomarkers is provided in the supplementary material (Supplementary Table 2 and 3).

Used metabolic syndrome definitions

In the included studies, a variety of MetS definitions was used of which the most common are described in Table 1 and the applied definition per study is depicted in Supplementary Table 4. The applied biomarker thresholds are summarized in Supplementary Table 5.

Evidence for newer, enlisted biomarkers as (additional) diagnostic criterion for metabolic syndrome

Twenty-nine studies reported on the diagnostic value of one or more of the nine enlisted newer biomarkers. These were all performed in the general population without a history of cancer. Six studies had a Caucasian study population.[63-68] The number of studies per biomarker ranged between zero [IL-1 and lp(a)] and twelve (adiponectin). The biomarker studied in the largest total number of participants was uric acid (73,190 participants). The relevant data extracted from each study, as well as the summary of evidence scored with the GRADE tool for each biomarker, are provided in the supplementary material (Supplementary Table 4). For each biomarker, a description of the number of studies and participants, and a summary of the several diagnostic outcomes, are provided in Table 2.

Whereas, ideally, the additional diagnostic value of a biomarker would be tested by comparing the AUC, sensitivity and specificity for a model containing only relevant covariates, versus a model containing

covariates and the newer biomarker, this method was used in only two of the 29 studies.[65, 69] One study compared the AUC of the biomarker with the AUC of waist circumference.[69] Most studies, however, only reported the AUC of the biomarker, either unadjusted or adjusted for age, sex, and sometimes BMI or waist circumference. Therefore, interpretation of the additional value is limited by detection and confounding bias for most of the biomarkers.

The overall summary of our findings, with a conclusion about the diagnostic value of each biomarker in the general population and in survivors based on the GRADE assessment, is shown in Figure 2. Of the nine investigated biomarkers, four were identified as valuable diagnostic biomarkers for MetS: leptin (high quality of evidence), uric acid (moderate quality), adiponectin, and hsCRP (both low quality). In addition, apoB may be valuable, although based on only one study with moderate quality of evidence. TNF-alpha and IL-6 appeared to be unusable, based on one low quality study testing both biomarkers. For IL-1 and lp(a), no studies were found.

Evidence for newer, enlisted biomarkers as independent predictor of metabolic syndrome

In total, 162 general population studies, and 5 survivor studies [two in acute lymphoblastic leukemia (ALL) survivors, two in survivors of hematological malignancies, one in survivors of heterogeneous tumors],[58-62] investigated the role of one or more of the nine enlisted, newer biomarkers as independent predictors of MetS. Twenty-six of the general population studies had a Western/Caucasian study population.[65, 67, 68, 70-92] The number of general population studies per biomarker ranged between 3 (TNF-alpha, 1,458 participants in total) and 78 (uric acid, 447,559 participants in total). Two of the survivors studies had a Western/Caucasian study population,[58, 59] the others were performed in Japan,[60] Malaysia,[61] and Mexico.[62] The number of survivors studies per biomarker ranged between zero [IL-1, apoB, and lp(a)] and 3 (adiponectin and leptin). The biomarker studied in the largest total number of survivors was uric acid (390 survivors). The

relevant data extracted from each study, as well as the summary of evidence scored with the GRADE tool for each biomarker, are provided in the supplementary material (Supplementary Table 4). For each biomarker, a description of the number of studies and participants, and a summary of the several prognostic outcomes, are provided in Table 2.

A common analysis strategy in these studies was to divide the biomarker value in quantiles, with thresholds that may differ per study. Not all participants in the highest or lowest quantile always had a biomarker value that would be classified as abnormal according to reference values. This may attenuate its value in predicting MetS. On the other hand, this bias towards the null hypothesis increases the effect of true positive findings. Also, several studies tested a dose-response effect by comparing the effect on MetS across the quantiles. Studies can be compared on whether a dose-response effect was observed or not.

Figure 2 Summary of conclusions: predictive and diagnostic value of novel 13 biomarkers for the MetS

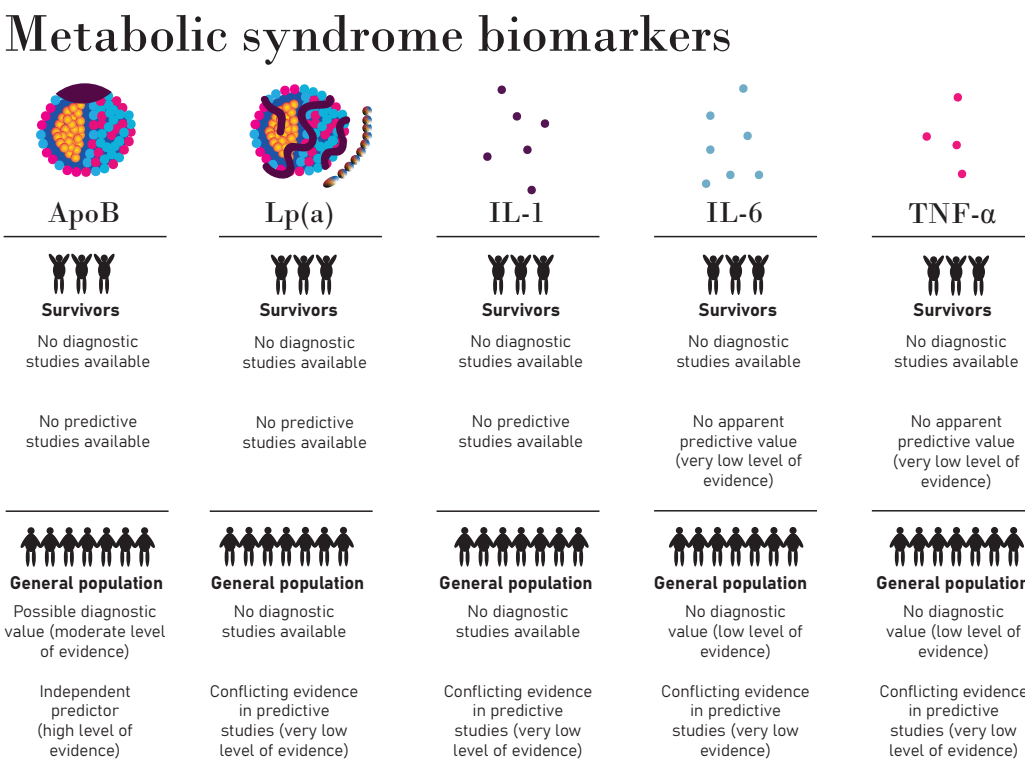


Figure 2 shows the overall summary of our findings, consisting of a conclusion about the role of each biomarker as independent predictor of MetS in the general population and in survivors, after GRADE assessment. Five biomarkers were identified as independent predictors of MetS in the general population: uric acid, adiponectin, hsCRP, apoB (all high quality of evidence), and leptin (moderate quality). There is conflicting evidence for the value of TNF-alpha, IL-1, IL-6, and lp(a) (very low quality of evidence). Among survivors, uric acid and hsCRP may be valuable as prognostic biomarkers, based on two and one studies, respectively, with very low quality of evidence. There is conflicting evidence for the prognostic value of adiponectin and leptin (very low quality). TNF-alpha and IL-6 appear not to be independent predictors, based on one very low quality study testing both biomarkers. For IL-1, apoB, and lp(a), no studies were found.

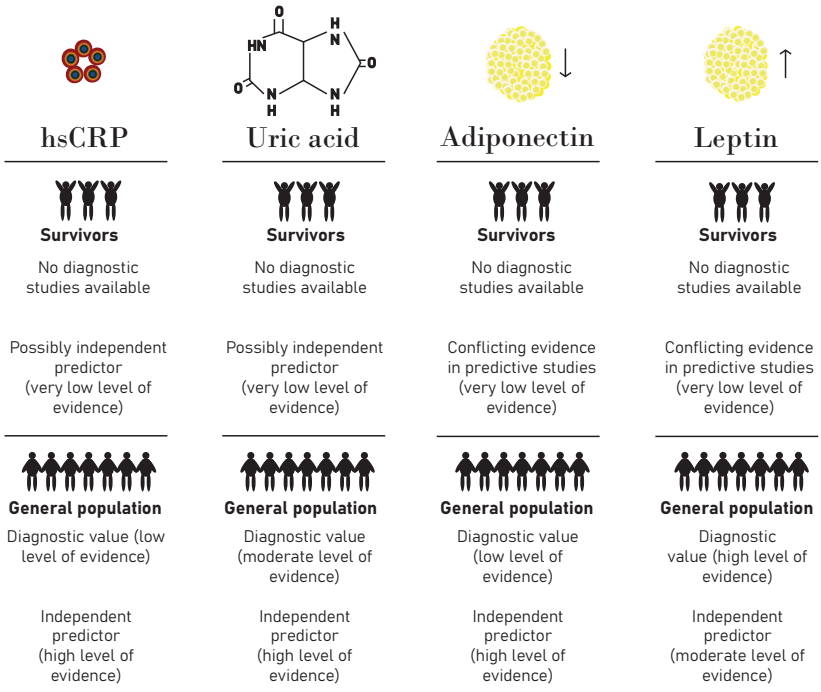
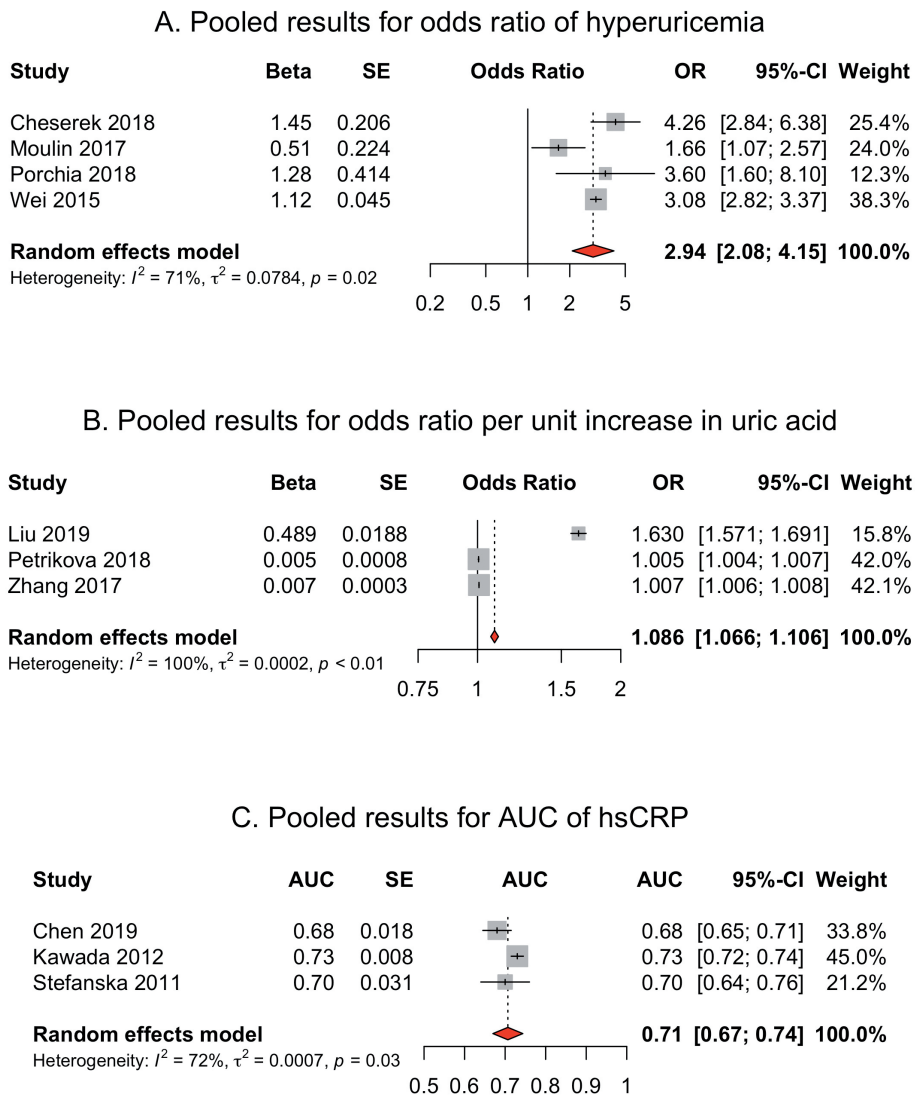


Figure 3 Forest plots for different study-specific outcomes



A. Odds ratio (OR) for hyperuricemia. B. OR for per unit increase in uric acid. C. Area under the curve (AUC) of hsCRP. The sizes of the square boxes on the forest plots are proportional to the total number of patients in the selected trials.

Meta-analysis of most relevant findings of enlisted biomarkers

We aimed to perform a meta-analysis of the most promising biomarkers: uric acid, adiponectin, leptin, hsCRP, and apoB. For diagnostic studies, only the AUC is suitable for meta-analysis, due to different thresholds used for sensitivity and specificity (Supplementary Table 6). For predictor studies, only dichotomous and continuous (per unit, or per unit log-transformed) studies are useful. Many studies use quantiles but these are unsuited for meta-analysis: cut-offs between the quantiles depend on the range and distribution in each study population, and are therefore insufficiently comparable between studies to perform a meta-analysis.

A wide variety of outcome measures was used in the studies, and many studies performed an analysis that was unsuited for meta-analysis. Also, there was variance in thresholds used for dichotomous outcomes, as well as in covariates in multivariable models. Therefore, we were unable to retain at least three sufficiently comparable studies for most biomarkers, and for most outcomes, in order to perform a meta-analysis. For a few biomarkers, enough studies were eligible for meta-analysis, because the authors also published crude outcomes, and outcomes that were only age- and sex-adjusted (Supplementary Table 6).

We were able to perform a meta-analysis for the prognostic value of uric acid (hyperuricemia and continuous uric acid levels), and for the diagnostic value of hsCRP. We estimated the pooled OR for the association between hyperuricemia and MetS, adjusted for age and sex (four studies,[93-96] with threshold variability accepted of 10%,[97] OR 2.94, 95% CI 2.08-4.15), the pooled OR per unit increase in uric acid, unadjusted (three studies,[90, 98, 99] OR 1.086, 95% CI 1.066-1.106), and the pooled AUC for hsCRP, also unadjusted (three studies,[91, 100, 101] AUC 0.71, 95% CI 0.67-0.74).[90, 99] Forest plots are shown in Figure 3. Unfortunately, many studies could not be included, and the reported estimators are not adjusted for relevant covariates, in particular age and sex for some, and overweight, insulin resistance, and smoking for all.

Other, non-enlisted biomarkers

In Supplementary Table 7, [179] articles for all other biomarkers for diagnosis or prognosis of MetS are enlisted and the main data is summarized. These included ratios of our studied biomarkers. All studies investigating leptin/adiponectin ratio as prognostic[62, 68, 73, 102-105] or diagnostic study[68, 102, 104-107] showed a possible relevance. Apolipoprotein A1 (apoA1) and apoB/apoA1 ratio seem valuable in predicting the MetS (6 studies with a protective effect of apoA1,[82, 86, 87, 108, 109] and 8 studies with an effect of increasing risk of increasing apoB/apoA1 ratio82,[108-114]). There are two studies reporting a diagnostic value of apoB/apoA1 ratio.[108, 111] Other recurrently reported, potentially useful biomarkers were Gamma GT, (non-high sensitivity) CRP, ferritin, leukocyte count, hemoglobin and urine pH and sodium excretion.

Discussion

This is the first systematic literature review investigating newer biomarkers for metabolic syndrome (MetS) in CCS, with the aim to obtain the highest level of evidence by including validated tools for risk of bias assessment and summary of evidence, and by performing a meta-analysis.

For five biomarkers, numerous studies with moderate to high quality of evidence were found for diagnosing and predicting MetS: uric acid, adiponectin, leptin, hsCRP, and apoB. The evidence was not sufficient to confirm the value of candidate biomarkers lp(a), IL-1, IL-6, and TNF-alpha.

Meta-analysis of eligible studies showed a predictive value of uric acid for MetS, with a positive association, and a diagnostic value for hsCRP.

These findings suggest that uric acid, adiponectin, leptin, hsCRP, and apoB may be used in a screening setting for CCS, in addition to standard MetS criteria, in order to provide better diagnosis and prediction of MetS (risk). Systematic reviews in other populations have identified not only elevated leptin,[115] uric acid,[115-118] and low (HWM) adiponectin,[115, 119, 120] but also Il-6[115] and TNF-alpha[115] as potential MetS biomarkers.

As anticipated, the number of publications for survivors on this topic was rather limited: we identified only five studies in CCS specifically, which found a possible predictive value for hsCRP and uric acid, and conflicting or no evidence for the value of adiponectin, leptin and TNF-alpha. Disadvantages of these survivor studies were low patient numbers and moderate to high (detection and confounding) bias risk. No studies investigated the diagnostic value of newer biomarkers. Survivor studies with information on altered biomarker values but no direct comparison between biomarker and MetS occurrence, were excluded.[60, 121-130] We expected to miss many relevant studies when designing the study, if we based our conclusions only on survivor studies. Therefore, evidence in the younger general adult population without childhood cancer history was included in our search as well, leading to 175 general population studies with relevant data which were generalizable to young adult survivors.

CCS can have an increased risk to develop MetS, in particular after treatment with cranial and/or abdominal radiotherapy, intensive chemotherapy, nephrectomy, adrenalectomy, or stem cell transplantation.[43, 131-139] These therapies can lead to several underlying conditions that can increase the risk for (components of) MetS, such as hypothalamic damage, growth hormone deficiency, pancreatic beta cell dysfunction, hypogonadism, hypothyroidism, and altered body composition with increased abdominal fat.[43, 131-139]

Furthermore, it is well acknowledged, that in CCS the biological age progresses faster than their true age, as can be derived from their high level of frailty.[4-9] Previous studies have shown, that the physiologic reserve of CCS with a median age of 33 is similar to that of adults in the general population who are aged 65 years.[6] For this reason, we included studies investigating biomarkers for MetS in the general population, with >75% of participants aged below 65 years, as may be very well applicable to CCS. We excluded studies investigating MetS biomarkers among elderly people on purpose, since they have an even higher level of frailty than CCS, comorbidities and aging factors, which may be confounders in the association between the newer biomarker and metabolic syndrome. We considered that extrapolating

conclusions from a general elderly population to CCS could draw invalid conclusions. Based on this approach, all available literature applicable to survivors is now discussed in this review, as it includes both survivor studies as well as all generalizable data from a reasoned selection of the general population studies.

On the other hand, several studies excluded people with certain chronic illnesses.[73, 101, 140-150] This may limit applicability of results to the population of CCS, in which the prevalence of comorbidities is high.[3, 25, 126, 151] This was taken into account when scoring the risk of bias. Additionally, childhood cancer (treatment) related long-term side effects, such as altered fat distribution, sarcopenic obesity, and hormonal disbalances, may play a survivor specific role in the pathogenesis of MetS;[11] development of future studies that apply the use of biomarkers in large cohorts of CCS is therefore important.

Due to differences in study designs and statistical analyses, a wide variety of outcome measures was used. There was also substantial diversity in follow-up time in longitudinal studies. By employing the GRADE tool for summarizing evidence, we were able to draw conclusions for each biomarker from this heterogeneity of results. The meta-analysis was based on few studies, as many studies could not be included. Also, heterogeneity was high in the meta-analysis on uric acid per unit increase, as the study of Liu et al. had a remarkably higher OR than the other two studies.[98]

Furthermore, although the ability of different MetS definitions to predict diabetes and CVD appears to be similar,[152, 153] the use of different definitions (Table 1) can lead to differences in occurrence of MetS. There are subtle differences between the definitions that were mostly used in the included studies (Table 1). The potential consequence of choice of definition is illustrated by studies that tested the biomarker use in diagnosing or predicting MetS according to multiple definitions, and sometimes found different results depending on the definition used.[67, 143, 154, 155] Therefore, comparing different studies and interpreting results of the meta-analysis requires some caution, as a full comparison of the studies is often not possible.

Adiposity, and hence the MetS, can be underdiagnosed in survivors, due to altered body composition after radiotherapy, stem cell transplantation, or amputations. For clinical applicability to survivors, it is important that newer biomarkers play an independent role in MetS, and measurement of newer biomarkers is only useful when their effect is not yet captured by established MetS components. Therefore, we did not investigate routine dyslipidemia and insulin resistance markers in our search (e.g., LDL, HOMA-IR). Although apoB and lp(a) are also lipid markers, they are of interest because they are better predictors of atherogenicity than triglycerides, HDL and LDL – particularly apoB, because it gives an estimate of the total number of circulating atherogenic particles.[156-158]

In this light, it is also favorable that studies adjust for MetS components, such as adiposity and insulin resistance, in order to adjust for potentially major correlations and interactions,[159-162] and to yield the independent/additional diagnostic and predictive value of the biomarker. Furthermore, it remains important to evaluate other traditional risk factors, including smoking, physical activity, socio-economic status, and family history.[78,163] In addition, genetic profile may still be relevant for MetS risk, although so far this is not included in standard screening.[164-166] Risk of detection and confounding bias remains high, especially in the diagnostic studies, as many studies did not adjust for MetS components and traditional risk factors. In particular for the diagnostic studies, a risk of (detection and) confounding bias remained.

The MetS is defined as a cluster of symptoms such as obesity, hypertension, impaired glucose tolerance and dyslipidemia.[11] These clustered symptoms are related to each other: an imbalance in energy intake and consumption causes a cascade of increased (visceral) adiposity, increased circulating free fatty acids and decreased adiponectin (which causes also an increase in insulin resistance), and high levels of pro-inflammatory and pro-thrombotic mediators, such as TNF-alpha, IL-1 and IL-6.[11, 34] Insulin resistance is associated with a lowered excretion of uric acid by the kidneys, and higher uric acid

production.[167, 168] The adipokines leptin and adiponectin are produced by adipocytes.[169] Low leptin values trigger metabolic, behavioral and endocrine responses that aim at a preservation of the fuel reserves of the body.[170] Adiponectin enhances insulin sensitization and suppresses inflammation and cell death.[170-173] Another important molecule is apoB: all atherogenic lipoproteins carry one single apoB molecule as their structural protein, and therefore apoB represents the atherogenic burden.[174] Serum apoB is a strong predictor of cardiovascular risks[156, 175, 176] and comes in as an important player for the MetS in this review as well. One of the low density lipoproteins carrying an apoB molecule, is Lp(a).[177] The interpretation of Lp(a) values in an individual can be difficult due to a high heterogeneity and wide distribution of Lp(a) concentrations.[178] Although evidence for relevance of Lp(a) for MetS evaluation in survivors was unavailable, it remains a marker of interest, since elevated Lp(a) levels were an independent predictor for cardio- and cerebrovascular outcomes[179-187] and were inversely associated with T2DM.[188]

An important inflammatory marker is (hs)CRP, which is synthesized by hepatocytes[189, 190] in response to infection, inflammation, tissue damage and malignant neoplasia.[189, 190] CRP binds to LDL[189, 191] and may have a causal role in atherogenesis,[189] as it is present in atherosclerotic plaques.[189, 192] Inflammatory markers may reflect a transient state instead of chronic state of inflammation.[193] Still, in the study of Oda et al., the diagnostic value of hsCRP was reproducible when the measurement was repeated after one year.[194] Many studies had a high CRP, or infection[100, 197-200] as exclusion criterion. Regarding inflammation; smooth muscle cells, endothelial cells and macrophages produce cytokines such as IL-1 and IL-6[201-203] in reaction to metabolic stress,[203, 204] by other inflammatory mediators such as interferon-gamma and TNF, and cholesterol itself.[203] Still, the evidence for the usefulness as marker for the MetS is rather limited.

Due to the systemic nature of MetS, our secondary objective to reveal other interesting biomarkers yielded many markers. Interesting markers for further research include Gamma GT, ferritin, leukocytes and hemoglobin. In several studies biomarkers were related to each other,

as MetS components are related as well.[205] In one study, leptin was inversely associated with uric acid excretion;[205] in another, a synergistic effect between hsCRP and high molecular weight adiponectin was found.[206] Also, ratios of biomarkers (e.g. leptin/adiponectin, apoB/apoA1) include extra information and may be better diagnostic or prognostic agents than single biomarkers. Future studies may investigate the value of combining biomarkers.

Some limitations are present in this systematic literature.

Many of the included studies had a cross-sectional design, which is suboptimal to investigate causality; this was taken into account for the GRADE and level of evidence. Some authors conducted prospective longitudinal studies[81, 193, 207, 208] and associated MetS risk at end of follow-up with baseline and/or change in biomarker level. Study designs even more suitable for determining prediction and causality include prediction models and Mendelian randomization.[209-212] These study designs require more time and financial resources, and large cohorts. These types of studies where either not performed or unsuitable for our research question.

Many studies were performed among Asian cohorts. Asian people are more susceptible to insulin resistance,[213, 214] which is accounted for in lower waist circumference thresholds. Additionally, there may be an ethnicity specific component in the relationship between biomarker and MetS.[215-222] This may limit the applicability to a Caucasian population.

For this literature study, we focused on diagnosis and prediction of the full MetS; other outcomes such as resolution of the MetS,[223] components of the MetS, CVD or T2DM were out of scope.[186, 224-235] Therefore, our findings do not provide a complete overview of the use of the newer biomarkers in diagnosing and predicting cardiovascular risk factors in CCS.

We have two suggestions for future research that are relevant for the implementations of our findings in the follow-up of CCS. The newer biomarkers could be added as a sixth criterion for MetS. This application

can be especially of value in cases of doubt of MetS diagnosis for individuals who had abdominal irradiation: it may be valuable to replace waist circumference with the adipokines leptin or adiponectin. This may identify MetS in more survivors, and can potentially improve the predictive ability for T2DM and CVD.[231]

An important requirement for the applicability of these newer biomarkers in such a screening setting for MetS (risk) in CCS, is the determination of a threshold. For uric acid, this is relatively well-established (Supplementary Table 5); for other biomarkers, this is less clear, as is illustrated by the range of applied thresholds (Supplementary Table 5). This is partly because of the use of different assays and testing of subfractions of a biomarker, such as high molecular weight adiponectin. Also, a tradeoff between sensitivity and specificity may influence the determination of an optimal threshold.

In conclusion, based on this systematic literature search, we suggest to consider the additional use of uric acid, adiponectin, hsCRP, leptin, and apoB in the screening setting for metabolic syndrome in CCS. As our conclusions are largely based on general population studies, studies in CCS are needed. Furthermore, future studies may specifically test the use of newer biomarkers as additional MetS components, and define optimal thresholds. The addition of one or more of these newer biomarkers as a criterion for MetS may lead to a newer and better classification and enhanced identification of risk of developing T2DM and CVD, especially in CCS in whom components are difficult to evaluate in the currently applied definitions. Early intervention can delay or prevent complications, and hence improve very long-term survival outcomes and quality of life.

References

1. Howlader N, Noone AM, Krapcho M, Miller D, Bishop K, Kosary CL, Yu M, Ruhl JTZ, Mariotto A, Lewis DR, Chen HS, Feuer EJ, Cronin KA, SEER Cancer Statistics Review (CSR) 1975-2014. National Cancer Institute: Bethesda, MD 2017.
2. Gibson TM, Mostoufi-Moab S, Stratton KL, Leisenring WM, Barnea D, Chow EJ, et al. Temporal patterns in the risk of chronic health conditions in survivors of childhood cancer diagnosed 1970-99: a report from the Childhood Cancer Survivor Study cohort. The Lancet Oncology. 2018; 19: 1590-601.
3. Hudson MM, Ness KK, Gurney JG, Mulrooney DA, Chemaitilly W, Krull KR, et al. Clinical ascertainment of health outcomes among adults treated for childhood cancer. Jama. 2013; 309: 2371-81.
4. Ness KK, Howell CR, Bjornard KL. Frailty and quality of life in adult survivors of childhood cancer. Expert review of quality of life in cancer care. 2017; 2: 79-85.
5. Ness KK, Armstrong GT, Kundu M, Wilson CL, Tchkonia T, Kirkland JL. Frailty in childhood cancer survivors. Cancer. 2015; 121: 1540-7.
6. Ness KK, Kirkland JL, Gramatges MM, Wang Z, Kundu M, McCastlain K, et al. Premature Physiologic Aging as a Paradigm for Understanding Increased Risk of Adverse Health Across the Lifespan of Survivors of Childhood Cancer. Journal of clinical oncology: official journal of the American Society of Clinical Oncology. 2018; 36: 2206-15.
7. Ness KK, Wogksch MD. Frailty and aging in cancer survivors. Translational research : the journal of laboratory and clinical medicine. 2020; 221: 65-82.
8. Armenian SH, Gibson CJ, Rockne RC, Ness KK. Premature Aging in Young Cancer Survivors. Journal of the National Cancer Institute. 2019; 111: 226-32.
9. Vatanen A, Hou M, Huang T, Söder O, Jahnukainen T, Kurimo M, et al. Clinical and biological markers of premature aging after autologous SCT in childhood cancer. Bone marrow transplantation. 2017; 52: 600-05.
10. Bhakta N, Liu Q, Ness KK, Baassiri M, Eissa H, Yeo F, et al. The cumulative burden of surviving childhood cancer: an initial report from the St Jude Lifetime Cohort Study (SJLIFE). Lancet (London, England). 2017; 390: 2569-82.
11. Pluimakers VG, van Waas M, Neggers S, van den Heuvel-Eibrink MM. Metabolic syndrome as cardiovascular risk factor in childhood cancer survivors. Critical reviews in oncology/hematology. 2019; 133: 129-41.

12. Reulen RC, Winter DL, Frobisher C, Lancashire ER, Stiller CA, Jenney ME, et al. Long-term cause-specific mortality among survivors of childhood cancer. Jama. 2010; 304: 172-9.

13. Tukenova M, Guibout C, Oberlin O, Doyon F, Mousannif A, Haddy N, et al. Role of cancer treatment in long-term overall and cardiovascular mortality after childhood cancer. Journal of clinical oncology: official journal of the American Society of Clinical Oncology. 2010; 28: 1308-15.

14. Castellino SM, Geiger AM, Mertens AC, Leisenring WM, Tooze JA, Goodman P, et al. Morbidity and mortality in long-term survivors of Hodgkin lymphoma: a report from the Childhood Cancer Survivor Study. Blood. 2011; 117: 1806-16.

15. Prasad PK, Signorello LB, Friedman DL, Boice JD, Jr., Pukkala E. Long-term non-cancer mortality in pediatric and young adult cancer survivors in Finland. Pediatric blood & cancer. 2012; 58: 421-7.

16. Armstrong GT, Oeffinger KC, Chen Y, Kawashima T, Yasui Y, Leisenring W, et al. Modifiable risk factors and major cardiac events among adult survivors of childhood cancer. Journal of clinical oncology: official journal of the American Society of Clinical Oncology. 2013; 31: 3673-80.

17. Perkins SM, Fei W, Mitra N, Shinohara ET. Late causes of death in children treated for CNS malignancies. Journal of neuro-oncology. 2013; 115: 79-85.

18. Kero AE, Järvelä LS, Arola M, Malila N, Madanat-Harjuoja LM, Matomäki J, et al. Cardiovascular morbidity in long-term survivors of early-onset cancer: a population-based study. International journal of cancer. 2014; 134: 664-73.

19. van Laar M, Feltbower RG, Gale CP, Bowen DT, Oliver SE, Glaser A. Cardiovascular sequelae in long-term survivors of young peoples' cancer: a linked cohort study. British journal of cancer. 2014; 110: 1338-41.

20. Olsen M, Schmidt M, Lash TL, Sorensen K, Pedersen L, Sorensen HT. Cardiovascular disease risk in childhood cancer survivors. American journal of epidemiology. 2014; 180: 120-3.

21. Kero AE, Järvelä LS, Arola M, Malila N, Madanat-Harjuoja LM, Matomäki J, et al. Late mortality among 5-year survivors of early onset cancer: a population-based register study. International journal of cancer. 2015; 136: 1655-64.

22. Gudmundsdottir T, Winther JF, de Fine Licht S, Bonnesen TG, Asdahl PH, Tryggvadottir L, et al. Cardiovascular disease in Adult Life after Childhood Cancer in Scandinavia: A population-based cohort study of 32,308 one-year survivors. International journal of cancer. 2015; 137: 1176-86.

23. Bhakta N, Liu Q, Yeo F, Baassiri M, Ehrhardt MJ, Srivastava DK, et al. Cumulative burden of cardiovascular morbidity in paediatric, adolescent, and young adult survivors of Hodgkin's lymphoma: an analysis from the St Jude Lifetime Cohort Study. The Lancet Oncology. 2016; 17: 1325-34.

24. Schindler M, Spycher BD, Ammann RA, Ansari M, Michel G, Kuehni CE. Cause-specific long-term mortality in survivors of childhood cancer in Switzerland: A population-based study. International journal of cancer. 2016; 139: 322-33.

25. Kero AE, Madanat-Harjuoja LM, Järvelä LS, Malila N, Matomäki J, Lähteenmäki PM. Cardiovascular medication after cancer at a young age in Finland: A nationwide registry linkage study. International journal of cancer. 2016; 139: 683-90.

26. Armstrong GT, Liu Q, Yasui Y, Neglia JP, Leisenring W, Robison LL, et al. Late mortality among 5-year survivors of childhood cancer: a summary from the Childhood Cancer Survivor Study. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2009; 27: 2328-38.

27. Reaven GM. Banting lecture 1988. Role of insulin resistance in human disease. Diabetes. 1988; 37: 1595-607.

28. Wittcopp C, Conroy R. Metabolic Syndrome in Children and Adolescents. Pediatrics in review. 2016; 37: 193-202.

29. Kaplan NM. The deadly quartet. Upper-body obesity, glucose intolerance, hypertriglyceridemia, and hypertension. Archives of internal medicine. 1989; 149: 1514-20.

30. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, Hammarstrand C, Janssen J, Van der Lely AJ, et al. The metabolic syndrome and its components in 178 patients treated for craniopharyngioma after 16 years of follow-up. European journal of endocrinology. 2017; 178: 11-22.

31. Alberti KG, Zimmet PZ. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. Diabetic medicine: a journal of the British Diabetic Association. 1998; 15: 539-53.

32. Balkau B, Charles MA. Comment on the provisional report from the WHO consultation. European Group for the Study of Insulin Resistance (EGIR). Diabetic medicine : a journal of the British Diabetic Association. 1999; 16: 442-3.

33. Executive Summary of The Third Report of The National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, And Treatment of High Blood Cholesterol In Adults (Adult Treatment Panel III). Jama. 2001; 285: 2486-97.

34. Grundy SM, Cleeman JI, Daniels SR, Donato KA, Eckel RH, Franklin BA, et al. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute Scientific Statement. Circulation. 2005; 112: 2735-52.

35. Alberti KG, Zimmet P, Shaw J. Metabolic syndrome--a new world-wide definition. A Consensus Statement from the International Diabetes Federation. Diabetic medicine: a journal of the British Diabetic Association. 2006; 23: 469-80.

36. Alberti KG, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation. 2009; 120: 1640-5.

37. Oda E. Metabolic syndrome: its history, mechanisms, and limitations. Acta diabetologica. 2012; 49: 89-95.

38. Ju SY, Lee JY, Kim DH. Association of metabolic syndrome and its components with all-cause and cardiovascular mortality in the elderly: A meta-analysis of prospective cohort studies. Medicine. 2017; 96: e8491.

39. Alberti G; Zimmet P; Shaw J GS. The IDF consensus worldwide definition of the Metabolic Syndrome. International Diabetes Federation: 2006.

40. Wilson PW, D'Agostino RB, Parise H, Sullivan L, Meigs JB. Metabolic syndrome as a precursor of cardiovascular disease and type 2 diabetes mellitus. Circulation. 2005; 112: 3066-72.

41. Ford ES. The metabolic syndrome and mortality from cardiovascular disease and all-causes: findings from the National Health and Nutrition Examination Survey II Mortality Study. Atherosclerosis. 2004; 173: 309-14.

42. Blijdorp K, van den Heuvel-Eibrink MM, Pieters R, Boot AM, Delhanty PJ, van der Lely AJ, et al. Obesity is underestimated using body mass index and waist-hip ratio in long-term adult survivors of childhood cancer. PLoS One. 2012; 7: e43269.

43. van Waas M, Neggers SJ, Raat H, van Rij CM, Pieters R, van den Heuvel-Eibrink MM. Abdominal radiotherapy: a major determinant of metabolic syndrome in nephroblastoma and neuroblastoma survivors. PLoS One. 2012; 7: e52237.

44. van Waas M, Neggers SJ, Uitterlinden AG, Blijdorp K, van der Geest IM, Pieters R, et al. Treatment factors rather than genetic variation determine metabolic syndrome in childhood cancer survivors. European journal of cancer (Oxford, England : 1990). 2013; 49: 668-75.

45. Ward LC. Bioelectrical impedance analysis for body composition assessment: reflections on accuracy, clinical utility, and standardisation. European journal of clinical nutrition. 2019; 73: 194-99.

46. Horber FF, Thomi F, Casez JP, Fonteille J, Jaeger P. Impact of hydration status on body composition as measured by dual energy X-ray absorptiometry in normal volunteers and patients on haemodialysis. The British journal of radiology. 1992; 65: 895-900.

47. O'Brien C, Young AJ, Sawka MN. Bioelectrical impedance to estimate changes in hydration status. International journal of sports medicine. 2002; 23: 361-6.

48. van Santen SS, Olsson DS, Hammarstrand C, Wijnen M, van den Heuvel-Eibrink MM, Van der Lely AJ, et al. Diagnosing Metabolic Syndrome in Craniopharyngioma Patients: Body Composition versus BMI. European journal of endocrinology. 2019.

49. Perreault L. Determining body composition in adults. In: Pi-Sunyer F, Sullivan D (eds.). UpToDate: Waltham, MA 2017.

50. Sommer I, Teufer B, Szelag M, Nussbaumer-Streit B, Titscher V, Klerings I, et al. The performance of anthropometric tools to determine obesity: a systematic review and meta-analysis. Scientific reports. 2020; 10: 12699.

51. Coffman E, Richmond-Bryant J. Multiple biomarker models for improved risk estimation of specific cardiovascular diseases related to metabolic syndrome: a cross-sectional study. Population health metrics. 2015; 13: 7.

52. Shea BJ, Reeves BC, Wells G, Thuku M, Hamel C, Moran J, et al. AMSTAR 2: a critical appraisal tool for systematic reviews that include randomised or non-randomised studies of healthcare interventions, or both. BMJ (Clinical research ed). 2017; 358: j4008.

53. Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. Annals of internal medicine. 2011; 155: 529-36.

54. Hayden JA, van der Windt DA, Cartwright JL, Côté P, Bombardier C. Assessing bias in studies of prognostic factors. Annals of internal medicine. 2013; 158: 280-6.

55. Guyatt G, Oxman AD, Akl EA, Kunz R, Vist G, Brozek J, et al. GRADE guidelines: 1. Introduction-GRADE evidence profiles and summary of findings tables. Journal of clinical epidemiology. 2011; 64: 383-94.

56. DerSimonian R, Laird N. Meta-analysis in clinical trials. Controlled clinical trials. 1986; 7: 177-88.

57. R Core Team (2020). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-project.org/.

58. Bielorai B, Weintraub Y, Hutt D, Hemi R, Kanety H, Modan-Moses D, et al. The metabolic syndrome and its components in pediatric survivors of allogeneic hematopoietic stem cell transplantation. Clin Transplant. 2017; 31.

59. Cheung YT, Edelmann MN, Mulrooney DA, Green DM, Chemaitilly W, John N, et al. Uric Acid and Neurocognitive Function in Survivors of Childhood Acute Lymphoblastic Leukemia Treated with Chemotherapy Only. Cancer epidemiology, biomarkers & prevention : a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology. 2016; 25: 1259-67.

60. Kojima C, Kubota M, Nagai A, Adachi S, Watanabe K, Nakahata T. Adipocytokines in childhood cancer survivors and correlation with metabolic syndrome components. Pediatr Int. 2013; 55: 438-42.

61. Ariffin H, Azanan MS, Abd Ghafar SS, Oh L, Lau KH, Thirunavakarasu T, et al. Young adult survivors of childhood acute lymphoblastic leukemia show evidence of chronic inflammation and cellular aging. Cancer. 2017; 123: 4207-14.

62. Barbosa-Cortes L, Lopez-Alarcon M, Mejia-Arangure JM, Klunder-Klunder M, Del Carmen Rodriguez-Zepeda M, Rivera-Marquez H, et al. Adipokines, insulin resistance, and adiposity as a predictors of metabolic syndrome in child survivors of lymphoma and acute lymphoblastic leukemia of a developing country. BMC cancer. 2017; 17: 125.

63. Eglit T, Lember M, Ringmets I, Rajasalu T. Gender differences in serum high-molecular-weight adiponectin levels in metabolic syndrome. European journal of endocrinology. 2013; 168: 385-91.

64. González-Jiménez E, Schmidt-Riovalle J, Sinausía L, Carmen Valenza M, Perona JS. Predictive value of ceruloplasmin for metabolic syndrome in adolescents. BioFactors (Oxford, England). 2016; 42: 163-70.

65. Henneman P, Janssens AC, Zillikens MC, Frolich M, Frants RR, Oostra BA, et al. Menopause impacts the relation of plasma adiponectin levels with the metabolic syndrome. Journal of internal medicine. 2010; 267: 402-9.

66. Krzystek-Korpacka M, Patryn E, Kustrzeba-Wojcicka I, Chrzanowska J, Gamian A, Noczynska A. Gender-specific association of serum uric acid with metabolic syndrome and its components in juvenile obesity. Clinical chemistry and laboratory medicine. 2011; 49: 129-36.

67. Ntzouvani A, Fragopoulou E, Panagiotakos D, Pitsavos C, Antonopoulou S. Reduced circulating adiponectin levels are associated with the metabolic syndrome independently of obesity, lipid indices and serum insulin levels: a cross-sectional study. Lipids in health and disease. 2016; 15: 140.

68. Stefanska A, Ponikowska I, Cwiklinska-Jurkowska M, Sypniewska G. Association of FSH with metabolic syndrome in postmenopausal women: a comparison with CRP, adiponectin and leptin. Biomarkers in medicine. 2014; 8: 921-30.

69. Kim JY, Ahn SV, Yoon JH, Koh SB, Yoon J, Yoo BS, et al. Prospective study of serum adiponectin and incident metabolic syndrome: the ARIRANG study. Diabetes care. 2013; 36: 1547-53.

70. Ahonen TM, Saltevo JT, Kautiainen HJ, Kumpusalo EA, Vanhala MJ. The association of adiponectin and low-grade inflammation with the course of metabolic syndrome. Nutrition, metabolism, and cardiovascular diseases: NMCD. 2012; 22: 285-91.

71. Bombelli M, Quarti-Trevano F, Tadic M, Facchetti R, Cuspidi C, Mancia G, et al. Uric acid and risk of new-onset metabolic syndrome, impaired fasting glucose and diabetes mellitus in a general Italian population: data from the Pressioni Arteriose Monitorate E Loro Associazioni study. J Hypertens. 2018; 36: 1492-98.

72. Cohen E, Krause I, Fraser A, Goldberg E, Garty M. Hyperuricemia and metabolic syndrome: lessons from a large cohort from Israel. Isr Med Assoc J. 2012; 14: 676-80.

73. Delitala AP, Scuteri A, Fiorillo E, Lakatta EG, Schlessinger D, Cucca F. Role of Adipokines in the Association between Thyroid Hormone and Components of the Metabolic Syndrome. Journal of Clinical Medicine. 2019; 8.

74. Fernandez-Berges D, Consuegra-Sanchez L, Penafiel J, Cabrera de Leon A, Vila J, Felix-Redondo FJ, et al. Metabolic and inflammatory profiles of biomarkers in obesity, metabolic syndrome, and diabetes in a Mediterranean population. DARIOS Inflammatory study. Rev Esp Cardiol (Engl Ed). 2014; 67: 624-31.

75. Goncalves JP, Oliveira A, Severo M, Santos AC, Lopes C. Cross-sectional and longitudinal associations between serum uric acid and metabolic syndrome. Endocrine. 2012; 41: 450-7.

76. Juonala M, Saarikoski LA, Viikari JS, Oikonen M, Lehtimaki T, Lyytikainen LP, et al. A longitudinal analysis on associations of adiponectin levels with metabolic syndrome and carotid artery intima-media thickness. The Cardiovascular Risk in Young Finns Study. Atherosclerosis. 2011; 217: 234-9.

77. Kanagasabai T, Alkhalaqi K, Churilla JR, Ardern CI. The Association Between Metabolic Syndrome and Serum Concentrations of Micronutrients, Inflammation, and Oxidative Stress Outside of the Clinical Reference Ranges: A Cross-Sectional Study. Metabolic syndrome and related disorders. 2019; 17: 29-36.

78. Khan RJ, Gebreab SY, Sims M, Riestra P, Xu R, Davis SK. Prevalence, associated factors and heritabilities of metabolic syndrome and its individual components in African Americans: the Jackson Heart Study. BMJ open. 2015; 5: e008675.

79. Koskinen J, Magnussen CG, Wurtz P, Soininen P, Kangas AJ, Viikari JS, et al. Apolipoprotein B, oxidized low-density lipoprotein, and LDL particle size in predicting the incidence of metabolic syndrome: the Cardiovascular Risk in Young Finns study. European journal of preventive cardiology. 2012; 19: 1296-303.

80. Kosola J, Vaara JP, Ahotupa M, Kyrolainen H, Santtila M, Oksala N, et al. Elevated concentration of oxidized LDL together with poor cardiorespiratory and abdominal muscle fitness predicts metabolic syndrome in young men. Metabolism: clinical and experimental. 2013; 62: 992-9.

81. Lindberg S, Jensen JS, Bjerre M, Frystyk J, Flyvbjerg A, Jeppesen J, et al. Low adiponectin levels at baseline and decreasing adiponectin levels over 10 years of follow-up predict risk of the metabolic syndrome. Diabetes & metabolism. 2017; 43: 134-39.

82. Luptáková L, Siváková D, Cvíčelová M, Wsólová L, Danková Z, Michnová A, et al. Power of biomarkers and their relative contributions to metabolic syndrome in Slovak adult women. Annals of human biology. 2013; 40: 132-8.

83. Mani P, Ren HY, Neeland IJ, McGuire DK, Ayers CR, Khera A, et al. The association between HDL particle concentration and incident metabolic syndrome in the multi-ethnic Dallas Heart Study. Diabetes & metabolic syndrome. 2017; 11 Suppl 1: S175-s79.

84. Mazidi M, Toth PP, Banach M. C-reactive Protein Is Associated With Prevalence of the Metabolic Syndrome, Hypertension, and Diabetes Mellitus in US Adults. Angiology. 2018; 69: 438-42.

85. Musani SK, Vasan RS, Bidulescu A, Liu J, Xanthakis V, Sims M, et al. Aldosterone, C-reactive protein, and plasma B-type natriuretic peptide are associated with the development of metabolic syndrome and longitudinal changes in metabolic syndrome components: findings from the Jackson Heart Study. Diabetes care. 2013; 36: 3084-92.

86. Onat A, Komurcu-Bayrak E, Can G, Kucukdurmaz Z, Hergenc G, Erginel-Unaltuna N. Apolipoprotein A-I positively associated with diabetes in women independently of apolipoprotein E genotype and apolipoprotein B levels. Nutrition (Burbank, Los Angeles County, Calif). 2010; 26: 975-80.

87. Onat A, Can G, Ornek E, Ayhan E, Erginel-Ünaltuna N, Murat SN. High serum apolipoprotein E determines hypertriglyceridemic dyslipidemias, coronary disease and apoA-I dysfunctionality. Lipids. 2013; 48: 51-61.

88. Onat A, Can G, Cakr H, Ozpamuk-Karadeniz F, Karadeniz Y, Yuksel H, et al. Sex-Specific Predictors of Metabolic Syndrome Independent of Its Components. Journal of investigative medicine : the official publication of the American Federation for Clinical Research. 2015; 63: 796-801.

89. Onat A, Can G, Çoban N, Dönmez İ, Çakır H, Ademoğlu E, et al. Lipoprotein(a) level and MIF gene variant predict incident metabolic syndrome and mortality. Journal of investigative medicine: the official publication of the American Federation for Clinical Research. 2016; 64: 392-9.

90. Petrikova J, Janicko M, Fedacko J, Drazilova S, Madarasova Geckova A, Marekova M, et al. Serum Uric Acid in Roma and Non-Roma-Its Correlation with Metabolic Syndrome and Other Variables. International journal of environmental research and public health. 2018; 15.

91. Stefanska A, Sypniewska G, Blaszkiewicz B, Ponikowska I, Cwiklinska-Jurkowska M. Comparison between C-reactive protein and adipocyte fatty acid-binding protein as a component of metabolic syndrome in middle-aged women. Clinical biochemistry. 2011; 44: 304-6.

92. Sun D, Li S, Zhang X, Fernandez C, Chen W, Srinivasan SR, et al. Uric acid is associated with metabolic syndrome in children and adults in a community: the Bogalusa Heart Study. PLoS One. 2014; 9: e89696.

93. Cheserek MJ, Shi Y, Le G. Association of hyperuricemia with metabolic syndrome among university workers: sex and occupational differences. Afr Health Sci. 2018; 18: 842-51.

94. Moulin SR, Baldo MP, Souza JB, Luchi WM, Capingana DP, Magalhaes P, et al. Distribution of Serum Uric Acid in Black Africans and Its Association With Cardiovascular Risk Factors. J Clin Hypertens (Greenwich). 2017; 19: 45-50.

95. Wei CY, Sun CC, Wei JC, Tai HC, Sun CA, Chung CF, et al. Association between Hyperuricemia and Metabolic Syndrome: An Epidemiological Study of a Labor Force Population in Taiwan. BioMed research international. 2015; 2015: 369179.

96. Porchia LM, Gonzalez-Mejia ME, Torres-Rasgado E, Ruiz-Vivanco G, Perez-Fuentes R. Low serum uric acid concentration augments insulin effects on the prevalence of metabolic syndrome. Diabetes & metabolic syndrome. 2018; 12: 325-31.

97. Yu KH, Luo SF, Tsai WP, Huang YY. Intermittent elevation of serum urate and 24-hour urinary uric acid excretion. Rheumatology (Oxford, England). 2004; 43: 1541-5.

98. Liu CW, Chen KH, Tseng CK, Chang WC, Wu YW, Hwang JJ. The dose-response effects of uric acid on the prevalence of metabolic syndrome and electrocardiographic left ventricular hypertrophy in healthy individuals. Nutrition, metabolism, and cardiovascular diseases: NMCD. 2019; 29: 30-38.

99. Zhang Y, Fu J, Yang S, Yang M, Liu A, Wang L, et al. Prevalence of metabolically obese but normal weight (MONW) and metabolically healthy but obese (MHO) in Chinese Beijing urban subjects. Bioscience trends. 2017; 11: 418-26.

100. Chen H, Xiong C, Shao X, Ning J, Gao P, Xiao H, et al. Lymphocyte To High-Density Lipoprotein Ratio As A New Indicator Of Inflammation And Metabolic Syndrome. Diabetes, metabolic syndrome and obesity: targets and therapy. 2019; 12: 2117-23.

101. Kawada T. Relationship between several markers and presence of metabolic syndrome or components of the metabolic syndrome in Japanese workers. Journal of occupational and environmental medicine. 2012; 54: 984-8.

102. Li G, Xu L, Zhao Y, Li L, Fu J, Zhang Q, et al. Leptin-adiponectin imbalance as a marker of metabolic syndrome among Chinese children and adolescents: The BCAMS study. PLoS One. 2017; 12: e0186222.

103. Chearskul S, Homsanit M, Chearskul S, Homsanit M, Udol K, Kooptiwut S, et al. Certain hormonal markers in urban Thai adults with metabolic syndrome. Journal of the Medical Association of Thailand = Chotmaihet thangphaet. 2014; 97: 77-84.

104. Yoon JH, Park JK, Oh SS, Lee KH, Kim SK, Cho IJ, et al. The ratio of serum leptin to adiponectin provides adjunctive information to the risk of metabolic syndrome beyond the homeostasis model assessment insulin resistance: the Korean Genomic Rural Cohort Study. Clinica chimica acta; international journal of clinical chemistry. 2011; 412: 2199-205.

105. Yun JE, Won S, Mok Y, Cui W, Kimm H, Jee SH. Association of the leptin to high-molecular-weight adiponectin ratio with metabolic syndrome. Endocrine journal. 2011; 58: 807-15.

106. Adejumo EN, Adejumo OA, Azenabor A, Ekun AO, Enitan SS, Adebola OK, et al. Leptin: Adiponectin ratio discriminated the risk of metabolic syndrome better than adiponectin and leptin in Southwest Nigeria. Diabetes & metabolic syndrome. 2019; 13: 1845-49.

107. Mirza S, Qu HQ, Li Q, Martinez PJ, Rentfro AR, McCormick JB, et al. Adiponectin/leptin ratio and metabolic syndrome in a Mexican American population. Clinical and investigative medicine Medecine clinique et experimentale. 2011; 34: E290.

108. Chou YC, Kuan JC, Bai CH, Yang T, Chou WY, Hsieh PC, et al. Predictive value of serum apolipoprotein B/apolipoprotein A-I ratio in metabolic syndrome risk: a Chinese cohort study. Endocrine. 2015; 49: 404-14.

109. Yang MH, Sung J, Gwak GY. The associations between apolipoprotein B, A1, and the B/A1 ratio and nonalcoholic fatty liver disease in both normal-weight and overweight Korean population. Journal of clinical lipidology. 2016; 10: 289-98.

110. Belfki H, Ben Ali S, Bougatef S, Ben Ahmed D, Haddad N, Jmal A, et al. The Apolipoprotein B/Apolipoprotein A 1 ratio in relation to metabolic syndrome and its components in a sample of the Tunisian population. Experimental and molecular pathology. 2011; 91: 622-5.

111. Jing F, Mao Y, Guo J, Zhang Z, Li Y, Ye Z, et al. The value of Apolipoprotein B/Apolipoprotein A1 ratio for metabolic syndrome diagnosis in a Chinese population: a cross-sectional study. Lipids in health and disease. 2014; 13: 81.

112. Jung CH, Hwang JY, Yu JH, Shin MS, Bae SJ, Park JY, et al. The value of apolipoprotein B/A1 ratio in the diagnosis of metabolic syndrome in a Korean population. Clinical endocrinology. 2012; 77: 699-706.

113. Kim SW, Jee JH, Kim HJ, Jin SM, Suh S, Bae JC, et al. Non-HDL-cholesterol/HDL-cholesterol is a better predictor of metabolic syndrome and insulin resistance than apolipoprotein B/apolipoprotein A1. International journal of cardiology. 2013; 168: 2678-83.

114. Zhong L, Li Q, Jiang Y, Cheng D, Liu Z, Wang B, et al. The ApoB/ApoA1 ratio is associated with metabolic syndrome and its components in a Chinese population. Inflammation. 2010; 33: 353-8.

115. Srikanthan K, Feyh A, Visweshwar H, Shapiro JI, Sodhi K. Systematic Review of Metabolic Syndrome Biomarkers: A Panel for Early Detection, Management, and Risk Stratification in the West Virginian Population. International journal of medical sciences. 2016; 13: 25-38.

116. Goli P, Riahi R, Daniali SS, Pourmirzaei M, Kelishadi R. Association of serum uric acid concentration with components of pediatric metabolic syndrome: A systematic review and meta-analysis. J Res Med Sci. 2020; 25: 43.

117. Yuan H, Yu C, Li X, Sun L, Zhu X, Zhao C, et al. Serum Uric Acid Levels and Risk of Metabolic Syndrome: A Dose-Response Meta-Analysis of Prospective Studies. The Journal of clinical endocrinology and metabolism. 2015; 100: 4198-207.

118. Liu Z, Que S, Zhou L, Zheng S. Dose-response Relationship of Serum Uric Acid with Metabolic Syndrome and Non-alcoholic Fatty Liver Disease Incidence: A Meta-analysis of Prospective Studies. Scientific reports. 2015; 5: 14325.

119. Liu Z, Liang S, Que S, Zhou L, Zheng S, Mardinoglu A. Meta-Analysis of Adiponectin as a Biomarker for the Detection of Metabolic Syndrome. Frontiers in physiology. 2018; 9: 1238.

120. Falahi E, Khalkhali Rad AH, Roosta S. What is the best biomarker for metabolic syndrome diagnosis? Diabetes & metabolic syndrome. 2015; 9: 366-72.

121. Ketterl TG, Chow EJ, Leisenring WM, Goodman P, Koves IH, Petryk A, et al. Adipokines, Inflammation, and Adiposity in Hematopoietic Cell Transplantation Survivors. Biology of blood and marrow transplantation: journal of the American Society for Blood and Marrow Transplantation. 2018; 24: 622-26.

122. Tonorezos ES, Vega GL, Sklar CA, Chou JF, Moskowitz CS, Mo Q, et al. Adipokines, body fatness, and insulin resistance among survivors of childhood leukemia. Pediatric blood & cancer. 2012; 58: 31-6.

123. Argyrou C, Hatziagapiou K, Theodorakidou M, Nikola OA, Vlahopoulos S, Lambrou GI. The role of adiponectin, LEPTIN, and ghrelin in the progress and prognosis of childhood acute lymphoblastic leukemia. Leukemia & lymphoma. 2019; 60: 2158-69.

124. Annaloro C, Airaghi L, Giannarelli D, Mometto G, Orsatti A, Saporiti G, et al. Prospective evaluation of metabolic syndrome and its features in a single-center series of hematopoietic stem cell transplantation recipients. Annals of hematology. 2018; 97: 2471-78.

125. Moschovi M, Trimis G, Vounatsou M, Katsibardi K, Margeli A, Damianos A, et al. Serial plasma concentrations of adiponectin, leptin, and resistin during therapy in children with acute lymphoblastic leukemia. Journal of pediatric hematology/oncology. 2010; 32: e8-13.

126. Cepelova M, Kruseova J, Luks A, Capek V, Cepela P, Potockova J, et al. Accelerated atherosclerosis, hyperlipoproteinemia and insulin resistance in long-term survivors of Hodgkin lymphoma during childhood and adolescence. Neoplasma. 2019; 66: 978-87.

127. Chow EJ, Simmons JH, Roth CL, Baker KS, Hoffmeister PA, Sanders JE, et al. Increased cardiometabolic traits in pediatric survivors of acute lymphoblastic leukemia treated with total body irradiation. Biology of blood and marrow transplantation: journal of the American Society for Blood and Marrow Transplantation. 2010; 16: 1674-81.

128. Cooksey R, Wu SY, Klesse L, Oden JD, Bland RE, Hodges JC, et al. Metabolic syndrome is a sequela of radiation exposure in hypothalamic obesity among survivors of childhood brain tumors. Journal of investigative medicine: the official publication of the American Federation for Clinical Research. 2019; 67: 295-302.

129. Frisk P, Arvidson J, Larsson M, Naessén T. Risk factors for cardiovascular disease are increased in young adults treated with stem cell transplantation during childhood. Pediatric transplantation. 2012; 16: 385-91.

130. Grote S, Almstedt HC, Tarleton HP. Cardiometabolic Health Among Cancer Survivors: A 13-Week Pilot Study of a Combined Aerobic and Resistance Training Program. Oncology nursing forum. 2016; 43: 306-15.

131. Wei C, Crowne E. The impact of childhood cancer and its treatment on puberty and subsequent hypothalamic pituitary and gonadal function, in both boys and girls. Best practice & research Clinical endocrinology & metabolism. 2019; 33: 101291.

132. Pluimakers VG, van Waas M, Looman CWN, de Maat MP, de Jonge R, Delhanty P, et al. Metabolic syndrome detection with biomarkers in childhood cancer survivors. Endocrine connections. 2020; 9: 676-86.

133. Marouli E, Graff M, Medina-Gomez C, Lo KS, Wood AR, Kjaer TR, et al. Rare and low-frequency coding variants alter human adult height. Nature. 2017; 542: 186-90.

134. Chemaitilly W, Cohen LE. Diagnosis of endocrine disease: Endocrine late-effects of childhood cancer and its treatments. European journal of endocrinology. 2017; 176: R183-r203.

135. de Vathaire F, El-Fayech C, Ben Ayed FF, Haddy N, Guibout C, Winter D, et al. Radiation dose to the pancreas and risk of diabetes mellitus in childhood cancer survivors: a retrospective cohort study. The Lancet Oncology. 2012; 13: 1002-10.

136. Darzy KH, Shalet SM. Hypopituitarism following Radiotherapy Revisited. Endocrine development. 2009; 15: 1-24.

137. Oudin C, Simeoni MC, Sirvent N, Contet A, Begu-Le Coroller A, Bordigoni P, et al. Prevalence and risk factors of the metabolic syndrome in adult survivors of childhood leukemia. Blood. 2011; 117: 4442-8.

138. Rose SR, Horne VE, Howell J, Lawson SA, Rutter MM, Trotman GE, et al. Late endocrine effects of childhood cancer. Nature reviews Endocrinology. 2016; 12: 319-36.

139. de Haas EC, Oosting SF, Lefrandt JD, Wolffenbuttel BH, Sleijfer DT, Gietema JA. The metabolic syndrome in cancer survivors. The Lancet Oncology. 2010; 11: 193-203.

140. de Abreu VG, Martins CJM, de Oliveira PAC, Francischetti EA. High-molecular weight adiponectin/HOMA-IR ratio as a biomarker of metabolic syndrome in urban multiethnic Brazilian subjects. PLoS One. 2017; 12: e0180947.

141. Baudrand R, Campino C, Carvajal CA, Olivieri O, Guidi G, Faccini G, et al. High sodium intake is associated with increased glucocorticoid production, insulin resistance and metabolic syndrome. Clinical endocrinology. 2014; 80: 677-84.

142. de Oliveira A, Hermsdorff HH, Cocate PG, Santos EC, Bressan J, Natali AJ. Accuracy of plasma interleukin-18 and adiponectin concentrations in predicting metabolic syndrome and cardiometabolic disease risk in middle-age Brazilian men. Applied physiology, nutrition, and metabolism = Physiologie appliquee, nutrition et metabolisme. 2015; 40: 1048-55.

143. Ding Y, Li S, Ma RL, Guo H, Zhang J, Zhang M, et al. Association of homeostasis model assessment of insulin resistance, adiponectin, and low-grade inflammation with the course of the metabolic syndrome. Clinical biochemistry. 2015; 48: 503-7.

144. Hirose H, Takayama T, Hozawa S, Hibi T, Saito I. Prediction of metabolic syndrome using artificial neural network system based on clinical data including insulin resistance index and serum adiponectin. Computers in biology and medicine. 2011; 41: 1051-6.

145. Ju H, Zhou Z, Sun M, Chen H. Association of fetuin-A to adiponectin ratio with metabolic syndrome: a cross-sectional study. Endocrine. 2017; 58: 190-93.

146. Kawada T, Hasegawa M. Predictive ability of serum high-molecular-weight adiponectin in combination with serum insulin and serum C-reactive protein for the presence of metabolic syndrome. Annals of human biology. 2012; 39: 108-12.

147. Abu-Farha M, Behbehani K, Elkum N. Comprehensive analysis of circulating adipokines and hsCRP association with cardiovascular disease risk factors and metabolic syndrome in Arabs. Cardiovascular diabetology. 2014; 13: 76.

148. Cheng KH, Huang SP, Huang CN, Lee YC, Chu CS, Chang CF, et al. The impact of estradiol and 1,25(OH)2D3 on metabolic syndrome in middle-aged Taiwanese males. PLoS One. 2013; 8: e60295.

149. Esteghamati A, Khalilzadeh O, Anvari M, Rashidi A, Mokhtari M, Nakhjavani M. Association of serum leptin levels with homeostasis model assessment-estimated insulin resistance and metabolic syndrome: the key role of central obesity. Metabolic syndrome and related disorders. 2009; 7: 447-52.

150. Jeong J, Suh YJ. Association between Serum Uric Acid and Metabolic Syndrome in Koreans. Journal of Korean medical science. 2019; 34: e307.

151. Lipshultz SE, Landy DC, Lopez-Mitnik G, Lipsitz SR, Hinkle AS, Constine LS, et al. Cardiovascular status of childhood cancer survivors exposed and unexposed to cardiotoxic therapy. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2012; 30: 1050-7.

152. Meigs JB, Rutter MK, Sullivan LM, Fox CS, D'Agostino RB, Sr., Wilson PW. Impact of insulin resistance on risk of type 2 diabetes and cardiovascular disease in people with metabolic syndrome. Diabetes care. 2007; 30: 1219-25.

153. Mottillo S, Filion KB, Genest J, Joseph L, Pilote L, Poirier P, et al. The metabolic syndrome and cardiovascular risk a systematic review and meta-analysis. Journal of the American College of Cardiology. 2010; 56: 1113-32.

154. Esteghamati A, Zandieh A, Zandieh B, Khalilzadeh O, Meysamie A, Nakhjavani M, et al. Leptin cut-off values for determination of metabolic syndrome: third national surveillance of risk factors of non-communicable diseases in Iran (SuRFNCD-2007). Endocrine. 2011; 40: 117-23.

155. Bermúdez V, Rojas J, Salazar J, Calvo MJ, Morillo J, Torres W, et al. The Maracaibo city metabolic syndrome prevalence study: Primary results and agreement level of 3 diagnostic criteria. Revista Latinoamericana de Hipertension. 2014; 9: 20-32.

156. Lamarche B, Moorjani S, Lupien PJ, Cantin B, Bernard PM, Dagenais GR, et al. Apolipoprotein A-I and B levels and the risk of ischemic heart disease during a five-year follow-up of men in the Québec cardiovascular study. Circulation. 1996; 94: 273-8.

157. Wilson DP, Jacobson TA, Jones PH, Koschinsky ML, McNeal CJ, Nordestgaard BG, et al. Use of Lipoprotein(a) in clinical practice: A biomarker whose time has come. A scientific statement from the National Lipid Association. Journal of clinical lipidology. 2019; 13: 374-92.

158. Barter PJ, Ballantyne CM, Carmena R, Castro Cabezas M, Chapman MJ, Couture P, et al. Apo B versus cholesterol in estimating cardiovascular risk and in guiding therapy: report of the thirty-person/ten-country panel. Journal of internal medicine. 2006; 259: 247-58.

159. Huffman FG, Knight-Sepulveda K, McLean M, Vaccaro JA, Zarini GG. Serum Adiponectin and Ghrelin, Metabolic Syndrome and Diabetes Status in Cuban Americans Int J Health Res. 2010; 3: 93-103.

160. Norvik JV, Storhaug HM, Ytrehus K, Jenssen TG, Zykova SN, Eriksen BO, et al. Overweight modifies the longitudinal association between uric acid and some components of the metabolic syndrome: The Tromsø Study. BMC cardiovascular disorders. 2016; 16: 85.

161. Dallmeier D, Larson MG, Vasan RS, Keaney JF, Jr., Fontes JD, Meigs JB, et al. Metabolic syndrome and inflammatory biomarkers: a community-based cross-sectional study at the Framingham Heart Study. Diabetology & metabolic syndrome. 2012; 4: 28.

162. Côté M, Cartier A, Reuwer AQ, Arsenault BJ, Lemieux I, Després JP, et al. Adiponectin and risk of coronary heart disease in apparently healthy men and women (from the EPIC-Norfolk Prospective Population Study). The American journal of cardiology. 2011; 108: 367-73.

163. Goldberg JF, Ness KK, Chi X, Santucci AK, Plana JC, Joshi VM, et al. Cardiovascular Family History Increases Risk for Late-Onset Adverse Cardiovascular Outcomes in Childhood Cancer Survivors: A St. Jude Lifetime Cohort Report. Cancer epidemiology, biomarkers & prevention: a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology. 2020.

164. Povel CM, Boer JM, Reiling E, Feskens EJ. Genetic variants and the metabolic syndrome: a systematic review. Obesity reviews: an official journal of the International Association for the Study of Obesity. 2011; 12: 952-67.

165. Monda KL, North KE, Hunt SC, Rao DC, Province MA, Kraja AT. The genetics of obesity and the metabolic syndrome. Endocrine, metabolic & immune disorders drug targets. 2010; 10: 86-108.

166. Fenwick PH, Jeejeebhoy K, Dhaliwal R, Royall D, Brauer P, Tremblay A, et al. Lifestyle genomics and the metabolic syndrome: A review of genetic variants that influence response to diet and exercise interventions. Critical reviews in food science and nutrition. 2019; 59: 2028-39.

167. Al-Meshaweh AF, Jafar Y, Asem M, Akanji AO. Determinants of Blood Uric Acid Levels in a Dyslipidemic Arab Population. Medical Principles and Practice. 2012; 21: 209-16.

168. Tsouli SG, Liberopoulos EN, Mikhailidis DP, Athyros VG, Elisaf MS. Elevated serum uric acid levels in metabolic syndrome: an active component or an innocent bystander? Metabolism: clinical and experimental. 2006; 55: 1293-301.

169. Zhang F, Basinski MB, Beals JM, Briggs SL, Churgay LM, Clawson DK, et al. Crystal structure of the obese protein leptin-E100. Nature. 1997; 387: 206-9.

170. Funcke JB, Scherer PE. Beyond adiponectin and leptin: adipose tissue-derived mediators of inter-organ communication. Journal of lipid research. 2019; 60: 1648-84.

171. Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. The Journal of biological chemistry. 1995; 270: 26746-9.

172. van Andel M, Heijboer AC, Drent ML. Adiponectin and Its Isoforms in Pathophysiology. Advances in clinical chemistry. 2018; 85: 115-47.

173. Ronsley R, Rassekh SR, Fleming A, Empringham B, Jennings W, Portwine C, et al. High molecular weight adiponectin levels are inversely associated with adiposity in pediatric brain tumor survivors. Scientific reports. 2020; 10: 18606.

174. Young SG. Recent progress in understanding apolipoprotein B. Circulation. 1990; 82: 1574-94.

175. Kastelein JJ, van der Steeg WA, Holme I, Gaffney M, Cater NB, Barter P, et al. Lipids, apolipoproteins, and their ratios in relation to cardiovascular events with statin treatment. Circulation. 2008; 117: 3002-9.

176. Sniderman AD, Williams K, Contois JH, Monroe HM, McQueen MJ, de Graaf J, et al. A meta-analysis of low-density lipoprotein cholesterol, non-high-density lipoprotein cholesterol, and apolipoprotein B as markers of cardiovascular risk. Circulation Cardiovascular quality and outcomes. 2011; 4: 337-45.

177. Devaraj S, Semaan JR, Jialal I. Biochemistry, Apolipoprotein B. StatPearls. StatPearls Publishing Copyright © 2020, StatPearls Publishing LLC.: Treasure Island (FL) 2020.

178. Clouet-Foraison N, Marcovina SM, Guerra E, Aarsand AK, Coşkun A, Díaz-Garzón J, et al. Analytical Performance Specifications for Lipoprotein(a), Apolipoprotein B-100, and Apolipoprotein A-I Using the Biological Variation Model in the EuBIVAS Population. Clinical chemistry. 2020; 66: 727-36.

179. Danesh J, Collins R, Peto R. Lipoprotein(a) and coronary heart disease. Meta-analysis of prospective studies. Circulation. 2000; 102: 1082-5.

180. Kar S. Elevated Lipoprotein A in South Asians and the Associated Risk of Cardiovascular Disease: A Systematic Review. Current problems in cardiology. 2020: 100581.

181. Craig WY, Neveux LM, Palomaki GE, Cleveland MM, Haddow JE. Lipoprotein(a) as a risk factor for ischemic heart disease: metaanalysis of prospective studies. Clinical chemistry. 1998; 44: 2301-6.

182. Forbes CA, Quek RG, Deshpande S, Worthy G, Wolff R, Stirk L, et al. The relationship between Lp(a) and CVD outcomes: a systematic review. Lipids in health and disease. 2016; 15: 95.

183. Smolders B, Lemmens R, Thijs V. Lipoprotein (a) and stroke: a meta-analysis of observational studies. Stroke. 2007; 38: 1959-66.

184. Nave AH, Lange KS, Leonards CO, Siegerink B, Doehner W, Landmesser U, et al. Lipoprotein (a) as a risk factor for ischemic stroke: a meta-analysis. Atherosclerosis. 2015; 242: 496-503.

185. Wang Z, Zhai X, Xue M, Cheng W, Hu H. Prognostic value of lipoprotein (a) level in patients with coronary artery disease: a meta-analysis. Lipids in health and disease. 2019; 18: 150.

186. Clarke R, Peden JF, Hopewell JC, Kyriakou T, Goel A, Heath SC, et al. Genetic variants associated with Lp(a) lipoprotein level and coronary disease. The New England journal of medicine. 2009; 361: 2518-28.

187. van Iperen EP, Sivapalaratnam S, Holmes MV, Hovingh GK, Zwinderman AH, Asselbergs FW. Genetic analysis of emerging risk factors in coronary artery disease. Atherosclerosis. 2016; 254: 35-41.

188. Paige E, Masconi KL, Tsimikas S, Kronenberg F, Santer P, Weger S, et al. Lipoprotein(a) and incident type-2 diabetes: results from the prospective Bruneck study and a meta-analysis of published literature. Cardiovascular diabetology. 2017; 16: 38.

189. Kaptoge S, Di Angelantonio E, Lowe G, Pepys MB, Thompson SG, Collins R, et al. C-reactive protein concentration and risk of coronary heart disease, stroke, and mortality: an individual participant meta-analysis. Lancet (London, England). 2010; 375: 132-40.

190. Pepys MB, Hirschfield GM. C-reactive protein: a critical update. The Journal of clinical investigation. 2003; 111: 1805-12.

191. Pepys MB, Rowe IF, Baltz ML. C-reactive protein: binding to lipids and lipoproteins. International review of experimental pathology. 1985; 27: 83-111.

192. Zhang YX, Cliff WJ, Schoefl GI, Higgins G. Coronary C-reactive protein distribution: its relation to development of atherosclerosis. Atherosclerosis. 1999; 145: 375-9.

193. Yoon K, Ryu S, Lee J, Park JD. Higher and increased concentration of hs-CRP within normal range can predict the incidence of metabolic syndrome in healthy men. Diabetes & metabolic syndrome. 2018; 12: 977-83.

194. Oda E, Kawai R. Reproducibility of high-sensitivity C-reactive protein as an inflammatory component of metabolic syndrome in Japanese. Circulation journal: official journal of the Japanese Circulation Society. 2010; 74: 1488-93.

195. Hillman AJ, Lohsoonthorn V, Hanvivatvong O, Jiamjarasrangsi W, Lertmaharit S, Williams MA. Association of High Sensitivity C-Reactive Protein Concentrations and Metabolic Syndrome among Thai Adults. Asian Biomed (Res Rev News). 2010; 4: 385-93.

196. Kim J, Pyo S, Yoon DW, Lee S, Lim JY, Heo JS, et al. The co-existence of elevated high sensitivity C-reactive protein and homocysteine levels is associated with increased risk of metabolic syndrome: A 6-year follow-up study. PLoS One. 2018; 13: e0206157.

197. Abu-Farha M, Abubaker J, Al-Khairi I, Cherian P, Noronha F, Kavalakatt S, et al. Circulating angiopoietin-like protein 8 (betatrophin) association with HsCRP and metabolic syndrome. Cardiovascular diabetology. 2016; 15: 25.

198. Chen T, Chen H, Xiao H, Tang H, Xiang Z, Wang X, et al. Comparison of the Value of Neutrophil to High-Density Lipoprotein Cholesterol Ratio and Lymphocyte to High-Density Lipoprotein Cholesterol Ratio for Predicting Metabolic Syndrome Among a Population in the Southern Coast of China. Diabetes, metabolic syndrome and obesity: targets and therapy. 2020; 13: 597-605.

199. Mirhafez SR, Pasdar A, Avan A, Esmaily H, Moezzi A, Mohebati M, et al. Cytokine and growth factor profiling in patients with the metabolic syndrome. The British journal of nutrition. 2015; 113: 1911-9.

200. Orsatti CL, Petri Nahas EA, Nahas-Neto J, Orsatti FL, Giorgi VI, Witkin SS. Evaluation of Toll-Like receptor 2 and 4 RNA expression and the cytokine profile in postmenopausal women with metabolic syndrome. PLoS One. 2014; 9: e109259.

201. Fearon WF, Fearon DT. Inflammation and cardiovascular disease: role of the interleukin-1 receptor antagonist. Circulation. 2008; 117: 2577-9.

202. Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. Annual review of immunology. 2009; 27: 519-50.

203. Ridker PM, Howard CP, Walter V, Everett B, Libby P, Hensen J, et al. Effects of interleukin-1 inhibition with canakinumab on hemoglobin A1c, lipids, C-reactive protein, interleukin-6, and fibrinogen: a phase IIb randomized, placebo-controlled trial. Circulation. 2012; 126: 2739-48.

204. Popovic M, Ebrahimi F, Urwyler SA, Donath MY, Christ-Crain M. The role of IL-1 in the regulation of copeptin in patients with metabolic syndrome. Endocrine connections. 2020; 9: 715-23.

205. D'Elia L, Giaquinto A, Cappuccio FP, Iacone R, Russo O, Strazzullo P, et al. Circulating leptin is associated with serum uric acid level and its tubular reabsorption in a sample of adult middle-aged men. Journal of endocrinological investigation. 2020; 43: 587-93.

206. Tabara Y, Osawa H, Kawamoto R, Tachibana-Iimori R, Yamamoto M, Nakura J, et al. Reduced high-molecular-weight adiponectin and elevated high-sensitivity C-reactive protein are synergistic risk factors for metabolic syndrome in a large-scale middle-aged to elderly population: the Shimanami Health Promoting Program Study. The Journal of clinical endocrinology and metabolism. 2008; 93: 715-22.

207. Oda E. Serum uric acid is an independent predictor of metabolic syndrome in a Japanese health screening population. Heart Vessels. 2014; 29: 496-503.

208. Oda E. High-sensitivity C-reactive protein and white blood cell count equally predict development of the metabolic syndrome in a Japanese health screening population. Acta diabetologica. 2013; 50: 633-8.

209. Dai X, Yuan J, Yao P, Yang B, Gui L, Zhang X, et al. Association between serum uric acid and the metabolic syndrome among a middle- and old-age Chinese population. European journal of epidemiology. 2013; 28: 669-76.

210. Wang L, Zhang T, Liu Y, Tang F, Xue F. Association of Serum Uric Acid with Metabolic Syndrome and Its Components: A Mendelian Randomization Analysis. BioMed research international. 2020; 2020: 6238693.

211. White J, Sofat R, Hemani G, Shah T, Engmann J, Dale C, et al. Plasma urate concentration and risk of coronary heart disease: a Mendelian randomisation analysis. The lancet Diabetes & endocrinology. 2016; 4: 327-36.

212. Mente A, Meyre D, Lanktree MB, Heydarpour M, Davis AD, Miller R, et al. Causal relationship between adiponectin and metabolic traits: a Mendelian randomization study in a multiethnic population. PLoS One. 2013; 8: e66808.

213. Kodama K, Tojjar D, Yamada S, Toda K, Patel CJ, Butte AJ. Ethnic Differences in the Relationship Between Insulin Sensitivity and Insulin Response. A systematic review and meta-analysis. 2013; 36: 1789-96.

214. WHO. Appropriate body-mass index for Asian populations and its implications for policy and intervention strategies. Lancet (London, England). 2004; 363: 157-63.

215. DeBoer MD, Dong L, Gurka MJ. Racial/ethnic and sex differences in the relationship between uric acid and metabolic syndrome in adolescents: an analysis of National Health and Nutrition Survey 1999-2006. Metabolism: clinical and experimental. 2012; 61: 554-61.

216. Tsimikas S, Clopton P, Brilakis ES, Marcovina SM, Khera A, Miller ER, et al. Relationship of oxidized phospholipids on apolipoprotein B-100 particles to race/ethnicity, apolipoprotein(a) isoform size, and cardiovascular risk factors: results from the Dallas Heart Study. Circulation. 2009; 119: 1711-9.

217. Chen W, Srinivasan SR, Bao W, Wattigney WA, Berenson GS. Sibling aggregation of low- and high-density lipoprotein cholesterol and apolipoproteins B and A-I levels in black and white children: the Bogalusa Heart Study. Ethnicity & disease. 1997; 7: 241-9.

218. Iso H, Harada S, Shimamoto T, Folsom AR, Koike K, Sato S, et al. Polymorphism of the apolipoprotein B gene and blood lipid concentrations in Japanese and Caucasian population samples. Atherosclerosis. 1996; 126: 233-41.

219. Vermaak WJ, Ubbink JB, Delport R, Becker PJ, Bissbort SH, Ungerer JP. Ethnic immunity to coronary heart disease? Atherosclerosis. 1991; 89: 155-62.

220. Hackler E, 3rd, Lew J, Gore MO, Ayers CR, Atzler D, Khera A, et al. Racial Differences in Cardiovascular Biomarkers in the General Population. Journal of the American Heart Association. 2019; 8: e012729.

221. Osei K, Gaillard T. Disparities in Cardiovascular Disease and Type 2 Diabetes Risk Factors in Blacks and Whites: Dissecting Racial Paradox of Metabolic Syndrome. Frontiers in endocrinology. 2017; 8: 204.

222. Jacobs S, Kroeger J, Schulze MB, Frank LK, Franke AA, Cheng I, et al. Dietary Patterns Derived by Reduced Rank Regression Are Inversely Associated with Type 2 Diabetes Risk across 5 Ethnic Groups in the Multiethnic Cohort. Current developments in nutrition. 2017; 1: e000620.

223. Kim JY, Yadav D, Ahn SV, Koh SB. A prospective study of serum adiponectin and regression of metabolic syndrome: The ARIRANG study. Biochemical and biophysical research communications. 2015; 466: 201-5.

224. Kramer CK, von Mühlen D, Jassal SK, Barrett-Connor E. Serum uric acid levels improve prediction of incident type 2 diabetes in individuals with impaired fasting glucose: the Rancho Bernardo Study. Diabetes care. 2009; 32: 1272-3.

225. Feig DI, Kang DH, Johnson RJ. Uric acid and cardiovascular risk. The New England journal of medicine. 2008; 359: 1811-21.

226. Chuang SY, Chen JH, Yeh WT, Wu CC, Pan WH. Hyperuricemia and increased risk of ischemic heart disease in a large Chinese cohort. International journal of cardiology. 2012; 154: 316-21.

227. Sattar N, Wannamethee G, Sarwar N, Tchernova J, Cherry L, Wallace AM, et al. Adiponectin and coronary heart disease: a prospective study and meta-analysis. Circulation. 2006; 114: 623-9.

228. Li S, Shin HJ, Ding EL, van Dam RM. Adiponectin levels and risk of type 2 diabetes: a systematic review and meta-analysis.The Journal of the American Medical Association. 2009; 302: 179-88.

229. Meier U, Gressner AM. Endocrine regulation of energy metabolism: review of pathobiochemical and clinical chemical aspects of leptin, ghrelin, adiponectin, and resistin. Clinical chemistry. 2004; 50: 1511-25.

230. Sabatine MS, Morrow DA, Jablonski KA, Rice MM, Warnica JW, Domanski MJ, et al. Prognostic significance of the Centers for Disease Control/American Heart Association high-sensitivity C-reactive protein cut points for cardiovascular and other outcomes in patients with stable coronary artery disease. Circulation. 2007; 115: 1528-36.

231. Povel CM, Beulens JW, van der Schouw YT, Dolle ME, Spijkerman AM, Verschuren WM, et al. Metabolic syndrome model definitions predicting type 2 diabetes and cardiovascular disease. Diabetes care. 2013; 36: 362-8.

232. Yusuf S, Hawken S, Ounpuu S, Dans T, Avezum A, Lanas F, et al. Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case-control study. Lancet (London, England). 2004; 364: 937-52.

233. Gianfagna F, Veronesi G, Guasti L, Chambless LE, Brambilla P, Corrao G, et al. Do apolipoproteins improve coronary risk prediction in subjects with metabolic syndrome? Insights from the North Italian Brianza cohort study. Atherosclerosis. 2014; 236: 175-81.

234. Swerdlow DI, Holmes MV, Kuchenbaecker KB, Engmann JE, Shah T, Sofat R, et al. The interleukin-6 receptor as a target for prevention of coronary heart disease: a mendelian randomisation analysis. Lancet (London, England). 2012; 379: 1214-24.

235. Zinman B, Hanley AJ, Harris SB, Kwan J, Fantus IG. Circulating tumor necrosis factor-alpha concentrations in a native Canadian population with high rates of type 2 diabetes mellitus. The Journal of clinical endocrinology and metabolism. 1999; 84: 272-8.

236. Li WC, Hsiao KY, Chen IC, Chang YC, Wang SH, Wu KH. Serum leptin is associated with cardiometabolic risk and predicts metabolic syndrome in Taiwanese adults. Cardiovascular diabetology. 2011; 10: 36.

237. Obeidat AA, Ahmad MN, Haddad FH, Azzeh FS. Leptin and uric acid as predictors of metabolic syndrome in jordanian adults. Nutr Res Pract. 2016; 10: 411-7.

238. Chiou WK, Wang MH, Huang DH, Chiu HT, Lee YJ, Lin JD. The relationship between serum uric acid level and metabolic syndrome: differences by sex and age in Taiwanese. J Epidemiol. 2010; 20: 219-24.

239. Chen YY, Kao TW, Yang HF, Chou CW, Wu CJ, Lai CH, et al. The association of uric acid with the risk of metabolic syndrome, arterial hypertension or diabetes in young subjects- An observational study. Clinica chimica acta; international journal of clinical chemistry. 2018; 478: 68-73.

240. Lee JG, Park SW, Cho BM, Lee S, Kim YJ, Jeong DW, et al. Serum amylase and risk of the metabolic syndrome in Korean adults. Clinica chimica acta; international journal of clinical chemistry. 2011; 412: 1848-53.

241. Lee YJ, Cho S, Kim SR. A possible role of serum uric acid as a marker of metabolic syndrome. Intern Med J. 2014; 44: 1210-6.

242. Sun HL, Pei D, Lue KH, Chen YL. Uric Acid Levels Can Predict Metabolic Syndrome and Hypertension in Adolescents: A 10-Year Longitudinal Study. PLoS One. 2015; 10: e0143786.

243. Fujikawa R, Ito C, Tsuboi A. Is the screening of metabolic syndrome using adiponectin possible? Diabetol Int. 2015; 6: 313-20.

244. Oh J, Kim JY, Park S, Youn JC, Son NH, Shin DJ, et al. The relationship between insulin-like growth factor-1 and metabolic syndrome, independent of adiponectin. Clinica chimica acta; international journal of clinical chemistry. 2012; 413: 506-10.

245. Saisho Y, Hirose H, Roberts R, Abe T, Kawabe H, Itoh H. C-reactive protein, high-molecular-weight adiponectin and development of metabolic syndrome in the Japanese general population: a longitudinal cohort study. PLoS One. 2013; 8: e73430.

246. Jeong H, Baek SY, Kim SW, Park EJ, Lee J, Kim H, et al. C reactive protein level as a marker for dyslipidaemia, diabetes and metabolic syndrome: results from the Korea National Health and Nutrition Examination Survey. BMJ open. 2019; 9: e029861.

247. Oda E, Kawai R. Comparison between high-sensitivity C-reactive protein (hs-CRP) and white blood cell count (WBC) as an inflammatory component of metabolic syndrome in Japanese. Internal medicine (Tokyo, Japan). 2010; 49: 117-24.

248. Abbasian M, Ebrahimi H, Delvarianzadeh M, Norouzi P, Fazli M. Association between serum uric acid (SUA) levels and metabolic syndrome (MetS) components in personnel of Shahroud University of Medical Sciences. Diabetes & metabolic syndrome. 2016; 10: 132-6.

249. Ahmadnezhad M, Arefhosseini SR, Parizadeh MR, Tavallaie S, Tayefi M, Darroudi S, et al. Association between serum uric acid, high sensitive C-reactive protein and pro-oxidant-antioxidant balance in patients with metabolic syndrome. BioFactors (Oxford, England). 2018; 44: 263-71.

250. Ali N, Miah R, Hasan M, Barman Z, Mou AD, Hafsa JM, et al. Association between serum uric acid and metabolic syndrome: a cross-sectional study in Bangladeshi adults. Scientific reports. 2020; 10: 7841.

251. Baygi F, Herttua K, Sheidaei A, Ahmadvand A, Jensen OC. Association of Serum Uric Acid with cardio-metabolic risk factors and metabolic syndrome in seafarers working on tankers. BMC public health. 2020; 20: 442.

252. Chen D, Zhang H, Gao Y, Lu Z, Yao Z, Jiang Y, et al. Cross-sectional and longitudinal associations between serum uric acid and metabolic syndrome: Results from Fangchenggang Area Male Health and Examination Survey in China. Clinica chimica acta; international journal of clinical chemistry. 2015; 446: 226-30.

253. Cho MH, Kim YM, Yoon JH, Kim DH, Lim JS. Serum uric acid in Korean children and adolescents: reference percentiles and association with metabolic syndrome. Annals of pediatric endocrinology & metabolism. 2020; 25: 104-11.

254. de Oliveira A, Hermsdorff HHM, Guedes Cocate P, Bressan J, Azevedo Novello A, Cardoso dos Santos E, et al. The impact of serum uric acid on the diagnostic of metabolic syndrome in apparently healthy brazilian middle-aged men. Nutricion hospitalaria. 2014; 30: 562-9.

255. Ferrara LA, Wang H, Umans JG, Franceschini N, Jolly S, Lee ET, et al. Serum uric acid does not predict incident metabolic syndrome in a population with high prevalence of obesity. Nutrition, metabolism, and cardiovascular diseases. 2014; 24: 1360-4.

256. Fu YQ, Yang H, Zheng JS, Zeng XY, Zeng W, Fan ZF, et al. Positive association between metabolic syndrome and serum uric acid in Wuhan. Asia Pacific journal of clinical nutrition. 2017; 26: 343-50.

257. Hara S, Tsuji H, Ohmoto Y, Amakawa K, Hsieh SD, Arase Y, et al. High serum uric acid level and low urine pH as predictors of metabolic syndrome: a retrospective cohort study in a Japanese urban population. Metabolism: clinical and experimental. 2012; 61: 281-8.

258. He SJ, Chan C, Xie ZD, Shi D, Hu XB, Li HY, et al. The relationship between serum uric acid and metabolic syndrome in premenopausal and postmenopausal women in the Jinchang Cohort. Gynecological endocrinology: the official journal of the International Society of Gynecological Endocrinology. 2017; 33: 141-44.

259. Huang S, Liu X, Li H, Xu W, Jia H. Sex difference in the association of serum uric acid with metabolic syndrome and its components: a cross-sectional study in a Chinese Yi population. Postgrad Med. 2017; 129: 828-33.

260. Huang LL, Dou DM, Liu N, Wang XX, Fu LY, Wu X, et al. Association of erythrocyte parameters with metabolic syndrome in the Pearl River Delta region of China: a cross sectional study. BMJ open. 2018; 8: e019792.

261. Jaipakdee J, Jiamjarasrangsri W, Lohsoonthorn V, Lertmaharit S. Prevalence of metabolic syndrome and its association with serum uric acid levels in Bangkok Thailand. The Southeast Asian journal of tropical medicine and public health. 2013; 44: 512-22.

262. Kawada T, Otsuka T, Inagaki H, Wakayama Y, Li Q, Li YJ, et al. Association of smoking status, insulin resistance, body mass index, and metabolic syndrome in workers: A 1-year follow-up study. Obesity research & clinical practice. 2010; 4: e163-246.

263. Kawada T, Otsuka T. Factor structure of indices of the second derivative of the finger photoplethysmogram with metabolic components and other cardiovascular risk indicators. Diabetes & metabolism journal. 2013; 37: 40-5.

264. Kawada T. Insulin-related biomarkers to predict the risk of metabolic syndrome. International journal of endocrinology and metabolism. 2013; 11: e10418.

265. Kawada T, Otsuka T, Inagaki H, Wakayama Y, Katsumata M. Biological markers, lifestyles and metabolic syndrome in workers. Diabetes & metabolic syndrome. 2015; 9: 71-3.

266. Lan Y, Mai Z, Zhou S, Liu Y, Li S, Zhao Z, et al. Prevalence of metabolic syndrome in China: An up-dated cross-sectional study. PLoS One. 2018; 13: e0196012.

267. Lee HJ, Park HT, Cho GJ, Yi KW, Ahn KH, Shin JH, et al. Relationship between uric acid and metabolic syndrome according to menopausal status. Gynecological endocrinology: the official journal of the International Society of Gynecological Endocrinology. 2011; 27: 406-11.

268. Lee JM, Kim HC, Cho HM, Oh SM, Choi DP, Suh I. Association between serum uric acid level and metabolic syndrome. J Prev Med Public Health. 2012; 45: 181-7.

269. Lee JK, Ryoo JH, Choi JM, Park SK. Serum uric acid level and the incidence of metabolic syndrome in middle-aged Korean men: a 5-year follow-up study. J Prev Med Public Health. 2014; 47: 317-26.

270. Lee YB, Jun JE, Lee SE, Ahn J, Kim G, Jee JH, et al. Utility of Serum Albumin for Predicting Incident Metabolic Syndrome according to Hyperuricemia. Diabetes & metabolism journal. 2018; 42: 529-37.

271. Li Y, Chen S, Shao X, Guo J, Liu X, Liu A, et al. Association of uric acid with metabolic syndrome in men, premenopausal women and postmenopausal women. International journal of environmental research and public health. 2014; 11: 2899-910.

272. Lim JH, Kim YK, Kim YS, Na SH, Rhee MY, Lee MM. Relationship between serum uric Acid levels, metabolic syndrome, and arterial stiffness in korean. Korean Circ J. 2010; 40: 314-20.

273. Lin CC, Liu CS, Li CI, Lin WY, Lai MM, Lin T, et al. The relation of metabolic syndrome according to five definitions to cardiovascular risk factors--a population-based study. BMC public health. 2009; 9: 484.

274. Lin YC, Chen JD, Lo SH, Chen PC. Worksite health screening programs for predicting the development of Metabolic Syndrome in middle-aged employees: a five-year follow-up study. BMC public health. 2010; 10: 747.

275. Liu PW, Chang TY, Chen JD. Serum uric acid and metabolic syndrome in Taiwanese adults. Metabolism: clinical and experimental. 2010; 59: 802-7.

276. Liu Y, Fan Y, Liu Q, Liu K, Chen F, Tang X, et al. Sex-specific association of serum uric acid dynamics with the incidence of metabolic syndrome in a health check-up Chinese population: a prospective cohort study. BMJ open. 2020; 10: e035289.

277. Meshkani R, Zargari M, Larijani B. The relationship between uric acid and metabolic syndrome in normal glucose tolerance and normal fasting glucose subjects. Acta diabetologica. 2011; 48: 79-88.

278. Nagahama K, Inoue T, Kohagura K, Ishihara A, Kinjo K, Ohya Y. Hyperuricemia predicts future metabolic syndrome: a 4-year follow-up study of a large screened cohort in Okinawa, Japan. Hypertens Res. 2014; 37: 232-8.

279. Ni W, Wang R, Liu Z, Yuan X, Chi H, Lv D, et al. Association of Serum Uric Acid with Metabolic Syndrome and Its Components: A Cross-Sectional Study in Chinese Coastal Population. Metabolic syndrome and related disorders. 2020; 18: 103-09.

280. Obokata M, Negishi K, Ohyama Y, Okada H, Imai K, Kurabayashi M. A risk score with additional four independent factors to predict the incidence and recovery from metabolic syndrome: Development and validation in large Japanese cohorts. PLoS ONE. 2015; 10.

281. Rhee C, Kim J, Kim JY, Chang E, Park SY, Lee W, et al. Clinical Markers Associated with Metabolic Syndrome Among Military Aviators. Aerosp Med Hum Perform. 2015; 86: 970-5.

282. Safiri S, Qorbani M, Heshmat R, Tajbakhsh R, Eslami Shahr Babaki A, Djalalinia S, et al. Association of Serum Uric Acid With Cardiometabolic Risk Factors and Metabolic Syndrome in Iranian Adolescents: the CASPIAN-III Study. Iranian journal of kidney diseases. 2016; 10: 126-34.

283. Salehidoost R, Aminorroaya A, Zare M, Amini M. Is uric acid an indicator of metabolic syndrome in the first-degree relatives of patients with type 2 diabetes? J Res Med Sci. 2012; 17: 1005-10.

284. Schmitt AC, Cardoso MR, Lopes H, Pereira WM, Pereira EC, de Rezende DA, et al. Prevalence of metabolic syndrome and associated factors in women aged 35 to 65 years who were enrolled in a family health program in Brazil. Menopause (New York, NY). 2013; 20: 470-6.

285. Song YM, Lee K. Genetic and Environmental Influences on the Associations Between Uric Acid Levels and Metabolic Syndrome Over Time. Metabolic syndrome and related disorders. 2018; 16: 299-304.

286. Sumiyoshi H, Ohyama Y, Imai K, Kurabayashi M, Saito Y, Nakamura T. Association of Uric Acid with Incident Metabolic Syndrome in a Japanese General Population. Int Heart J. 2019; 60: 830-35.

287. Tani S, Matsuo R, Imatake K, Suzuki Y, Takahashi A, Yagi T, et al. The serum uric acid level in females may be a better indicator of metabolic syndrome and its components than in males in a Japanese population. J Cardiol. 2020; 76: 100-08.

288. Tao LX, Li X, Zhu HP, Huo D, Zhou T, Pan L, et al. Association of hematological parameters with metabolic syndrome in Beijing adult population: a longitudinal study. Endocrine. 2014; 46: 485-95.

289. Tsai TY, Cheng JF, Lai YM. Prevalence of metabolic syndrome and related factors in Taiwanese high-tech industry workers. Clinics (Sao Paulo, Brazil). 2011; 66: 1531-5.

290. Wakabayashi I. Inverse associations between serum urate and glycemic status in a general population and in persons with diabetes mellitus. Diabetology & metabolic syndrome. 2020; 12: 21.

291. Wang JY, Chen YL, Hsu CH, Tang SH, Wu CZ, Pei D. Predictive value of serum uric acid levels for the diagnosis of metabolic syndrome in adolescents. The Journal of pediatrics. 2012; 161: 753-6.e2.

292. Yadav D, Lee ES, Kim HM, Choi E, Lee EY, Lim JS, et al. Prospective study of serum uric acid levels and incident metabolic syndrome in a Korean rural cohort. Atherosclerosis. 2015; 241: 271-7.

293. Yang T, Chu CH, Bai CH, You SL, Chou YC, Chou WY, et al. Uric acid level as a risk marker for metabolic syndrome: a Chinese cohort study. Atherosclerosis. 2012; 220: 525-31.

294. Yu TY, Jee JH, Bae JC, Jin SM, Baek JH, Lee MK, et al. Serum uric acid: A strong and independent predictor of metabolic syndrome after adjusting for body composition. Metabolism: clinical and experimental. 2016; 65: 432-40.

295. Yu TY, Jin SM, Jee JH, Bae JC, Lee MK, Kim JH. The Protective Effects of Increasing Serum Uric Acid Level on Development of Metabolic Syndrome. Diabetes & metabolism journal. 2019; 43: 504-20.

296. You L, Liu A, Wuyun G, Wu H, Wang P. Prevalence of hyperuricemia and the relationship between serum uric acid and metabolic syndrome in the Asian Mongolian area. J Atheroscler Thromb. 2014; 21: 355-65.

297. Zhang Q, Lou S, Meng Z, Ren X. Gender and age impacts on the correlations between hyperuricemia and metabolic syndrome in Chinese. Clinical rheumatology. 2011; 30: 777-87.

298. Zhang Z, Bian L, Choi Y. Serum uric acid: a marker of metabolic syndrome and subclinical atherosclerosis in Korean men. Angiology. 2012; 63: 420-8.

299. Zhang Q, Zhang C, Song X, Lin H, Zhang D, Meng W, et al. A longitudinal cohort based association study between uric acid level and metabolic syndrome in Chinese Han urban male population. BMC public health. 2012; 12: 419.

300. Zhang ML, Gao YX, Wang X, Chang H, Huang GW. Serum uric acid and appropriate cutoff value for prediction of metabolic syndrome among Chinese adults. J Clin Biochem Nutr. 2013; 52: 38-42.

301. Zhang H, Li Y, Mao Z, Liu X, Zhang X, Yang K, et al. Sex-specific associations of serum uric acid with metabolic syndrome in Chinese rural population: The RuralDiab study. Clinica chimica acta; international journal of clinical chemistry. 2018; 480: 119-25.

302. Zhao Y, Yu Y, Li H, Li M, Zhang D, Guo D, et al. The Association between Metabolic Syndrome and Biochemical Markers in Beijing Adolescents. International journal of environmental research and public health. 2019; 16.

303. Ziaee A, Esmailzadehha N, Ghorbani A, Asefzadeh S. Association between Uric Acid and Metabolic Syndrome in Qazvin Metabolic Diseases Study (QMDS), Iran. Glob J Health Sci. 2012; 5: 155-65.

304. Lin KP. The relationship between serum uric acid concentration and metabolic syndrome in university freshmen. J Nurs Res. 2009; 17: 286-92.

305. Ahn SV, Jung DH, Yadav D, Kim JY, Koh SB. Relative contribution of obesity and menopause to the association between serum adiponectin and incident metabolic syndrome. Menopause (New York, NY). 2018; 25: 154-59.

306. Ayina CNA, Endomba FTA, Mandengue SH, Noubiap JJN, Ngoa LSE, Boudou P, et al. Association of the leptin-to-adiponectin ratio with metabolic syndrome in a sub-Saharan African population. Diabetology & metabolic syndrome. 2017; 9: 66.

307. Ben Ali S, Jemaa R, Ftouhi B, Kallel A, Feki M, Slimene H, et al. Adiponectin and metabolic syndrome in a Tunisian population. Inflammation. 2012; 35: 828-33.

308. Cho SA, Joo HJ, Cho JY, Lee SH, Park JH, Hong SJ, et al. Visceral Fat Area and Serum Adiponectin Level Predict the Development of Metabolic Syndrome in a Community-Based Asymptomatic Population. PLoS One. 2017; 12: e0169289.

309. Jean-Luc Gradidge P, Norris SA, Jaff NG, Crowther NJ. Metabolic and Body Composition Risk Factors Associated with Metabolic Syndrome in a Cohort of Women with a High Prevalence of Cardiometabolic Disease. PLoS One. 2016; 11: e0162247.

310. Hata A, Yonemoto K, Shikama Y, Aki N, Kosugi C, Tamura A, et al. Cut-off value of total adiponectin for managing risk of developing metabolic syndrome in male Japanese workers. PLoS One. 2015; 10: e0118373.

311. Huh JH, Yoon TW, Kang DR, Kim JY. Prospective Study of Sex-Specific Adiponectin Changes and Incident Metabolic Syndrome: The ARIRANG Study. Journal of Clinical Medicine. 2019; 8.

312. Klünder-Klünder M, Flores-Huerta S, García-Macedo R, Peralta-Romero J, Cruz M. Adiponectin in eutrophic and obese children as a biomarker to predict metabolic syndrome and each of its components. BMC public health. 2013; 13: 88.

313. Koh SB, Park JK, Yoon JH, Chang SJ, Oh SS, Kim JY, et al. Preliminary report: a serious link between adiponectin levels and metabolic syndrome in a Korean nondiabetic population. Metabolism: clinical and experimental. 2010; 59: 333-7.

314. Lee SK, Yoon DW, Kim J, Lee SW, Kim JY, Jee SH, et al. Association of adiponectin, ghrelin, and leptin with metabolic syndrome and its metabolic components in Sasang constitutional type. Eur J Integr Med. 2018; 22: 16-21.

315. Lee KW, Shin D. Prospective Associations of Serum Adiponectin, Leptin, and Leptin-Adiponectin Ratio with Incidence of Metabolic Syndrome: The Korean Genome and Epidemiology Study. International journal of environmental research and public health. 2020; 17.

316. Li P, Jiang R, Li L, Liu C, Yang F, Qiu Y. Correlation of serum adiponectin and adiponectin gene polymorphism with metabolic syndrome in Chinese adolescents. European journal of clinical nutrition. 2015; 69: 62-7.

317. Song YM, Lee K, Sung J. Adiponectin Levels and Longitudinal Changes in Metabolic Syndrome: The Healthy Twin Study. Metabolic syndrome and related disorders. 2015; 13: 312-8.

318. Tanihara S, Imatoh T, Momose Y, Miyazaki M, Une H. Inverse correlation between adiponectin and the risk of metabolic syndrome in middle-aged Japanese male workers. Acta Med Okayama. 2009; 63: 325-30.

319. Yu D, Yu Z, Sun Q, Sun L, Li H, Song J, et al. Effects of body fat on the associations of high-molecular-weight adiponectin, leptin and soluble leptin receptor with metabolic syndrome in Chinese. PLoS One. 2011; 6: e16818.

320. Chatterjee B, Mehta M, Shah T, Mahant H, Katwa V, Gosai K. Association of Lipoprotein(a) and hsCRP Levels with Metabolic Syndrome and its Components. Indian Journal of Clinical Biochemistry. 2015; 30: 394-402.

321. Darroudi S, Fereydouni N, Tayefi M, Ahmadnezhad M, Zamani P, Tayefi B, et al. Oxidative stress and inflammation, two features associated with a high percentage body fat, and that may lead to diabetes mellitus and metabolic syndrome. BioFactors (Oxford, England). 2019; 45: 35-42.

322. Hong GB, Gao PC, Chen YY, Xia Y, Ke XS, Shao XF, et al. High-Sensitivity C-Reactive Protein Leads to Increased Incident Metabolic Syndrome in Women but Not in Men: A Five-Year Follow-Up Study in a Chinese Population. Diabetes, metabolic syndrome and obesity : targets and therapy. 2020; 13: 581-90.

323. Lai MM, Li CI, Kardia SL, Liu CS, Lin WY, Lee YD, et al. Sex difference in the association of metabolic syndrome with high sensitivity C-reactive protein in a Taiwanese population. BMC public health. 2010; 10: 429.

324. Mahajan A, Jaiswal A, Tabassum R, Podder A, Ghosh S, Madhu SV, et al. Elevated levels of C-reactive protein as a risk factor for metabolic syndrome in Indians. Atherosclerosis. 2012; 220: 275-81.

325. Meng G, Zhu Q, Shao J, Zhang Q, Liu L, Wu H, et al. Comparing the diagnostic ability of inflammatory markers in metabolic syndrome. Clinica chimica acta; international journal of clinical chemistry. 2017; 475: 1-6.

326. Perez CM, Ortiz AP, Guzman M, Suarez E. Distribution and correlates of the metabolic syndrome in adults living in the San Juan Metropolitan Area of Puerto Rico. Puerto Rico health sciences journal. 2012; 31: 114-22.

327. Song Y, Yang SK, Kim J, Lee DC. Association between C-Reactive Protein and Metabolic Syndrome in Korean Adults. Korean J Fam Med. 2019; 40: 116-23.

328. Thompson AM, Zhang Y, Tong W, Xu T, Chen J, Zhao L, et al. Association of inflammation and endothelial dysfunction with metabolic syndrome, prediabetes and diabetes in adults from Inner Mongolia, China. BMC endocrine disorders. 2011; 11: 16.

329. Yang T, Chu CH, Hsieh PC, Hsu CH, Chou YC, Yang SH, et al. C-reactive protein concentration as a significant correlate for metabolic syndrome: a Chinese population-based study. Endocrine. 2013; 43: 351-9.

330. Ying X, Jiang Y, Qin G, Qian Y, Shen X, Jiang Z, et al. Association of body mass index, waist circumference, and metabolic syndrome with serum cystatin C in a Chinese population. Medicine. 2017; 96: e6289.

331. Du R, Wu X, Peng K, Lin L, Li M, Xu Y, et al. Serum apolipoprotein B is associated with increased risk of metabolic syndrome among middle-aged and elderly Chinese: A cross-sectional and prospective cohort study. J Diabetes. 2019; 11: 752-60.

332. Ryoo JH, Park SK. Association of apolipoprotein B and incidence of metabolic syndrome in Korean men: a 5-years' follow-up study. Atherosclerosis. 2013; 226: 496-501.

333. Yi DW, Jeong DW, Lee SY, Son SM, Kang YH. The Association between Apolipoprotein A-II and Metabolic Syndrome in Korean Adults: A Comparison Study of Apolipoprotein A-I and Apolipoprotein B. Diabetes & metabolism journal. 2012; 36: 56-63.

334. Choi JR, Kim JY, Huh JH, Kim SH, Koh SB. Contribution of obesity as an effect regulator to an association between serum leptin and incident metabolic syndrome. Clinica chimica acta; international journal of clinical chemistry. 2018; 487: 275-80.

335. Esteghamati A, Noshad S, Khalilzadeh O, Morteza A, Nazeri A, Meysamie A, et al. Contribution of serum leptin to metabolic syndrome in obese and nonobese subjects. Archives of medical research. 2011; 42: 244-51.

336. Suriyaprom K, Tungtrongchitr R, Thawnasom K. Measurement of the levels of leptin, BDNF associated with polymorphisms LEP G2548A, LEPR Gln223Arg and BDNF Val66Met in Thai with metabolic syndrome. Diabetology & metabolic syndrome. 2014; 6: 6.

337. Yun JE, Kimm H, Jo J, Jee SH. Serum leptin is associated with metabolic syndrome in obese and nonobese Korean populations. Metabolism: clinical and experimental. 2010; 59: 424-9.

338. Bermudez V, Rojas J, Salazar J, Bello L, Anez R, Toledo A, et al. Variations of lipoprotein(a) levels in the metabolic syndrome: a report from the Maracaibo City Metabolic Syndrome Prevalence Study. Journal of diabetes research. 2013; 2013: 416451.

339. Jain SR, Shah KH, Acharya HN, Barot K, Sharma KH. Prevalence and Predictors of Metabolic Syndrome in Young Asymptomatic Gujarati Population. Int J Chronic Dis. 2015; 2015: 365217.

340. Wu XY, Lin L, Qi HY, Du R, Hu CY, Ma LN, et al. Association between Lipoprotein (a) Levels and Metabolic Syndrome in a Middle-aged and Elderly Chinese Cohort. Biomed Environ Sci. 2019; 32: 477-85.

341. Abdel-Moneim A, Mahmoud B, Sultan E, Mahmoud R. Association of erythrocytes indices and interleukin-1 beta with metabolic syndrome components. University of Toronto Medical Journal. 2020 97: 6-13.

Supplementary material

Appendix A

Search terms
Criteria for article selection and risk of bias assessment
Critical appraisal of studies
Data extraction and GRADE for each research question per predefined biomarker
Data extraction non-predefined biomarkers

https://doi.org/10.1111/obr.133129

7

Bariatric Surgery for Hypothalamic Obesity in Craniopharyngioma Patients: A Retrospective, Matched Case-Control Study

Selveta S. van Santen,[1,2] Peter Wolf,[3] Natalia Kremenevski,[4] Cesar L. Boguszewski,[5] Hannes Beiglböck,[3] Marta Fiocco,[2,6,7] Mark Wijnen,[1] Ville R. Wallenius,[8] Marry M. van den Heuvel-Eibrink,[2] Aart J. van der Lely,[1] Gudmundur Johannsson,[9,10] Anton Luger,[3] Michael Krebs,[3] Michael Buchfelder,[4] Patric J. D. Delhanty,[1] Sebastian J. C. M. M. Neggers[1,2,*], and Daniel S. Olsson.[9,10,*]

[1]Department of Endocrinology; Erasmus Medical Center, Rotterdam, The Netherlands;
[2]Princess Máxima Center for Paediatric Oncology, Utrecht, The Netherlands;
[3]Department of Endocrinology and Metabolism; Medical University of Vienna, Austria;
[4]Department of Neurosurgery; University Hospital Erlangen, Germany;
[5]Department of Internal Medicine; Universidade Federal do Paraná, Curitiba, Brazil;
[6]Mathematical Institute, Leiden University, Leiden, The Netherlands;
[7]Department of Biomedical Data Sciences, Medical Statistics Section, Leiden University Medical Center, Leiden, The Netherlands;
[8]Department of Gastrosurgical Research and Education; Sahlgrenska University Hospital, Gothenburg, Sweden;
[9]Department of Internal Medicine and Clinical Nutrition, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden;
[10]Department of Endocrinology at Sahlgrenska University Hospital, Gothenburg, Sweden.
*S.J.C.M.M.N. and D.S.O. provided equal contributions to the published work.

The Journal of Clinical Endocrinology & Metabolism

Abstract

Context: Craniopharyngioma is a sellar tumor associated with high rates of pituitary deficiencies (~98%) and hypothalamic obesity (~50%).

Objective: To determine the efficacy regarding long-term weight loss after bariatric surgery in obese craniopharyngioma patients with hypothalamic dysfunction.

Design: Retrospective case control study.

Setting: Multicenter international study.

Patients: Obese craniopharyngioma patients (N = 16; of which 12 women) with a history of bariatric surgery [12 Roux-en-Y gastric bypass, 4 sleeve gastrectomy; median age of 21 years (range 15-52), median follow-up 5.2 years (range 2.0-11.3)] and age/sex/surgery/BMI-matched obese controls (N = 155).

Main outcome measures: Weight loss and obesity-related comorbidities up to 5 years after bariatric surgery were compared and changes in hormonal replacement therapy evaluated.

Results: Mean weight loss at 5-year follow-up was 22.0% (95% CI 16.1, 27.8) in patients versus 29.5% (28.0, 30.9) in controls ($P = 0.02$), which was less after Roux-en-Y gastric bypass (22.7% [16.9, 28.5] vs. 32.0% [30.4, 33.6]; $P = 0.003$) but at a similar level after sleeve gastrectomy (21.7% [–1.8, 45.2] vs. 21.8% [18.2, 25.5]; $P = 0.96$). No major changes in endocrine replacement therapy were observed after surgery. One patient died (unknown cause). One patient had long-term absorptive problems.

Conclusions: Obese patients with craniopharyngioma had a substantial mean weight loss of 22% at 5-year follow-up after bariatric surgery, independent of type of bariatric surgery procedure. Weight loss was lower than in obese controls after Roux-en-Y gastric bypass. Bariatric surgery appears effective and relatively safe in the treatment of obese craniopharyngioma patients.

Keywords: craniopharyngioma, hypothalamic obesity, hypothalamic dysfunction, bariatric surgery, weight loss, case-control study

Introduction

Craniopharyngiomas are rare brain tumors, which mostly affect children or older adults.[1-3] They are typically located in the sellar and suprasellar regions.[1,2] State-of-the- art treatment for craniopharyngiomas is tumor resection with or without radiotherapy.[2] Although craniopharyngiomas usually have a benign histology, treated patients often suffer from severe long-term sequelae as a result of hypothalamic dysfunction due to tumor localization or therapeutic interventions.[2-4]

Besides the necessity for life-long hormone replacement therapy due to hypopituitarism, hypothalamic dysfunction can also cause eating disorders such as hyperphagia, leading to obesity in 50% to 75% of the patients.[2,5,6] Energy expenditure may also be decreased and cognitive performance can be weak, which can interfere with conservative weight loss strategies.[2,7] Morbid hypothalamic obesity and associated complications like type 2 diabetes, hypertension, obstructive sleep apnea syndrome, hypersomnia and increased daytime sleepiness have a major impact on quality of life in patients with craniopharyngioma.[3,6,8-11] The morbid obesity and related comorbidities contribute to an increased cardiovascular mortality.[3,6,8-11]

Since pharmacologic treatment options are limited,[12] bariatric surgery might be a promising treatment strategy in combating obesity in patients with craniopharyngioma. Key to the effectiveness of bariatric surgery in the general obese population is a decrease in appetite, which is caused by changes in gastrointestinal hormones such as glucagon-like peptide 1 (GLP-1).[13,14] GLP-1 activates neurons in the hypothalamus that influence satiety, feeding, the sympathetic nervous system, and the pituitary.[12,13] Bariatric surgical procedures are not all equally effective – gastric bypass surgery seems to be the most promising option in the general population.[15-17] However, the question remains whether bariatric surgery is still effective if the hypothalamus is damaged, as is often the case in patients with craniopharyngioma.[2] Previously, studies that were hampered by a follow-up of ≤2 years reported a significant weight reduction following gastric bypass surgery in patients with severe hypothalamic obesity without postoperative impairment of oral

hormone replacement therapy for pituitary insufficiencies.[16-18]

In patients suffering from morbid obesity in the general population, bariatric surgery is an efficient treatment with sustained long-term weight loss.[19,20] However, data on the long-term effects of bariatric surgery in morbid hypothalamic obesity due to a craniopharyngioma are limited. This is a major barrier in providing evidence-based advice to patients. This study is therefore aimed at analyzing (medium) long-term weight reduction following gastric bypass and sleeve gastrectomy surgery in obese craniopharyngioma patients compared to a matched control group from a general obese population treated with bariatric surgery and at describing safety aspects regarding pituitary hormone replacement therapy.

Materials and methods

Study Design and Participants

In this international, multicenter, matched case-control study, patients with a history of craniopharyngioma and bariatric surgery to treat hypothalamic obesity were compared to bariatric surgery patients from a general obese population. Sixteen patients with histopathology-proven craniopharyngioma with ≥2 years follow-up were included from the Erasmus Medical Center, Rotterdam, The Netherlands (n = 4),[17] the Sahlgrenska University Hospital, Gothenburg, Sweden (n = 4),[17] the Medical University Hospital of Vienna, Austria (n = 5), University Hospital Erlangen, Erlangen, Germany (n = 2), and the Federal University of Parana, Curitiba, Brazil (n = 1). The study methods and the Dutch/Swedish patients have been previously described;[17] this report increased patient numbers by recruiting from additional centers and increased follow-up duration. This international study followed all national laws and recommendations in the country where the patient was treated concerning ethical approval and written consent. The study was approved by all local ethics committees. Two patients underwent a second bariatric surgery procedure; one had the Roux-en-Y gastric bypass ~20 months after a sleeve gastrectomy due to insufficient

weight loss and abdominal pain, and one patient had a Roux-en-Y gastric bypass ~6 years after gastric banding (the gastric banding was removed after 1 week due to abdominal complaints). Data from the second bariatric surgery onwards were applied in this analysis. Hypothalamic damage was defined as injury to the hypothalamus and/or third ventricle, diagnosed by neuroimaging and/or neurosurgery reports.[8] Patients with a history of radiotherapy were considered at high risk of hypothalamic dysfunction and therefore also included in the study if they met all other inclusion criteria.

Matching Procedure for Controls

Controls were acquired from the Scandinavian Obesity Surgery Registry (SOReg), a Swedish nationwide registry. The patients were matched to controls from a sample of 69,672 individuals who were pre-selected from the total cohort of 75,600 SOReg participants after exclusion of subjects who were not Swedish, were reoperated, had surgery before 2007, or had a procedure other than gastric bypass surgery or sleeve gastrectomy. Controls had follow-up data on body weight (kg) available at 6 weeks, and at 1, 2, and 5 years after bariatric surgery. Patients had a considerable variation in follow-up duration due to the retrospective design. Missing data for 3- and 4-year follow-up in controls and 1- to 5-year follow-up in patients was interpolated linearly between the two closest available time points. The matching procedure was extensive: potential controls were first selected according to sex, type of bariatric surgery (Roux-en-Y gastric bypass or sleeve gastrectomy), pre-operative type 2 diabetes, and preoperative hypertension. Further matching was performed by year of obesity operation (10-year span category), age at obesity operation (10-year span category), and preoperative body mass index (BMI) (maximum of ±5 kg/m^2 different from the control). Controls were included only once. If less than ten controls were found, the criteria for matching age at bariatric surgery were extended to ±10 years of the patient's age instead of a certain age category, which was required in five patients. For one patient, the criteria for BMI were extended as this patient was an outlier due to extremely high BMI: the best-matched controls were chosen without a limit to BMI criteria. Ultimately,

all patients were each matched with ten controls except for two patients: nine controls were found for one and six controls were found for the extreme outlier. This resulted in the selection of 155 optimally matched controls (mean BMI difference –1.1 kg/m^2; maximum BMI difference between patient and control ranging from 0.04 to 12.6 kg/m^2)

Outcomes of Interest

Data were gathered retrospectively. Outcomes of interest were percentage weight change at 6 weeks, at 1, 2, 3, 4, and 5 years, and at last available follow-up after bariatric surgery. The presence of type 2 diabetes, hypertension, and dyslipidemia as comorbidities before bariatric surgery and during follow-up, and complications of the bariatric procedure were evaluated. We studied alterations in hormone replacement therapy for pituitary deficiency in patients with craniopharyngioma.[17] For insulin-like growth factor 1 (IGF-1) values, standardized deviation scores (SDS) were calculated if the applied assay and normative data were known.[21-24]

Statistical Procedure

Statistical analysis was performed using the Statistical Package for Social Sciences (IBM SPSS Statistics 25, Chicago, IL). Continuous data are represented as mean ± standard deviation (SD), or median and range; categorical data are represented as frequencies and percentages. Baseline statistics were compared between patients with obesity after craniopharyngioma and the matched controls from the SOReg database by Mann-Whitney U-test and Fisher's exact test for continuous and categorical data, respectively. Related continuous data were evaluated with Wilcoxon's rank test. A two-way analysis of variance was used to compare percentage weight change between patients and controls. In this procedure, a one-factor generalized randomized block design was applied with matched case-control units included as blocks. Percentage weight change was applied as the dependent variable and type of subject (patient with

craniopharyngioma or control) as the independent variable in the two-way ANOVA. Bootstrapping with 1,000 replicates was performed to meet the normality assumption related to the two-way analysis of variance test.[17]

Results

Patient Characteristics

The characteristics of the 16 craniopharyngioma patients treated with bariatric surgery are shown in Table 1. Twelve (75%) patients were female and 13 (81%) had childhood (<18 years) onset of disease. Initial treatment for craniopharyngioma was surgery (n = 13 [81%]), surgery and radiotherapy (n = 2 [13%]), and cyst aspiration (n = 1 [6%]). Two patients (13%) had a history of recurrence of craniopharyngioma twice and nine patients (56%) had residual tumor tissue on their last magnetic resonance imaging (MRI) scan. All patients had signs of either hypothalamic damage or third-ventricle involvement on MRI (n = 13 [87%, data missing for 1 patient]), or a history of radiotherapy (n = 7 [44%]). Two patients (13%) had used medication to treat obesity before bariatric surgery: sibutramine with no results, and sibutramine and orlistat with no results and no results/side effects, respectively. Twelve patients (75%) underwent Roux-en-Y gastric bypass and four patients (25%) underwent sleeve gastrectomy. Median follow-up duration of the patients since bariatric surgery was 5.8 years (range 2.0–11.3), hereafter being referred to as last follow-up.

A comparison of baseline characteristics of patients with craniopharyngioma and their controls with 'common' obesity matched for age, sex, comorbidity, BMI, and bariatric surgery is shown in Table 2. Baseline characteristics were comparable between patients and controls except for age at bariatric surgery (with a slightly lower mean age of 26 years [SD 12] in patients vs. 31 years [12] in controls; P = 0.03) and more frequent presence of dyslipidemia before surgery (4/16 [25%] in patients vs. 6/155 [4%] in controls; P = 0.008); the difference in the presence of dyslipidemia between patients and controls before

surgery was more pronounced in those undergoing sleeve gastrectomy (3/4 [75%] vs. 1/39 [3%]; $P = 0.001$]. The occurrence of dyslipidemia before bariatric surgery was higher in craniopharyngioma patients who had a gastric sleeve than in those who underwent gastric bypass (3/4 [75%] vs. 1/12 [8%]; $P = 0.03$).

Table 1 Baseline demographic and clinical characteristics of patients with craniopharyngioma

Characteristic	Craniopharyngioma patients (N = 16)
Sex, n (%)	
Women	12 (75)
Men	4 (25)
Median (range) age at first craniopharyngioma treatment (years)	12 (4–48)
Median age (range) at last follow-up (years)	33 (17–61)
Mean (SD) follow-up duration since craniopharyngioma surgery at last follow-up (years)	11.9 ± 3.8
Treatment for craniopharyngioma, n (%)	
Surgery	
Initially*	15 (94)
Ever	15 (94)
Median (range) number of craniopharyngioma surgeries	1 (1–6)
Radiotherapy	
Initially (in addition to surgery)	2 (13)
Ever	7 (44)
Mean (SD) cumulative radiotherapy dose (mGy)	4,225 ± 1,801
Hypothalamic damage, n (%)	9 (60)
Third-ventricle involvement, n (%)	9 (60)
Hypothalamic damage and/or third-ventricle involvement, n (%)	13 (87)

Hypothalamic damage and/or third-ventricle involvement and/or radiotherapy, n (%)	16 (100)
Pituitary deficiencies	
GH deficiency	
Frequency, n (%)	16 (100)
Median (range) age at occurrence (years)	13 (6–49)
GH replacement therapy at last follow-up, n (%)	14 (88)
TSH deficiency	
Frequency, n (%)	15 (94)
Median (range) age at occurrence (years)	14 (4–48)
Gonadal axis deficiency	
Frequency, n (%)	14 (88)
Median (range) age at occurrence (years)	13 (6–48)
Gonadal replacement therapy at last follow-up, n (%)	12 (75)
ACTH deficiency	
Frequency, n (%)	12 (75)
Median (range) age at occurrence (years)	12 (4–48)
ADH deficiency	
Frequency, n (%)	14 (88)
Median (range) age at occurrence (years)	12 (4–48)
Use of antiepileptic drugs, n (%)	2 (13)
Bariatric procedure	
Median (range) age at bariatric surgery (years)	21 (15–52)
Mean (SD) BMI before bariatric surgery (kg/m^2)	46.0 ± 8.0
Median (range) follow-up since bariatric procedure (years)	5.8 (2.0–11.3)

ACTH, adenocorticotropic hormone; ADH, antidiuretic hormone; BMI, body mass index; GH, growth hormone; TSH, thyroid-stimulating hormone. *Excluding one patient treated with cyst aspiration initially.

Table 2 Baseline characteristics of patients treated with bariatric surgery: craniopharyngioma-related hypothalamic obesity versus controls with 'common' obesity

Characteristic	Craniopharyngioma patients (N = 16)	Matched controls (N = 155)
Sex, n (%)		
Female	12 (75)	119 (77)
Male	4 (25)	36 (23)
Mean (SD) age at bariatric surgery (years)	26.4 ± 12.1	30.5 ± 11.5*
Bariatric procedure, n (%)		
Roux-en-Y gastric bypass	12 (75)	116 (75)
Sleeve gastrectomy	4 (25%)	39 (25)
Mean (SD) preoperative BMI (kg/m^2)	46.0 ± 8.0	45.1 ± 6.9
Roux-en-Y gastric bypass	45.4 ± 6.0	44.9 ± 5.6
Sleeve gastrectomy	48.0 ± 13.5	45.6 ± 9.7
Pre-operative diabetes mellitus, n (%)	1 (6)	10 (6)
Pre-operative hypertension, n (%)	4 (25)	35 (23)
Pre-operative dyslipidemia, n (%)	4 (25)	6 (4)†

BMI, body mass index. *P = 0.03, †P = 0.008.

Weight Change after Bariatric Surgery

Mean weight loss at 5 years after surgery was 22.0% (95% CI 16.1, 27.8) in craniopharyngioma patients compared to 29.5% (28.0, 30.9) in controls ($P = 0.02$; Table 3 and Figure 1). Patients had significantly less weight loss compared to controls from 1- to 5-year follow-up after any bariatric surgery procedure and from 2- to 5-year follow-up after Roux-en-Y gastric bypass specifically, but not after sleeve gastrectomy specifically. Mean weight loss at 5-year follow-up after Roux-en-Y surgery was less in patients compared to controls (22.7% [95% CI 16.9, 28.5] vs. 32.0% [30.4, 33.6]; $P = 0.003$) but was comparable at 5-year follow-up after sleeve gastrectomy comparing patients and controls, respectively (21.7% [−1.8, 45.2] vs. 21.8% [18.2, 25.5], $P = 0.96$). If the type of bariatric surgery is compared in patients or controls as a group, mean percentage weight loss at 4-year follow-up in controls was higher after Roux-en-Y gastric bypass compared to sleeve gastrectomy (33.0 [SD 7.8] vs. 24.0 [9.9]; $P < 0.001$), but not different when comparing these bariatric procedures in craniopharyngioma patients (21.8 [12.0] vs. 19.1 [5.7], $P = 1.00$). Figure 2 shows the percentage of patients and controls in 5% weight loss categories at 2- and 4-year follow-up. At last follow-up, eight (50%) of 16 patients had lost at least 20% of their original body weight, three (19%) between 10% and 15%, three (19%) between 5% and 10%, and one (6%) <5% weight loss, while one (6%) showed weight increase. Among the patients who used sibutramine, one had at least 20% weight reduction and the other had 5–10% weight loss at last follow-up. There was no significant difference in mean percentage weight loss at last follow-up comparing patients with (n = 13) and without (n = 2) hypothalamic damage or third-ventricle involvement (23.2% [SD 16.6] vs. 10.5% [5.4]; $P = 0.23$, one missing) and those with (n = 7) and without (n = 9) radiotherapy (16.6% [16.9] vs. 23.5% [15.7]; $P = 0.41$), respectively.

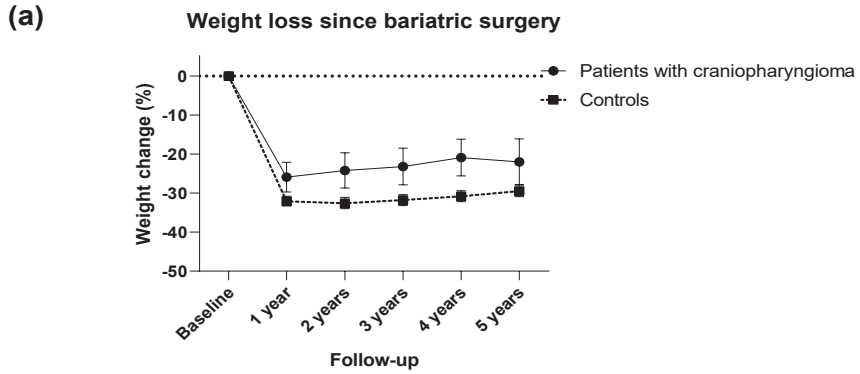
Table 3 Percentage weight loss after bariatric surgery of patients with craniopharyngioma and matched controls

Time after surgery	% Weight loss (95% CI)		
	Sleeve gastrectomy		
	Patients	Controls	*P*-value
6 weeks	18.1 (12.5, 23.7)	14.7 (12.9, 16.5)	0.25
1 year	25.3 (17.4, 33.2)	27.5 (25.0, 30.0	0.60
2 years	23.8 (14.6, 33.0)	27.1 (24.1, 30.0)	0.50
3 years	28.9 (18.4, 39.5)	25.5 (22.6, 28.4)	0.53
4 years	22.5 (8.9, 36.0)	24.0 (21.0, 26.7)	0.82
5 years	21.7 (–1.8, 45.2)	21.8 (18.2, 25.5)	0.96

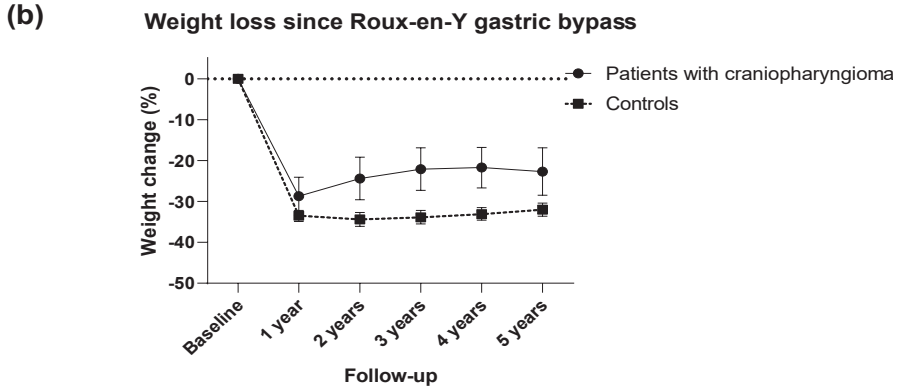
Time after surgery	% Weight loss (95% CI)		
	Roux-en-Y gastric bypass		
	Patients	Controls	*P*-value
6 weeks	15.0 (12.1, 18.0)	15.5 (14.6, 16.4)	0.74
1 year	28.7 (24.1, 33.4)	33.5 (32.2, 34.9)	0.05
2 years	24.4 (19.2, 29.6)	34.4 (32.7, 36.1)	<0.001
3 years	22.1 (16.9, 27.3)	33.9 (32.2, 35.5)	<0.001
4 years	21.7 (16.8, 26.7)	33.1 (31.5, 34.6)	<0.001
5 years	22.7 (16.9, 28.5)	32.0 (30.4, 33.6)	0.003

Time after surgery	% Weight loss (95% CI)		
	Any bariatric surgery procedure		
	Patients	Controls	*P*-value
6 weeks	15.8 (13.2, 18.4)	15.3 (14.5, 16.1)	0.71
1 year	25.9 (22.1, 29.7)	32.1 (30.8, 33.3)	0.003
2 years	24.2 (19.7, 28.7)	32.6 (31.1, 34.0)	0.001
3 years	23.2 (18.5, 27.9)	31.8 (30.4, 33.2)	0.001
4 years	20.9 (16.2, 25.6)	30.8 (29.4, 32.2)	<0.001
5 years	22.0 (16.1, 27.8)	29.5 (28.0, 30.9)	0.02

Figure 1 Weight loss up to 5-year follow-up after bariatric surgery



Patients with craniopharyngioma	n=16	16	16	15	13	10
Controls	n=155	155	155	125	124	96



Patients with craniopharyngioma	n=12	12	12	11	11	9
Controls	n=116	116	116	105	105	86

Patients with craniopharyngioma	n=4	4	4	3	2	1
Controls	n=39	39	39	29	19	10

Mean (SD) percentage weight loss in obese craniopharyngioma patients and matched controls after any bariatric surgery (A), Roux-en-Y gastric bypass specifically (B), and sleeve gastrectomy specifically (C).

Cardiometabolic Features

The prevalence of type 2 diabetes before bariatric surgery was similar in patients and controls (1/16 [6%] vs. 10/155 [6%]; *P* = 1.00). At last follow-up, type 2 diabetes had resolved in all subjects except for one control, resulting in similar percentages in patients and controls (0/16 [0%] vs. 1/155 [1%]; *P* = 1.00). Dyslipidemia occurred more often in patients than controls before bariatric surgery (4/16 [25%] vs. 6/155 [4%]; *P* =0.008) and at last follow-up (2/15 [13%; one missing data] vs. 2/155 [1%]; *P* = 0.04). Hypertension occurred at a similar prevalence in patients and controls before bariatric surgery (4/16 [25%] vs. 35/155 [23%]; *P* = 0.76) and there was no significant difference at last follow-up (0/16 [0%] vs. 18/155 [12%]; *P* = 0.22), although all cases of hypertension in patients were resolved at last follow-up.

Complications after Bariatric Surgery

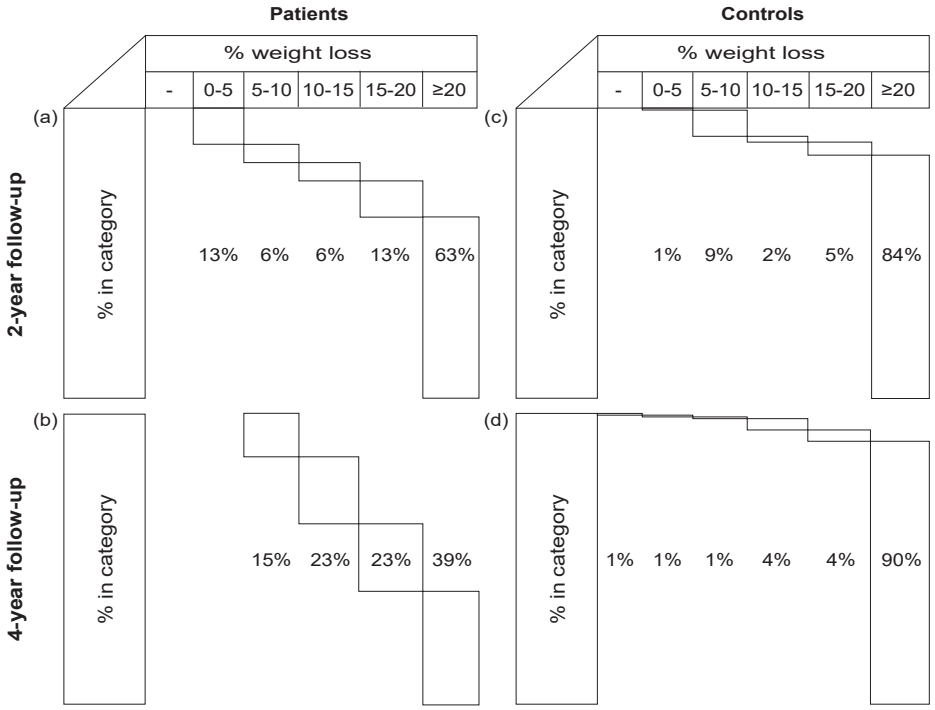
Five patients experienced short-term peri- and post-operative complaints or issues: post-operative abdominal pain (n = 1), dumping syndrome (n = 1), inability to eat solid food due to abdominal fullness (n = 1), stenosis of the anastomosis (n = 1), a generally complicated post-operative course with nephrolithiasis, pulmonary embolism, and post-infarction pneumonia (n = 1). Regarding long-term complications, one patient suffered from long-term severe absorptive problems ever since the bariatric surgery, which was accompanied by malnutrition and a low quality of life. One patient died at the age of 32 years, approximately 2.5 years after their second bariatric surgery; the cause of death was unknown. Since the cause of death could not be determined, an adrenal insufficiency cannot be excluded as a contributing factor to the fatality.

Replacement Therapies for Pituitary Deficiencies

Fifteen (95%) of the 16 patients needed minor-to-moderate changes of pituitary hormone replacement therapy during follow-up, the only exception being a patient only receiving growth hormone replacement

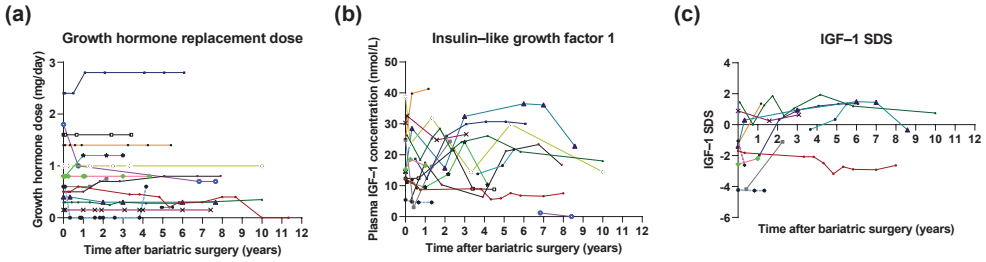
therapy (see Table 1 for baseline pituitary deficiencies, and Figures 3-4 for individual changes of replacement therapy). All patients were growth hormone deficient. Mean daily growth hormone dose was not significantly different before bariatric surgery versus last follow-up (0.92 [SD 0.65] vs.0.72 [0.88] mg; *P* = 0.50); similarly, mean IGF-1 (19.4 [10.1] vs. 34.0 [57.6] nmol/L; *P* = 0.72) and mean IGF-1 SDS values (–1.6 [1.6] vs. –0.7 [2.0]; *P* = 0.61) did not change significantly (Figure 3). One patient did not initially receive growth hormone replacement therapy due to fear of tumor growth but the patient reconsidered and decided to start during follow-up. Three patients stopped using growth hormone replacement therapy at some point during follow-up, one of which was due to diagnosis of a malignancy (Figure 3).

Figure 2 Waterfall plots of weight loss categories at 2- and 4-year follow-up after bariatric surgery.



Percentage of obese craniopharyngioma patients (A, B, n=16) and matched controls (C, D, n=155) in 5% weight loss categories for interpolated data at 2-year (A, C) and 4-year (B, D) follow-up after bariatric surgery. Percentages may not add up to 100% exactly due to rounding. Missing data at 4-year follow-up in three patients and one control.

Figure 3 Change in growth hormone replacement therapy and IGF-1 in obese craniopharyngioma patients following bariatric surgery

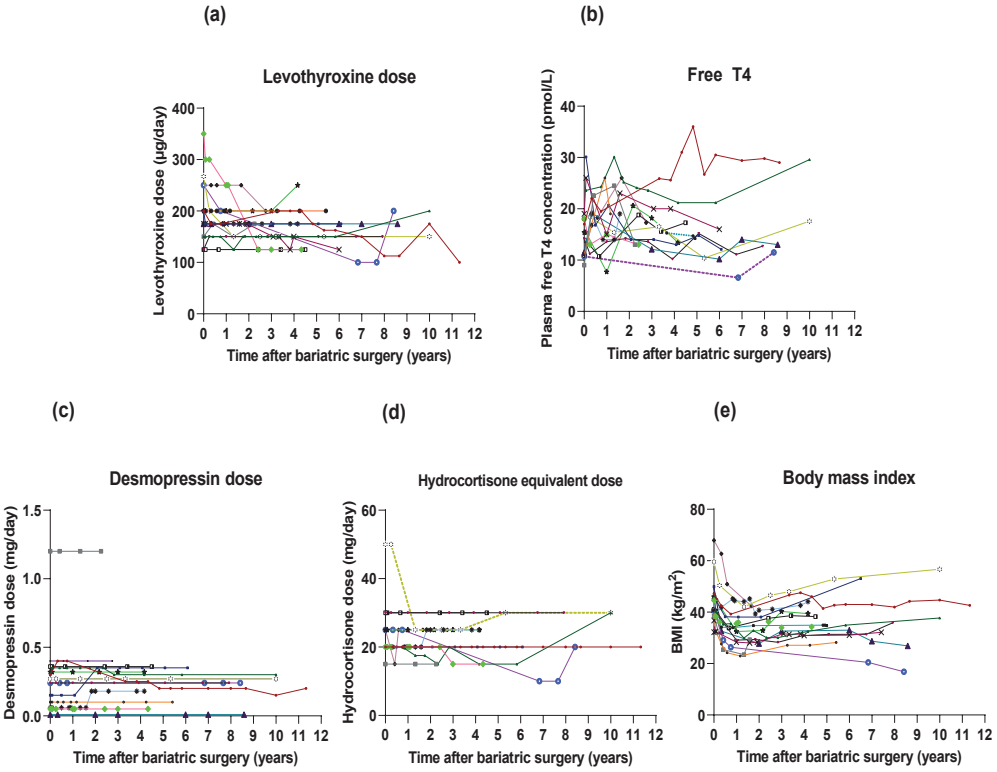


Spaghetti plots of changes in growth hormone replacement therapy daily dose (A), plasma IGF-1 (B), and IGF-1 SDS (C) for individual patients following bariatric surgery. A continuous line between observations represents patients using growth hormone replacement therapy and an interrupted line indicates patients who did not use growth hormone at last follow-up. IGF-1=insulin-like growth factor 1; SDS=standardised deviation score.

Fourteen (88%) of 16 patients had a gonadal hormone deficiency; eight had no change in their replacement therapy and one did not use any during follow-up. Three patients switched type of gonadal hormone replacement therapy (estradiol/dydrogesterone to ethinylestradiol/levonorgestrel at different doses; multiple esters of testosterone every 15 days intramuscular to testosterone undecanoate every 3 months; and gel application to injection). One patient required only a minor dose change. One patient stopped treatment due to a liver adenoma.

Thyroid hormone replacement therapy was needed in 15 (94%) of 16 patients, of which five had no changes in thyroid medication at all (Figure 4); mean daily levothyroxine dose was comparable before bariatric surgery and at last follow-up (199.5 [SD 59.8] vs. 171.2 [40.6] µg; $P = 0.23$). Mean cumulative daily 1-desamino-8-d-arginine vasopressin (DDAVP) doses for central diabetes insipidus remained similar comparing before surgery to last follow-up (0.29 [0.29] vs. 0.24 [0.10] mg; $P = 0.29$). DDAVP dose was unchanged in ten patients (Figure 4). Mean cumulative daily hydrocortisone dose did not change from before bariatric surgery to last follow-up (25.5 [9.3] vs. 24.2 [5.1] mg; $P = 0.85$).

Figure 4 Change in pituitary hormone replacement therapy, free T4, and BMI in obese craniopharyngioma patients following bariatric surgery



Spaghetti plots of changes in daily levothyroxine dose (A), plasma free T4 concentration (B), daily desmopressin dose (C), daily hydrocortisone dose (D), and BMI (E) for individual patients following bariatric surgery. A continuous line between observations represents patients with a deficiency for the particular pituitary hormone axis using hormone replacement therapy at last follow-up; an interrupted line is shown for patients who have a deficiency but do not use hormone replacement therapy at last follow-up; and a dotted line represents patients not known with this particular pituitary deficiency at last follow-up. BMI=body mass index, T4=thyroxine.

Discussion

This is the first long-term case-control study on the outcome of bariatric surgery in the largest cohort of patients with craniopharyngioma and hypothalamic dysfunction to date. It showed a mean ~22% weight loss 5 years after bariatric surgery. All patients required minor adjustments of hormonal replacement therapy after bariatric surgery, which were anticipated as most patients underwent a serious change in body weight. Although these results are encouraging, weight loss was significantly less pronounced, but still clinically relevant, compared to obese controls without a history of craniopharyngioma.

This less dramatic weight loss for craniopharyngioma patients compared to controls was, however, only observed after Roux-en-Y gastric bypass. This is in contrast to the results of our previous study in a smaller cohort,[17] when we found similar weight reduction after Roux-en-Y gastric bypass in patients with craniopharyngioma and controls, and less weight reduction in patients than controls after sleeve gastrectomy. The current study now includes the same Dutch and Swedish patients,[17] but the sample size has been enlarged by international cooperation, leading to a larger cohort of patients with a considerably longer follow-up after the bariatric procedure. The number of patients who underwent sleeve gastrectomy is, however, still relatively small (n = 4).

In a comparative effectiveness study in the general population, patients with Roux-en-Y gastric bypass had a greater mean weight loss (25.5%) than patients with sleeve gastrectomy (18.8%) at 5-year follow-up.[20] A previous meta-analysis showed that there were also better results at 5-year follow-up for Roux-en-Y gastric bypass compared to sleeve gastrectomy regarding not only weight loss but also remission of comorbidities such as hypertension, dyslipidemia, and type 2 diabetes.[19] The number of events in patients was too low in our study to compare decline in comorbidities between the two types of surgery. In the general population, Roux-en-Y gastric bypass patients had, on the other hand, a higher 30-day rate of major adverse events than those undergoing sleeve gastrectomy (5.0% vs. 2.6%).[20]

It seems that craniopharyngioma patients have a higher risk of post-operative adverse events than the general population, as five (31%) patients had problems shortly after surgery, of which two (13% of craniopharyngioma bariatric surgery patients) were serious adverse events. The risk-benefit ratio must especially be taken into account when applying bariatric surgery to underaged patients who may be unable to make a proper informed discussion. Considering our patients had similar weight loss compared to controls after sleeve gastrectomy and their weight reduction was similar after sleeve gastrectomy compared to Roux-en-Y gastric bypass as well as the lower adverse event rate with sleeve gastrectomy in the general population, sleeve gastrectomy may be considered a more advantageous strategy in patients with craniopharyngioma. Our study did not include laparoscopic gastric banding, which is another bariatric surgery procedure.[16] One patient included in our study was subject to a previous unsuccessful laparoscopic gastric banding (LAGB). Weismann et al. reported on 3 out of 6 described patients with LAGB that needed another bariatric surgery procedure.[16,25] LABG appears a less effective option in these patients.[25] Adjustable gastric banding is less effective in the general population as well: the PCORnet study described a mean five-year weight loss of 12% for LAGB, versus 26% after Roux-en-Y gastric bypass, and 19% for sleeve gastrectomy.[20] Only one patient was described to have had a biliopancreatic diversion.[25] In the general population, biliopancreatic diversion is not often performed as it is accompanied with very high rates of severe nutritional deficiencies and high rates of revisions.[26]

Although individual patients required adjustments of hormone replacement therapy during follow-up, mean doses did not change significantly. Despite growth hormone being administered subcutaneously and thus not being absorbed in the intestine, it is unsurprising some dose changes were needed during follow-up as growth hormone doses are known to be influenced by age and BMI.[27] Weight-based regimens have been proposed for glucocorticoid replacement, growth hormone replacement, and thyroid hormone replacement.[27,28] Changes in hormone replacement therapy can be considered as part of long-term practice in the care of patients with

hypopituitarism and are expected in the case of weight change; this did not lead to any confirmed major adverse events such as acute adrenal crisis. Our results suggest that bariatric surgery can be regarded as safe for patients with hypopituitarism and complementary replacement therapy, which is in line with previous research that found no major negative effects regarding hormone replacement therapy.[17,18,29,30] Wolf and colleagues[18] performed an oral thyroid/hydrocortisone/paracetamol absorption test in a patient who had undergone gastric bypass surgery and found sufficient gastrointestinal drug absorption. Hence, additional emphasis on individual drug management and adjustments, especially shortly after bariatric surgery, seems important.[29]

A limitation of our study is the retrospective design. Nevertheless, the study has several strengths. For such a rare disease, we report the largest sample size in the history of studies investigating bariatric surgery after craniopharyngioma and our study is unique in its duration of long-term follow-up. In addition, the cases were matched almost perfectly to controls from an average obese non-craniopharyngioma bariatric surgery population, thereby enabling an optimal comparison. Future research could investigate whether our data can be generalized to subjects with other causes of hypothalamic dysfunction.

Future studies that include even more patients and a longer follow-up as well as investigating differences between responders and non-responders to bariatric surgery with respect to weight loss will be able to show whether the observations from our study are sustained over time. Sleeve gastrectomy combines restriction of food intake with favorable hormonal alterations and Roux-en-Y gastric bypass adds a component of mild malabsorption to that in the general obese population.[29] As our study shows a similar weight loss effect in patients compared to controls with respect to sleeve gastrectomy but not after Roux-en-Y gastric bypass, it would be of interest to measure hormonal changes, such as GLP-1, related to bariatric surgery in future studies.[14] This would not only provide insight into the changed pathophysiology in patients with craniopharyngioma and hypothalamic dysfunction but also contribute to the exploration of other weight loss strategies for obesity in these

patients, such as GLP- 1 analogues.[31] In a small study of ten patients with different causes of hypothalamic obesity, treatment with the GLP-1 analogue exenatide resulted in weight stabilization or decrease.[31] It might also contribute to strategies to maintain weight loss after bariatric surgery: for example, the GLP-1 analogue liraglutide provided weight loss in a patient with craniopharyngioma whose bariatric surgery failed to be effective long term.[32]

In conclusion, patients with craniopharyngioma had a mean weight loss of 21% up to 5 years after bariatric surgery. Although weight loss was significantly less compared to matched obese controls, this was a clinically relevant reduction. Weight loss was only less in patients who underwent Roux-en-Y gastric bypass surgery while it was similar in those who underwent sleeve gastrectomy, compared to controls. Weight loss appears independent of bariatric surgery type. As for all patients with hypopituitarism, craniopharyngioma patients needed endocrinological follow-up and guidance with special attention for dose adjustment of their hormonal replacement therapy after bariatric surgery, but in this retrospective analysis no major changes were made in terms of dose and type of treatment. There were no confirmed cases of adrenal insufficiency. Bariatric surgery can therefore be regarded as a relatively safe and efficacious option in patients with craniopharyngioma who have no or mild cognitive decline and can cope with lifestyle restrictions after bariatric procedure. The possible downsides of bariatric surgery in craniopharyngioma patients are suboptimal weight loss compared to controls and the potential for absorption problems and perhaps more postoperative problems, which should be thoroughly discussed with patients before proceeding with this invasive intervention.[3]

References

1. Drapeau A, Walz PC, Eide JG, et al. Pediatric craniopharyngioma. Childs Nerv Syst 2019;35:2133–2145.
2. Müller HL, Merchant TE, Warmuth-Metz M, Martinez-Barbera JP, Puget S. Craniopharyngioma. Nat Rev Dis Primers 2019;5:75.
3. Olsson DS, Andersson E, Bryngelsson IL, Nilsson AG, Johannsson G. Excess mortality and morbidity in patients with craniopharyngioma, especially in patients with childhood onset: a population-based study in Sweden. J Clin Endocrinol Metab 2015;100:467–474.
4. Wijnen M, van den Heuvel-Eibrink MM, Janssen J, et al. Very long-term sequelae of craniopharyngioma. Eur J Endocrinol 2017;176:755–767.
5. van Santen SS, Olsson DS, Hammarstrand C, et al. Diagnosing metabolic syndrome in craniopharyngioma patients: body composition versus BMI. Eur J Endocrinol 2019;181:173–183.
6. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. The metabolic syndrome and its components in 178 patients treated for craniopharyngioma after 16 years of follow-up. Eur J Endocrinol 2017;178:11–22.
7. Roth CL. Hypothalamic obesity in patients with craniopharyngioma: Profound changes of several weight regulatory circuits. Front Endocrinol (Lausanne) 2011;2:49.
8. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. Excess morbidity and mortality in patients with craniopharyngioma: a hospital-based retrospective cohort study. Eur J Endocrinol 120;1878:95–104.
9. Müller HL. Consequences of craniopharyngioma surgery in children. J Clin Endocrinol Metab 2011;96:1981–1991.
10. Crowley RK, Woods C, Fleming M, et al. Somnolence in adult craniopharyngioma patients is a common, heterogeneous condition that is potentially treatable. Clin Endocrinol (Oxf) 2011;74:750–755.
11. Iyigun I, Alikasifoglu A, Gonc N, et al. Obstructive sleep apnea in children with hypothalamic obesity: evaluation of possible related factors. Pediatr Pulmonol 2020;55:3532–3540.
12. van Iersel L, Brokke KE, Adan RAH, Bulthuis LCM, van den Akker ELT, van Santen HM. Pathophysiology and individualized treatment of hypothalamic obesity following craniopharyngioma and other suprasellar tumors: a systematic review. Endocr Rev 2019;40:193–235.
13. Rao RS. Bariatric surgery and the central nervous system. Obes Surg 2012;22:967–978.

14. Casimiro I, Sam S, Brady MJ. Endocrine implications of bariatric surgery: a review on the intersection between incretins, bone, and sex hormones. Physiol Rep 2019;7:e14111.

15. Bingham NC, Rose SR, Inge TH. Bariatric surgery in hypothalamic obesity. Front Endocrinol (Lausanne) 2012;3:23.

16. Weismann D, Pelka T, Bender G, et al. Bariatric surgery for morbid obesity in craniopharyngioma. Clin Endocrinol (Oxf) 2013;78:385–390.

17. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. Efficacy and safety of bariatric surgery for craniopharyngioma-related hypothalamic obesity: a matched case-control study with 2 years of follow-up. Int J Obes (Lond) 2017;41:210–216.

18. Wolf P, Winhofer Y, Smajis S, et al. Hormone substitution after gastric bypass surgery in patients with hypopituitarism secondary to craniopharyngioma. Endocr Pract 2016;22:595–601.

19. Gu L, Huang X, Li S, et al. A meta-analysis of the medium- and long-term effects of laparoscopic sleeve gastrectomy and laparoscopic Roux-en-Y gastric bypass. BMC Surg 2020;20:30.

20. Arterburn D, Wellman R, Emiliano A, et al. Comparative effectiveness and safety of bariatric procedures for weight loss: a PCORnet Cohort Study. Ann Intern Med 2018;169:741–750.

21. Bidlingmaier M, Friedrich N, Emeny RT, et al. Reference intervals for insulin-like growth factor-1 (IGF-1) from birth to senescence: results from a multicenter study using a new automated chemiluminescence IGF-1 immunoassay conforming to recent international recommendations. J Clin Endocrinol Metab 2014;99:1712–1721.

22. Elmlinger MW, Kühnel W, Weber MM, Ranke MB. Reference ranges for two automated chemiluminescent assays for serum insulin-like growth factor I (IGF-I) and IGF-binding protein 3 (IGFBP-3). Clin Chem Lab Med 2004;42:654–664.

23. Svensson J, Johannsson G, Bengtsson BA. Insulin-like growth factor-I in growth hormone-deficient adults: relationship to population-based normal values, body composition and insulin tolerance test. Clin Endocrinol (Oxf) 1997;46:579–586.

24. Varewijck AJ, Lamberts SW, van der Lely AJ, Neggers SJ, Hofland LJ, Janssen JA. The introduction of the IDS-iSYS total IGF-1 assay may have far-reaching consequences for diagnosis and treatment of GH deficiency. J Clin Endocrinol Metab 2015;100:309–316.

25. Boguszewski CL. Individual sensitivity to growth hormone replacement in adults. Rev Endocr Metab Disord 2021;22:117–124.

26. Fleseriu M, Hashim IA, Karavitaki N, et al. Hormonal replacement in hypopituitarism in adults: an Endocrine Society Clinical Practice Guideline. J Clin Endocrinol Metab 2016;101:3888–3921.

27. Trotta M, Da Broi J, Salerno A, Testa RM, Marinari GM. Sleeve gastrectomy leads to easy management of hormone replacement therapy and good weight loss in patients treated for craniopharyngioma. Updates Surg 2017;69:95–99.

28. Garrez I, Lapauw B, Van Nieuwenhove Y. Bariatric surgery for treatment of hypothalamic obesity after craniopharyngioma therapy: a matched case-control study. Obes Surg 2020;30:2439–2444.

29. Lomenick JP, Buchowski MS, Shoemaker AH. A 52-week pilot study of the effects of exenatide on body weight in patients with hypothalamic obesity. Obesity (Silver Spring) 2016;24:1222–1225.

30. Bretault M, Carette C, Zaharia R, et al. Liraglutide 3mg as a weight-loss strategy after failed bariatric surgery in a patient with hypothalamic obesity following craniopharyngioma. Diabetes Metab 2020;46:514–515.

8

General Discussion

General discussion

The outcome of treatment of patients with tumors has improved over the past decades, which went hand in hand with a growing awareness for very long-term consequences regarding comorbidities and quality of life. This is especially true for children with cancer: they generally have an excellent short-term survival rates, and still have many years of living ahead thereafter. This awareness led to large cohorts of childhood cancer survivors being examined on a wide variety of long-term effects, such as the LATER study in the Netherlands.[1-3] A group for which appriate awareness is perhaps still lacking, although the attention is growing, are patients suffering from relatively benign neoplasms such as craniopharyngiomas, who often suffer from a compromised health.[4] Patients with craniopharyngioma have experienced similar therapeutic regimens as patients with a malignant disease, such as neurosurgery and radiotherapy. Also, this benign tumor lies very close to important neurological and vascular structures such as the hypothalamus, pituitary, carotid arteries and optic nerves, which can be damaged by the tumor and by treatment.[4] The hypothalamus and pituitary are important organs for the endocrine system and metabolism. Previous Dutch and Swedish studies already revealed, that craniopharyngioma patients had high rates of long-term sequalae like pituitary deficiencies (81-98%), visual deficiencies (75%), neurological deficits (34%) and obesity (56%).[5,6] The aim of this thesis was to further expand on identification of long-term endocrine, bone-related and metabolic alterations in those who suffer(ed) from a tumor, with a special focus on patients with benign tumors of the pituitary (craniopharyngioma and macro-adenoma). We aimed to determine risk factors as well as protective factors related to metabolic health and bone health, and evaluated the efficacy of current common methods for diagnosis and treatment of metabolic consequences.

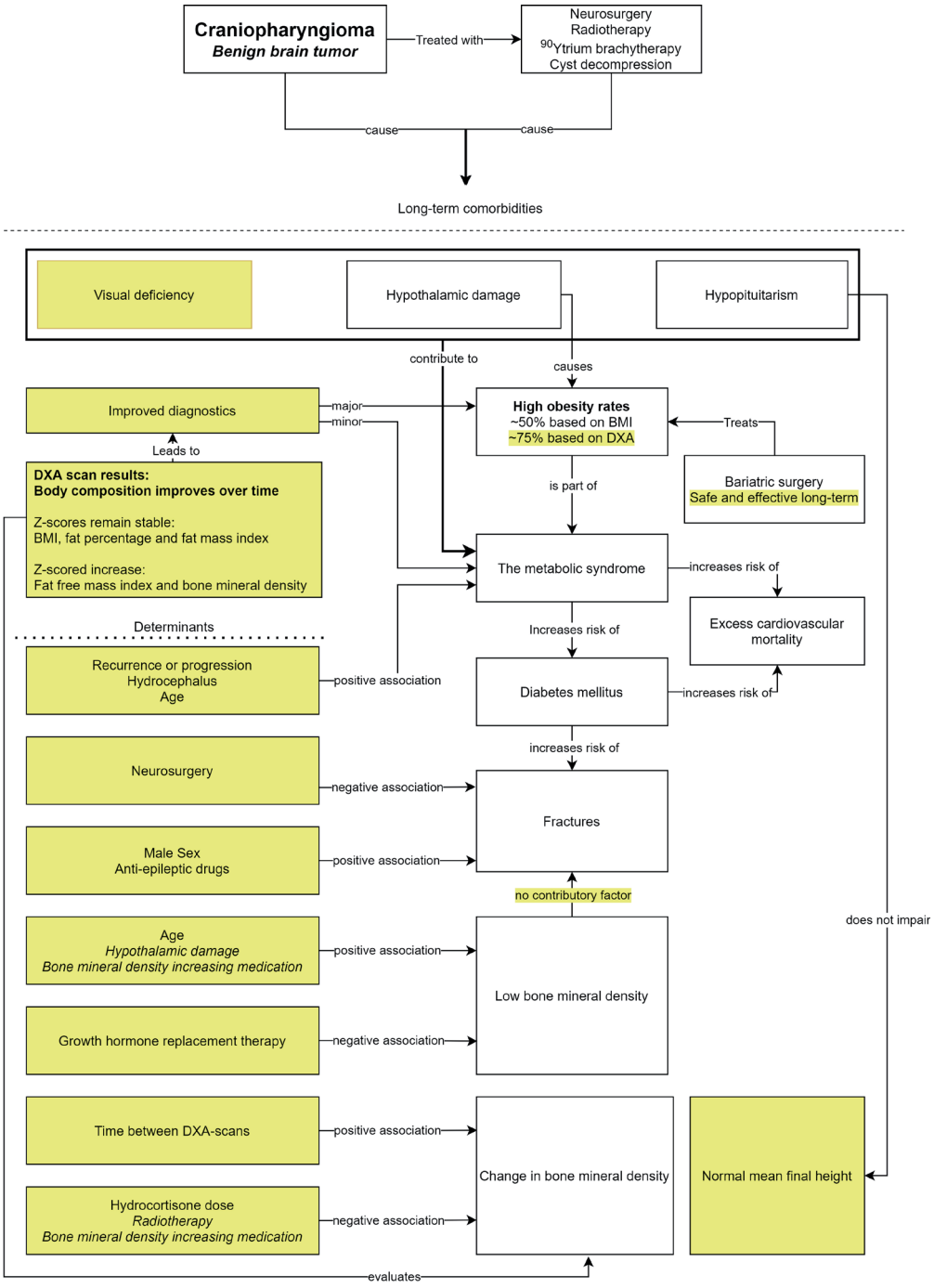
Tumors and bone

Non-malignant tumors can have a major effect on development and sustainment of bone. This is expecially the case after tumors in de sellar region that cause an increase or decrease in the secretion of pituitary hormones. First, we described the discovery of a novel *AIP* mutation, causing accelerated growth due to a growth hormone secreting adenoma with a complicated treatment in an exceptionally young boy. Thereafter, we described the impact on bone health in a large cohort of patients with craniopharyngioma and frequent occurrence of hypopituitarism, including fracture risk, bone mineral density (changes) and growth, and related risk- and protective factors (Figure 1).

Pituitary adenoma and growth

An example of a tumor in the sellar region with high secretion of hormones and obvious influence on bone, is a growth hormone secreting adenoma. **Chapter 2** describes the discovery of a novel truncating *AIP* mutation (p.Tyr202x) in a case of growth hormone secreting adenoma with an unexpected course of disease, and with an onset at an extraordinarily young age. The 7-year old boy presented with a strongly reduced intake of nutrients, which could have led to failure to thrive and decelerated growth. Instead, the tumor caused an excess growth hormone before closure of the epiphysis and led to an accelerated growth.[7] In all pituitary adenomas, only 5% of the cases relate to a genetic alteration; therefore not all cases need genetic screening.[8] However, the chances of identifying a genetic cause increase especially in case of familial predisposition, and in pediatric acromegaly.[8] In case of gigantism or familial predisposition in familial acromegaly in Familial Isolated Pituitary Adenomas (FIPA) kindreds, 50% of the patients have a causative mutation.[8,9] It is important to screen high-risk populations of mutations, as it allows earlier diagnosis in relatives and a better choice of therapy.[9,10] *AIP* mutated adenomas seem to have a suboptimal response to treatment with 1st generation somatostatin analogues, but a good response to 2nd generation somatostatin analogues.[11,12] Second generation somatostatin analogue pasireotide has the highest affinity for

Figure 1 Summary of the findings in craniopharyngioma patients of this thesis



Findings are marked in yellow if they are newly discovered by the projects presented in this thesis. Scentences in italic represent findings in univariable analysis or with unlikely causal relationship.

somatostatin receptor (SSTR) 5, followed by SSTR2, SSTR3 and SSTR1 thereafter.[13] Despite the highest affinity for SSTR5, pasireotide resistance is mostly related to SSTR2 expression instead of SSTR5 in the general acromegalic population.[14,15] In the patient of **Chapter 2**, tumor growth continued despite treatment with neurosurgery and second-generation somatostatin analogues. The pathology reported presence of SSTR2 expression, and loss of SSTR5 expression at the third operation – a possible decline starting from the first surgery could not be demonstrated due to lack of available tissue for examination. Furthermore, *in vitro* experiments showed a resistance for pasireotide. This suggests that it could be preferable to test the *in vitro* response of tumor tissue to growth hormone secretion instead of assessing somatostatin receptor expression to improve the predicted reponse *in vivo*.[15] This is because the functional test combines the presence of the receptors with their down-stream function. The aberrant course of disease in this patient underlines importance of individualized, tumor-specific treatment.

Craniopharyngioma and bone health

Growing towards an adequate height

While the patient in **Chapter 2** had an increased growth due to high growth hormone levels, on the other side of the spectrum a growth hormone deficiency can cause an impaired growth. Growth is an important aspect of bone health. Infant growth partially determines the acquired bone mass of the adult.[16] Size, or final height, also matters for metabolic properties of an individual.[17] Healthy taller persons have larger total resting energy expenditure, but a relatively lower mass-specific resting energy expenditure compared to smaller persons.[17-19] Lastly, height is of importance in the evaluation of bone mineral density: smaller bones with a comparable volumetric bone mineral density appear to have a lower areal bone mineral density, as a two-dimensional image does not incorporate the depth of bone.[20,21]

Impaired growth due to growth hormone deficiency in the context of hypopituitarism can occur when a benign tumor grows near the

pituitary, or when this tumor is treated with surgery or radiotherapy.[22,23] In **Chapter 3**, we performed one of the largest studies investigating bone health in patients with craniopharyngioma with a median follow-up of 16 years. In this retrospective study, growth appeared normal, as both childhood and adulthood onset patients achieved a mean final height despite high frequencies of pituitary deficiencies: a pituitary deficiency of at least one axis occurred in 92% of the cases, with growth hormone deficiency in 85% in this cohort. In the literature, certain studies[24] were consistent with finding a normal mean final height. However, one study still showed short stature in 31% of the cases,[25] and in a very large cohort of 393 craniopharyngioma patients, patients with childhood-onset disease were slightly (0.6 SDS) shorter.[26] The size of this cohort was even larger than our cohort, which enables to find a significant difference when the difference between two groups are relatively small.

Proper treatment with growth hormone therapy is likely the most important contributor to achieving a normal final height,[27] as growth acceleration is observed after initiating growth hormone therapy.[28,29] Still, patients with craniopharyngiomaq seem to have a better growth after growth hormone therapy than growth hormone deficient patients with other types of brain tumors, regardless of cranial radiotherapy.[30] Perhaps, the 'growth without growth hormone syndrome', plays a role in adding extra centimeters of achieved height on top of the results from growth hormone treatment. This 'growth without growth hormone syndrome' supposedly relates to hypothalamic damage, which occur at a high rate (~45%) in patients with craniopharyngioma.[31] In the International Registry of Hypothalamic Obesity Disorders (IRHOD), craniopharyngioma was even the most common type of brain tumor causing hypothalamic obesity.[32] Craniopharyngioma patients with grade 2 hypothalamic damage were taller than patients with grade 0 or 1 hypothalamic damage.[25] The precise mechanism of 'growth without growth hormone syndrome' is not yet elucidated, but it appears that hypothalamic damage and related obesity play a central role, as they lead to insulin resistance, initiating high insulin levels and a mitogenic effect,[33,34] and to elevated leptin levels which affects the bone growth center.[35-40] Besides hypothalamic damage, another theory suggests the

existence of yet undiscovered hormones and growth factors, other than growth hormone, which cause the unexpected pursued growth.[35-40] Growth without any growth hormone replacement indeed occurred in patients with craniopharyngioma.[33,34,36,41-44]

Bones breaking bad: fractures and bone density in patients with craniopharyngioma

Risk factors for fractures in the general population were low bone density, hypogonadism, poor physical function, chronic health problems, anticonvulsants, corticosteroid use and estrogen replacement therapy.[45] These risk factors are all potentially applicable in patients with craniopharyngioma.[6,31] **Chapter 3** describes the first study investigating determinants of fracture risk of patients with craniopharyngioma, and one of few studies investigating determinants of low bone mineral density.

The occurrence of bone fractures in 18% of the craniopharyngioma patients in **Chapter 3** appears much higher than the cumulative incidence of 8-10% in a general European population.[26] Fracture risk was especially high in men and patients who received medication for epilepsy, while previous surgery as craniopharyngioma therapy was a protective factor. The mean bone density values of the cohort were in the normal range, but 24% still had values in the osteoporotic range due to a wide spread. Factors associated with higher risk of osteoporotic bone mineral density values were age and bone mineral density increasing medication.

Despite the apparent high fracture rate in this study, patients with craniopharyngioma did not have more fractures than healthy controls in a previous comparison.[31] The craniopharyngioma patients were of mixed Dutch and Swedish descent, while the reference population was a general Swedish population.[31] However, an increased fracture risk was found in a cohort with solely Swedish patients compared to a Swedish reference group.[5] This difference may be explained by the choice of the reference cohort: people from Scandinavian countries

were reported to generally have a higher fracture risk than people from countries in Western Europe.[46] A reference cohort from the same geographic location would be preferable, but a suitable Dutch healthy reference cohort was unfortunately unavailable for the study of Wijnen et al.,[31] and for the study described in **Chapter 3**. The cause of high geographical heterogeneity is unknown, but may relate to lack of sun exposure and relating vitamin D deficiency risks,[47] differences in diet or physical activity, or registration bias. Vitamin D is formed in the skin under sun exposure (UV-B).[47] A higher fracture risk of hip fracture risk in the winter was observed in Sweden, especially at higher latitudes.[47] In this study, there was a significant interaction with seasonal effect and latitude for fracture risk, which suggested the vitamin D levels to relate to the fracture risk.[47] Vitamin D combined with calcium suppletion gives a small reduction in fracture risk in elderly.[48] This may not be a causal relationship, as vitamin D correlates inversely with body mass index (BMI) as well.[49] As many patients with craniopharyngioma are obese, they may be at high risk of vitamine D deficiency. To our knowledge, vitamin D deficiency was not evaluated in cohorts of patients with craniopharyngioma, nor is the effect of vitamine D suppletion evaluated in a randomized controlled trial in this population.

In a population with growth hormone deficient subjects due to causes other than craniopharyngiomas, the fracture rate was even higher, at 28-29%.[50] However, fracture rate was equal in craniopharyngioma patients and non-functioning pituitary adenoma patients who were both using growth hormone replacement therapy for two years.[51] The fracture rate described in **Chapter 3** is still likely an underestimation due to the retrospective design, which could partially explain the apparent lower fracture rate compared to other growth hormone deficient subjects.

Another factor that potentially protects patients with craniopharyngioma to fractures compared with other growth hormone deficient populations, is the high rate of obesity.[52] The risk of fractures in case of obesity in the general population is controversial: mostly, obesity is regarded as protective factor against fractures,[53] particularly hip and vertebral fractures in adults.[54] On the other hand, fracture risk

may be site and subpopulation dependent, as obesity was found as a risk factor[55,56] for humerus, finger and ankle fractures,[53,54,57] and in older men.[58] Obesity increases the risk of type II diabetes, which may increase the fracture risk despite a high bone mineral density, as it increases the risk of falling due to retinopathy and autonomic dysfunction.[59] In **Chapter 3**, BMI was not significantly protective against fractures in patients with craniopharyngioma, although a trend for a mild protective association was apparent in univariable analysis. Probably, the protective factors of high BMI are annuled by the risk factors. The protective factors may related to higher absorption of falling forces by soft tissue padding and increased bone density due to skeletal loading, but high BMI is accompanied with an increased risk of type II diabetes mellitus and increased forces of impact after a fall.[60]

People with a low bone density may have a high risk of fractures of nearly all types.[61-64] Osteoporotic fractures are most typically located at the hip, spine, distal radius and (proximal) humerus, but it can also occur at other sites such as the ankle, ribs or pelvis.[61] In **Chapter 3**, the fractures of craniopharyngioma patients were most often situated in the long pipebones or vertebrae (in 52%). Surprisingly, osteoporotic bone density values did not relate to an increased fracture risk in these patients when evaluated in a logistic regression mode. Possibly, the risk is overrulled by other determinants.

Also, no association was found between fractures and pituitary deficiencies, which is likely due to an appropriate hormonal replacement. This assumption is further substantiated by the described normal growth regardless of high rates of growth hormone deficiency in childhood onset disease, and high rates of pituitary replacement therapy. In a female general population, premenopausal women still had a lower risk of fractures than postmenopausal women using hormonal replacement therapy and calcium and vitamin D suppletion.[65] This suggests that in general, age still plays an important role besides hormonal status. Age did however not show as an important factor in our cohort, probably because other factors have a bigger impact.

Age could also explain higher fracture risk of men compared

to women in **Chapter 3**. The median age in the craniopharyngioma cohort was 45 years. In the general population, the lifetime risk of fractures is much higher in women than in men.[66] However, in the age category below 50 years, men have a higher fracture risk due to risk of trauma.[66-69] A higher fracture risk in men than women with craniopharyngioma can therefore be regarded as 'normal'. The difference between men and women of the craniopharyngioma may turn around when the cohort increases further in follow-up duration: in a general population in a postmenopausal age category, women have a higher fracture risk than men of a similar age.[67-69] Also, although not significantly different, men in our craniopharyngioma study had a 10% higher rate of corticotrophic deficiency. In the general population, oral corticosteroid use had a dose-dependent positive relationship with fractures.[70] However, in patients with hypopituitarism and loss of endogenous production of glucocorticoids, replacement therapy did not relate to an increased fracture risk.[71] This small potential difference in corticotropic deficiency therefore unlikely explains the different fracture rate between men and women with craniopharyngioma. A better explanation may be formed by a difference in behaviour potentially leading to traumatic injury from factors such as use of alcohol, work activity, recreational or sports activity, and even in making choices when crossing a road.[66,72,73] Unfortunately, we were unable to investigate behavioral factors in patients with craniopharyngioma within the current design of the study.

Despite a higher fracture risk in men, women in the cohort more often received medication aimed to improve bone density. One could argue, that this may have reduced the fracture risk. However, in the logistic regression model, bone mineral density improving medication was not a significant factor for determining the fracture risk. Although bone mineral increasing medication was prescribed, our study did not evaluate the compliance of the use. In the general population, women tended use more unique medications, but were less compliant than men when using medication chronically.[74]

In **Chapter 3**, craniopharyngioma patients using antiepileptic drugs had tripled odds of having a fracture compared to patients who did

not use any antiepileptic drugs. Generally, patients with epilepsy are at high risk of fractures. The fractures are caused by falls and seizures; the falls may or not may relate to the seizure, and the seizures may or not may relate to a fall.[75,76] Even strong muscular contractions during a seizure without a fall can infrequently cause fractures, which occurred more often in men.[75] Falling may be caused by seizures, coexisting neurological deficits, and by ataxia, clumsiness and poor balance due to antiepileptic drugs.[75] Furthermore, antiepileptic drugs can give an accelerated bone loss, low bone density and hyponatremia, and increase fracture risk.[77-79] In our study, the presence of epilepsy itself was not a significant factor in predicting fracture risk. Craniopharyngioma patients with epilepsy who do not use antiepileptic drugs may represent a subset of patients who suffer little from seizures and therefore not require therapy.

It is not entirely clear why in **Chapter 3** surgical craniopharyngioma treatment was observed as protective factor for fractures. Perhaps patients had a generally low performance due to severe comorbidities, which could form a contra-indication for major surgery and relate to a higher fracture risk as well. Another theory is that this is related to treatment with cyst aspiration; initial treatment with cyst aspiration was associated with lower recurrence- or progression-free survival in a previous study of the cohort.[6] This may lead to more damage to neurovascular structures. Also, previous treatment with radiotherapy tended to relate with lower fracture risk. In the past years, a shift occurred from extensive surgery (gross total resection) towards more partial resection, combined with radiotherapy, to avoid hypothalamic obesity.[6] Radiotherapy could be a related to a more limited surgery, resulting in less damage of neurovascular structures, and therefore diminishing risks of falling and diabetes mellitus type 2.

For low bone density, mineral density increasing medication was identified as 'risk factor', but that is unlikely to be causally related: probably the patients with low bone density were starting treatment for the purpose of increasing their bone density. Also, in univariable analysis, hypothalamic damage was confirmed as a risk factor for low bone density. This is in line with the findings of Holmer et al., who

found hypothalamic involvement of the tumor with consequently leptin resistance, and additionally, insufficient gonadal replacement therapy were previously found as risk factors for reduced bone mineral density.[80,81] In our analysis, insufficient gonadal replacement therapy was not confirmed as risk factor for low bone density; however, almost all patients received gonadal replacement therapy when applicable. Leptin resistence may explain why in our study BMI did not have a significant negative relationship with osteoporotic bone density values, while in the general population obese adults have a lower osteoporosis risk than health weight individuals.[82] Leptin correlates positively with BMI as it is produced by adipose tissue.[83-85] Leptin has a direct anabolic effect on bone, but the overall effect may be bimodal as the hypothalamus exerts a central effect as well.[86] This balance may be disrupted in patients with craniopharyngioma and hypothalamic damage.

In the general population, the fracture risk is assessed with the FRAX to estimate the 10-year probability of major osteoporotic fractures or hip fractures.[87] Suggestions have been made to improve the estimation by FRAX in men, by adding comorbidities, medication and behavioral factors. [87] Many of the classical risk factors for osteoporosis of which some are implemented in FRAX, such as smoking, high alcohol consumption, nutrition, inactivity and family history,[81,88] could not be evaluated in the craniopharyngioma cohort described in **Chapter 3**, as the study had a retrospective design and these factors were often poorly described in the medical files. Our research however shows that non-classical risk factors are involved. Application of the FRAX in craniopharyngioma patients could therefore not give a good estimation.

Future studies could be aimed at prospectively investigating fracture rates and influential factors for fractures including classical risk factors such as smoking and alcohol use, but also bone quality and leptin levels. The validity of fracture risk assessment by FRAX could be evaluated in patients with craniopharyngioma, and perhaps adjusted with novel found risk factors to evaluate the risk in patients of this population specifically. Studies would preferably be performed multiple, different cohorts of patients with craniopharyngioma to validate the results of

the current study. The effect of bone density increasing medication on fracture risk could be evaluated in these patients, especially in men.

Longitudinal changes in bone density of patients with craniopharyngioma

Chapter 4 is the first longitudinal study investigating bone density changes over time as evaluated with DXA-scan and determinants bone density changes in a large cohort of patients with craniopharyngioma. This retrospective study showed that 60% of the patients had a low bone mineral density at first evaluation, but fortunately a significant increase in bone mineral density Z-scores is shown after a median time of 10 years between the first and last DXA-scan. The improvement in bone density was stimulated by use of growth hormone therapy, while radiotherapy and medication to improve bone density related to a decrease in bone density.

Our findings were for the most part in line of expectation of previous studies in craniopharyngioma or patients with hypopituitarism. The high rates of low bone density were similar to a smaller study of Andereggen et al., who found pathological bone density in 53% of patients with craniopharyngioma.[89] In the Pfizer International Metabolic Database study, growth hormone deficient patients with craniopharyngioma or pituitary adenoma were observed, who were receiving growth hormone replacement therapy.[90] Here, patients with radiotherapy had a lower bone mineral density than patients without radiotherapy, despite a higher fat mass.[90] This probably relates to hypothalamic dysfunction after radiotherapy.[91,92]

Other studies studies indeed showed an increase in bone density if growth hormone replacement therapy was given in case of a growth hormone deficiency (not specifically in craniopharyngioma).[93-97] Aside from the importance of growth hormone for bone mass acquisition during growth, it plays an important role in bone mass maintenance due to bone remodeling.[98] Additionally, growth hormone has an anabolic effect on muscle mass,[99] which also improved as is discussed in the next

paragraph. These findings underline the importance of growth hormone replacement therapy. Although malignant neoplasms were a concern for the application of growth hormone therapy for a long time, this does not appear to be an important concern: the incidence of malignant neoplasms in case of hormone replacement therapy were low, and reported between 6-27 per 100,000 person-years.[28,100,101]

The negative association of medication to improve bone density and change in bone density appeared odd at first; however, bone density improving medication is more often given to patients with impaired bone health, and again this association is not likely a causal relationship.

As is further elaborated below, the general improvement of bone mass was accompanied by an improvement of muscle mass in the cohort. This improvement of bone and muscle mass may correlate: high-impact exercise, such as jumping, causes an increase in bone density.[102] Our study did not evaluate lifestyle aspects such as exercise. Patients with craniopharyngioma have a low aerobic capacity, potentially related to hypothalamic involvement.[103,104] This low aerobic capacity may be a barrier in the performance of physical activity, as well as many other factors including psychosocial difficulties, increased daytime sleepiness, growth hormone and gonadal deficiency, neurological and visual deficits, decreased motor profiency and working memory.[44,104] Other studies have indeed observed a decreased physical activity in children with craniopharyngioma compared to obese and normal weight controls.[105-107] Physical activity in patients with craniopharyngioma is often not a sustainable way to achieve a weight reduction, not even when combined with a diet and pharmacotherapy.[108]

It would be of interest to investigate whether physical activity does improve bone strength and reduce fracture risk, as is the case in the general population.[109] These and other potential factors such as neurological deficits, exercise and diet, and potential weight loss (i.e. after bariatric surgery) that may influence bone health could be investigated, and previous findings may be validated in another cohort.

Consecutive bone density measurements should then preferably be performed with the same DXA-scanner to avoid a potential systematic bias, as the DXA-scanner type may influence the measurements[110] (although agreement between DXA-scans is still high).[111]

Metabolic long-term effects

Fat and muscle: body composition changes in patients with craniopharyngioma

Chapter 4 is the first study evaluating body composition changes in patients with craniopharyngioma, by comparing DXA-scan results, which are the gold standard to evaluate body composition.[112-114] Previous studies investigating body composition in patients with craniopharyngioma were only small cohorts or did not use the DXA-scan,[26,115,116] and barely evaluated change over time.[51] Although most patients in **Chapter 4** had a high mean fat percentage and fat mass index, change in these adiposity indicators over a median time of ten years was comparable to an age- and sex-matched reference population, while their fat free mass improved over time. Still 55% of the craniopharyngioma population of **Chapter 4** is obese, based on their BMI at last follow-up.

The increase in muscle mass could be explained by the frequent application of growth hormone replacement therapy. Body composition is influenced by all pituitary hormones, but particularly growth hormone.[117-119] In another study with craniopharyngioma patients, growth hormone replacement therapy for a duration of two years did not induce a decrease in fat percentage, while patients with non-functioning pituitary adenoma lowered in fat percentage.[51] The increase in fat-free mass and lipids after growth hormone replacement was however visible in craniopharyngioma patients as well.[51]

Other observations show an increase in BMI (Z-scores) which occur mostly before or in the first years after treatment.[24,120] In **Chapter 4**, an increase in BMI is observed as well, but BMI Z-scores did not change,

nor did the fat percentage Z-score. The evaluations in our cohort are however presumably performed in long-term follow-up. The observed increase in fat-free mass could potentially contribute to ceasing a further increase of fat mass. In the general population, resting energy rate is dependent on fat-free mass, fat mass, sex and height;[17,18,121] the fat-free mass is perhaps the most important indicator, and relates to skeletal muscle mass.[122-126] Low resting energy expenditure is a major contributor to obesity in patients with craniopharyngioma.[107,116,127] Multiple studies showed this low resting energy expenditure occurred regardless their high fat-free mass.[44,106,107,115,116,127] The low resting energy expenditure appears most influenced by the degree of hypothalamic damage[4,128] which may give a dysregulation of the autonomous nervous system.[129-131] It is unclear whether an increase in muscle mass would result to an increase in resting energy expenditure in patients with craniopharyngioma, as the previous studies did not evaluate a change in time. Nor has it been investigated whether the improved fat-free mass relates to a change in habits of physical activity. Future studies could focus on these research questions, with sequential use of the same DXA-scanner preferably. This is especially important when evaluating body composition, as especially lean body mass measurements are dependent on the carbohydrate intake and (related) hydration status of the person.[110,132] The hydration status of craniopharyngioma patients may be altered due to their risk of hypopituitarism,[133-136] including growth hormone deficiency and diabetes insipidus. These deficiencies require replacement therapy that may not match the normal physiology and may differ per patient due to individualized therapeutic schedules.

Application of the DXA-scanner in diagnosis of obesity and the metabolic syndrome

Obesity and low relative skeletal muscle mass are important factors in metabolic dysregulation and the metabolic syndrome.[123,137-143] **Chapter 5** is the first study in patients with craniopharyngioma that shows a major improvement in the diagnostics of obesity by evaluating body composition with the DXA-scanner. The obesity rate was as high as 75% based on fat percentage. Apparently other metabolic

alterations resulting in metabolic syndrome components were very frequent, as the improved obesity diagnostics only led to a minor improvement in the metabolic syndrome evaluation.

In **Chapter 5**, childhood onset patients had a higher obesity rate based on body fat percentage than adulthood onset patients. This probably relates to previous findings of Wijnen et al.[6] and Gautier et al.,[144] who observed that childhood onset patients have higher rates of panhypopituitarism, diabetes insidipidus.[144] and are more frequently treated with radiotherapy[144] than adulthood onset patients. Diabetes insipidus and radiotherapy can relate to hypothalamic dysfunction.[91,92,145]

Risk factors for the metabolic syndrome in uni- and multivariable logistic regresson analysis in **Chapter 3** were visual impairment, and age at last DXA-scan, recurrence or progression of disease, and hydrocephalus. Wijnen et al. found visual impairment as risk factor for the metabolic syndrome as well in a larger cohort of which the current study is a subset.[52] Visual impairment relates to low levels of physical activity.[146] Recurrence of disease will require more medical interventions such as neurosurgery and radiotherapy, which may cause a further damage of neurological structures. Radiotherapy and hydrocephalus may relate to a decline in IQ, which occurred in survivors of brain tumors.[147] It may be harder for patients with a low IQ to adhere to lifestyle advices. Low IQ was also related to development of obesity in a general population.[148]

BMI misclassified obesity if compared to fat percentage in 15% of men and 9% of women in one study in Denmark, and 25% of men and 48% of women in another study in the general population of the United States.[149,150] Both studies handled a threshold of fat percentage of 25% for men and 30% for women to identify obesity; there may however be a difference in the studied population. The population from Denmark is probably more suitable to compare with Dutch patients, as it is a European population as well, with a similarly described obesity rate of 17% in a cohort in 2015.[151] In **Chapter 5**, craniopharyngioma patients were misclassified in 29%, which appears to be in between the study from Denmark and the United States. Perhaps the different percentage

is due to the geographic difference, due to the high prevalence of hypopituitarism in patients with craniopharyngioma or the different definition for obesity. Currently, the consensus for the threshold of fat percentage for diagnosing obesity is not entirely formed.[152] This is still one of the drawbacks of the application of the DXA-scan for evaluation of body composition. Furthermore, DXA-scans are relatively expensive and time-consuming.[153] Future studies could therefore compare evaluation of obesity by DXA-scan measured fat percentage with other modalities, for example by muscle to visceral fat ratios by DXA-scan,[142] bio-impedance, perhaps even the simple waist circumference, or biomarkers (as is elaborated on in the next paragraph). The cost-effectiveness of performing DXA-scans remains to be determined before general clinical use in CP survivors can be recommended for this application. It may be useful to measure body composition components before performing interventions for obesity or low muscle mass such as growth hormone replacement to evaluate the effect, and in cases who have a BMI in an uncertain range, and if the DXA-scan is performed for another indication (to evaluate bone health).

Application of biomarkers in the diagnosis of the metabolic syndrome in childhood cancer survivors

The DXA-scan is the gold standard when an altered body composition is suspected. An altered body composition occurs in patients with craniopharyngioma, as described in **Chapter 4** and **Chapter 5**, but **Chapter 6** shifts the focus to another group at risk of an altered body composition: the childhood cancer survivors. Body composition is especially altered in childhood cancer survivors with metabolic alterations due to pituitary deficiency after high doses of cranial radiotherapy, abdominal radiotherapy and corticosteroids,[117,154-160] which contributes to their high risk of cardiovascular death.[153,161] Frail obese childhood cancer survivors with inadequate muscle mass are subject to the doubled jeopardy of sarcopenic obesity.[162]

The obesity rate is underestimated in childhood cancer survivors when conventional measuring methods such as BMI and waist hip ratio are used instead of fat percentage, measured by DXA-scan.[112] Therefore, the metabolic syndrome risk may be underestimated as well, as obesity is one of the criteria for the metabolic syndrome. For childhood cancer survivors it can be argued as well, that although the DXA-scan could provide information on body composition and give a better estimate of obesity, a routine application remains costly and time-consuming.[153] As alternative, we investigated the role of biomarkers to identify the metabolic syndrome in survivors in **Chapter 6**. We performed the first systematic review and meta-analysis to find novel biomarkers to diagnose or predict the metabolic syndrome in childhood cancer survivors, or a young general population from which the results were applicable to survivors. Although studies in childhood cancer survivors were very rare,[163-167] still 180 studies performed in the general population reported information on potentially useful biomarkers. For the prediction of the metabolic syndrome, identified valuable biomarkers were uric acid, adiponectin, hsCRP, leptin and apoB. For the diagnosis of the metabolic syndrome, uric acid, leptin, adiponectin and hsCRP were considered valuable. This study is a very important first step towards early identification of the metabolic syndrome in childhood cancer survivors. In the next step, optimal thresholds should be determined for most biomarkers, and the biomarkers should be tested specifically in this population and included in a prediction model. Therafter, the model can be validated in another survivor cohort, and the effect of early treatment of the metabolic syndrome on long-term vascular complications and mortality can be determined. It may offer a major improvement in long-term survival of childhood cancer survivors, as patients with metabolic syndrome have twice the risk of dying from cardio- and cerebrovascular disease compared to patients without the metabolic syndrome.[153,168] Application of these biomarkers for determination of the metabolic syndrome could be of interest as well for survivors of pediatric benign tumors, such as craniopharyngioma.

Metabolic surgery in patients with craniopharyngioma and severe obesity

The medical society is still in search of treatment options to lower risk of cardiovascular mortality in patients with craniopharyngioma, by lessening risk factors such as obesity.[4,31,169] Treatment of obesity is an very challenging in patients with craniopharyngioma and related hypothalamic damage. There is only limited evidence for the efficacy of pharmaceutical therapies.[170] Generally, severe obesity can be treated with metabolic surgery, also called bariatric surgery.[170-174] These types of operations are performed on the intestine and stomach and cause an decreased volume of the stomach, increased satiety, mild malabsorption of nutrients and consequently significant long-term weight loss in the general obese population. The changes in appetite are caused by hormonal changes, such as gut hormone glucagon-like peptide-1 (GLP-1), which acts on the hypothalamus.[170,171,175] However, the hypothalamus and pituitary are very frequently dysfunctioning in patients with craniopharyngioma, making the efficacy of bariatric surgery questionable.

In **Chapter 7** we present a study with the largest population of 16 craniopharyngioma patients who underwent bariatric surgery, with the longest follow-up to date. The study showed a mean loss of 22% after 5 years in craniopharyngioma patients, which is considerable but was less than 155 matched controls if patients received a Roux-en-Y gastric bypass.

In previous studies, including one with the Dutch and Swedish patients of our cohort and two years follow-up, the result was the opposite, with equal weight loss after Roux-en-Y gastric bypass and less weight loss after sleeve gastrectomy.[169] The current study contains more patients and has a longer follow-up. This study is therefore a better representation of the craniopharyngioma population and suggests that craniopharyngioma patients gain more weight long-term than a general population after bariatric surgery. The weight loss between patients with craniopharyngioma was comparable for Roux-en-Y gastric bypass and sleeve gastrectomy (23% versus 22%) in

Chapter 7, while in another study in a general obese population Roux-en-Y gastric bypass causes a higher weight loss than sleeve gastrectomy (26% versus 19%).[174] The weight loss therefore appears independent of the type of bariatric surgery in craniopharyngioma patients. Sleeve gastrectomy and Roux-en-Y gastric bypass both give a restriction in food intake with favorable hormonal alterations, but the Roux-en-Y gastric bypass has also a component of mild malabsorption.[172] It is possible that this malabsorption component does not add much in patients with craniopharyngioma because the hypothalamic damage dysfunction prevents an even larger effect. Still, the observation of 22% weight loss after five years suggests that hormonal alterations still exert an effect, probably in different brain centers. GLP-1 receptors are for example also located in the medulla and parietal cortex.[176] GLP-1 analogue liraglutide resulted in a decreased response to desirable food with high content of fat or sugar in the parietal cortex on functional MRI in a diabetic population.[176] It would be of interest to evaluate GLP-1 changes in patients with craniopharyngioma after bariatric surgery, and their effect in the brain.

Safety remains an issue in surgical procedures. The fear of complications is even larger when the surgery induces malabsorptions in patients with hypopituitarism, who depend on their replacement therapy with their lives. Short-term, 13% of craniopharyngioma patients with any type of bariatric surgery procedure had serious adverse events in **Chapter 7**. This appears higher than in the general population, who have a 30-day rate of major adverse events of 5% after Roux-en-Y gastric bypass, and of 3% after sleeve gastrectomy.[174] Perhaps the sleeve gastrectomy should be of small preference, as the safety profile in general is better and our patients had as much weight loss as after Roux-en-Y gastric bypass. The apparent good efficacy of sleeve gastrectomy must however be regarded with caution, as this studied subgroup was much smaller than the subgroup who had a Roux-en-Y gastric bypass.

Regarding long-term safety, one patient had severe absorptive issues and another died of unknown cause. The mean doses of pituitary replacement therapy did not change. All necessary changes in pituitary replacement therapy were within expectation of long-term care in

patients with changing age and weight. Other studies have indeed found no large negative effects on hormone replacement therapy.[169,172,177,178] One study even performed an oral thyroid/hydrocortisone/paracetamol absorption test after Roux-en-Y gastric bypass in one patient, which showed sufficient absorption of each drug.[178] Therefore, bariatric surgery was considered relatively safe if evaluated long-term in these patients. Future studies can focus on achieving even larger cohorts with a longer follow-up duration, and evaluate the differences between responders and non-responders of bariatric surgery, including evaluation of gastric hormones and the response of the brain by functional MRI.

Future perspectives for patients with craniopharyngioma

Targeted therapy before neurosurgery to protect neurovascular structures

This thesis underlines the importance of avoiding further damage of the hypothalamus and other brain structures due to treatment. A way of avoiding surgical damage, is to apply novel targeted molecular therapy to reach tumor shrinkage in advance of neurosurgery. In oncology, neo-adjuvant and targeted therapies are already often applied.[179] As explained in **Chapter 1**, there are two different histopathological subtypes of craniopharyngioma: the papillary and adamantinomatous subtype.[4] It is still unclear whether histological subtype and/or biological characteristics such as age of onset have any influence on survival rates. It seems that papillary craniopharyngioma patients have a better outcome than adamantinomatous craniopharyngioma patients regarding tumor size and recurrence rate, but this remains controversial.[180,181] The histopathological background may also be important for application of targeted therapy. For papillary craniopharyngioma, there is limited experience with *BRAF* inhibitors such as dabrafenib and bemurafenib, often combined with MEK inhibitors such as trametinib, in tumors carrying *BRAF*^{V600E} mutations.[182-189] The choice for dual therapy with *BRAF* and MEK

inhibitors, is made due to better survival when applied for melanoma compared to monotherapy.[187,190] Dual BRAF/MEK inhibitory therapy is proposed as neoadjuvant therapy or treatment option in complicated cases of papillary craniopharyngioma, but was only applied in a few cases so far.[182] Randomized controlled trials for evaluating the value of neoadjuvant application are still lacking. Also, papillary craniopharyngioma only account for 1 out of 10 craniopharyngiomas, which means this targeted therapy is only an option in 10% of the cases at best.[4,191] Targeted molecular therapies for adamantinomatous craniopharyngioma are being developed and relate to IL-6, PD1/PD-L1, MEK, IDO-1, but need further research as well before they can be applied in daily clinical practice.[192] New treatment modalities such as monoclonal antibodies may co-occur with currently unknown late effects.[193]

GLP-1 analogues for hypothalamic obesity

Still, the development of targeted therapies will be too late for many patients that have already been diagnosed. They may already suffer from hypothalamic obesity. Although **Chapter 7** shows promising results of bariatric surgery for hypothalamic obesity, not all obese patients will opt for this: patients may not comply with the indications for surgery, and importantly every surgery comes with a risk of complications, while not all patients had beneficient results in our research. Application of GLP-1 analogues may be a viable pharmaceutical alternative in these patients as primary treatment or after ineffective bariatric surgery. A study with one year open-label exenatide showed no significant weight loss, but still stabilisation or decreasing weight in 75% of a small group of patients with hypothalamic obesity.[194] Ultimately, even more effective GLP-1 analogues with more advantageous administration such as semaglutide[195] could be tested in patients with hypothalamic damage in a randomized controlled trial. It could as well be investigated whether GLP-1 analogue response predicts the efficacy of bariatric surgery in these patients. Since craniopharyngioma is a rare disease, international cooperation is essential in finding solutions for the health problems of the patients.

COVID-19

Current affairs bring new challenges: in 2020, the world news is dominated by a pandemic of coronavirus disease 2019 (COVID-19). Long-term health care had to make way for the acute COVID-19 issues; for example, osteoporosis care in the Netherlands temporarily suspended.[196] Health care professionals involved in endocrinological care were unavailable for active medical practice related to the COVID-19, due to illness or due to work related to COVID-19.[197] Patients with craniopharyngioma may have a more frequent occurrence of serious COVID-19 disease, as people with severe obesity more easily evolve towards respiratory failure.[197] It is still uncertain to what extend the COVID-19 pandemic has an influence on survival rates in these high risk patients. Regarding childhood cancer survivors, there did not seem to be a large effect so far on survival.[198] Still, the social isolation could potentially aggravate mental issues in survivors.[199,200] Only time will tell what the long-term effects of the pandemic are on the lives of these vulnerable people.

Conclusion

Mortality from cancer has declined in the past decades,[201] with the largest victory in childhood onset cancer, of which the 5-year survival rate is now 80%.[202,203] Still, the previous disease and treatment (especially radiotherapy) remain a continuous threat due to late effects and shorter lifespans.[204] Radiotherapy is also applied in patients with craniopharyngioma, who are generally regarded as subject to a benign tumor.[4] Despite the benign aspect of this rare tumor, patients are still at risk of a wide variety of long-term health issues and increased mortality.[31] The health issues presented in this dissertation relate to impaired endocrine, metabolic and bone-related health. Craniopharyngioma should therefore be regarded as a chronic disease, which requires life-long follow-up from a multidisciplinary team in expert centers. The performance of research in this rare disease requires international collaboration, for which we have found a great partner

in the Sahlgrenska University Hospital in Gothenborg, Sweden, and extended this collaboration in the past years with Erlangen University Hospital in Erlangen, Germany, the Medical University of Vienna in Vienna, Austria, and the Federal University of Parana in Curitiba, Brazil. We are very thankful for this co-operation and hope the future will bring an even more extensive collaboration. Ultimately, research should have the purpose of serving the patient a tailored therapy, to achieve an optimal life span and quality of life.

References

1. Kok JL, Teepen JC, van der Pal HJ, et al. Incidence of and Risk Factors for Histologically Confirmed Solid Benign Tumors Among Long-term Survivors of Childhood Cancer. JAMA Oncol 2019; 5(5): 671-80.
2. Streefkerk N, Heins MJ, Teepen JC, et al. The involvement of primary care physicians in care for childhood cancer survivors. Pediatr Blood Cancer 2019; 66(8): e27774.
3. Howell CR, Bjornard KL, Ness KK, et al. Cohort Profile: The St. Jude Lifetime Cohort Study (SJLIFE) for paediatric cancer survivors. Int J Epidemiol 2021; 50(1): 39-49.
4. Muller HL, Merchant TE, Warmuth-Metz M, Martinez-Barbera JP, Puget S. Craniopharyngioma. Nat Rev Dis Primers 2019; 5(1): 75.
5. Olsson DS, Andersson E, Bryngelsson IL, Nilsson AG, Johannsson G. Excess mortality and morbidity in patients with craniopharyngioma, especially in patients with childhood onset: a population-based study in Sweden. J Clin Endocrinol Metab 2015; 100(2): 467-74.
6. Wijnen M, van den Heuvel-Eibrink MM, Janssen J, et al. Very long-term sequelae of craniopharyngioma. Eur J Endocrinol 2017; 176(6): 755-67.
7. Vilar L, Vilar CF, Lyra R, Lyra R, Naves LA. Acromegaly: clinical features at diagnosis. Pituitary 2017; 20(1): 22-32.
8. Rostomyan L, Beckers A. Screening for genetic causes of growth hormone hypersecretion. Growth Horm IGF Res 2016; 30-31: 52-7.
9. Gadelha MR, Kasuki L, Korbonits M. The genetic background of acromegaly. Pituitary 2017; 20(1): 10-21.
10. Dénes J, Korbonits M. The clinical aspects of pituitary tumour genetics. Endocrine 2021; 71(3): 663-74.
11. Bogner EM, Daly AF, Gulde S, et al. miR-34a is upregulated in AIP-mutated somatotropinomas and promotes octreotide resistance. Int J Cancer 2020; 147(12): 3523-38.
12. Korbonits M, Kumar AV. AIP Familial Isolated Pituitary Adenomas. In: Adam MP, Ardinger HH, Pagon RA, et al., eds. GeneReviews(®). Seattle (WA): University of Washington, Seattle Copyright © 1993-2020, University of Washington, Seattle. GeneReviews is a registered trademark of the University of Washington, Seattle. All rights reserved.; 1993.
13. Bruns C, Lewis I, Briner U, Meno-Tetang G, Weckbecker G. SOM230: a novel somatostatin peptidomimetic with broad somatotropin release inhibiting factor (SRIF) receptor binding and a unique antisecretory profile. Eur J Endocrinol 2002; 146(5): 707-16.

14. Muhammad A, Coopmans EC, Gatto F, et al. Pasireotide Responsiveness in Acromegaly Is Mainly Driven by Somatostatin Receptor Subtype 2 Expression. J Clin Endocrinol Metab 2019; 104(3): 915-24.

15. Hofland LJ, van der Hoek J, van Koetsveld PM, et al. The novel somatostatin analog SOM230 is a potent inhibitor of hormone release by growth hormone- and prolactin-secreting pituitary adenomas in vitro. J Clin Endocrinol Metab 2004; 89(4): 1577-85.

16. Oliver H, Jameson KA, Sayer AA, Cooper C, Dennison EM. Growth in early life predicts bone strength in late adulthood: the Hertfordshire Cohort Study. Bone 2007; 41(3): 400-5.

17. Heymsfield SB, Childers D, Beetsch J, Allison DB, Pietrobelli A. Body size and human energy requirements: reduced mass-specific resting energy expenditure in tall adults. J Appl Physiol (1985) 2007; 103(5): 1543-50.

18. Heymsfield SB, Pietrobelli A. Body size and human energy requirements: Reduced mass-specific total energy expenditure in tall adults. Am J Hum Biol 2010; 22(3): 301-9.

19. Nishimoto Y, Ida S, Etani Y, Miyatani S. Resting energy expenditure in short-stature children. Endocr J 2012; 59(3): 265-71.

20. Zemel BS, Leonard MB, Kelly A, et al. Height adjustment in assessing dual energy x-ray absorptiometry measurements of bone mass and density in children. J Clin Endocrinol Metab 2010; 95(3): 1265-73.

21. Leonard MB, Zemel BS. Current concepts in pediatric bone disease. Pediatr Clin North Am 2002; 49(1): 143-73.

22. Doknić M, Pekić S, Miljić D, et al. Etiology of Hypopituitarism in Adult Patients: The Experience of a Single Center Database in the Serbian Population. International journal of endocrinology 2017; 2017: 6969286.

23. Higham CE, Johannsson G, Shalet SM. Hypopituitarism. Lancet 2016; 388(10058): 2403-15.

24. Tosta-Hernandez PDC, Siviero-Miachon AA, da Silva NS, Cappellano A, Pinheiro MM, Spinola-Castro AM. Childhood Craniopharyngioma: A 22-Year Challenging Follow-Up in a Single Center. Horm Metab Res 2018 ;50(9):675-682.

25. Huang CC, Lin KL, Wu CT, et al. Clinical and endocrinological manifestations of childhood-onset craniopharyngioma before surgical removal: A report from one medical center in Taiwan. Pediatr Neonatol 2021; 62(2): 181-6.

26. Kendall-Taylor P, Jonsson PJ, Abs R, et al. The clinical, metabolic and endocrine features and the quality of life in adults with childhood-onset craniopharyngioma compared with adult-onset craniopharyngioma. Eur J Endocrinol 2005; 152(4): 557-67.

27. Tanner JM, Whitehouse RH, Hughes PC, Carter BS. Relative importance of growth hormone and sex steroids for the growth at puberty of trunk length, limb length, and muscle width in growth hormone-deficient children. J Pediatr 1976; 89(6): 1000-8.

28. Rhie YJ, Yoo JH, Choi JH, et al. Long-term safety and effectiveness of growth hormone therapy in Korean children with growth disorders: 5-year results of LG Growth Study. PLoS One 2019; 14(5): e0216927.

29. Boekhoff S, Bogusz A, Sterkenburg AS, Eveslage M, Müller HL. Long-term Effects of Growth Hormone Replacement Therapy in Childhood-onset Craniopharyngioma: Results of the German Craniopharyngioma Registry (HIT-Endo). Eur J Endocrinol 2018; 179(5): 331-41.

30. Rose SR, Carlsson M, Grimberg A, et al. Response to GH Treatment After Radiation Therapy Depends on Location of Irradiation. J Clin Endocrinol Metab 2020; 105(10): e3730-41.

31. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. Excess morbidity and mortality in patients with craniopharyngioma: a hospital-based retrospective cohort study. Eur J Endocrinol 2018; 178(1): 95-104.

32. Rose SR, Horne VE, Bingham N, Jenkins T, Black J, Inge T. Hypothalamic Obesity: 4 Years of the International Registry of Hypothalamic Obesity Disorders. Obesity (Silver Spring) 2018; 26(11): 1727-32.

33. Geffner ME. The growth without growth hormone syndrome. Endocrinol Metab Clin North Am 1996; 25(3): 649-63.

34. Makras P, Papadogias D, Kaltsas G, Kaklas N, Piaditis G. Growth without growth hormone (GH): a case report. Hormones (Athens) 2004; 3(4): 259-65.

35. Phillip M, Moran O, Lazar L. Growth without growth hormone. J Pediatr Endocrinol Metab 2002; 15 Suppl 5: 1267-72.

36. Lazar L, Dan S, Phillip M. Growth without growth hormone: growth pattern and final height of five patients with idiopathic combined pituitary hormone deficiency. Clin Endocrinol (Oxf) 2003; 59(1): 82-8.

37. Geffner ME, Lippe BM, Bersch N, et al. Growth without growth hormone: evidence for a potent circulating human growth factor. Lancet 1986; 1(8477): 343-7.

38. Bistritzer T, Chalew SA, Lovchik JC, Kowarski AA. Growth without growth hormone: the "invisible" GH syndrome. Lancet 1988; 1(8581): 321-3.

39. Shtaif B, Dror N, Bar-Maisels M, Phillip M, Gat-Yablonski G. Growth without growth hormone: can growth and differentiation factor 5 be the mediator? Growth Factors 2015; 33(4): 309-18.

40. Venken K, Schuit F, Van Lommel L, et al. Growth without growth hormone receptor: estradiol is a major growth hormone-independent regulator of hepatic IGF-I synthesis. J Bone Miner Res 2005; 20(12): 2138-49.

41. O'Gorman CS, Simoneau-Roy J, Pencharz P, et al. Sleep-disordered breathing is increased in obese adolescents with craniopharyngioma compared with obese controls. J Clin Endocrinol Metab 2010; 95(5): 2211-8.

42. Simoneau-Roy J, O'Gorman C, Pencharz P, Adeli K, Daneman D, Hamilton J. Insulin sensitivity and secretion in children and adolescents with hypothalamic obesity following treatment for craniopharyngioma. Clin Endocrinol (Oxf) 2010; 72(3): 364-70.

43. Srinivasan S, Ogle GD, Garnett SP, Briody JN, Lee JW, Cowell CT. Features of the metabolic syndrome after childhood craniopharyngioma. J Clin Endocrinol Metab 2004; 89(1): 81-6.

44. Cohen M, Guger S, Hamilton J. Long term sequelae of pediatric craniopharyngioma - literature review and 20 years of experience. Front Endocrinol (Lausanne) 2011; 2: 81.

45. Ross PD. Risk factors for osteoporotic fracture. Endocrinol Metab Clin North Am 1998; 27(2): 289-301.

46. Kanis JA, McCloskey EV, Johansson H, et al. European guidance for the diagnosis and management of osteoporosis in postmenopausal women. Osteoporosis international: a journal established as result of cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA 2013; 24(1): 23-57.

47. Odén A, Kanis JA, McCloskey EV, Johansson H. The effect of latitude on the risk and seasonal variation in hip fracture in Sweden. J Bone Miner Res 2014; 29(10): 2217-23.

48. Rosen CJ. Clinical practice. Vitamin D insufficiency. N Engl J Med 2011; 364(3): 248-54.

49. Abdullah Thani NSI, Khairudin R, Ho JJ, Muhamad NA, Ismail H. Vitamin D supplementation for overweight or obese adults. The Cochrane Database of Systematic Reviews 2019; 2019(5): CD011629.

50. Wuster C, Abs R, Bengtsson BA, et al. The influence of growth hormone deficiency, growth hormone replacement therapy, and other aspects of hypopituitarism on fracture rate and bone mineral density. J Bone Miner Res 2001; 16(2): 398-405.

51. Verhelst J, Kendall-Taylor P, Erfurth EM, et al. Baseline characteristics and response to 2 years of growth hormone (GH) replacement of hypopituitary patients with GH deficiency due to adult-onset craniopharyngioma in comparison with patients with nonfunctioning pituitary adenoma: data from KIMS (Pfizer International Metabolic Database). J Clin Endocrinol Metab 2005; 90(8): 4636-43.

52. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. The metabolic syndrome and its components in 178 patients treated for craniopharyngioma after 16 years of follow-up. Eur J Endocrinol 2017; 178(1): 11-22.

53. Court-Brown CM, Duckworth AD, Ralston S, McQueen MM. The relationship between obesity and fractures. Injury 2019; 50(8): 1423-8.

54. Dimitri P, Bishop N, Walsh JS, Eastell R. Obesity is a risk factor for fracture in children but is protective against fracture in adults: a paradox. Bone 2012; 50(2): 457-66.

55. Adami G, Gatti D, Rossini M, et al. Risk of fragility fractures in obesity and diabetes: a retrospective analysis on a nation-wide cohort. Osteoporos Int 2020; 31(11): 2113-22.

56. Johansson H, Kanis JA, Odén A, et al. A meta-analysis of the association of fracture risk and body mass index in women. J Bone Miner Res 2014; 29(1): 223-33.

57. Prieto-Alhambra D, Premaor MO, Fina Avilés F, et al. The association between fracture and obesity is site-dependent: a population-based study in postmenopausal women. J Bone Miner Res 2012; 27(2): 294-300.

58. Nielson CM, Marshall LM, Adams AL, et al. BMI and fracture risk in older men: the osteoporotic fractures in men study (MrOS). J Bone Miner Res 2011; 26(3): 496-502.

59. Moseley KF. Type 2 diabetes and bone fractures. Current opinion in endocrinology, diabetes, and obesity 2012; 19(2): 128-35.

60. Ishii S, Cauley JA, Greendale GA, et al. Pleiotropic effects of obesity on fracture risk: the Study of Women's Health Across the Nation. Journal of bone and mineral research: the official journal of the American Society for Bone and Mineral Research 2014; 29(12): 2561-70.

61. Yoo J-H, Moon S-H, Ha Y-C, et al. Osteoporotic Fracture: 2015 Position Statement of the Korean Society for Bone and Mineral Research. Journal of bone metabolism 2015; 22(4): 175-81.

62. Nguyen TV, Eisman JA, Kelly PJ, Sambrook PN. Risk factors for osteoporotic fractures in elderly men. Am J Epidemiol 1996; 144(3): 255-63.

63. Johnell O, Kanis J. Epidemiology of osteoporotic fractures. Osteoporos Int 2005; 16 Suppl 2: S3-7.

64. Johnell O, Kanis JA. An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. Osteoporos Int 2006; 17(12): 1726-33.

65. Sullivan SD, Lehman A, Nathan NK, Thomson CA, Howard BV. Age of menopause and fracture risk in postmenopausal women randomized to calcium + vitamin D, hormone therapy, or the combination: results from the Women's Health Initiative Clinical Trials. Menopause (New York, NY) 2017; 24(4): 371-8.

66. Alswat KA. Gender Disparities in Osteoporosis. Journal of clinical medicine research

2017; 9(5): 382-7.

67. Lobo E, Marcos G, Santabarbara J, et al. Gender differences in the incidence of and risk factors for hip fracture: A 16-year longitudinal study in a southern European population. Maturitas 2017; 97: 38-43.

68. Duyvendak M, Naunton M, Atthobari J, van den Berg PB, Brouwers JR. Corticosteroid-induced osteoporosis prevention: longitudinal practice patterns in The Netherlands 2001-2005. Osteoporos Int 2007; 18(10): 1429-33.

69. Lagro-Janssen T, Grosicar J. Morbidity figures from general practice: sex differences in traumatology. J Eval Clin Pract 2010; 16(4): 673-7.

70. Van Staa TP, Leufkens HG, Abenhaim L, Zhang B, Cooper C. Use of oral corticosteroids and risk of fractures. J Bone Miner Res 2000; 15(6): 993-1000.

71. Wüster C, Abs R, Bengtsson BA, et al. The influence of growth hormone deficiency, growth hormone replacement therapy, and other aspects of hypopituitarism on fracture rate and bone mineral density. J Bone Miner Res 2001; 16(2): 398-405.

72. Johnell O, Kanis J, Gullberg G. Mortality, morbidity, and assessment of fracture risk in male osteoporosis. Calcif Tissue Int 2001; 69(4): 182-4.

73. Tom A, Granié MA. Gender differences in pedestrian rule compliance and visual search at signalized and unsignalized crossroads. Accid Anal Prev 2011; 43(5): 1794-801.

74. Manteuffel M, Williams S, Chen W, Verbrugge RR, Pittman DG, Steinkellner A. Influence of patient sex and gender on medication use, adherence, and prescribing alignment with guidelines. J Womens Health (Larchmt) 2014; 23(2): 112-9.

75. Mattson RH, Gidal BE. Fractures, epilepsy, and antiepileptic drugs. Epilepsy Behav 2004; 5 Suppl 2: S36-40.

76. Vestergaard P, Tigaran S, Rejnmark L, Tigaran C, Dam M, Mosekilde L. Fracture risk is increased in epilepsy. Acta Neurol Scand 1999; 99(5): 269-75.

77. Koppel BS, Harden CL, Nikolov BG, Labar DR. An analysis of lifetime fractures in women with epilepsy. Acta Neurol Scand 2005; 111(4): 225-8.

78. Upala S, Sanguankeo A. Association Between Hyponatremia, Osteoporosis, and Fracture: A Systematic Review and Meta-analysis. J Clin Endocrinol Metab 2016; 101(4): 1880-6.

79. Fraser LA, Burneo JG, Fraser JA. Enzyme-inducing antiepileptic drugs and fractures in people with epilepsy: A systematic review. Epilepsy Res 2015; 116: 59-66.

80. Holmer H, Popovic V, Ekman B, Follin C, Siversson AB, Erfurth EM. Hypothalamic involvement and insufficient sex steroid supplementation are associated with low bone mineral density in women with childhood onset craniopharyngioma. Eur J Endocrinol 2011; 165(1): 25-31.

81. Erfurth EM, Holmer H, Fjalldal SB. Mortality and morbidity in adult craniopharyngioma. Pituitary 2013; 16(1): 46-55.

82. Qiao D, Li Y, Liu X, et al. Association of obesity with bone mineral density and osteoporosis in adults: a systematic review and meta-analysis. Public Health 2020; 180: 22-8.

83. Pandit R, Beerens S, Adan RAH. Role of leptin in energy expenditure: the hypothalamic perspective. Am J Physiol Regul Integr Comp Physiol 2017; 312(6): R938-r47.

84. Enriori PJ, Evans AE, Sinnayah P, Cowley MA. Leptin resistance and obesity. Obesity (Silver Spring) 2006; 14 Suppl 5: 254s-8s.

85. Izquierdo AG, Crujeiras AB, Casanueva FF, Carreira MC. Leptin, Obesity, and Leptin Resistance: Where Are We 25 Years Later? Nutrients 2019; 11(11).

86. Cirmanová V, Bayer M, Stárka L, Zajícková K. The effect of leptin on bone: an evolving concept of action. Physiol Res 2008; 57 Suppl 1: S143-51.

87. Byberg L, Gedeborg R, Cars T, et al. Prediction of fracture risk in men: a cohort study. Journal of bone and mineral research: the official journal of the American Society for Bone and Mineral Research 2012; 27(4): 797-807.

88. Wark JD. Osteoporotic fractures: background and prevention strategies. Maturitas 1996; 23(2): 193-207.

89. Andereggen L, Hess B, Andres R, et al. A ten-year follow-up study of treatment outcome of craniopharyngiomas. Swiss Med Wkly 2018; 148: w14521.

90. Maiter D, Abs R, Johannsson G, et al. Baseline characteristics and response to GH replacement of hypopituitary patients previously irradiated for pituitary adenoma or craniopharyngioma: data from the Pfizer International Metabolic Database. Eur J Endocrinol 2006; 155(2): 253-60.

91. Constine LS, Woolf PD, Cann D, et al. Hypothalamic-Pituitary Dysfunction after Radiation for Brain Tumors. New England Journal of Medicine 1993; 328(2): 87-94.

92. Lustig RH, Post SR, Srivannaboon K, et al. Risk factors for the development of obesity in children surviving brain tumors. J Clin Endocrinol Metab 2003; 88(2): 611-6.

93. Snyder PJ, Biller BM, Zagar A, et al. Effect of growth hormone replacement on BMD in adult-onset growth hormone deficiency. J Bone Miner Res 2007; 22(5): 762-70.

94. Attie KM. The importance of growth hormone replacement therapy for bone mass in young adults with growth hormone deficiency. J Pediatr Endocrinol Metab 2000; 13 Suppl 2: 1011-21.

95. Tritos NA. Focus on growth hormone deficiency and bone in adults. Best Pract Res Clin Endocrinol Metab 2017; 31(1): 49-57.

96. Clanget C, Seck T, Hinke V, Wüster C, Ziegler R, Pfeilschifter J. Effects of 6 years of growth hormone (GH) treatment on bone mineral density in GH-deficient adults. Clin Endocrinol (Oxf) 2001; 55(1): 93-9.

97. Götherström G, Bengtsson BA, Bosaeus I, Johannsson G, Svensson J. Ten-year GH replacement increases bone mineral density in hypopituitary patients with adult onset GH deficiency. Eur J Endocrinol 2007; 156(1): 55-64.

98. Olney RC. Regulation of bone mass by growth hormone. Med Pediatr Oncol 2003; 41(3): 228-34.

99. Klefter O, Feldt-Rasmussen U. Is increase in bone mineral content caused by increase in skeletal muscle mass/strength in adult patients with GH-treated GH deficiency? A systematic literature analysis. Eur J Endocrinol 2009; 161(2): 213-21.

100. Maneatis T, Baptista J, Connelly K, Blethen S. Growth hormone safety update from the National Cooperative Growth Study. J Pediatr Endocrinol Metab 2000; 13 Suppl 2: 1035-44.

101. Wilton P, Mattsson AF, Darendeliler F. Growth hormone treatment in children is not associated with an increase in the incidence of cancer: experience from KIGS (Pfizer International Growth Database). J Pediatr 2010; 157(2): 265-70.

102. Guadalupe-Grau A, Fuentes T, Guerra B, Calbet JA. Exercise and bone mass in adults. Sports Med 2009; 39(6): 439-68.

103. Piguel X, Abraham P, Bouhours-Nouet N, et al. Impaired aerobic exercise adaptation in children and adolescents with craniopharyngioma is associated with hypothalamic involvement. Eur J Endocrinol 2012; 166(2): 215-22.

104. Conklin HM, Ness KK, Ashford JM, et al. Cognitive Performance, Aerobic Fitness, Motor Proficiency, and Brain Function Among Children Newly Diagnosed With Craniopharyngioma. J Int Neuropsychol Soc 2019; 25(4): 413-25.

105. Harz KJ, Müller HL, Waldeck E, Pudel V, Roth C. Obesity in patients with craniopharyngioma: assessment of food intake and movement counts indicating physical activity. J Clin Endocrinol Metab 2003; 88(11): 5227-31.

106. Shaikh MG, Grundy RG, Kirk JM. Reductions in basal metabolic rate and physical activity contribute to hypothalamic obesity. J Clin Endocrinol Metab 2008; 93(7): 2588-93.

107. Holmer H, Pozarek G, Wirfalt E, et al. Reduced energy expenditure and impaired feeding-related signals but not high energy intake reinforces hypothalamic obesity in adults with childhood onset craniopharyngioma. J Clin Endocrinol Metab 2010; 95(12): 5395-402.

108. Page-Wilson G, Wardlaw SL, Khandji AG, Korner J. Hypothalamic obesity in patients with craniopharyngioma: treatment approaches and the emerging role of gastric bypass surgery. Pituitary 2012; 15(1): 84-92.

109. Suominen H. Muscle training for bone strength. Aging Clin Exp Res 2006; 18(2): 85-93.

110. Horber FF, Thomi F, Casez JP, Fonteille J, Jaeger P. Impact of hydration status on body composition as measured by dual energy X-ray absorptiometry in normal volunteers and patients on haemodialysis. Br J Radiol 1992; 65(778): 895-900.

111. Hull H, He Q, Thornton J, et al. iDXA, Prodigy, and DPXL dual-energy X-ray absorptiometry whole-body scans: a cross-calibration study. Journal of clinical densitometry: the official journal of the International Society for Clinical Densitometry 2009; 12(1): 95-102.

112. Blijdorp K, van den Heuvel-Eibrink MM, Pieters R, et al. Obesity is underestimated using body mass index and waist-hip ratio in long-term adult survivors of childhood cancer. PLoS One 2012; 7(8): e43269.

113. van Santen SS, Olsson DS, Hammarstrand C, et al. Diagnosing Metabolic Syndrome in Craniopharyngioma Patients: Body Composition versus BMI. Eur J Endocrinol 2019.

114. Perreault L. Determining body composition in adults. July 10, 2017. https://www-uptodate-com.eur.idm.oclc.org/contents/determining-body-composition-in-adults?search=body%20composition%20adults&source=search_result&selectedTitle=1~116&usage_type=default&display_rank=1#H3702267332 (accessed July 4th, 2018 2018).

115. Caminiti C, Saure C, Bomer I, Brea M, Gonzalez Ramos J. Nutritional assessment of a population with a history of childhood craniopharyngioma seen at Hospital "Prof. Dr. Juan P. Garrahan". Arch Argent Pediatr 2017; 115(1): 43-9.

116. Bomer I, Saure C, Caminiti C, et al. Comparison of energy expenditure, body composition, metabolic disorders, and energy intake between obese children with a history of craniopharyngioma and children with multifactorial obesity. J Pediatr Endocrinol Metab 2015; 28(11-12): 1305-12.

117. Oeffinger KC, Mertens AC, Sklar CA, et al. Obesity in adult survivors of childhood acute lymphoblastic leukemia: a report from the Childhood Cancer Survivor Study. J Clin Oncol 2003; 21(7): 1359-65.

118. Green DM, Cox CL, Zhu L, et al. Risk factors for obesity in adult survivors of childhood cancer: a report from the Childhood Cancer Survivor Study. J Clin Oncol 2012; 30(3): 246-55.

119. Binnerts A, Swart GR, Wilson JH, et al. The effect of growth hormone administration in growth hormone deficient adults on bone, protein, carbohydrate and lipid homeostasis, as well as on body composition. Clin Endocrinol (Oxf) 1992; 37(1): 79-87.

120. Muller HL, Emser A, Faldum A, et al. Longitudinal study on growth and body mass index before and after diagnosis of childhood craniopharyngioma. J Clin Endocrinol Metab 2004; 89(7): 3298-305.

121. Goran MI, Kaskoun M, Johnson R. Determinants of resting energy expenditure in young children. J Pediatr 1994; 125(3): 362-7.

122. Zurlo F, Larson K, Bogardus C, Ravussin E. Skeletal muscle metabolism is a major determinant of resting energy expenditure. J Clin Invest 1990; 86(5): 1423-7.

123. Orsso CE, Tibaes JRB, Rubin DA, et al. Metabolic implications of low muscle mass in the pediatric population: a critical review. Metabolism 2019; 99: 102-12.

124. Illner K, Brinkmann G, Heller M, Bosy-Westphal A, Müller MJ. Metabolically active components of fat free mass and resting energy expenditure in nonobese adults. Am J Physiol Endocrinol Metab 2000; 278(2): E308-15.

125. Frontera WR, Ochala J. Skeletal muscle: a brief review of structure and function. Calcif Tissue Int 2015; 96(3): 183-95.

126. Taguchi M, Ishikawa-Takata K, Tatsuta W, et al. Resting energy expenditure can be assessed by fat-free mass in female athletes regardless of body size. J Nutr Sci Vitaminol (Tokyo) 2011; 57(1): 22-9.

127. Kim RJ, Shah R, Tershakovec AM, et al. Energy expenditure in obesity associated with craniopharyngioma. Childs Nerv Syst 2010; 26(7): 913-7.

128. Nogueira MC, Berbel Junior AS, Koenigkam-Santos M, Moreira AC, Nonino CB, de Castro M. Nutritional and endocrinologic evaluation of patients with craniopharyngioma. Clin Nutr ESPEN 2015; 10(6): e213-e8.

129. Jung HW, Kim HY, Kim JY, et al. Cardiac autonomic dysfunction is associated with hypothalamic damage in patients with childhood-onset craniopharyngioma. PLoS One 2021; 16(2): e0246789.

130. Thompson CJ, Costello RW, Crowley RK. Management of hypothalamic disease in patients with craniopharyngioma. Clin Endocrinol (Oxf) 2019; 90(4): 506-16.

131. Cohen M, Syme C, McCrindle BW, Hamilton J. Autonomic nervous system balance in children and adolescents with craniopharyngioma and hypothalamic obesity. Eur J Endocrinol 2013; 168(6): 845-52.

132. Toomey CM, McCormack WG, Jakeman P. The effect of hydration status on the measurement of lean tissue mass by dual-energy X-ray absorptiometry. Eur J Appl Physiol 2017; 117(3): 567-74.

133. Christ ER, Cummings MH, Westwood NB, et al. The importance of growth hormone in the regulation of erythropoiesis, red cell mass, and plasma volume in adults with growth hormone deficiency. J Clin Endocrinol Metab 1997; 82(9): 2985-90.

134. Vokes TJ, Robertson GL. Disorders of antidiuretic hormone. Endocrinol Metab Clin North Am 1988; 17(2): 281-99.

135. Newsome HH, Jr. Vasopressin: deficiency, excess and the syndrome of inappropriate antiduretic hormone secretion. Nephron 1979; 23(2-3): 125-9.

136. Fleseriu M, Hashim IA, Karavitaki N, et al. Hormonal Replacement in Hypopituitarism in Adults: An Endocrine Society Clinical Practice Guideline. The Journal of Clinical Endocrinology & Metabolism 2016; 101(11): 3888-921.

137. McPherron AC, Guo T, Bond ND, Gavrilova O. Increasing muscle mass to improve metabolism. Adipocyte 2013; 2(2): 92-8.

138. Scott D, Park MS, Kim TN, et al. Associations of Low Muscle Mass and the Metabolic Syndrome in Caucasian and Asian Middle-aged and Older Adults. J Nutr Health Aging 2016; 20(3): 248-55.

139. Kim G, Kim JH. Impact of Skeletal Muscle Mass on Metabolic Health. Endocrinol Metab (Seoul) 2020; 35(1): 1-6.

140. Lee J, Hong YP, Shin HJ, Lee W. Associations of Sarcopenia and Sarcopenic Obesity With Metabolic Syndrome Considering Both Muscle Mass and Muscle Strength. J Prev Med Public Health 2016; 49(1): 35-44.

141. Atlantis E, Martin SA, Haren MT, Taylor AW, Wittert GA. Inverse associations between muscle mass, strength, and the metabolic syndrome. Metabolism 2009; 58(7): 1013-22.

142. Kim TN, Park MS, Lim KI, et al. Skeletal muscle mass to visceral fat area ratio is associated with metabolic syndrome and arterial stiffness: The Korean Sarcopenic Obesity Study (KSOS). Diabetes Res Clin Pract 2011; 93(2): 285-91.

143. Vega GL. Obesity and the metabolic syndrome. Minerva Endocrinol 2004; 29(2): 47-54.

144. Gautier A, Godbout A, Grosheny C, et al. Markers of recurrence and long-term morbidity in craniopharyngioma: a systematic analysis of 171 patients. J Clin Endocrinol Metab 2012; 97(4): 1258-67.

145. Maghnie M. Diabetes insipidus. Horm Res 2003; 59 Suppl 1: 42-54.

146. Ong SR, Crowston JG, Loprinzi PD, Ramulu PY. Physical activity, visual impairment, and eye disease. Eye (Lond) 2018; 32(8): 1296-303.

147. Mulhern RK, Merchant TE, Gajjar A, Reddick WE, Kun LE. Late neurocognitive sequelae in survivors of brain tumours in childhood. Lancet Oncol 2004; 5(7): 399-408.

148. Belsky DW, Caspi A, Goldman-Mellor S, et al. Is obesity associated with a decline in intelligence quotient during the first half of the life course? American journal of epidemiology 2013; 178(9): 1461-8.

149. Pedersen SD, Astrup AV, Skovgaard IM. Reduction of misclassification rates of obesity by body mass index using dual-energy X-ray absorptiometry scans to improve subsequent prediction of per cent fat mass in a Caucasian population. Clin Obes 2011; 1(2-3): 69-76.

150. Shah NR, Braverman ER. Measuring adiposity in patients: the utility of body mass index (BMI), percent body fat, and leptin. PLoS One 2012; 7(4): e33308.

151. von Ruesten A, Steffen A, Floegel A, et al. Trend in obesity prevalence in European adult cohort populations during follow-up since 1996 and their predictions to 2015. PLoS One 2011; 6(11): e27455.

152. Sommer I, Teufer B, Szelag M, et al. The performance of anthropometric tools to determine obesity: a systematic review and meta-analysis. Scientific Reports 2020; 10(1): 12699.

153. Pluimakers VG, van Waas M, Neggers S, van den Heuvel-Eibrink MM. Metabolic syndrome as cardiovascular risk factor in childhood cancer survivors. Crit Rev Oncol Hematol 2019; 133: 129-41.

154. Razzouk BI, Rose SR, Hongeng S, et al. Obesity in survivors of childhood acute lymphoblastic leukemia and lymphoma. J Clin Oncol 2007; 25(10): 1183-9.

155. Lipshultz SE, Lipsitz SR, Sallan SE, et al. Chronic progressive cardiac dysfunction years after doxorubicin therapy for childhood acute lymphoblastic leukemia. J Clin Oncol 2005; 23(12): 2629-36.

156. Blanco JG, Sun CL, Landier W, et al. Anthracycline-related cardiomyopathy after childhood cancer: role of polymorphisms in carbonyl reductase genes--a report from the Children's Oncology Group. J Clin Oncol 2012; 30(13): 1415-21.

157. Hudson MM, Rai SN, Nunez C, et al. Noninvasive evaluation of late anthracycline cardiac toxicity in childhood cancer survivors. J Clin Oncol 2007; 25(24): 3635-43.

158. Oeffinger KC, Adams-Huet B, Victor RG, et al. Insulin resistance and risk factors for cardiovascular disease in young adult survivors of childhood acute lymphoblastic leukemia. J Clin Oncol 2009; 27(22): 3698-704.

159. Visscher H, Ross CJ, Rassekh SR, et al. Pharmacogenomic prediction of anthracycline-induced cardiotoxicity in children. J Clin Oncol 2012; 30(13): 1422-8.

160. Hudson MM, Link MP, Simone JV. Milestones in the curability of pediatric cancers. J Clin Oncol 2014; 32(23): 2391-7.

161. Armstrong GT, Liu Q, Yasui Y, et al. Late mortality among 5-year survivors of childhood cancer: a summary from the Childhood Cancer Survivor Study. J Clin Oncol 2009; 27(14): 2328-38.

162. Marriott CJC, Beaumont LF, Farncombe TH, et al. Body composition in long-term survivors of acute lymphoblastic leukemia diagnosed in childhood and adolescence: A focus on sarcopenic obesity. Cancer 2018; 124(6): 1225-31.

163. Bielorai B, Weintraub Y, Hutt D, et al. The metabolic syndrome and its components in pediatric survivors of allogeneic hematopoietic stem cell transplantation. Clin Transplant 2017; 31(3).

164. Cheung YT, Edelmann MN, Mulrooney DA, et al. Uric Acid and Neurocognitive Function in Survivors of Childhood Acute Lymphoblastic Leukemia Treated with Chemotherapy Only. Cancer Epidemiol Biomarkers Prev 2016; 25(8): 1259-67.

165. Kojima C, Kubota M, Nagai A, Adachi S, Watanabe K, Nakahata T. Adipocytokines in childhood cancer survivors and correlation with metabolic syndrome components. Pediatr Int 2013; 55(4): 438-42.

166. Ariffin H, Azanan MS, Abd Ghafar SS, et al. Young adult survivors of childhood acute lymphoblastic leukemia show evidence of chronic inflammation and cellular aging. Cancer 2017; 123(21): 4207-14.

167. Barbosa-Cortes L, Lopez-Alarcon M, Mejia-Arangure JM, et al. Adipokines, insulin resistance, and adiposity as a predictors of metabolic syndrome in child survivors of lymphoma and acute lymphoblastic leukemia of a developing country. BMC Cancer 2017; 17(1): 125.

168. Ju SY, Lee JY, Kim DH. Association of metabolic syndrome and its components with all-cause and cardiovascular mortality in the elderly: A meta-analysis of prospective cohort studies. Medicine (Baltimore) 2017; 96(45): e8491.

169. Wijnen M, Olsson DS, van den Heuvel-Eibrink MM, et al. Efficacy and safety of bariatric surgery for craniopharyngioma-related hypothalamic obesity: a matched case-control study with 2 years of follow-up. Int J Obes (Lond) 2017; 41(2): 210-6.

170. van Iersel L, Brokke KE, Adan RAH, Bulthuis LCM, van den Akker ELT, van Santen HM. Pathophysiology and Individualized Treatment of Hypothalamic Obesity Following Craniopharyngioma and Other Suprasellar Tumors: A Systematic Review. Endocr Rev 2019; 40(1): 193-235.

171. Rao RS. Bariatric surgery and the central nervous system. Obes Surg 2012; 22(6): 967-78.

172. Trotta M, Da Broi J, Salerno A, Testa RM, Marinari GM. Sleeve gastrectomy leads to easy management of hormone replacement therapy and good weight loss in patients treated for craniopharyngioma. Updates Surg 2017; 69(1): 95-9.

173. Gu L, Huang X, Li S, et al. A meta-analysis of the medium- and long-term effects of laparoscopic sleeve gastrectomy and laparoscopic Roux-en-Y gastric bypass. BMC Surg 2020; 20(1): 30.

174. Arterburn D, Wellman R, Emiliano A, et al. Comparative Effectiveness and Safety of Bariatric Procedures for Weight Loss: A PCORnet Cohort Study. Ann Intern Med 2018; 169(11): 741-50.

175. Casimiro I, Sam S, Brady MJ. Endocrine implications of bariatric surgery: a review on the intersection between incretins, bone, and sex hormones. Physiol Rep 2019; 7(10): e14111.

176. Farr OM, Sofopoulos M, Tsoukas MA, et al. GLP-1 receptors exist in the parietal cortex, hypothalamus and medulla of human brains and the GLP-1 analogue liraglutide alters brain activity related to highly desirable food cues in individuals with diabetes: a crossover, randomised, placebo-controlled trial. Diabetologia 2016; 59(5): 954-65.

177. Garrez I, Lapauw B, Van Nieuwenhove Y. Bariatric Surgery for Treatment of Hypothalamic Obesity After Craniopharyngioma Therapy: a Matched Case-Control Study. Obes Surg 2020.

178. Wolf P, Winhofer Y, Smajis S, et al. Hormone substitution after gastric bypass surgery in patients with hypopituitarism secondary to craniopharyngioma. Endocr Pract 2016; 22(5): 595-601.

179. Korde LA, Somerfield MR, Carey LA, et al. Neoadjuvant Chemotherapy, Endocrine Therapy, and Targeted Therapy for Breast Cancer: ASCO Guideline. J Clin Oncol 2021: Jco2003399.

180. Adamson TE, Wiestler OD, Kleihues P, Yaşargil MG. Correlation of clinical and pathological features in surgically treated craniopharyngiomas. J Neurosurg 1990; 73(1): 12-7.

181. Tavangar SM, Larijani B, Mahta A, Hosseini SM, Mehrazine M, Bandarian F. Craniopharyngioma: a clinicopathological study of 141 cases. Endocr Pathol 2004; 15(4): 339-44.

182. Khaddour K, Chicoine MR, Huang J, Dahiya S, Ansstas G. Successful Use of BRAF/MEK Inhibitors as a Neoadjuvant Approach in the Definitive Treatment of Papillary Craniopharyngioma. J Natl Compr Canc Netw 2020; 18(12): 1590-5.

183. Brastianos PK, Shankar GM, Gill CM, et al. Dramatic Response of BRAF V600E Mutant Papillary Craniopharyngioma to Targeted Therapy. J Natl Cancer Inst 2016; 108(2).

184. Aylwin SJ, Bodi I, Beaney R. Pronounced response of papillary craniopharyngioma to treatment with vemurafenib, a BRAF inhibitor. Pituitary 2016; 19(5): 544-6.

185. Himes BT, Ruff MW, Van Gompel JJ, et al. Recurrent papillary craniopharyngioma with BRAF V600E mutation treated with dabrafenib: case report. J Neurosurg 2018: 1-5.

186. Rao M, Bhattacharjee M, Shepard S, Hsu S. Newly diagnosed papillary craniopharyngioma with BRAF V600E mutation treated with single-agent selective BRAF inhibitor dabrafenib: a case report. Oncotarget 2019; 10(57): 6038-42.

187. Roque A, Odia Y. BRAF-V600E mutant papillary craniopharyngioma dramatically responds to combination BRAF and MEK inhibitors. CNS Oncol 2017; 6(2): 95-9.

188. Rostami E, Witt Nystrom P, Libard S, Wikstrom J, Casar-Borota O, Gudjonsson O. Recurrent papillary craniopharyngioma with BRAFV600E mutation treated with neo-adjuvant-targeted therapy. Acta Neurochir (Wien) 2017; 159(11): 2217-21.

189. Bernstein A, Mrowczynski OD, Greene A, et al. Dual BRAF/MEK therapy in BRAF V600E-mutated primary brain tumors: a case series showing dramatic clinical and radiographic responses and a reduction in cutaneous toxicity. J Neurosurg 2019: 1-6.

190. Grob JJ, Amonkar MM, Karaszewska B, et al. Comparison of dabrafenib and trametinib combination therapy with vemurafenib monotherapy on health-related quality of life in patients with unresectable or metastatic cutaneous BRAF Val600-mutation-positive melanoma (COMBI-v): results of a phase 3, open-label, randomised trial. Lancet Oncol 2015; 16(13): 1389-98.

191. Borrill R, Cheesman E, Stivaros S, Kamaly-Asl ID, Gnanalingham K, Kilday JP. Papillary craniopharyngioma in a 4-year-old girl with BRAF V600E mutation: a case report and review of the literature. Childs Nerv Syst 2019; 35(1): 169-73.

192. Whelan R, Prince E, Gilani A, Hankinson T. The Inflammatory Milieu of Adamantinomatous Craniopharyngioma and Its Implications for Treatment. J Clin Med 2020; 9(2).

193. Phelan R, Eissa H, Becktell K, et al. Upfront Therapies and Downstream Effects: Navigating Late Effects in Childhood Cancer Survivors in the Current Era. Curr Oncol Rep 2019; 21(12): 104.

194. Lomenick JP, Buchowski MS, Shoemaker AH. A 52-week pilot study of the effects of exenatide on body weight in patients with hypothalamic obesity. Obesity (Silver Spring) 2016; 24(6): 1222-5.

195. Wilding JPH, Batterham RL, Calanna S, et al. Once-Weekly Semaglutide in Adults with Overweight or Obesity. N Engl J Med 2021; 384(11): 989.

196. Peeters JJM, van den Berg P, van den Bergh JP, et al. Osteoporosis care during the COVID-19 pandemic in the Netherlands: A national survey. Arch Osteoporos 2021; 16(1): 11.

197. Puig-Domingo M, Marazuela M, Giustina A. COVID-19 and endocrine diseases. A statement from the European Society of Endocrinology. Endocrine 2020; 68(1): 2-5.

198. Jimenez-Kurlander L, Antal Z, DeRosa A, et al. COVID-19 in pediatric survivors of childhood cancer and hematopoietic cell transplantation from a single center in New York City. Pediatr Blood Cancer 2021; 68(3): e28857.

199. Forster VJ, Schulte F. Unique needs of childhood cancer survivors during the COVID-19 pandemic. Support Care Cancer 2021; 29(1): 17-9.

200. Yeh JM, Ward ZJ, Chaudhry A, et al. Life Expectancy of Adult Survivors of Childhood Cancer Over 3 Decades. JAMA Oncol 2020; 6(3): 350-7.

201. Santucci C, Carioli G, Bertuccio P, et al. Progress in cancer mortality, incidence, and survival: a global overview. Eur J Cancer Prev 2020; 29(5): 367-81.

202. Howlader N, Noone AM, Krapcho M, Miller D, Bishop K, Kosary CL, Yu M, Ruhl JTZ, Mariotto A, Lewis DR, Chen HS, Feuer EJ, Cronin KA SEER, Cancer Statistics Review (CSR) 1975-2014. April 2, 2018 2017. https://seer.cancer.gov/archive/csr/1975_2014/ (accessed 29-08-2019).

203. Gibson TM, Mostoufi-Moab S, Stratton KL, et al. Temporal patterns in the risk of chronic health conditions in survivors of childhood cancer diagnosed 1970-99: a report from the Childhood Cancer Survivor Study cohort. Lancet Oncol 2018; 19(12): 1590-601.

204. Syed IA, Klassen AF, Barr R, et al. Factors associated with childhood cancer survivors' knowledge about their diagnosis, treatment, and risk for late effects. J Cancer Surviv 2016; 10(2): 363-74.

9

Summary
Samenvatting

Summary

A major improvement in survival rates after tumors has been accomplished by the medical field, especially in pediatric cancer. This led to an increased awareness for the still suboptimal very long-term survival, and for frequently occuring late effects after treatment of neoplasms. The aim of this thesis was to further examine long-term endocrine, bone-related and metabolic alterations in those who suffer(ed) from a tumor, with a particular focus on patients with craniopharyngioma.

Accelerated growth after acromegaly due to *AIP* mutation

We describe a case of a 7-year old boy with accelerated growth, despite decreased food intake, due to excessive growth hormone production of a pituitary adenoma. Patients presenting with acromegaly at a young age or family members with acromegaly, are tested for genetic mutations. This should lead to early diagnosis of relatives and a better choice of therapy. In this exceptionally young patient, a new germline, paternally inherited *AIP* mutation (p.Tyr202x) was identified. *AIP* mutated somatotropinomas present mostly in young adolescent males and typically have an aggressive phenotype. Multiple surgeries and 1st and 2nd generation somatostatin analogues did not lead to cure, and the patient will require radiotherapy. In general, *AIP* mutated somatotropinomas may be more sensitive to 2nd generation somatostatin analogues, but it proves ineffective in our patient, which corresponded with in vitro insusceptability of the tumor to pasireotide. In vitro susceptibility of tumor tissue to somatostatin analogues may serve as a guide for the expected efficacy in the patient. This underlines the importance of targeted, tumor-specific therapy and combining treatment options.

Fractures, bone mineral density, growth, body composition and the metabolic syndrome in patients with craniopharyngioma

Craniopharyngioma is a benign neoplasm that lies close to the pituitary, hypothalamus and optic nerve. It often requires surgery and radiotherapy, which can lead to major long-term complications, including pituitary deficiences, visual impairments, neurological damage and obesity due to hypothalamic dysfunction. Together, this may influence bone health of these patients. We retrospectively studied bone health of Dutch and Swedish patients with craniopharyngioma. This population (with a median age at follow-up of 45 years) reached a normal mean final height, despite a high frequency of growth hormone deficiency (85%) and even after childhood-onset of disease. The cumulative incidence of fractures in patients with craniopharyngioma was 18%, as opposed to 8-10% in a general European population. Osteoporotic bone mineral density values occurred in 24%. Osteoporotic bone density values and pituitary deficiencies did not predict fracture risk; growth hormone replacement therapy protected against low bone mineral density. Fracture risk was especially high in patients using anti-epileptic drugs, and in men, while men received interventions (i.e. vitamin D or bisphosphonates) to improve bone mineral density less often than women. In addition, we evaluated the change in bone mineral density and body composition in patients with craniopharyngioma with a median time of 10 years between the first and last DXA-scan. The bone mineral density Z-scores of total body, L2-L4 and femur neck and fat-free mass index Z-scores increased significantly, while Z-scores of BMI and fat-related measures remained stable. This suggests bone density and body composition improve over time. A positive association with increasing bone density was found for growth hormone replacement therapy, while a negative association was found for radiotherapy, glucocorticoid dose and medication to improve bone mineral density. Obesity was better evaluated when applying fat mass index or fat percentage as derived by DXA-scan compared to body mass index. Obesity is one of the components of the metabolic syndrome (together with dyslipidemia (low HDL and high triglycerides), high glucose levels and hypertension). The metabolic syndrome is

associated with cardiovascular disease and is diagnosed if patients meet at least three out of five criteria of an altered metabolic profile. Even though more patients were diagnosed with obesity by DXA-scan, many patients had already met enough criteria to be already diagnosed with the metabolic syndrome. Their metabolic profile was so impaired that improved diagnosis of obesity did not give a major improvement the diagnosis of the metabolic syndrome. Risk factors for the metabolic syndrome were recurrence or progression of disease, hydrocephalus and age.

Biomarkers for the metabolic syndrome in childhood cancer survivors

Childhood cancer survivors have a high risk of metabolic alterations due to their past cancer treatment, while assessment of the metabolic syndrome can be difficult. This is especially the case after abdominal radiotherapy, which increases the risk of the metabolic syndrome while introducing a bias in waist circumference evaluation. In a systematic review and meta-analysis in childhood cancer survivors and a representative general population, hsCRP, adiponectin, uric acid and leptin were identified as useful biomarkers to diagnose and predict the metabolic syndrome, and apoB as additional predictive biomarker.

Long-term efficacy and safety of bariatric surgery in obese patients with craniopharyngioma

Treatment with bariatric surgery for obese patients with craniopharyngioma may have unsatisfactory results, as the hypothalamus is often damaged and insusceptible to the change of appetite-regulating hormones that occur after bariatric surgery. In this thesis we describe the first long-term case-control study, showing patients with craniopharyngioma had a mean weight loss of 22% at 5 years after bariatric surgery. Although average weight loss was lower in patients than in controls, especially in case of Roux-en-Y gastric bypass, the achieved weight loss was clinically meaningful. Bariatric surgery appeared relatively safe. Dose adjustments to hormonal replacement therapy were within the limits of expectation in the light of changing BMI and age.

In summary, craniopharyngioma patients are at high risk of impaired bone health, obesity and the metabolic syndrome; awareness and interventions to overcome the metabolic syndrome and consequent co-morbidities are warranted. For that purpose, international co-operation is needed to develop systemic therapies, for treatment of hypothalamic obesity and metabolic consequences, and for neo-adjuvant therapy targeted at the histopathological background of the craniopharyngioma.

Samenvatting

De verbeterde behandeling van tumoren heeft in de afgelopen decennia tot een duidelijke toename in de overleving geleid, met name onder kinderen. Daardoor is er in toegenomen mate aandacht gekomen voor de gezondheid van overlevenden van kanker op de langere termijn; late effecten komen frequent voor. In dit proefschrift zijn de botgezondheid, en endocriene en metabole status bij patiënten met een tumor onderzocht. Hierbij werden risicofactoren en beschermende factoren voor langetermijngevolgen en de wijze van diagnostiek bestudeerd. De onderzoeken werden met name gericht op patiënten met een craniopharyngioom.

Versnelde groei bij acromegalie door een *AIP* mutatie

We beschrijven de casus van een 7-jarige jongen met een versnelde lengtegroei, ondanks dat hij weinig eetlust heeft, door excessieve productie van groeihormoon vanuit een hypofyse-adenoom op een uitzonderlijk jonge leeftijd. Bij acromegalie patiënten die erg jong zijn bij ontstaan van de ziekte, of die familieleden met acromegalie hebben, dient te worden getest op genetische mutaties zodat familieleden vroegtijdig gediagnosticeerd kunnen worden en er gerichtere keuzes voor therapie gemaakt worden. Bij onze patiënt was sprake van een nieuwe, vanuit vaders zijde overgeërfde truncerende *AIP* mutatie (pTyr202x). *AIP* gemuteerde adenomen komen vooral voor bij mannelijke adolescenten en worden gekenmerkt door een agressief phenotype. Meerdere hersenoperaties en 1e en 2e generatie somatostatine analogen leidden bij onze patiënt niet tot genezing, en deze patiënt zal radiotherapie nodig hebben. Tweede generatie somatostatine analogen lijken in het algemeen effectief bij *AIP* gemuteerde adenomen, maar waren ineffectief in deze casus, wat overeen kwam met de ongevoeligheid van het tumorweefsel voor pasireotide bij vitro analyse. Deze casus onderschrijft het belang van een specifieke, tumorgerichte therapie en het combineren van behandelopties.

Botbreuken, botdichtheid, groei, lichaamscompositie en het metabool syndroom in patiënten met een craniopharyngioom

Het craniopharyngioom is een zeldzame goedaardige hersentumor, gelegen nabij de hypofyse, hypothalamus en optische zenuw. Deze tumor wordt meestal behandeld met een hersenoperatie en radiotherapie en wordt gekenmerkt door een breed spectrum aan complicaties op lange termijn, waaronder hormoonuitval door hypofysebeschadiging, visuele beperkingen, neurologische schade en obesitas door schade aan de hypothalamus. Dit zijn factoren die van invloed kunnen zijn op de botgezondheid. In een retrospectieve cohortstudie hebben we de botgezondheid onderzocht in Nederlandse en Zweedse patiënten met een craniopharyngioom (mediane leeftijd bij follow-up 45 jaar). Deze patiënten bereikten gemiddeld een normale lichaamslengte, ook wanneer de ziekte op de kinderleeftijd geconstateerd was. Botbreuken kwamen voor in 18% van de patiënten, terwijl dit percentage in de algemene Europese bevolking op 8-10% ligt. Gebruikers van medicijnen tegen epilepsie en mannen liepen een hoger risico op botbreuken, terwijl mannen minder medicatie gebruikten om de botdensiteit te verhogen (bijv. vitamine D en bisfosfonaten). Zeer lage botmineraaldensiteit, passend bij botontkalking, kwam voor bij 24%. Groeihormoonbehandeling leek beschermend tegen een lage botdichtheid. Een verbetering van botdichtheid en lichaamscompositie werd geobserveerd met DXA-scans met een mediane tijd van 10 jaar tussen de metingen. Er was een significante toename zichtbaar van de Z-scores voor botdichtheid van de botten in het hele lichaam, in de lage rugwervels (L2-L4) en de femurhals, en voor vetvrije massa index, terwijl Z-scores voor body mass index en vetgerelateerde waarden stabiel bleven. Er was een positieve associatie met botdichtheidstoename voor groeihormoontherapie, en een negatieve associatie met radiotherapie, glucocorticoïd dosering en botdichtheidsverhogende medicatie. Obesitas bleef echter een veelvuldig voorkomend probleem, en is een van de onderdelen van het metabool syndroom (naast afwijkende vetten in het bloed (laag HDL en hoog triglyceriden), een hoge bloeddruk en verhoogde bloedglucosewaarden).

Het metabool syndroom wordt gediagnosticeerd als aan minimaal drie van de vijf onderdelen voldaan wordt. Het metabool syndroom geeft een verhoogd risico op hart- en vaatziekten en diabetes mellitus. Het meten van vetpercentage en vet massa index met een DXA-scan verbeterde de diagnostiek van obesitas ten opzichte van body mass index. Echter, veel patiënten hadden al veel afwijkende bevindingen voor de andere onderdelen van het metabool syndroom, waardoor deze verbeterde diagnose van de component obesitas slechts beperkte waarde had voor de diagnose van het metabool syndroom. Risicofactoren voor het metabool syndroom waren progressie van ziekte, hydrocephalus (ophoping hersenvloeistof) en leeftijd.

Diagnostiek en voorspelling van het metabool syndroom met biomarkers in overlevenden van kinderkanker

Kinderen die kanker overleefd hebben, hebben ook een verhoogd risico op het metabool syndroom, terwijl de diagnose van obesitas en daarmee metabool syndroom bemoeilijkt wordt, met name na radiotherapie op de buik. Hierbij kan de buikgroei beperkt zijn, waardoor de buikomvang klein is ondanks een verhoogd vetpercentage. Bij een systematisch literatuuronderzoek naar het toepassen van biomarkers voor de diagnose en voorspelling van het metabool syndroom, werden studies onder overlevenden van kinderkanker en een representatieve gezonde populatie gebruikt. Aan de diagnose van het metabool syndroom konden de biomarkers hsCRP, adiponectine, urinezuur en leptine bijdragen. Het voorspellen van het ontstaan van het metabool syndroom werd voorspeld door dezelfde biomarkers, en daarnaast ook apoB.

Effectiviteit en veiligheid op de lange termijn van bariatrische chirurgie voor obese patiënten met een craniopharyngioom

De behandeling van obesitas bij patiënten met een craniopharyngioom met bariatrische chirurgie (maagverkleiningsoperaties) leidt potentieel tot matige resultaten. De hypothalamus reguleert het lichaamsgewicht

maar is frequent beschadigd bij deze patiënten, en kan hierdoor verminderd gevoelig zijn voor hongergevoel-regulerende hormonen. We presenteren de eerste studie die het gewichtsverlies van craniopharyngioom patiënten op lange termijn onderzoekt. Na vijf jaar hebben patiënten gemiddeld een gewichtsverlies van 22%. Dit is een goed resultaat, maar het is minder dan in gematchte controlepatiënten (bariatrische chirurgiepatiënten zonder craniopharyngioom). Er waren geen grote wijzigingen noodzakelijk in de medicatie voor de hypofyse-uitval. Bariatrische chirurgie lijkt een veilige therapeutische optie voor obesitas in deze patiënten.

De toekomst brengt hopelijk systemische therapieën die gericht zijn op het verminderen van hypothalame obesitas en op tumorverkleining voorafgaand aan operatie, gebaseerd op de histopathologie van het craniopharyngioom. Bij het onderzoek naar deze zeldzame ziekte is internationale samenwerking noodzakelijk.

10

About the Author

About the author

Selveta Sanne (Selvetta) van Santen was born on April 24th in Dordrecht, the Netherlands. She graduated from high school in 2010 (CSG De Lage Waard, Papendrecht). She immediately enrolled in the Bachelor study in Medicine, followed by her Master's, for which she received her degree in 2017. During her training period, she performed research into cardiovascular risk related to apoB at the Department of Internal Medicine/Vascular diseases of Sint Franciscus Gasthuis, Rotterdam, under the direction of Doctor M. Castro Cabezas and Doctor M. de Vries. After her graduation, she worked as a physician for the inpatient clinic at the department of Internal Medicine of the Albert Schweitzer Ziekenhuis, Dordrecht for 7 months, until she decided to focus on research again. She started her PhD in September 2017, of which the results are presented in this thesis. Selvetta thankfully received guidance from her promotor A.J. Professor van der Lelij and copromotor Doctor S.J.C.M.M. Neggers, both specialized in endocrinology (Department of Medicine – Endocrinology, Erasmus Medical /Pituitary Center Rotterdam), and her promotor Professor van den Heuvel-Eibrink, who is specialized in pediatric oncology (Princess Máxima Center for Pediatric Oncology and Department of Pediatric Oncology/Hematology Erasmus Medical Center). During her training, she worked as a physician at the Late Effects clinic of the Erasmus University Medical Center and Princess Máxima Center for Pediatric Oncology, and treated adult survivors of childhood cancer for long-term complications of their disease and previous treatment. After finishing her PhD training in March 2021, she returned to the Albert Schweitzer Ziekenhuis to work as a physician at the Department of Internal Medicine again under the supervision of Doctor P.J.H. Smak Gregoor, with the ultimate goal of entering a traineeship in Internal Medicine.

PhD Portfolio

Name PhD student: Selveta Sanne van Santen

Erasmus MC department: Medicine, section Endocrinology – Pituitary Center Rotterdam
Research school: Netherlands Institute for Health Sciences
Other affiliation: Princess Máxima Center for Pediatric Oncology

PhD period: September 2017 – March 2021

Promotors: Prof. dr. A.J. van der Lelij
Prof. dr. M.M. van den Heuvel-Eibrink
Copromotor: Dr. S.J.C.M.M. Neggers

	Year	ECTS
General courses		
Basic course for Clinical Investigators (BROK)	2017	2.0
NIHES Biostatistical Methods I: Basic Principles (CC02) (grade 9.0)	2017	5.7
Systematic literature research Pubmed I and II, Medical Library Erasmus MC	2017	0.33
Systematic literature research Multiple databases, Medical Library Erasmus MC	2017	0.33
Endnote course, Medical Library Erasmus MC	2017	0.33
Open Clinica, Erasmus MC	2017	0.3
Logframe course, Erasmus MC	2017	0.2
Masterclass acromegaly (partially completed)	2017	0.2
Course basic life support (pediatrics), Princess Máxima Center for Pediatric Oncology, Utrecht, the Netherlands	2018	0.2

Excel basic course, Medical Library Erasmus MC	2018	0.3
Excel advanced course, Medical Library Erasmus MC	2018	0.4
Access basic course, Medical Library Erasmus MC	2018	0.3
Access advanced course, Medical Library Erasmus MC	2018	0.4
NIHES ESP66 Logistic Regression (summer course)	2018	1.4
NIHES ESP09 Regression Analysis (summer course)	2018	1.4
Scientific writing (partially completed)	2018	-
Time management and negotiating, Erasmus MC	2018	1
Postgraduate Course, GH and Growth Factors – Metabolic Disorders, Gothenburg, Marstrand, Sweden (with case presentation)	2018	2
25th ESE Postgraduate Training course on Endocrinology, Diabetes and Metabolism, Rotterdam, the Netherlands	2019	0.9
Research Integrity Course, Erasmus MC	2019	0.3
Pituitary Masterclass, Milan, Italy	2019	0.5
NIHES Survival Analysis for Clinicians (EWP24) (grade 9.5)	2020	1.9
NIHES Advanced Topics in Clinical Trials (EWP10)	2020	1.9
Pituitary Society fellowship course (online)	2020	0.3
Postgraduate Craniopharyngioma Course: New developments and challenges in the treatment of Childhood Craniopharyngioma; striving for optimal balance	2021	0.6
Research skills		

Weekly research meeting Endocrinology Laboratory (with 10 oral presentations)	2017-2021	4.5
Weekly research meeting Princess Máxima center for Pediatric Oncology - van den Heuvel-groep and Late Effects of Childhood Cancer (with 2 oral presentations)	2020-2021	2.0
Seminars, meetings and workshops		
Scientific Endocrinology meetings (Internal Medicine Research Meeting, Erasmus Lectures on Endocrinology and Workdiscussions with once an oral presentation)	2017-2020	1.5
PhD day: a Healthy PhD, Rotterdam, the Netherlands	2018	0.2
ESP64 Summer lectures (summer course)	2018	0.2
Van den Heuvel group retreat "Fier op Schier", Schiermonnikoog, the Netherlands	2019	1.0
Van den Heuvel group retreat (online)	2020	0.5
National and international conferences		
JNVE meeting, Nijmegen, the Netherlands (with oral presentation)	2018	2.0
European Congress of Endocrinology, Barcelona, Spain (poster presentation)	2018	1.0
Pancare, Prague, Czech Republic	2018	1.0
Wetenschapsdagen (with oral presentation)	2019	2.0
Dutch Endocrine Meeting, Noordwijkerhout, the Netherlands (poster presentation with oral pitch)	2019	1.0
ENDO, New Orleans, USA (poster presentation, nominated for presidential poster)	2019	1.0
European Congress of Endocrinology, Lyon, France (poster presentation)	2019	1.0
EYES, Athens, Greece (oral presentation)	2019	2.0

Wetenschapsdagen, Sint-Michielsgestel, the Netherlands (poster presentation)	2020	1.0
Dutch Endocrine Meeting, Noordwijkerhout, the Netherlands (poster presentation with oral pitch)	2020	1.1
ENDO (Online)	2020	1.0
European Congress of Endocrinology (online, with poster presentation)	2020	1.0
SIOP (with poster presentation; online)	2020	1.0
ICCBH (with poster presentation)	2020	1.0
Dutch Endocrine Meeting (online)	2021	0.3
Clinical meetings and participation		
Outpatient clinic for long-term complications of childhood cancer treatment	2017-2020	40
Teaching activities		
Lecture "Late effects after treatment of childhood cancer" for medical students in the second year of their study.	2018	1.0
Skills education "Hypercortisolism" for medical students in the first year of their study.	2018	1.0
Lecture "Late effects after treatment of childhood cancer" for medical students in the second year of their study.	2019	1.0
Lecture "Late effects after treatment of childhood cancer" for medical students in the second year of their study.	2020	1.0

List of publications

1. Klop B, van der Pol P, van Bruggen R, Wang Y, de Vries MA, **van Santen S**, O'Flynn J, van de Geijn GJ, Njo TL, Janssen HW, de Man P, Jukema JW, Rabelink TJ, Rensen PC, van Kooten C, Cabezas MC. Differential complement activation pathways promote C3b deposition on native and acetylated LDL thereby inducing lipoprotein binding to the complement receptor 1. Journal of Biological Chemistry 2014; 289(51): 35421-30.

2. de Vries MA, Alipour A, Birnie E, Westzaan A, **van Santen S**, van der Zwan E, Liem AH, van der Meulen N, Cabezas MC. Coronary leukocyte activation in relation to progression of coronary artery disease. Frontiers in Medicine 2016; 10(1): 85-90.

3. de Vries MA, **van Santen SS**, Klop B, van der Meulen N, van Vliet M, van de Geijn GM, van der Zwan-van Beek EM, Birnie E, Liem AH, de Herder WW, Castro Cabezas M. Erythrocyte-bound apolipoprotein B in atherosclerosis and mortality. European Journal of Clinical Investigation 2017; 47(4): 289-96.

4. **van Santen SS**, Olsson DS, Hammarstrand C, Wijnen M, van den Heuvel-Eibrink MM, Van der Lely AJ, Johannsson G, Janssen J, Neggers S. Diagnosing Metabolic Syndrome in Craniopharyngioma Patients: Body Composition versus BMI. European Journal of Endocrinology 2019;181(2):173-183

5. **van Santen SS**, Olsson DS, van den Heuvel-Eibrink MM, Wijnen M, Hammarstrand C, Janssen J, Johansson G, van der Lely AJ, Neggers S. Fractures, Bone Mineral Density, and Final Height in Craniopharyngioma Patients with a Follow-up of 16 Years. The Journal of Clinical Endocrinology & Metabolism 2020;105(4):e1397-e1407

6. **van Santen SS**, Olsson DS, Hammarstrand C, Wijnen M, Fiocco M, van den Heuvel-Eibrink MM, Johannsson G, Janssen JAMJL, van der Lely AJ, Neggers SJCMM. Body Composition and Bone Mineral Density in Craniopharyngioma Patients: A Longitudinal Study Over 10 Years. The Journal of Clinical Endocrinology & Metabolism 2020;105(12):dgaa607.

7. Pluimakers VG, **van Santen SS**, Fiocco M, Bakker MCE, van der Lelij AJ, van den Heuvel-Eibrink MM, Neggers SJCMM. Can biomarkers be used to improve diagnosis and prediction of metabolic syndrome in childhood cancer survivors? A systematic review. Obesity Reviews 2021; 22(11): e13312.

8. **van Santen SS**, Wolf P, Kremenevski N, Boguszewski CL, Beiglböck H, Fiocco M, Wijnen M, Wallenius VR, van den Heuvel-Eibrink MM, van der Lely AJ, Johannsson G, Luger A, Krebs M, Buchfelder M, Delhanty PJD, Neggers SJCMM, Olsson DSO. Bariatric Surgery for Hypothalamic Obesity in Craniopharyngioma Patients: A Retrospective, Matched Case-Control Study. Journal of Clinical Endocrinology & Metabolism 2021; 106(11): e4734-e4745.

9. **van Santen SS**, Daly AF, Buchfelder M, Coras R, Zhao Y, Beckers A, van der Lely AJ, Hofland LJ, Balvers RK, van den Heuvel-Eibrink MM, Neggers SJCMM. Complicated clinical course in incipient gigantism due to a treatment resistant, *AIP*-mutated pediatric somatotropinoma. *Submitted.*

Affiliations of co-authors

Author	Affiliation
Marie-Christine E. Bakker, BSc	Princess Máxima Center for Pediatric Oncology, Utrecht, The Netherlands Department of Medicine, University Medical Center Utrecht, The Netherlands
Rutger K. Balvers, MD, PhD	Department of Medicine, Neurosurgery; Erasmus Medical Center, Rotterdam, The Netherlands
Albert Beckers, MD, PhD	Department of Endocrinology, Liège University Hospital Center, Liège, Belgium
Hannes Beiglböck, MD	Department of Endocrinology and Metabolism; Medical University of Vienna, Austria
Cesar L. Boguszewski, MD, PhD	Department of Internal Medicine; Universidade Federal do Paraná, Curitiba, Brazil
Michael Buchfelder, MD, PhD	Department of Neurosurgery; University Hospital Erlangen, Germany
Roland Coras, MD	Department of Pathology; University Hospital Erlangen, Erlangen, Germany
Adrian F. Daly, MD, PhD	Department of Endocrinology, Liège University Hospital Center, Liège, Belgium
Patric J.D. Delhanty, PhD	Department of Endocrinology; Erasmus Medical Center, Rotterdam, The Netherlands The Metabolism and Reproduction laboratory; Erasmus Medical Center, Rotterdam, The Netherlands

Marta Fiocco, PhD	Princess Máxima Center for Paediatric Oncology, Utrecht, The Netherlands Mathematical Institute, Leiden University, Leiden, The Netherlands Department of Biomedical Data Sciences, Medical Statistics Section, Leiden University Medical Center, Leiden, The Netherlands
Casper Hammarstrand, MD	Department of Endocrinology, Sahlgrenska University Hospital, Gothenburg, Sweden Department of Internal Medicine and Clinical Nutrition, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden
Leo J. Hofland, MD, PhD	Department of Medicine, Endocrinology; Erasmus Medical Center, Rotterdam, The Netherland
Marry M. van den Heuvel-Eibrink, MD, PhD	Princess Maxima Center for Paediatric Oncology, Utrecht, the Netherlands Department of Paediatric Oncology/ Haematology, Erasmus MC – Sophia Children's Hospital, Rotterdam, the Netherlands
Joseph A.M.J.L. Janssen, MD, PhD	Department of Medicine, Section Endocrinology, Pituitary Center Rotterdam, Erasmus University Medical Center, Rotterdam, the Netherlands
Gudmundur Johannsson, MD, PhD	Department of Endocrinology, Sahlgrenska University Hospital, Gothenburg, Sweden Department of Internal Medicine and Clinical Nutrition, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden
Michael Krebs, MD, PhD	Department of Endocrinology and Metabolism, Medical University of Vienna, Austria

Natalia Kremenevski, MD, PhD	Department of Neurosurgery, University Hospital Erlangen, Germany
Aart-Jan van der Lelij, MD, PhD	Department of Medicine, Section Endocrinology, Pituitary Center Rotterdam, Erasmus University Medical Center, Rotterdam, the Netherlands
Anton Luger, MD, PhD	Department of Endocrinology and Metabolism; Medical University of Vienna, Austria
Sebastian J.C.M.M. Neggers, MD, PhD	Department of Medicine, Section Endocrinology, Pituitary Center Rotterdam, Erasmus University Medical Center, Rotterdam, the Netherlands Department of Paediatric Oncology/ Haematology, Erasmus MC – Sophia Children's Hospital, Rotterdam, the Netherlands Princess Máxima Center for Pediatric Oncology, Utrecht, the Netherlands
Daniel S. Olsson, MD, PhD	Department of Medicine, Endocrinology; Sahlgrenska University Hospital, Gothenburg, Sweden Department of Internal Medicine and Clinical Nutrition, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden
Vincent G. Pluimakers, MD, MSc	Princess Máxima Center for Pediatric Oncology, Utrecht, The Netherlands
Ville Wallenius, MD, PhD	Department of Gastrosurgical Research and Education, Institute of Clinical Sciences, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden

Mark Wijnen, MD, PhD	Department of Medicine, Endocrinology, Erasmus Medical Center, Rotterdam, The Netherlands Department of Paediatric Oncology/ Haematology, Erasmus MC – Sophia Children's Hospital, Rotterdam, The Netherlands
Peter Wolf, MD, PhD	Department of Endocrinology and Metabolism; Medical University of Vienna, Austria
Yining Zhao, MD, MSc	Department of Neurosurgery; University Hospital Erlangen, Erlangen, Germany

Dankwoord (acknowledgements)

Dit werk was nooit tot stand gekomen zonder het enthousiasme, input, hulp en steun van heel veel mensen, die ik bij deze wil bedanken.

Allereerst en in het bijzonder, mijn promotoren en copromotor:

Professor M.M. van den Heuvel-Eibrink (mijn promotor), beste **Marry**, hartelijk dank voor al je input en werk aan mijn promotie! Het was heel verfrissend om naast een hoofdzakelijk endocrinologische basis ook gebruik te kunnen maken van jouw uitstekende kennis van pediatrische oncologie en bijbehorende late effecten. Je gaf altijd met enthousiasme grote prioriteit aan beoordelen van manuscripten ('dat vind ik heerlijk, lezen onder de kerstboom!'), en bracht meer variatie en een andere kijk op mijn schrijfstijl. Naast je professionele kennis had je altijd oog voor de mensen met wie je werkte. Je bent zeer attent en hebt bij het organiseren van de retraites je promovendi nader tot elkaar gebracht en uitgedaagd tot persoonlijke ontwikkeling. Ik heb ook veel respect voor het harde werk dat je in LATER hebt gestoken. Je mag trots zijn dat dankzij jouw leiderschap en doorzettingsvermogen, en ondanks de nodige tegenslagen mooie vruchten heeft afgeworpen.

Doctor S.J.C.M.M. Neggers (mijn copromotor), beste **Sebastian**, als dagelijkse begeleider heb jij misschien wel de grootste stempel op dit werk gedrukt. Ik wil je hartelijk bedanken voor je begeleiding en inzichten bij het werken aan de manuscripten. Je zeer uitgebreide endocrinologische kennis hielp enorm bij het verkrijgen van de inzichten tijdens de studies die we verrichten, en je kon bevindingen uiterst goed onder woorden brengen. Tijdens mijn promotie had ik soms weinig vertrouwen in het slagen van sommige dingen; gelukkig kon ik dan altijd op jouw positieve instelling rekenen. Zoals ook Mark in zijn dankwoord schreef, is jouw reactie meestal: "Dat komt wel goed!" Je hebt wel eens verteld dat je in het 'grootbrengen' van je promovendi dezelfde tactiek hanteert als bij je eigen kinderen, namelijk: leren door zelf te doen! Dat betekende dat ik in alle vrijheid zelf een plan van aanpak op kon stellen, en in alle benodigde factoren voor succes werd gefaciliteerd. Bij problemen kon ik gelukkig altijd bij je terecht, en je besprak met geduld

zowel zaken gerelateerd aan onderzoek als aan de LATER-polikliniek. Ik heb hierdoor geweldig veel geleerd in de afgelopen jaren. Dit alles waardeer ik zeer.

Professor A.J. van der Lelij (mijn promotor), beste **Aart-Jan**, hartelijke dank dat ik op jouw afdeling mijn onderzoek heb mogen doen! Bij de besprekingen op maandagochtend werd jouw kundigheid en wijsheid in basaal, translationeel en klinisch metabool-endocrinologisch onderzoek zeer duidelijk. Je vriendelijkheid droeg bij aan het positieve klimaat en de veilige leeromgeving. Ons directe contact over mijn projecten was niet heel uitgebreid tijdens mijn periode als PhD student, maar je was desondanks duidelijk erg betrokken. Dit werd bevestigd door je enthousiaste emails en door je input op onze bevindingen via Sebastian. Heel erg bedankt voor je inzichten en commentaren op de manuscripten.

Dear **Doctor D.S. Olsson** and **professor G. Johannsson**, dear **Daniel** and **Gudmundur**, with Mark's PhD you have laid the foundation for a great and extensive international collaboration, which I am proud and thankful to be part of. These joined forces extensively broadened our knowledge on this rare disease. **Daniel**, you are one of the most hard-working, brilliant and eager researchers that I have ever met. Thank you for your guidance in the past years. **Gudmundur**, thank you for your wisdom and for applying your experience in the advice for our manuscripts. You are one of the most experienced professors in the field; you always put a lot of effort in the articles and had sharp observations which I appreciated very much. Dear **Casper Hammarstrand**, thank you for all your hard work on the data collection for our joint projects.

Ook wil ik mijn commissieleden **professor E.F.S. van Rossum**, **professor A.M. Pereira Arias**, **dr. A.J.P.E. Vincent**, **associate professor D.S. Olsson**, **doctor H.M. van Santen**, **professor W.W. de Herder**, **professor B. Lapauw** en **doctor E.H.G. Oei** hartelijk danken voor het beoordelen van mijn proefschrift.

Doctor M. Wijnen, **beste Mark**, jouw databases en daaruit voortgevloeide prachtige artikelen hebben een hele mooie basis gelegd

voor mijn promotie. Hartelijk dank daarvoor, en ook voor het wegwijs maken in het eerste jaar van mijn promotie en helpen met de statistiek. **Doctor K. Blijdorp**, **beste Karin**, ook dank voor jouw hulp na je eerdere ervaringen bij het onderzoek naar lichaamssamenstelling.

Professor M. Fiocco, beste **Marta**, met grote trots mag ik je nu met professor aanspreken. Dat is geheel terecht, gezien je kunde en de uitmuntende begeleiding die ik en andere PhD studenten hebben ontvangen op het gebied van statistiek. Jouw enthousiasme, warme uitstraling en geruststellende woorden bij het bespreken van alle statistische kwesties hebben me veel goed gedaan en de kwaliteit van mijn artikelen absoluut verder verhoogd, ik wil je daar heel erg voor bedanken.

Also, I would like to express my gratitude to all other collaborators. In particular, **professor A. Daly**, **professor M. Buchfelder**, **professor R. Coras**, **doctor Y. Zhao**, **professor A. Beckers**, **professor L.J. Hofland**, **doctor R.K. Balvers**, **doctor P. Wolf, doctor N. Kremenevski**, **professor C.L. Boguszewski**, **doctor Hannes Beiglböck**, **doctor V.R. Wallenius**, **professor A. Luger**, **professor M. Krebs**, **Peter Todd** and **doctor J.A.M.L. Janssen**. (International) collaborations are an absolute necessity when performing research aimed at a rare disease, and I could not have performed these studies without your help. Thank you very, very much! **Professor C. de Block**, **professor B. Lapauw**, **Patric, doctor H.M. van Santen**, **doctor E.L.T. van den Akker**, **mr. De Graaf**, **mrs. Braspenning**, although we were not so lucky when applying for funding for a randomized controlled trial, we still laid a base of strong coherence to develop future trials. Thank you for your hard work!

Doctor J.A.M.L. Janssen, beste **Joop**, hartelijk dank voor de wijze raad die u heeft gegeven als ervaren onderzoeker aan een beginnende onderzoeker. Ik heb daar veel van geleerd. Daarnaast wil ik graag professor **J.M. Kros** bedanken voor zijn inzet voor het histopathologische onderzoek naar het craniopharyngioom, dat waarschijnlijk door een opvolger verder uitgewerkt zal worden. **Professor M.C. Zillikens** en **professor F. Rivadeneira**, ook veel dank aan u beiden voor het overleg over de botgerelateerde stukken. **Doctor S.M.F. Pluijm**, hartelijk dank voor uw adviezen omtrent de statistiek in mijn eerste stuk.

Aan mijn paranimfen, **Myrthe van Santen** (mijn jongere zus) en **Vincent Pluimakers.** Superbedankt voor het mogelijk maken van deze mooie dag! **Myrthe**, het lezen van jouw thesis bracht me daarnaast weer even in de positie lezer van onbekende informatie, wat me hernieuwde punten voor verbetering voor het schrijven van mijn eigen stukken gaf. **Vincent**, het was me een waar genoegen om met jou samen te werken! Jij straalt een enorme rust en vriendelijkheid uit en kan je altijd supergoed verwoorden. Dat kwam zeker goed uit toen we samen werkten aan ons gemeenschappelijke metabool syndroom project! Aan mijn favoriete collega **Ariadne Bosman**, superblij was ik toen jij mij achterna kwam, vanuit het ASZ naar het EMC om ook onderzoek te doen! De beste kamergenoot die ik me maar kan wensen, fijn om een klankbord te hebben en iemand om mee te kunnen lachen! Je had een geweldige paranimf geweest, maar bent in blijde verwachting en daarom helaas niet bij mijn verdediging. Desondanks heel veel dank voor alle hulp bij de voorbereidingen!

Graag wil ik mijn dank uiten aan de Raad van Bestuur van het Erasmus MC (**professor E.J. Kuipers**, **professor J.P.T.M. van Leeuwen**, **doctor J.G. Boonstra** en **doctor ingenieur P.A.M. Boomkamp**) en het Princes Máxima Center voor Pediatrische Oncologie (**professor Alexander Eggermont**, **doctorandus Gita Gallé**, **professor Rob Pieters**) voor het mogelijk maken van mijn promotietraject. Ook mijn dank aan **doctor S. Klein Nagelvoort – Schuit** en **professor R. Peeters** die zich als hoofden van de Interne Geneeskunde over mijn arbeidscontracten hebben gebogen tijdens mijn promotietraject. **Doctor A.A.M. Zandbergen**, dank u voor het fijne overleg over de vervolgstappen in mijn carrière. **Doctor R.A. Feelders**, ook dank aan u voor het bespreken van mijn ambities.

Bovenal wil ik mijn dank uitspreken naar de patiënten die deelnamen aan de onderzoeken. Ook veel dank aan de artsen bij het Hypofyse Centrum in het Erasmus MC en bij LATER. Zonder hun werk was mijn proefschrift niet tot stand gekomen.

Heel graag wil ik ook mijn collega's bij mijn studie en werk bij het Sint Franciscus Gasthuis en Albert Schweitzer Ziekenhuis bedanken.

Allereerst **Marijke de Vries**, **Gert-Jan van de Geijn**, **Boudewijn Klop**, **Noëlle Vermeulen** en bovenal **Manuel Castro Cabezas**; dankzij jullie is mijn enthousiasme voor de medische wetenschap ontstaan. Jullie hebben me tijdens mijn onderzoeksstage de beste begeleiding gegeven die een medisch student maar kan wensen, en het was heel mooi om ook ervaring in het laboratorium op te doen. **Marijke** en **Minet (Kosman)**, het intensieve samenwerken en samen studeren heeft uiteindelijk tot een mooie vriendschap geleid; ik ben heel blij met onze gezellige terrasmomenten, die de nodige ontspanning hebben geboden en ook heerlijk de ruimte bood om van alles, van werk tot privé te bespreken. Dank jullie wel daarvoor! Marijkes gezinnetje met **Nikolaj, Annemijn en Floris** droegen ook zeker bij aan de gezelligheid.

Andere supervisoren en collega's die ik wil bedanken voor de fijne samenwerking en begeleiding zijn onder meer **dr. Indhiradjanti-Tomasoa, dr. Wiebolt, dr. Schrama** en **dr. Rietveld** uit het Sint Franciscus Gasthuis, en **dr. Smak Gregoor, dr. Van Bommel, dr. Schoofs, dr. Kiewiet-Kemper, dr. Massolt, dr. Meijer,** en de andere internisten, longartsen, MDL-artsen, SEH-artsen en cardiologen bij het Albert Schweitzer ziekenhuis, en daarnaast **Siham, Gijs, Bob, Hanah, Hardjit, Henk Jan, Inge, Khathera, Ingrid, Rashaad, Sven, Anne, Dorothy, Elske, Eva, Jelena, Marilyn, Mick, Maarten, Milou, Nicolien, Floor, Bastiaan, Claire, Victor, Michelle, Sindhu, Mathé, Lindsay, Marjolein, Eske, Faryel, Annemarie, Lucia, Evelien, Gideon, Eva, Mustafa, Laura, Shauny, Stephanie, Eline, Annejet, Annemarie, Lisanne, Mark, Rugina, Geerte, Joep, Odile, Elise, Mirna, Tim, Mirna, Nina, Dani, Maartje, Charlotte, Liza, Eva, Kelly, Anne-Marie, Naomi, Renske, Sanne, Milaine, Jojanneke, Kasia, Esma, Eva, Emily, Evi, Merve, Diana, Cynthia** en **Lichelle**. Deze lijst personen is vast niet compleet, ook dank aan de anderen en aan alle collega's die ik tijdens mijn tweede periode bij het ASZ nog moet leren kennen! Ik heb heel veel geleerd van het werken in de kliniek.

Tijdens mijn onderzoek heb ik enkele jaren polikliniek voor LATER-patiënten gedaan (lange termijn effecten na kinderkanker). Graag wil ik mijn collega's bij LATER heel erg bedanken voor de fijne tijd. Allereerst dank voor deze positie aan **doctor W.J.W. Kollen**, managing director

care. **Manita**, jij en je gezin hebben natuurlijk een heel bijzondere rol gespeeld door jullie betrokkenheid bij mijn bruiloft! Dank voor de mooie herinneringen. **Doctor van der Pal, doctor de Vries, doctor Koopman** en **doctor Batenburg**, beste **Heleen, Andrica, Rianne** en **Lilian**, dank voor de supervisie ten tijde van de afwezigheid van Sebastian. Dank ook voor de plantechnische, secretariële en overige samenwerking, en de gezellige kletspraatjes aan **Maaike Lafeber, Magda van Korlaar**, en daarnaast **Marjolein, Marjolein, Willemijn, Diana, Suzanne, Lucienne, Herma, Wendy, Lisa, Elze, wederom Shauny, Carola, Carine, Felice, Gerda, Joke, Suzanne, Diana, Judith** en **Yvonne**.

Daarnaast wil ik ook **Trudy, Sjaan, Kirsten, Lotte, Judith van Eck** en de andere collega's van de polikliniek Endocrinologie in het Erasmus MC bedanken voor alle hulp en gezelligheid in de afgelopen jaren. Tevens veel dank aan **Karin van der Zwaan, Monique Langenberg** en **Marieke Slikker** die me altijd met veel enthousiasme met het plannen van afspraken en administratieve frustraties hebben geholpen! Much thanks to **Frau Speck** for helping out with administrative issues relating to the University Hospital Erlangen.

During all my travels for courses and national and international congresses, I have met many enthusiastic colleagues doing research and caring for patients in the field of endocrinology. I would like to thank all of them for the great times of learning together, and especially mention **professor Bengtsson, Sten Renstad, Dayakshi, Viral, Daniela, Mark, Eider, Liro, Jesper, Paul, Waiel, Pedro, Eline** and **Zografia**. This list is definitely far from complete of course, thank you all!

Mijn collega's van het endocrinologielaboratorium hebben me veel geleerd op het gebied van basale en translationeel onderzoek. Geachte **doctor Visser**, beste **Jenny**, veel respect heb ik voor jouw scherpe observaties. Je was altijd bereid om mee te denken en gaf feedback waar ik iets aan had in duidelijke taal. Tegelijkertijd zorgde je dat er sprake was van een veilige leeromgeving; dat stel(de) ik erg op prijs! **Doctor Delhanty**, beste **Patric**, wat heb ik goed 'misbruik' kunnen maken van jouw kennis en daarnaast ook schrijfadviezen, zijnde een native speaker! Ook dank aan **doctor J. Hofland** en **doctor L.C.G. de Graaf,**

Martin, Cobie, Anke, Bas en de andere collega's bij de endocrinologie die aansloten bij het overleg en de benodigde input gaven.

Beste **Gido, Loes, Keng, Karina, Margreet, Alejandra, Melitza**, ook heel erg bedankt voor alle input bij de presentaties en voor de gezellige tijd bij ENDO. Het was erg interessant om een beeld te krijgen van jullie basaal-translationele onderzoek binnen de endocrinologie. **Keng**, thank you for all your input during your time in Rotterdam. **Li** and **Yini**, I do not know you very well as you started when I was finalizing my PhD, but I hope you will have a great time of learning as well.

Natuurlijk ook veel dank aan de andere (PhD) studenten in het Erasmus MC: **Anna, Rosanne, Carlijn, Anela, Mathé, Karlijn, Deborah, Lisanne, Evert, Jelle, Myrthe, Sebastian, Eva, Wouter, Albert, Arvind, Denise, Nour**, en mijn kamergenootjes **Eva, Ammar, Ariadne** en **Komal**: wat was het fijn om te kunnen overleggen over de pro's en cons van de methodes, struggles te bespreken waaronder statistiek (met name met vraagbaak **Anna**), en samen te genieten van een kop koffie en van de congressen!

Ook met de (PhD) studenten van het Princes Máxima Centrum heb ik veel mooie momenten meegemaakt. Beste **Jiska, Camilla, Madeleine, Alissa, Vincent, Jenneke, Justine, Joeri, Mathilde, Annelot, Manon, Loes, Sanne, Demi, Eva, Pauline, Anne-Lotte, Boudewijn, Raquel, Sophie, Janna, Julie, Chris, Robin, Niki, Eline, Camilla, Winnie, Ariadne, Natanja, Emma, Annelienke, Prakriti, Julia, Lana, Marie-Christine, Richelle, Daphne** en supervisors **Martine en Saskia**. Heel erg bedankt voor de mooie tijd! Ondanks dat ik relatief weinig in Utrecht was, heb ik me toch altijd erg welkom gevoeld in de Van den Heuvel groep! (Ik hoop dat ik iedereen uit deze grote groep nu genoemd heb ;)) Alle mensen in de groep waren erg open en eerlijk, stonden altijd paraat om iemand te helpen en ideëen te opperen voor de vele onderzoeken.

Door mijn vriendengroep heb ik al lange tijd hoognodige goede afleiding kunnen vinden. Beste **Martin en Erica, Angelique en Bart, Suzanne en Robert**, dank jullie voor de gezellige avonden en weekenden weg. Jammer dat ik maar twee paranimfen mag kiezen,

anders hadden Liek en Suus met 'zjeu' de stellingen kunnen lezen! Hartelijk dank voor de hulp bij de organisatie. **Eri en Suus**, wat stoer dat jullie je ook waagden aan de wandeltrips die uitstekend waren voor onze conditie. Daarnaast heb ik ook veel gezellige momenten gehad met **Joanna en Jeffrey, Arianne, Aart, Eben en Millie, Jorrit, Annika en Sam, Remi en Anne, Eaman, Akeel, Amber, en Naciye, Daphne en Olivier en de kids**, en herinneringen op mogen halen met **Kayleigh, Sanne en Marleen**. **Joanna**, op de bruiloft heb jij je rot gerend om iedereen van lekkers te voorzien. Jij en **Liek** waren de Meesteressen van de ceremonie! Het is een heel bijzondere en mooie dag geworden.

Annika, naast veel gezelligheid heb jij ook mijn boekje vormgegeven. Wat een prachtig resultaat, dankjewel daarvoor!

Aan **Ivan Nogueira**, mijn cello docent: mijn proefschrift had vaak de prioriteit boven het oefenen met cello spelen, toch hield jij altijd eindeloos veel geduld en daagde je me uit om me ook buiten de wetenschap te ontwikkelen. Ook veel dank aan **Ingrid en Paul** die de moed hadden om een nieuwe crossfit box in de buurt op te richten (Gold Pact) en zo de mogelijkheid te creëren om samen met anderen ook aan de eigen gezondheid te blijven werken! Ook dank aan de trainers voor hun inzet daarvoor en de persoonlijke begeleiding. **Marjolein van Zanen**, naast onze vaste kapster ben jij ook onze officieuze psychotherapeut!

Aan mijn ooms en tantes (met name **tante Wil** en wijlen **ome Joop, oom Engelbert** en **tante Gerda**), nichtjes en neefjes (in het bijzonder **Kirsten**) van de families **van Santen en Jonker, Dinie van Graafeiland**, en bevriende families **Cehajic, Dzafic, van Eck/Wijnia, van Genderen/ Pander en Terborg**. Jullie hebben me een geborgen jeugd gegeven, dank daarvoor. **Joop en Arry, Jane, Dyon en Michelle**, wat was het bijzonder om onze bruiloft op jullie landgoed te mogen vieren! Het is heel belangrijk om ook buiten het werk om dit soort mooie momenten te mogen kennen.

Ook heel veel dank aan mijn schoonfamilie, aan 'de koude kant' houd ik een heel warm gevoel over! Lieve **Lida, Klaas, Aaron en Rebecca**

en Nora, Kirsten en Kasper en mini, Matthias, Ruco en Lianne Suijker, Regine, Hammi, Elias, Aiden en mini Abdali, Jantine, Michiel, Rebecca, Daniel Bos, jullie hebben me van begin af aan welkom geheten in deze grote familie en altijd veel oprechte interesse getoond in mijn promotie.

Aan mijn opa, mijn opa, mijn opa, niemand is zo aardig als hij! Mijn **lieve opa Gerrit en oma Gré**, veel dank voor jullie steun en lieve woorden. Toen ik heel klein was zijn jullie al begonnen met goed voor mij zorgen en jullie zijn nooit gestopt. Dat zal ik nooit vergeten. Opa, bij onze verhuizing was u het hoofd van de verbouwing, waardoor ik mijn promotie in een fijne omgeving kon afronden tijdens de lockdown door de pandemie. Oma, u heeft ons voorzien van goede raad en een goedgevulde maag met allerlei heerlijke huisgemaakte producten. Voor dit alles ook mijn grote dank.

Aan mijn zusje en broertje **Myrthe en Mees** (en natuurlijk aanhang **Johannes en Natascha**), ook dank aan jullie voor de steun en gezelligheid in de afgelopen jaren.

Voor mijn liefste papa en mama, beste **Henny en Wilma**, ook jullie hebben natuurlijk van kleins af aan goed voor mij gezorgd, en zonder jullie had ik dit zeker nooit en te nimmer gehaald! Al op de basisschool wilde ik dokter worden, en jullie hebben me altijd gestimuleerd om mijn dromen na te streven. Ook tijdens de studie en bij het promotietraject had ik jullie onverminderde en onvoorwaardelijke hulp en steun. Heel erg bedankt daarvoor!

Aan de liefste **Arjen**, als mijn wederhelft ben jij de belangrijkste van allemaal in dit dankwoord: mijn klankbord, mijn motivator, mijn proeflezer, mijn ontwerper, mijn grapjesmaker, mijn klimbuddy, mijn reismaatje, mijn knuffelslachtoffer en 'mijn grootste fan' (quote van jou). Jij daagt mij uit om enge dingen, zoals promoveren, toch gewoon te doen! En haalt daarmee het beste in me naar boven. Na meer dan tien jaar gelukkig met jou te zijn mag ik je nu mijn man noemen. Ik hoop dat er nog vele tientallen jaren samen mogen volgen, met veel reizen, geluk en liefde. Ik kan me geen leven zonder jou voorstellen!