

# Metabolic investigations prevent liver transplantation in two young children with citrullinemia type I

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**Abstract** Acute liver failure may be caused by a variety of disorders including inborn errors of metabolism. In those cases, rapid metabolic investigations and adequate treatment may avoid the need for liver transplantation. We report two patients who presented with acute liver failure and were referred to our center for liver transplantation work-up. Urgent metabolic investigations revealed citrullinemia type I. Treatment for citrullinemia type I avoided the

need for liver transplantation. Acute liver failure as a presentation of citrullinemia type I has not previously been reported in young children. Although acute liver failure has occasionally been described in other urea cycle disorders, these disorders may be underestimated as a cause. Timely diagnosis and treatment of these disorders may avoid liver transplantation and improve clinical outcome. Therefore, urea cycle disorders should be included in the differential diagnosis in young children presenting with acute liver failure.

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## Introduction

Acute liver failure, for which liver transplantation is considered, may be caused by a wide variety of diseases. Among these are several inborn errors of metabolism (Table 1). Recent reviews on acute liver failure in children do not mention urea cycle disorders as a cause (Squires 2008; Dhawan 2008). We describe two unrelated patients presenting with acute liver failure, who were referred for liver transplantation. Urgent metabolic work-up revealed citrullinemia type I (OMIM 215700). Treatment resulted in clinical and biochemical improvement, thereby avoiding liver transplantation. These case reports show the importance of urgent and thorough metabolic investigations in young children with acute liver failure referred for liver transplantation.

## Patient 1

Patient 1 was a female Caucasian of healthy non-consanguineous parents. After an uneventful pregnancy, the patient's mother was referred to a district hospital when fetal decelerations were noted. Vaginal labor was accelerated with vacuum- and forceps extraction. The child was

**Table 1** Inborn errors of metabolism that may present with acute liver failure and associated investigations

Disorder	Suggested investigations
Tyrosinemia type I <sup>a-c</sup>	Blood succinylacetone; urinary OA, succinylacetone, $\delta$ -aminolevulinic acid
Urea cycle disorders <sup>a,d</sup>	Blood ammonia, AA; urinary OA, purines/pyrimidines (orotic acid excretion)
Wilson's disease <sup>b,c</sup>	Blood copper and ceruloplasmin, urinary copper; consult ophthalmologist (evaluation for Kayser-Fleischer ring)
Neonatal hemochromatosis <sup>a,b</sup>	Blood ferritin, transferrin, total iron binding capacity, iron
Galactosemia <sup>a,b</sup>	Blood galactose; galactose-1-phosphate in erythrocytes; urine sugar chromatogram
Hereditary fructose intolerance <sup>a</sup>	Urinary AA, sugar chromatogram
Fatty acid oxidation disorders <sup>b,c</sup>	Blood acylcarnitine analysis, urinary OA
Mitochondrial respiratory chain deficiency <sup>a,b</sup>	Blood lactate, pyruvate, AA; urinary lactate; DNA-analysis mtDNA depletion, POLG mutations
$\alpha$ 1-antitrypsin deficiency <sup>c</sup>	Blood $\alpha$ 1-antitrypsin

AA Amino acids, OA organic acids

Sources: <sup>a</sup> Durand et al. 2001; <sup>b</sup> Squires 2008; <sup>c</sup> Narkewicz et al. 2009; <sup>d</sup> Keskinen et al. 2008; <sup>e</sup> Fregonese and Stolk 2008. Full citations provided in the References section

born at 41<sup>+1</sup> weeks gestation (birth weight 2,190 g, below Z -2.5 SD; Apgar-scores 5-6-8). The umbilical cord pH of 7.19 quickly normalized after admission to the neonatal ward. A cerebral CT revealed a right parietal fracture with edema without intracerebral hemorrhages, compatible with cerebral contusion due to complicated delivery.

At 5 days of age, the child started vomiting, and became lethargic and hypotonic. At physical examination, slow symmetric movements and absent orienting and acoustical blink reflexes were noted. She was transferred to the neonatal intensive care unit of a University Medical Center. Laboratory evaluation showed liver parameter concentrations of ALT 96 U/L (<45 U/L) and  $\gamma$ GT 306 U/L (<50 U/L) (reference values in parentheses; AST not determined). After cessation of paracetamol, liver parameters decreased to ALT 13 U/L and  $\gamma$ GT 35 U/L, respectively. Metabolic laboratory evaluations were not performed. Brain MRI revealed bilateral frontal and frontoparietal hemorrhages, with normal myelinization and anatomy. She was discharged after 16 days.

At follow-up, she grew along the -2 SD curve, and was delayed in developmental milestones. At 17 months of age, she presented with fever without clear focus, refusal of feeding, persistent vomiting, and diffuse itching. She was alert, anicteric, and without abnormalities on physical examination. Laboratory analysis showed the following results (reference values in parentheses): leukocytes  $27.5 \times 10^9/L$  ( $4.5-12.0 \times 10^9/L$ ), ESR 50 mm/h (<3-13 mm/h), hemoglobin 5.7 mmol/L (6.5-8.4 mmol/L), glucose 5.4 mmol/L (4.0-7.8 mmol/L), creatinin 22  $\mu$ mol/L (18-35  $\mu$ mol/L), urea 1.2 mmol/L (1.8-6.7 mmol/L), total bilirubin 118  $\mu$ mol/L (3-14  $\mu$ mol/L), conjugated bilirubin 109  $\mu$ mol/L (1-4  $\mu$ mol/L), albumin 21 g/L (35-55 g/L), AST 1,581 U/L (<40 U/L), ALT 2,070 U/L (<45 U/L),  $\gamma$ GT 192 U/L (<50 U/L), alkaline phosphatase 601 U/L

(55-250 U/L), LDH 1,087 U/L (<200 U/L), PTT 62 s (9-12 s), and APTT 62 s (23-33 s). Arterial blood gas analysis showed pH 7.45 (7.35-7.45), pCO<sub>2</sub> 5.2 kPa (4.6-6.0 kPa), HCO<sub>3</sub><sup>-</sup> 26.6 mmol/L (21-25 mmol/L), and BE +2.7 (-3.0 to +3.0). NH<sub>3</sub> concentration was 114  $\mu$ mol/L (15-45  $\mu$ mol/L). Abdominal ultrasound revealed no abnormalities.

An underlying metabolic disorder was suspected and the child was eventually referred to the University Medical Center Groningen (UMCG), so that, if necessary, liver transplantation could be performed as soon as possible. Metabolic analysis revealed glutamine 1,184  $\mu$ mol/L (335-810  $\mu$ mol/L), citrulline 1,442  $\mu$ mol/L (<50  $\mu$ mol/L), and arginine 23  $\mu$ mol/L (10-110  $\mu$ mol/L), and an urinary orotic acid excretion of 656  $\mu$ mol/mmol creatinine (undetectable in healthy subjects). She was diagnosed with citrullinemia type I. Treatment was started with intravenous infusion of glucose, L-arginine, and oral sodium benzoate. Two days later, enteral feeding with restriction of natural protein intake was started. Concentrations of ammonia and transaminases normalised within a few days and 3 weeks, respectively. The child showed a marked clinical improvement. Incorporation of <sup>14</sup>C-citrulline in fibroblasts was reduced to 7% of intra-assay control. DNA analyses are pending.

## Patient 2

Patient 2 was a male Caucasian born to non-consanguineous parents. Patient 2 was born in the same district hospital as patient 1, at 38<sup>+1</sup> weeks after an uneventful gestation and delivery, with birth weight 2,680 g (Z -1.3 SD), and Apgar scores 9 and 10.

At 22 months of age, he presented with episodic vomiting since 3 months, not obviously related to feeding,

with a frequency varying from daily to weekly. Further history revealed increased fatigue, and a mild growth retardation since the age of 8 months. Concerns were raised about his development, since he was not able to walk without support nor to throw a ball without losing balance. At physical examination, the child was well and without abnormalities. Laboratory results revealed leukocytes  $10.7 \times 10^9/L$ , thrombocytes  $378 \times 10^9/L$  ( $150\text{--}350 \times 10^9/L$ ), Hb 7.2 mmol/L, glucose 4.9 mmol/L, Na + 138 mmol/L (138–146 mmol/L), K + 4.3 mmol/L (4.1–5.5 mmol/L), Ca<sup>2+</sup> + 2.38 mmol/L (2.20–2.60 mmol/L), phosphate 1.58 mmol/L (1.45–2.10 mmol/L), albumin 33 g/L, AST 206 U/L, ALT 323 U/L, and urea 2.5 mmol/L, capillary blood gas analysis pH 7.47 (7.32–7.42), pCO<sub>2</sub> 3.8 kPa (5.5–6.8 kPa), HCO<sub>3</sub><sup>-</sup> 20.2 mmol/L (24–28 mmol/L), and BE -2.1 mmol/L (-3.0 to +3.0), with NH<sub>3</sub> 210 μmol/L. Abdominal X-ray, barium enema, abdominal ultrasound, screening results of viral hepatitis and toxoplasmosis, and triple feces test were normal.

The patient was referred to an University Medical Center considering an urea cycle disorder. As liver function parameters deteriorated, the patient was referred the same day to the UMCG for liver transplantation. Laboratory evaluations revealed lactate 2.1 mmol/L (<2.0 mmol/L), albumin 37 g/L, AST 757 U/L, ALT 1,315 U/L, γGT 38 U/L, alkaline phosphatase 285 U/L, INR 1.6 (0.8–1.3), APTT 35 s, arterial blood gas analysis pH 7.38, with pCO<sub>2</sub> 4.8 kPa, and HCO<sub>3</sub><sup>-</sup> 21 mmol/L, NH<sub>3</sub> 210 μmol/L, glutamine 539 μmol/L, citrulline 1,963 μmol/L, and arginine 51 μmol/L. Urine analysis revealed an orotic acid excretion of 1,315 μmol/mmol creatinine. The boy was diagnosed with citrullinemia type I. Treatment was started with intravenous infusion of glucose and L-arginine, oral sodium benzoate, and restriction of natural protein intake. Within 2 days, the child improved markedly, and blood concentrations of ammonia and liver parameters normalized. Incorporation of <sup>14</sup>C-citrulline in fibroblasts was reduced to 14% of intra-assay control. DNA analysis revealed the c.685del8bp (fs232X) and c.815G > A (p.R272H) mutations in the *ASS*-gene.

## Discussion

To our knowledge, these two Caucasian patients with citrullinemia type I are the first two presenting with acute liver failure for which liver transplantation was considered. Thus far, acute liver failure as a presentation of citrullinemia type I has been reported once, in a patient diagnosed during pregnancy (Dimmock et al. 2008). Pediatric acute liver failure may be defined as “hepatic necrosis resulting in loss of liver function within weeks or a few months of the onset of clinical liver disease” (Whittington et al. 2008). In our patients, synthetic liver function was compromised, as evident from

the coagulopathy and hypoalbuminemia present in both, within the time frame mentioned in the definition. This validates the diagnosis of acute liver failure in our patients.

Citrullinemia type I is caused by argininosuccinate synthetase deficiency (E.C. 6.3.4.5), one of the six enzymes that constitute the urea cycle. Urea cycle disorders usually present with feeding problems, lethargy, convulsions, coma, and death during the first days of life (Brusilow and Horwich 2001). In addition, milder presentations at a later age with failure to thrive, neurological symptoms, developmental delay and/or behavioral disorders have also been described (Summar and Tuchman 2001).

In urea cycle disorders, acute liver failure is not generally considered to be a presenting symptom (Saudubray et al. 2002; Endo et al. 2004; Olpin 2010). However, in contrast to a statement made in a recent report (Olpin 2010), it should be acknowledged that urea cycle disorders can present with elevated transaminases and other biochemical signs of liver pathology. In urea cycle disorders other than citrullinemia type I, acute liver failure has occasionally been reported as the presenting symptom (Keskinen et al. 2008; Durand et al. 2001). The pathogenesis of liver dysfunction in urea cycle defects is unknown. It seems to be unrelated to the specific enzymatic defect, intermediary metabolites, or the number of hyperammonemic episodes, and may occur without concomitant hyperammonemia (Zamora et al. 1997; Kleijer et al. 2002; Mori et al. 2002).

Retrospectively, one might suggest that the neurological deterioration at day 5 in patient 1 was related to citrullinemia type I. Blood ammonia was never measured in this period, but she improved without any specific treatment, and also, retrospectively, the cerebral MRI performed at that time did not show abnormalities consistent with citrullinemia type I.

In this report, we emphasize urea cycle disorders as a cause of acute liver failure, particularly citrullinemia type I. In 40–50% of patients presenting with acute liver failure, an etiological diagnosis is not made (Squires 2008; Dhawan 2008). Urea cycle disorders may be underestimated as a cause of acute liver failure. Our case report shows that citrullinemia type I should be included in the differential diagnosis of children presenting with acute liver failure. The presentations of these two infants stress the importance of urgent and thorough metabolic investigations in such patients. Importantly, hyperammonemia in children with acute liver failure may be caused by urea cycle disorders, rather than be secondary to liver failure. An overview of inborn errors of metabolism that may lead to acute liver failure and associated investigations is presented in Table 1.

In conclusion, acute liver failure in children may be caused by citrullinemia type I and other inborn errors of metabolism. Adequate diagnosis and treatment may prevent progression of liver failure and liver transplantation.

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