

References

1. Darwin, E., *Phytologia*, 1800. **115**: p. 103.
2. Sarre, S.D., T. Ezaz, and A. Georges, *Transitions between sex-determining systems in reptiles and amphibians*. *Annu Rev Genomics Hum Genet*, 2011. **12**: p. 391-406.
3. Rhen, T. and A. Schroeder, *Molecular mechanisms of sex determination in reptiles*. *Sex Dev*, 2010. **4**(1-2): p. 16-28.
4. Ohno, S., *Sex chromosomes and Sex-linked genes*. 1967, Berlin: Springer.
5. Muller, H.J., *A gene for the fourth chromosome in Drosophila*. *J. Exp. Zool.*, 1914. **17**: p. 325-336.
6. Charlesworth, B., *The evolution of sex chromosomes*. *Science*, 1991. **251**(4997): p. 1030-3.
7. Graves, J.A., *The origin and function of the mammalian Y chromosome and Y-borne genes--an evolving understanding*. *Bioessays*, 1995. **17**(4): p. 311-20.
8. Graves, J.A., E. Koina, and N. Sankovic, *How the gene content of human sex chromosomes evolved*. *Curr Opin Genet Dev*, 2006. **16**(3): p. 219-24.
9. Graves, J.A., *Sex chromosome specialization and degeneration in mammals*. *Cell*, 2006. **124**(5): p. 901-14.
10. Marshall Graves, J.A., *Weird animal genomes and the evolution of vertebrate sex and sex chromosomes*. *Annu Rev Genet*, 2008. **42**: p. 565-86.
11. Sutton, E., et al., *Identification of SOX3 as an XX male sex reversal gene in mice and humans*. *J Clin Invest*, 2011. **121**(1): p. 328-41.
12. Skaletsky, H., et al., *The male-specific region of the human Y chromosome is a mosaic of discrete sequence classes*. *Nature*, 2003. **423**(6942): p. 825-37.
13. Koopman, P., et al., *Male development of chromosomally female mice transgenic for Sry*. *Nature*, 1991. **351**(6322): p. 117-21.
14. Ross, M.T., et al., *The DNA sequence of the human X chromosome*. *Nature*, 2005. **434**(7031): p. 325-37.
15. Wang, P.J., et al., *An abundance of X-linked genes expressed in spermatogonia*. *Nat Genet*, 2001. **27**(4): p. 422-6.
16. Zhang, Y.E., et al., *Chromosomal redistribution of male-biased genes in mammalian evolution with two bursts of gene gain on the X chromosome*. *PLoS Biol*, 2010. **8**(10).
17. Khil, P.P., et al., *The mouse X chromosome is enriched for sex-biased genes not subject to selection by meiotic sex chromosome inactivation*. *Nat Genet*, 2004. **36**(6): p. 642-6.
18. Mueller, J.L., et al., *The mouse X chromosome is enriched for multicopy testis genes showing postmeiotic expression*. *Nat Genet*, 2008. **40**(6): p. 794-9.
19. Saifi, G.M. and H.S. Chandra, *An apparent excess of sex- and reproduction-related genes on the human X chromosome*. *Proc Biol Sci*, 1999. **266**(1415): p. 203-9.
20. Bellott, D.W., et al., *Convergent evolution of chicken Z and human X chromosomes by expansion and gene acquisition*. *Nature*, 2010. **466**(7306): p. 612-6.
21. Delgado, C.L., et al., *Physical mapping of the elephant X chromosome: conservation of gene order over 105 million years*. *Chromosome Res*, 2009. **17**(7): p. 917-26.
22. Mohandas, T.K., et al., *Role of the pseudoautosomal region in sex-chromosome pairing during male meiosis: meiotic studies in a man with a deletion of distal Xp*. *Am J Hum Genet*, 1992. **51**(3): p. 526-33.
23. Burgoyne, P.S., et al., *Fertility in mice requires X-Y pairing and a Y-chromosomal "spermiogenesis" gene mapping to the long arm*. *Cell*, 1992. **71**(3): p. 391-8.
24. Helena Mangs, A. and B.J. Morris, *The Human Pseudoautosomal Region (PAR): Origin, Function and Future*. *Curr Genomics*, 2007. **8**(2): p. 129-36.
25. Freije, D., et al., *Identification of a second pseudoautosomal region near the Xq and Yq telomeres*. *Science*, 1992. **258**(5089): p. 1784-7.

References

26. Kuhl, H., et al., *Loss of the Y chromosomal PAR2-region in four familial cases of satellited Y chromosomes (Yqs)*. Chromosome Res, 2001. **9**(3): p. 215-22.
27. Birchler, J.A., et al., *Dosage balance in gene regulation: biological implications*. Trends Genet, 2005. **21**(4): p. 219-26.
28. Velitia, R.A., S. Bottani, and J.A. Birchler, *Cellular reactions to gene dosage imbalance: genomic, transcriptomic and proteomic effects*. Trends Genet, 2008. **24**(8): p. 390-7.
29. Deng, X. and C.M. Disteché, *Genomic responses to abnormal gene dosage: the X chromosome improved on a common strategy*. PLoS Biol, 2010. **8**(2): p. e1000318.
30. Lyon, M.F., *Gene action in the X-chromosome of the mouse (Mus musculus L.)*. Nature, 1961. **190**: p. 372-3.
31. Lyon, M.F., R.J. Phillips, and A.G. Searle, *A test for mutagenicity of caffeine in mice*. Z Vererbungsl, 1962. **93**: p. 7-13.
32. Lyon, M.F., *Sex chromatin and gene action in the mammalian X-chromosome*. Am J Hum Genet, 1962. **14**: p. 135-48.
33. Davidson, R.G., H.M. Nitowsky, and B. Childs, *Demonstration of Two Populations of Cells in the Human Female Heterozygous for Glucose-6-Phosphate Dehydrogenase Variants*. Proc Natl Acad Sci U S A, 1963. **50**: p. 481-5.
34. Shapiro, L.J., et al., *Non-inactivation of an x-chromosome locus in man*. Science, 1979. **204**(4398): p. 1224-6.
35. Prothero, K.E., J.M. Stahl, and L. Carrel, *Dosage compensation and gene expression on the mammalian X chromosome: one plus one does not always equal two*. Chromosome Res, 2009. **17**(5): p. 637-48.
36. Adler, D.A., et al., *Evidence of evolutionary up-regulation of the single active X chromosome in mammals based on Clc4 expression levels in Mus spretus and Mus musculus*. Proc Natl Acad Sci U S A, 1997. **94**(17): p. 9244-8.
37. Nguyen, D.K. and C.M. Disteché, *Dosage compensation of the active X chromosome in mammals*. Nat Genet, 2006. **38**(1): p. 47-53.
38. Lin, H., et al., *Dosage compensation in the mouse balances up-regulation and silencing of X-linked genes*. PLoS Biol, 2007. **5**(12): p. e326.
39. Birchler, J.A., H.R. Fernandez, and H.H. Kavi, *Commonalities in compensation*. Bioessays, 2006. **28**(6): p. 565-8.
40. Johnston, C.M., et al., *Large-scale population study of human cell lines indicates that dosage compensation is virtually complete*. PLoS Genet, 2008. **4**(1): p. e9.
41. Xiong, Y., et al., *RNA sequencing shows no dosage compensation of the active X-chromosome*. Nat Genet, 2010. **42**(12): p. 1043-7.
42. Casci, T., *What dosage compensation?* Nat Rev Genet, 2011. **12**(1): p. 2.
43. Deng, X., et al., *Evidence for compensatory upregulation of expressed X-linked genes in mammals, Caenorhabditis elegans and Drosophila melanogaster*. Nat Genet, 2011.
44. Lin, H., et al., *Relative overexpression of X-linked genes in mouse embryonic stem cells is consistent with Ohno's hypothesis*. Nat Genet, 2011. **43**(12): p. 1169-70.
45. Pessia, E., et al., *Mammalian X chromosome inactivation evolved as a dosage-compensation mechanism for dosage-sensitive genes on the X chromosome*. Proc Natl Acad Sci U S A, 2012. **109**(14): p. 5346-51.
46. Gelbart, M.E. and M.I. Kuroda, *Drosophila dosage compensation: a complex voyage to the X chromosome*. Development, 2009. **136**(9): p. 1399-410.
47. Straub, T. and P.B. Becker, *Dosage compensation: the beginning and end of generalization*. Nat Rev Genet, 2007. **8**(1): p. 47-57.
48. Macner, S., M. Muller, and P.B. Becker, *Roles of long, non-coding RNA in chromosome-wide transcription regulation: Lessons from two dosage compensation systems*. Biochimie, 2012.
49. Meyer, B.J., et al., *Sex and X-chromosome-wide repression in Caenorhabditis elegans*. Cold Spring Harb Symp Quant Biol, 2004. **69**: p. 71-9.
50. Meyer, B.J., *Targeting X chromosomes for repression*. Curr Opin Genet Dev, 2010. **20**(2): p. 179-89.
51. Huynh, K.D. and J.T. Lee, *Inheritance of a pre-inactivated paternal X chromosome in early mouse embryos*. Nature, 2003. **426**(6968): p. 857-62.
52. Okamoto, I., et al., *Epigenetic dynamics of imprinted X inactivation during early mouse development*. Science, 2004. **303**(5658): p. 644-9.

References

53. Takagi, N. and M. Sasaki, *Preferential inactivation of the paternally derived X chromosome in the extraembryonic membranes of the mouse*. *Nature*, 1975. **256**(5519): p. 640-2.
54. Mak, W., et al., *Reactivation of the paternal X chromosome in early mouse embryos*. *Science*, 2004. **303**(5658): p. 666-9.
55. Evans, M.J. and M.H. Kaufman, *Establishment in culture of pluripotent cells from mouse embryos*. *Nature*, 1981. **292**(5819): p. 154-6.
56. Martin, G.R., *Isolation of a pluripotent cell line from early mouse embryos cultured in medium conditioned by teratocarcinoma stem cells*. *Proc Natl Acad Sci U S A*, 1981. **78**(12): p. 7634-8.
57. Thomson, J.A., et al., *Embryonic stem cell lines derived from human blastocysts*. *Science*, 1998. **282**(5391): p. 1145-7.
58. Keller, G., *Embryonic stem cell differentiation: emergence of a new era in biology and medicine*. *Genes Dev*, 2005. **19**(10): p. 1129-55.
59. Leahy, A., et al., *Use of developmental marker genes to define temporal and spatial patterns of differentiation during embryoid body formation*. *J Exp Zool*, 1999. **284**(1): p. 67-81.
60. Monk, M., *A stem-line model for cellular and chromosomal differentiation in early mouse-development*. *Differentiation*, 1981. **19**(2): p. 71-6.
61. Takahashi, K. and S. Yamanaka, *Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors*. *Cell*, 2006. **126**(4): p. 663-76.
62. Nakagawa, M., et al., *Generation of induced pluripotent stem cells without Myc from mouse and human fibroblasts*. *Nat Biotechnol*, 2008. **26**(1): p. 101-6.
63. Wernig, M., et al., *In vitro reprogramming of fibroblasts into a pluripotent ES-cell-like state*. *Nature*, 2007. **448**(7151): p. 318-24.
64. Yu, J., et al., *Induced pluripotent stem cell lines derived from human somatic cells*. *Science*, 2007. **318**(5858): p. 1917-20.
65. Takahashi, K., et al., *Induction of pluripotent stem cells from adult human fibroblasts by defined factors*. *Cell*, 2007. **131**(5): p. 861-72.
66. Park, J.H., et al., *Reprogramming of human somatic cells to pluripotency with defined factors*. *Nature*, 2008. **451**(7175): p. 141-6.
67. Maherali, N., et al., *Directly reprogrammed fibroblasts show global epigenetic remodeling and widespread tissue contribution*. *Cell Stem Cell*, 2007. **1**(1): p. 55-70.
68. Stadtfeld, M., et al., *Defining molecular cornerstones during fibroblast to iPS cell reprogramming in mouse*. *Cell Stem Cell*, 2008. **2**(3): p. 230-40.
69. Tam, P.P., S.X. Zhou, and S.S. Tan, *X-chromosome activity of the mouse primordial germ cells revealed by the expression of an X-linked lacZ transgene*. *Development*, 1994. **120**(10): p. 2925-32.
70. Turner, J.M., *Meiotic sex chromosome inactivation*. *Development*, 2007. **134**(10): p. 1823-31.
71. Russell, L.B., *Mammalian X-chromosome action: inactivation limited in spread and region of origin*. *Science*, 1963. **140**: p. 976-8.
72. Rastan, S., *Non-random X-chromosome inactivation in mouse X-autosome translocation embryos--location of the inactivation centre*. *J Embryol Exp Morphol*, 1983. **78**: p. 1-22.
73. Rastan, S. and E.J. Robertson, *X-chromosome deletions in embryo-derived (EK) cell lines associated with lack of X-chromosome inactivation*. *J Embryol Exp Morphol*, 1985. **90**: p. 379-88.
74. Brown, C.J., et al., *A gene from the region of the human X inactivation centre is expressed exclusively from the inactive X chromosome*. *Nature*, 1991. **349**(6304): p. 38-44.
75. Therman, E., G.E. Sarto, and K. Patau, *Center for Barr body condensation on the proximal part of the human Xq: a hypothesis*. *Chromosoma*, 1974. **44**(4): p. 361-6.
76. Therman, E., et al., *Position of the human X inactivation center on Xq*. *Hum Genet*, 1979. **50**(1): p. 59-64.
77. Pettigrew, A.L., et al., *Isodicentric X chromosome in a patient with Turner syndrome--implications for localization of the X-inactivation center*. *Hum Genet*, 1991. **87**(4): p. 498-502.
78. Mattei, M.G., et al., *Structural anomalies of the X chromosome and inactivation center*. *Hum Genet*, 1981. **56**(3): p. 401-8.
79. Flejter, W.L., D.L. Van Dyke, and L. Weiss, *Bends in human mitotic metaphase chromosomes, including a bend marking the X-inactivation center*. *Am J Hum Genet*, 1984. **36**(1): p. 218-26.
80. Flejter, W.L., D.L. Van Dyke, and L. Weiss, *Location of the X inactivation center in primates and other mammals*. *Hum Genet*, 1986. **74**(1): p. 63-6.
81. Brown, C.J., et al., *Localization of the X inactivation centre on the human X chromosome in Xq13*. *Nature*, 1991. **349**(6304): p. 82-4.

References

82. Rastan, S. and S.D. Brown, *The search for the mouse X-chromosome inactivation centre*. Genet Res, 1990. **56**(2-3): p. 99-106.
83. Heard, E. and P. Avner, *Role play in X-inactivation*. Hum Mol Genet, 1994. **3 Spec No**: p. 1481-5.
84. Borsani, G., et al., *Characterization of a murine gene expressed from the inactive X chromosome*. Nature, 1991. **351**(6324): p. 325-9.
85. Brockdorff, N., et al., *Conservation of position and exclusive expression of mouse Xist from the inactive X chromosome*. Nature, 1991. **351**(6324): p. 329-31.
86. Brockdorff, N., et al., *The product of the mouse Xist gene is a 15 kb inactive X-specific transcript containing no conserved ORF and located in the nucleus*. Cell, 1992. **71**(3): p. 515-26.
87. Brown, C.J., et al., *The human XIST gene: analysis of a 17 kb inactive X-specific RNA that contains conserved repeats and is highly localized within the nucleus*. Cell, 1992. **71**(3): p. 527-42.
88. Sheardown, S.A., et al., *Stabilization of Xist RNA mediates initiation of X chromosome inactivation*. Cell, 1997. **91**(1): p. 99-107.
89. Sun, B.K., A.M. Deaton, and J.T. Lee, *A transient heterochromatic state in Xist preempts X inactivation choice without RNA stabilization*. Mol Cell, 2006. **21**(5): p. 617-28.
90. Panning, B., J. Dausman, and R. Jaenisch, *X chromosome inactivation is mediated by Xist RNA stabilization*. Cell, 1997. **90**(5): p. 907-16.
91. Clemson, C.M., et al., *XIST RNA paints the inactive X chromosome at interphase: evidence for a novel RNA involved in nuclear/chromosome structure*. J Cell Biol, 1996. **132**(3): p. 259-75.
92. Jonkers, I., et al., *Xist RNA is confined to the nuclear territory of the silenced X chromosome throughout the cell cycle*. Mol Cell Biol, 2008. **28**(18): p. 5583-94.
93. Penny, G.D., et al., *Requirement for Xist in X chromosome inactivation*. Nature, 1996. **379**(6561): p. 131-7.
94. Marahrens, Y., et al., *Xist-deficient mice are defective in dosage compensation but not spermatogenesis*. Genes Dev, 1997. **11**(2): p. 156-66.
95. Csankovszki, G., et al., *Conditional deletion of Xist disrupts histone macroH2A localization but not maintenance of X inactivation*. Nat Genet, 1999. **22**(4): p. 323-4.
96. Kay, G.F., et al., *Expression of Xist during mouse development suggests a role in the initiation of X chromosome inactivation*. Cell, 1993. **72**(2): p. 171-82.
97. Plath, K., et al., *Role of histone H3 lysine 27 methylation in X inactivation*. Science, 2003. **300**(5616): p. 131-5.
98. Silva, J., et al., *Establishment of histone h3 methylation on the inactive X chromosome requires transient recruitment of Eed-Enx1 polycomb group complexes*. Dev Cell, 2003. **4**(4): p. 481-95.
99. Zhao, J., et al., *Polycomb proteins targeted by a short repeat RNA to the mouse X chromosome*. Science, 2008. **322**(5902): p. 750-6.
100. Pillet, N., C. Bonny, and D.F. Schorderet, *Characterization of the promoter region of the mouse Xist gene*. Proc Natl Acad Sci U S A, 1995. **92**(26): p. 12515-9.
101. Sheardown, S.A., et al., *Regulatory elements in the minimal promoter region of the mouse Xist gene*. Gene, 1997. **203**(2): p. 159-68.
102. Komura, J., et al., *In vivo ultraviolet and dimethyl sulfate footprinting of the 5' region of the expressed and silent Xist alleles*. J Biol Chem, 1997. **272**(16): p. 10975-80.
103. Newall, A.E., et al., *Primary non-random X inactivation associated with disruption of Xist promoter regulation*. Hum Mol Genet, 2001. **10**(6): p. 581-9.
104. Nesterova, T.B., et al., *Skewing X chromosome choice by modulating sense transcription across the Xist locus*. Genes Dev, 2003. **17**(17): p. 2177-90.
105. Korotkova, A.M., E.A. Elisafenko, and S.M. Zakiiian, *[Functional analysis of the Xist promoter region in mouse Mus musculus]*. Genetika, 2011. **47**(1): p. 140-4.
106. Johnston, C.M., et al., *Developmentally regulated Xist promoter switch mediates initiation of X inactivation*. Cell, 1998. **94**(6): p. 809-17.
107. Warshawsky, D., N. Stavropoulos, and J.T. Lee, *Further examination of the Xist promoter-switch hypothesis in X inactivation: evidence against the existence and function of a P(0) promoter*. Proc Natl Acad Sci U S A, 1999. **96**(25): p. 14424-9.
108. Hendrich, B.D., R.M. Plenge, and H.F. Willard, *Identification and characterization of the human XIST gene promoter: implications for models of X chromosome inactivation*. Nucleic Acids Res, 1997. **25**(13): p. 2661-71.
109. Norris, D.P., et al., *Evidence that random and imprinted Xist expression is controlled by preemptive methylation*. Cell, 1994. **77**(1): p. 41-51.

References

110. Hendrich, B.D., C.J. Brown, and H.F. Willard, *Evolutionary conservation of possible functional domains of the human and murine XIST genes*. Hum Mol Genet, 1993. **2**(6): p. 663-72.
111. Navarro, P., et al., *Tsix transcription across the Xist gene alters chromatin conformation without affecting Xist transcription: implications for X-chromosome inactivation*. Genes Dev, 2005. **19**(12): p. 1474-84.
112. Ohhata, T., et al., *Crucial role of antisense transcription across the Xist promoter in Tsix-mediated Xist chromatin modification*. Development, 2008. **135**(2): p. 227-35.
113. Panning, B. and R. Jaenisch, *DNA hypomethylation can activate Xist expression and silence X-linked genes*. Genes Dev, 1996. **10**(16): p. 1991-2002.
114. Beard, C., E. Li, and R. Jaenisch, *Loss of methylation activates Xist in somatic but not in embryonic cells*. Genes Dev, 1995. **9**(19): p. 2325-34.
115. Wutz, A., T.P. Rasmussen, and R. Jaenisch, *Chromosomal silencing and localization are mediated by different domains of Xist RNA*. Nat Genet, 2002. **30**(2): p. 167-74.
116. Elisaphenko, E.A., et al., *A dual origin of the Xist gene from a protein-coding gene and a set of transposable elements*. PLoS ONE, 2008. **3**(6): p. e2521.
117. Nesterova, T.B., et al., *Characterization of the genomic Xist locus in rodents reveals conservation of overall gene structure and tandem repeats but rapid evolution of unique sequence*. Genome Res, 2001. **11**(5): p. 833-49.
118. Lee, J.T., L.S. Davidow, and D. Warshawsky, *Tsix, a gene antisense to Xist at the X-inactivation centre*. Nat Genet, 1999. **21**(4): p. 400-4.
119. Lee, J.T. and N. Lu, *Targeted mutagenesis of Tsix leads to nonrandom X inactivation*. Cell, 1999. **99**(1): p. 47-57.
120. Clerc, P. and P. Avner, *Role of the region 3' to Xist exon 6 in the counting process of X-chromosome inactivation*. Nat Genet, 1998. **19**(3): p. 249-53.
121. Shibata, S. and J.T. Lee, *Characterization and quantitation of differential Tsix transcripts: implications for Tsix function*. Hum Mol Genet, 2003. **12**(2): p. 125-36.
122. Marks, H., et al., *High-resolution analysis of epigenetic changes associated with X inactivation*. Genome Res, 2009. **19**(8): p. 1361-73.
123. Luikenhuis, S., A. Wutz, and R. Jaenisch, *Antisense transcription through the Xist locus mediates Tsix function in embryonic stem cells*. Mol Cell Biol, 2001. **21**(24): p. 8512-20.
124. Ohhata, T., et al., *Lineage-specific function of the noncoding Tsix RNA for Xist repression and Xi reactivation in mice*. Genes Dev, 2011. **25**(16): p. 1702-15.
125. Navarro, P., et al., *Tsix-mediated epigenetic switch of a CTCF-flanked region of the Xist promoter determines the Xist transcription program*. Genes Dev, 2006. **20**(20): p. 2787-92.
126. Sado, T., Y. Hoki, and H. Sasaki, *Tsix silences Xist through modification of chromatin structure*. Dev Cell, 2005. **9**(1): p. 159-65.
127. Vigneau, S., et al., *An essential role for the DXPas34 tandem repeat and Tsix transcription in the counting process of X chromosome inactivation*. Proc Natl Acad Sci U S A, 2006. **103**(19): p. 7390-5.
128. Cohen, D.E., et al., *The DXPas34 repeat regulates random and imprinted X inactivation*. Dev Cell, 2007. **12**(1): p. 57-71.
129. Debrand, E., et al., *Functional analysis of the DXPas34 locus, a 3' regulator of Xist expression*. Mol Cell Biol, 1999. **19**(12): p. 8513-25.
130. Prissette, M., et al., *Methylation profiles of DXPas34 during the onset of X-inactivation*. Hum Mol Genet, 2001. **10**(1): p. 31-8.
131. Boumil, R.M., et al., *Differential methylation of Xite and CTCF sites in Tsix mirrors the pattern of X-inactivation choice in mice*. Mol Cell Biol, 2006. **26**(6): p. 2109-17.
132. Ogawa, Y., B.K. Sun, and J.T. Lee, *Intersection of the RNA interference and X-inactivation pathways*. Science, 2008. **320**(5881): p. 1336-41.
133. Nesterova, T.B., et al., *Dicer regulates Xist promoter methylation in ES cells indirectly through transcriptional control of Dnmt3a*. Epigenetics Chromatin, 2008. **1**(1): p. 2.
134. Kanellopoulou, C., et al., *X chromosome inactivation in the absence of Dicer*. Proc Natl Acad Sci U S A, 2009. **106**(4): p. 1122-7.
135. Shibata, S. and J.T. Lee, *Tsix transcription- versus RNA-based mechanisms in Xist repression and epigenetic choice*. Curr Biol, 2004. **14**(19): p. 1747-54.
136. Ogawa, Y. and J.T. Lee, *Xite, X-inactivation intergenic transcription elements that regulate the probability of choice*. Mol Cell, 2003. **11**(3): p. 731-43.

References

137. Stavropoulos, N., R.K. Rowntree, and J.T. Lee, *Identification of developmentally specific enhancers for Tsix in the regulation of X chromosome inactivation*. Mol Cell Biol, 2005. **25**(7): p. 2757-69.
138. Rogner, U.C., et al., *Control of neurulation by the nucleosome assembly protein-1-like 2*. Nat Genet, 2000. **25**(4): p. 431-5.
139. Rougeulle, C. and P. Avner, *Cloning and characterization of a murine brain specific gene Bpx and its human homologue lying within the Xic candidate region*. Hum Mol Genet, 1996. **5**(1): p. 41-9.
140. Chureau, C., et al., *Comparative sequence analysis of the X-inactivation center region in mouse, human, and bovine*. Genome Res, 2002. **12**(6): p. 894-908.
141. Gamcr, L.W. and C.V. Wright, *Murine Cdx-4 bears striking similarities to the Drosophila caudal gene in its homeodomain sequence and early expression pattern*. Mech Dev, 1993. **43**(1): p. 71-81.
142. Simmler, M.C., et al., *Localization and expression analysis of a novel conserved brain expressed transcript, Brx/BRX, lying within the Xic/XIC candidate region*. Mamm Genome, 1997. **8**(10): p. 760-6.
143. Simmler, M.C., et al., *A 94 kb genomic sequence 3' to the murine Xist gene reveals an AT rich region containing a new testis specific gene Tsx*. Hum Mol Genet, 1996. **5**(11): p. 1713-26.
144. Cunningham, D.B., et al., *The mouse Tsx gene is expressed in Sertoli cells of the adult testis and transiently in premeiotic germ cells during puberty*. Dev Biol, 1998. **204**(2): p. 345-60.
145. Sebastiano, V., et al., *Cloned pre-implantation mouse embryos show correct timing but altered levels of gene expression*. Mol Reprod Dev, 2005. **70**(2): p. 146-54.
146. Johnston, C.M., et al., *Enox, a novel gene that maps 10 kb upstream of Xist and partially escapes X inactivation*. Genomics, 2002. **80**(2): p. 236-44.
147. Chow, J.C., et al., *Characterization of expression at the human XIST locus in somatic, embryonal carcinoma, and transgenic cell lines*. Genomics, 2003. **82**(3): p. 309-22.
148. Lafreniere, R.G., L. Carrel, and H.F. Willard, *A novel transmembrane transporter encoded by the XPCT gene in Xq13.2*. Hum Mol Genet, 1994. **3**(7): p. 1133-9.
149. Debrand, E., E. Heard, and P. Avner, *Cloning and localization of the murine Xpct gene: evidence for complex rearrangements during the evolution of the region around the Xist gene*. Genomics, 1998. **48**(3): p. 296-303.
150. Friesema, E.C., et al., *Identification of monocarboxylate transporter 8 as a specific thyroid hormone transporter*. J Biol Chem, 2003. **278**(41): p. 40128-35.
151. Schwartz, C.E., et al., *Allan-Herndon-Dudley syndrome and the monocarboxylate transporter 8 (MCT8) gene*. Am J Hum Genet, 2005. **77**(1): p. 41-53.
152. Friesema, E.C., et al., *Association between mutations in a thyroid hormone transporter and severe X-linked psychomotor retardation*. Lancet, 2004. **364**(9443): p. 1435-7.
153. Koina, E., et al., *Isolation, X location and activity of the marsupial homologue of SLC16A2, an XIST-flanking gene in eutherian mammals*. Chromosome Res, 2005. **13**(7): p. 687-98.
154. Sugiyama, M., et al., *Overexpression of MCT8 enhances the differentiation of ES cells into neural progenitors*. Biochem Biophys Res Commun, 2007. **360**(4): p. 741-5.
155. Trajkovic, M., et al., *Abnormal thyroid hormone metabolism in mice lacking the monocarboxylate transporter 8*. J Clin Invest, 2007. **117**(3): p. 627-35.
156. Heard, E., et al., *Methylation of histone H3 at Lys-9 is an early mark on the X chromosome during X inactivation*. Cell, 2001. **107**(6): p. 727-38.
157. Rougeulle, C., et al., *Differential histone H3 Lys-9 and Lys-27 methylation profiles on the X chromosome*. Mol Cell Biol, 2004. **24**(12): p. 5475-84.
158. Jacobs, P.A., et al., *Evidence for the existence of the human "super female"*. Lancet, 1959. **2**(7100): p. 423-5.
159. Maclean, N. and J.M. Mitchell, *A survey of sex-chromosome abnormalities among 4514 mental defectives*. Lancet, 1962. **1**(7224): p. 293-6.
160. Grumbach, M.M., A. Morishima, and J.H. Taylor, *Human Sex Chromosome Abnormalities in Relation to DNA Replication and Heterochromatinization*. Proc Natl Acad Sci U S A, 1963. **49**(5): p. 581-9.
161. Carr, D.H., *Chromosome studies in selected spontaneous abortions. 1. Conception after oral contraceptives*. Can Med Assoc J, 1970. **103**(4): p. 343-8.
162. Webb, S., T.J. de Vries, and M.H. Kaufman, *The differential staining pattern of the X chromosome in the embryonic and extraembryonic tissues of postimplantation homozygous tetraploid mouse embryos*. Genet Res, 1992. **59**(3): p. 205-14.
163. Hamden, D.G., *Nuclear sex in triploid XXY human cells*. Lancet, 1961. **2**(7200): p. 488.
164. Nicodemi, M. and A. Prisco, *Symmetry-breaking model for X-chromosome inactivation*. Phys Rev Lett, 2007. **98**(10): p. 108104.

References

165. Nicodemi, M. and A. Prisco, *Self-assembly and DNA binding of the blocking factor in x chromosome inactivation*. PLoS Comput Biol, 2007. **3**(11): p. e210.
166. Lee, J.T., et al., *A 450 kb transgene displays properties of the mammalian X-inactivation center*. Cell, 1996. **86**(1): p. 83-94.
167. Herzing, L.B., et al., *Xist has properties of the X-chromosome inactivation centre*. Nature, 1997. **386**(6622): p. 272-5.
168. Lee, J.T. and R. Jaenisch, *Long-range cis effects of ectopic X-inactivation centres on a mouse autosome*. Nature, 1997. **386**(6622): p. 275-9.
169. Lee, J.T., N. Lu, and Y. Han, *Genetic analysis of the mouse X inactivation center defines an 80-kb multifunction domain*. Proc Natl Acad Sci U S A, 1999. **96**(7): p. 3836-41.
170. Migeon, B.R., et al., *Human X inactivation center induces random X chromosome inactivation in male transgenic mice*. Genomics, 1999. **59**(2): p. 113-21.
171. Migeon, B.R., et al., *Low-copy-number human transgene is recognized as an X inactivation center in mouse ES cells, but fails to induce cis-inactivation in chimeric mice*. Genomics, 2001. **71**(2): p. 156-62.
172. Heard, E., et al., *Transgenic mice carrying an Xist-containing YAC*. Hum Mol Genet, 1996. **5**(4): p. 441-50.
173. Matsuura, S., et al., *Xist expression from an Xist YAC transgene carried on the mouse Y chromosome*. Hum Mol Genet, 1996. **5**(4): p. 451-9.
174. Jonkers, I., et al., *RNF12 is an X-Encoded dose-dependent activator of X chromosome inactivation*. Cell, 2009. **139**(5): p. 999-1011.
175. Heard, E., et al., *Xist yeast artificial chromosome transgenes function as X-inactivation centers only in multicopy arrays and not as single copies*. Mol Cell Biol, 1999. **19**(4): p. 3156-66.
176. Gribnau, J., et al., *X chromosome choice occurs independently of asynchronous replication timing*. J Cell Biol, 2005. **168**(3): p. 365-73.
177. Marahrens, Y., J. Loring, and R. Jaenisch, *Role of the Xist gene in X chromosome choosing*. Cell, 1998. **92**(5): p. 657-64.
178. Sado, T., E. Li, and H. Sasaki, *Effect of TSIX disruption on XIST expression in male ES cells*. Cytogenet Genome Res, 2002. **99**(1-4): p. 115-8.
179. Monkhorst, K., et al., *X inactivation counting and choice is a stochastic process: evidence for involvement of an X-linked activator*. Cell, 2008. **132**(3): p. 410-21.
180. Lee, J.T., *Homozygous Tsix mutant mice reveal a sex-ratio distortion and revert to random X-inactivation*. Nat Genet, 2002. **32**(1): p. 195-200.
181. Mlynarczyk-Evans, S., et al., *X chromosomes alternate between two states prior to random X-inactivation*. PLoS Biol, 2006. **4**(6): p. e159.
182. Bacher, C.P., et al., *Transient colocalization of X-inactivation centres accompanies the initiation of X inactivation*. Nat Cell Biol, 2006. **8**(3): p. 293-9.
183. Xu, N., C.L. Tsai, and J.T. Lee, *Transient homologous chromosome pairing marks the onset of X inactivation*. Science, 2006. **311**(5764): p. 1149-52.
184. Masui, O., et al., *Live-cell chromosome dynamics and outcome of X chromosome pairing events during ES cell differentiation*. Cell, **145**(3): p. 447-58.
185. Xu, N., et al., *Evidence that homologous X-chromosome pairing requires transcription and Ctfc protein*. Nat Genet, 2007. **39**(11): p. 1390-6.
186. Donohoe, M.E., et al., *The pluripotency factor Oct4 interacts with Ctfc and also controls X-chromosome pairing and counting*. Nature, 2009. **460**(7251): p. 128-32.
187. Augui, S., et al., *Sensing X chromosome pairs before X inactivation via a novel X-pairing region of the Xic*. Science, 2007. **318**(5856): p. 1632-6.
188. Takagi, N., *Variable X chromosome inactivation patterns in near-tetraploid murine EC x somatic cell hybrid cells differentiated in vitro*. Genetica, 1993. **88**(2-3): p. 107-17.
189. Monkhorst, K., et al., *The probability to initiate X chromosome inactivation is determined by the X to autosomal ratio and X chromosome specific allelic properties*. PLoS One, 2009. **4**(5): p. e5616.
190. Monk, M. and M.I. Harper, *Sequential X chromosome inactivation coupled with cellular differentiation in early mouse embryos*. Nature, 1979. **281**(5729): p. 311-3.
191. Navarro, P., et al., *Molecular coupling of Xist regulation and pluripotency*. Science, 2008. **321**(5896): p. 1693-5.
192. Navarro, P. and P. Avner, *An embryonic story: analysis of the gene regulative network controlling Xist expression in mouse embryonic stem cells*. Bioessays, 2010. **32**(7): p. 581-8.

References

193. Donohoe, M.E., et al., *Identification of a Ctf cofactor. Yyl, for the X chromosome binary switch*. Mol Cell, 2007. **25**(1): p. 43-56.
194. Ma, Z., et al., *Sequence-specific regulator Prdm14 safeguards mouse ESCs from entering extraembryonic endoderm fates*. Nat Struct Mol Biol, 2011. **18**(2): p. 120-7.
195. Marson, A., et al., *Wnt signaling promotes reprogramming of somatic cells to pluripotency*. Cell Stem Cell, 2008. **3**(2): p. 132-5.
196. Barr, M.L. and E.G. Bertram, *A morphological distinction between neurones of the male and female, and the behaviour of the nucleolar satellite during accelerated nucleoprotein synthesis*. Nature, 1949. **163**(4148): p. 676.
197. Ohno, S. and T.S. Hauschka, *Alloccy of the X-chromosome in tumors and normal tissues*. Cancer Res, 1960. **20**: p. 541-5.
198. Chaumeil, J., et al., *A novel role for Xist RNA in the formation of a repressive nuclear compartment into which genes are recruited when silenced*. Genes Dev, 2006. **20**(16): p. 2223-37.
199. Clemson, C.M., et al., *The X chromosome is organized into a gene-rich outer rim and an internal core containing silenced nongenic sequences*. Proc Natl Acad Sci U S A, 2006. **103**(20): p. 7688-93.
200. Goto, Y., et al., *Differential patterns of histone methylation and acetylation distinguish active and repressed alleles at X-linked genes*. Cytogenet Genome Res, 2002. **99**(1-4): p. 66-74.
201. Mak, W., et al., *Mitotically stable association of polycomb group proteins eed and enx1 with the inactive x chromosome in trophoblast stem cells*. Curr Biol, 2002. **12**(12): p. 1016-20.
202. Boggs, B.A., et al., *Differentially methylated forms of histone H3 show unique association patterns with inactive human X chromosomes*. Nat Genet, 2002. **30**(1): p. 73-6.
203. Mermoud, J.E., et al., *Histone H3 lysine 9 methylation occurs rapidly at the onset of random X chromosome inactivation*. Curr Biol, 2002. **12**(3): p. 247-51.
204. Peters, A.H., et al., *Histone H3 lysine 9 methylation is an epigenetic imprint of facultative heterochromatin*. Nat Genet, 2002. **30**(1): p. 77-80.
205. Kohlmaier, A., et al., *A chromosomal memory triggered by Xist regulates histone methylation in X inactivation*. PLoS Biol, 2004. **2**(7): p. E171.
206. de Napoles, M., et al., *Polycomb group proteins Ring1A/B link ubiquitylation of histone H2A to heritable gene silencing and X inactivation*. Dev Cell, 2004. **7**(5): p. 663-76.
207. Fang, J., et al., *Ring1b-mediated H2A ubiquitination associates with inactive X chromosomes and is involved in initiation of X inactivation*. J Biol Chem, 2004. **279**(51): p. 52812-5.
208. Costanzi, C. and J.R. Pehrson, *Histone macroH2A1 is concentrated in the inactive X chromosome of female mammals*. Nature, 1998. **393**(6685): p. 599-601.
209. Lock, L.F., N. Takagi, and G.R. Martin, *Methylation of the Hprt gene on the inactive X occurs after chromosome inactivation*. Cell, 1987. **48**(1): p. 39-46.
210. Norris, D.P., N. Brockdorff, and S. Rastan, *Methylation status of CpG-rich islands on active and inactive mouse X chromosomes*. Mamm Genome, 1991. **1**(2): p. 78-83.
211. Mohandas, T., R.S. Sparkes, and L.J. Shapiro, *Reactivation of an inactive human X chromosome: evidence for X inactivation by DNA methylation*. Science, 1981. **211**(4480): p. 393-6.
212. Senner, C.E., et al., *Disruption of a conserved region of Xist exon 1 impairs Xist RNA localisation and X-linked gene silencing during random and imprinted X chromosome inactivation*. Development, 2011. **138**(8): p. 1541-50.
213. Beletskii, A., et al., *PNA interference mapping demonstrates functional domains in the noncoding RNA Xist*. Proc Natl Acad Sci U S A, 2001. **98**(16): p. 9215-20.
214. Sarma, K., et al., *Locked nucleic acids (LNAs) reveal sequence requirements and kinetics of Xist RNA localization to the X chromosome*. Proc Natl Acad Sci U S A, 2010. **107**(51): p. 22196-201.
215. Fackelmayer, F.O., *A stable proteinaceous structure in the territory of inactive X chromosomes*. J Biol Chem, 2005. **280**(3): p. 1720-3.
216. Hasegawa, Y., et al., *The matrix protein hnRNP U is required for chromosomal localization of Xist RNA*. Dev Cell, 2010. **19**(3): p. 469-76.
217. Helbig, R. and F.O. Fackelmayer, *Scaffold attachment factor A (SAF-A) is concentrated in inactive X chromosome territories through its RGG domain*. Chromosoma, 2003. **112**(4): p. 173-82.
218. Pullirsch, D., et al., *The Trithorax group protein Ash2l and Saf-A are recruited to the inactive X chromosome at the onset of stable X inactivation*. Development, 2010. **137**(6): p. 935-43.
219. Roshon, M.J. and H.E. Ruley, *Hypomorphic mutation in hnRNP U results in post-implantation lethality*. Transgenic Res, 2005. **14**(2): p. 179-92.

References

220. Jeon, Y. and J.T. Lee, *YY1 tethers Xist RNA to the inactive X nucleation center*. Cell, 2011. **146**(1): p. 119-33.
221. Ganesan, S., et al., *BRCA1 supports XIST RNA concentration on the inactive X chromosome*. Cell, 2002. **111**(3): p. 393-405.
222. Silver, D.P., et al., *Further evidence for BRCA1 communication with the inactive X chromosome*. Cell, 2007. **128**(5): p. 991-1002.
223. Vincent-Salomon, A., et al., *X inactive-specific transcript RNA coating and genetic instability of the X chromosome in BRCA1 breast tumors*. Cancer Res, 2007. **67**(11): p. 5134-40.
224. Xiao, C., et al., *The XIST noncoding RNA functions independently of BRCA1 in X inactivation*. Cell, 2007. **128**(5): p. 977-89.
225. Russell, L.B. and C.S. Montgomery, *Comparative studies on X-autosome translocations in the mouse. II. Inactivation of autosomal loci, segregation, and mapping of autosomal breakpoints in five T (X;1) S. Genetics*, 1970. **64**(2): p. 281-312.
226. Duthie, S.M., et al., *Xist RNA exhibits a banded localization on the inactive X chromosome and is excluded from autosomal material in cis*. Hum Mol Genet, 1999. **8**(2): p. 195-204.
227. Keohane, A.M., et al., *H4 acetylation, XIST RNA and replication timing are coincident and define x-autosome boundaries in two abnormal X chromosomes*. Hum Mol Genet, 1999. **8**(2): p. 377-83.
228. Popova, B.C., et al., *Attenuated spread of X-inactivation in an X-autosome translocation*. Proc Natl Acad Sci U S A, 2006. **103**(20): p. 7706-11.
229. Gartler, S.M. and A.D. Riggs, *Mammalian X-chromosome inactivation*. Annu Rev Genet, 1983. **17**: p. 155-90.
230. Lyon, M.F., *X-chromosome inactivation: a repeat hypothesis*. Cytogenet Cell Genet, 1998. **80**(1-4): p. 133-7.
231. Lyon, M.F., *LINE-1 elements and X chromosome inactivation: a function for "junk" DNA?* Proc Natl Acad Sci U S A, 2000. **97**(12): p. 6248-9.
232. Lyon, M.F., *Do LINEs Have a Role in X-Chromosome Inactivation?* J Biomed Biotechnol, 2006. **2006**(1): p. 59746.
233. Lyon, M.F., *No longer 'all-or-none'*. Eur J Hum Genet, 2005. **13**(7): p. 796-7.
234. Wichman, H.A., et al., *Transposable elements and the evolution of genome organization in mammals*. Genetica, 1992. **86**(1-3): p. 287-93.
235. Furano, A.V., *The biological properties and evolutionary dynamics of mammalian LINE-1 retrotransposons*. Prog Nucleic Acid Res Mol Biol, 2000. **64**: p. 255-94.
236. Parish, D.A., et al., *Distribution of LINEs and other repetitive elements in the karyotype of the bat Carollia: implications for X-chromosome inactivation*. Cytogenet Genome Res, 2002. **96**(1-4): p. 191-7.
237. Waters, P.D., et al., *LINE-1 distribution in Afrotheria and Xenarthra: implications for understanding the evolution of LINE-1 in eutherian genomes*. Chromosoma, 2004. **113**(3): p. 137-44.
238. Bailey, J.A., et al., *Molecular evidence for a relationship between LINE-1 elements and X chromosome inactivation: the Lyon repeat hypothesis*. Proc Natl Acad Sci U S A, 2000. **97**(12): p. 6634-9.
239. Dobigny, G., et al., *Viability of X-autosome translocations in mammals: an epigenomic hypothesis from a rodent case-study*. Chromosoma, 2004. **113**(1): p. 34-41.
240. Carrol, L. and H.F. Willard, *X-inactivation profile reveals extensive variability in X-linked gene expression in females*. Nature, 2005. **434**(7031): p. 400-4.
241. Wang, Z., et al., *Evidence of influence of genomic DNA sequence on human X chromosome inactivation*. PLoS Comput Biol, 2006. **2**(9): p. e113.
242. White, W.M., et al., *The spreading of X inactivation into autosomal material of an x-autosome translocation: evidence for a difference between autosomal and X-chromosomal DNA*. Am J Hum Genet, 1998. **63**(1): p. 20-8.
243. Sharp, A., D.O. Robinson, and P. Jacobs, *Absence of correlation between late-replication and spreading of X inactivation in an X-autosome translocation*. Hum Genet, 2001. **109**(3): p. 295-302.
244. Solari, A.J., et al., *The behavior of sex chromosomes in two human X-autosome translocations: failure of extensive X-inactivation spreading*. Biocell, 2001. **25**(2): p. 155-66.
245. Hall, L.L., et al., *Unbalanced X-autosome translocations provide evidence for sequence specificity in the association of XIST RNA with chromatin*. Hum Mol Genet, 2002. **11**(25): p. 3157-65.
246. Sharp, A.J., et al., *Molecular and cytogenetic analysis of the spreading of X inactivation in X-autosome translocations*. Hum Mol Genet, 2002. **11**(25): p. 3145-56.

247. Ke, X. and A. Collins, *CpG islands in human X-inactivation*. *Ann Hum Genet*, 2003. **67**(Pt 3): p. 242-9.
248. Cantrell, M.A., B.C. Carstens, and H.A. Wichman, *X chromosome inactivation and Xist evolution in a rodent lacking LINE-1 activity*. *PLoS One*, 2009. **4**(7): p. e6252.
249. Tang, Y.A., et al., *Efficiency of Xist-mediated silencing on autosomes is linked to chromosomal domain organisation*. *Epigenetics Chromatin*, 2010. **3**(1): p. 10.
250. Chow, J.C., et al., *LINE-1 activity in facultative heterochromatin formation during X chromosome inactivation*. *Cell*, 2010. **141**(6): p. 956-69.
251. Hansen, R.S., *X inactivation-specific methylation of LINE-1 elements by DNMT3B: implications for the Lyon repeat hypothesis*. *Hum Mol Genet*, 2003. **12**(19): p. 2559-67.
252. Cao, R. and Y. Zhang, *SUZ12 is required for both the histone methyltransferase activity and the silencing function of the EED-EZH2 complex*. *Mol Cell*, 2004. **15**(1): p. 57-67.
253. de la Cruz, C.C., et al., *Developmental regulation of Suz 12 localization*. *Chromosoma*, 2005. **114**(3): p. 183-92.
254. Kaneko, S., et al., *Phosphorylation of the PRC2 component Ezh2 is cell cycle-regulated and up-regulates its binding to ncRNA*. *Genes Dev*, 2010. **24**(23): p. 2615-20.
255. Kanhere, A., et al., *Short RNAs are transcribed from repressed polycomb target genes and interact with polycomb repressive complex-2*. *Mol Cell*, 2010. **38**(5): p. 675-88.
256. Wang, J., et al., *Imprinted X inactivation maintained by a mouse Polycomb group gene*. *Nat Genet*, 2001. **28**(4): p. 371-5.
257. Kalantry, S. and T. Magnuson, *The Polycomb group protein EED is dispensable for the initiation of random X-chromosome inactivation*. *PLoS Genet*, 2006. **2**(5): p. e66.
258. Schoeffner, S., et al., *Recruitment of PRC1 function at the initiation of X inactivation independent of PRC2 and silencing*. *Embo J*, 2006. **25**(13): p. 3110-22.
259. Hoki, Y., et al., *A proximal conserved repeat in the Xist gene is essential as a genomic element for X-inactivation in mouse*. *Development*, 2009. **136**(1): p. 139-46.
260. Royce-Tolland, M.E., et al., *The A-repeat links ASF/SF2-dependent Xist RNA processing with random choice during X inactivation*. *Nat Struct Mol Biol*, 2010. **17**(8): p. 948-54.
261. Wutz, A. and R. Jaenisch, *A shift from reversible to irreversible X inactivation is triggered during ES cell differentiation*. *Mol Cell*, 2000. **5**(4): p. 695-705.
262. Hoki, Y., et al., *Incomplete X-inactivation initiated by a hypomorphic Xist allele in the mouse*. *Development*, 2011. **138**(13): p. 2649-59.
263. Landeira, D., et al., *Jarid2 is a PRC2 component in embryonic stem cells required for multi-lineage differentiation and recruitment of PRC1 and RNA Polymerase II to developmental regulators*. *Nat Cell Biol*, 2010. **12**(6): p. 618-24.
264. Peng, J.C., et al., *Jarid2/Jumonji coordinates control of PRC2 enzymatic activity and target gene occupancy in pluripotent cells*. *Cell*, 2009. **139**(7): p. 1290-302.
265. Shen, X., et al., *Jumonji modulates polycomb activity and self-renewal versus differentiation of stem cells*. *Cell*, 2009. **139**(7): p. 1303-14.
266. Casanova, M., et al., *Polycomblike 2 facilitates the recruitment of PRC2 Polycomb group complexes to the inactive X chromosome and to target loci in embryonic stem cells*. *Development*, 2011. **138**(8): p. 1471-82.
267. Fischle, W., et al., *Molecular basis for the discrimination of repressive methyl-lysine marks in histone H3 by Polycomb and HP1 chromodomains*. *Genes Dev*, 2003. **17**(15): p. 1870-81.
268. Min, J., Y. Zhang, and R.M. Xu, *Structural basis for specific binding of Polycomb chromodomain to histone H3 methylated at Lys 27*. *Genes Dev*, 2003. **17**(15): p. 1823-8.
269. Wang, L., et al., *Hierarchical recruitment of polycomb group silencing complexes*. *Mol Cell*, 2004. **14**(5): p. 637-46.
270. Leeb, M., et al., *Polycomb complexes act redundantly to repress genomic repeats and genes*. *Genes Dev*, 2010. **24**(3): p. 265-76.
271. Tavares, L., et al., *RYBP-PRC1 complexes mediate H2A ubiquitylation at polycomb target sites independently of PRC2 and H3K27me3*. *Cell*, 2012. **148**(4): p. 664-78.
272. Arrighi, R., et al., *The Polycomb-associated protein Rybp is a ubiquitin binding protein*. *FEBS Lett*, 2006. **580**(26): p. 6233-41.
273. Savarese, F., et al., *Hematopoietic precursor cells transiently reestablish permissiveness for X inactivation*. *Mol Cell Biol*, 2006. **26**(19): p. 7167-77.

References

274. Agrelo, R., et al., *SATB1 defines the developmental context for gene silencing by Xist in lymphoma and embryonic cells*. *Dev Cell*, 2009. **16**(4): p. 507-16.
275. Han, H.J., et al., *SATB1 reprogrammes gene expression to promote breast tumour growth and metastasis*. *Nature*, 2008. **452**(7184): p. 187-93.
276. Alvarez, J.D., et al., *The MAR-binding protein SATB1 orchestrates temporal and spatial expression of multiple genes during T-cell development*. *Genes Dev*, 2000. **14**(5): p. 521-35.
277. Britanova, O., et al., *Satb2 haploinsufficiency phenocopies 2q32-q33 deletions, whereas loss suggests a fundamental role in the coordination of jaw development*. *Am J Hum Genet*, 2006. **79**(4): p. 668-78.
278. Dobrev, G., et al., *SATB2 is a multifunctional determinant of craniofacial patterning and osteoblast differentiation*. *Cell*, 2006. **125**(5): p. 971-86.
279. Hernandez-Munoz, I., et al., *Stable X chromosome inactivation involves the PRC1 Polycomb complex and requires histone MACROH2A1 and the CULLIN3/SPOP ubiquitin E3 ligase*. *Proc Natl Acad Sci U S A*, 2005. **102**(21): p. 7635-40.
280. Nusinow, D.A., et al., *Poly(ADP-ribose) polymerase 1 is inhibited by a histone H2A variant, MacroH2A, and contributes to silencing of the inactive X chromosome*. *J Biol Chem*, 2007. **282**(17): p. 12851-9.
281. Kim, M.Y., T. Zhang, and W.L. Kraus, *Poly(ADP-ribosylation) by PARP-1: 'PAR-laying' NAD⁺ into a nuclear signal*. *Genes Dev*, 2005. **19**(17): p. 1951-67.
282. Rouleau, M., R.A. Aubin, and G.G. Poirier, *Poly(ADP-ribosyl)ated chromatin domains: access granted*. *J Cell Sci*, 2004. **117**(Pt 6): p. 815-25.
283. Hassa, P.O., et al., *Nuclear ADP-ribosylation reactions in mammalian cells: where are we today and where are we going?* *Microbiol Mol Biol Rev*, 2006. **70**(3): p. 789-829.
284. Kim, M.Y., et al., *NAD⁺-dependent modulation of chromatin structure and transcription by nucleosome binding properties of PARP-1*. *Cell*, 2004. **119**(6): p. 803-14.
285. Chadwick, B.P. and H.F. Willard, *Histone H2A variants and the inactive X chromosome: identification of a second macroH2A variant*. *Hum Mol Genet*, 2001. **10**(10): p. 1101-13.
286. Mietton, F., et al., *Weak but uniform enrichment of the histone variant macroH2A1 along the inactive X chromosome*. *Mol Cell Biol*, 2009. **29**(1): p. 150-6.
287. Rasmussen, T.P., et al., *Expression of Xist RNA is sufficient to initiate macrochromatin body formation*. *Chromosoma*, 2001. **110**(6): p. 411-20.
288. Costanzi, C., et al., *Histone macroH2A1 is concentrated in the inactive X chromosome of female preimplantation mouse embryos*. *Development*, 2000. **127**(11): p. 2283-9.
289. Changoikar, L.N., et al., *Developmental changes in histone macroH2A1-mediated gene regulation*. *Mol Cell Biol*, 2007. **27**(7): p. 2758-64.
290. Boulard, M., et al., *Histone variant macroH2A1 deletion in mice causes female-specific steatosis*. *Epigenetics Chromatin*, 2010. **3**(1): p. 8.
291. Costanzi, C. and J.R. Pehrson, *MACROH2A2, a new member of the MARCOH2A core histone family*. *J Biol Chem*, 2001. **276**(24): p. 21776-84.
292. Rasmussen, T.P., et al., *Messenger RNAs encoding mouse histone macroH2A1 isoforms are expressed at similar levels in male and female cells and result from alternative splicing*. *Nucleic Acids Res*, 1999. **27**(18): p. 3685-9.
293. Tanasijevic, B. and T.P. Rasmussen, *X chromosome inactivation and differentiation occur readily in ES cells doubly-deficient for macroH2A1 and macroH2A2*. *PLoS One*, 2011. **6**(6): p. e21512.
294. Cotton, A.M., et al., *Chromosome-wide DNA methylation analysis predicts human tissue-specific X inactivation*. *Hum Genet*, 2011. **130**(2): p. 187-201.
295. Weber, M., et al., *Chromosome-wide and promoter-specific analyses identify sites of differential DNA methylation in normal and transformed human cells*. *Nat Genet*, 2005. **37**(8): p. 853-62.
296. Hellman, A. and A. Chess, *Gene body-specific methylation on the active X chromosome*. *Science*, 2007. **315**(5815): p. 1141-3.
297. Sado, T., et al., *X inactivation in the mouse embryo deficient for Dnmt1: distinct effect of hypomethylation on imprinted and random X inactivation*. *Dev Biol*, 2000. **225**(2): p. 294-303.
298. Sado, T., et al., *De novo DNA methylation is dispensable for the initiation and propagation of X chromosome inactivation*. *Development*, 2004. **131**(5): p. 975-82.
299. Blewitt, M.E., et al., *An N-ethyl-N-nitrosourea screen for genes involved in variegation in the mouse*. *Proc Natl Acad Sci U S A*, 2005. **102**(21): p. 7629-34.
300. Blewitt, M.E., et al., *SmcHDI, containing a structural-maintenance-of-chromosomes hinge domain, has a critical role in X inactivation*. *Nat Genet*, 2008. **40**(5): p. 663-9.

References

301. Kanno, T., et al., *A structural-maintenance-of-chromosomes hinge domain-containing protein is required for RNA-directed DNA methylation*. Nat Genet, 2008. **40**(5): p. 670-5.
302. Garrick, D., et al., *Loss of Atrx affects trophoblast development and the pattern of X-inactivation in extraembryonic tissues*. PLoS Genet, 2006. **2**(4): p. e58.
303. Baumann, C. and R. De La Fuente, *ATR^X marks the inactive X chromosome (Xi) in somatic cells and during imprinted X chromosome inactivation in trophoblast stem cells*. Chromosoma, 2009. **118**(2): p. 209-22.
304. Brown, C.J. and H.F. Willard, *The human X-inactivation centre is not required for maintenance of X-chromosome inactivation*. Nature, 1994. **368**(6467): p. 154-6.
305. Al Nadaf, S., et al., *A cross-species comparison of escape from X inactivation in Eutheria: implications for evolution of X chromosome inactivation*. Chromosoma, 2012. **121**(1): p. 71-8.
306. Disteche, C.M., *Escape from X inactivation in human and mouse*. Trends Genet, 1995. **11**(1): p. 17-22.
307. Disteche, C.M., *Escapees on the X chromosome*. Proc Natl Acad Sci U S A, 1999. **96**(25): p. 14180-2.
308. Disteche, C.M., G.N. Filippova, and K.D. Tsuchiya, *Escape from X inactivation*. Cytogenet Genome Res, 2002. **99**(1-4): p. 36-43.
309. Brown, C.J. and J.M. Greally, *A stain upon the silence: genes escaping X inactivation*. Trends Genet, 2003. **19**(8): p. 432-8.
310. Yen, Z.C., et al., *A cross-species comparison of X-chromosome inactivation in Eutheria*. Genomics, 2007. **90**(4): p. 453-63.
311. Patrat, C., et al., *Dynamic changes in paternal X-chromosome activity during imprinted X-chromosome inactivation in mice*. Proc Natl Acad Sci U S A, 2009. **106**(13): p. 5198-203.
312. Yang, F., et al., *Global survey of escape from X inactivation by RNA-sequencing in mouse*. Genome Res, 2010. **20**(5): p. 614-22.
313. Carrel, L., et al., *Genomic environment predicts expression patterns on the human inactive X chromosome*. PLoS Genet, 2006. **2**(9): p. e151.
314. Gaszner, M. and G. Felsenfeld, *Insulators: exploiting transcriptional and epigenetic mechanisms*. Nat Rev Genet, 2006. **7**(9): p. 703-13.
315. Phillips, J.E. and V.G. Corces, *CTCF: master weaver of the genome*. Cell, 2009. **137**(7): p. 1194-211.
316. Filippova, G.N., et al., *Boundaries between chromosomal domains of X inactivation and escape bind CTCF and lack CpG methylation during early development*. Dev Cell, 2005. **8**(1): p. 31-42.
317. Ciavatta, D., et al., *A DNA insulator prevents repression of a targeted X-linked transgene but not its random or imprinted X inactivation*. Proc Natl Acad Sci U S A, 2006. **103**(26): p. 9958-63.
318. Li, N. and L. Carrel, *Escape from X chromosome inactivation is an intrinsic property of the Jarid1c locus*. Proc Natl Acad Sci U S A, 2008. **105**(44): p. 17055-60.
319. Chan, K.M., et al., *Diverse factors are involved in maintaining X chromosome inactivation*. Proc Natl Acad Sci U S A, 2011. **108**(40): p. 16699-704.
320. Bininda-Emonds, O.R., et al., *The delayed rise of present-day mammals*. Nature, 2007. **446**(7135): p. 507-12.
321. Rens, W., et al., *Resolution and evolution of the duck-billed platypus karyotype with an X1Y1X2Y2X3Y3X4Y4X5Y5 male sex chromosome constitution*. Proc Natl Acad Sci U S A, 2004. **101**(46): p. 16257-61.
322. Rens, W., et al., *The multiple sex chromosomes of platypus and echidna are not completely identical and several share homology with the avian Z*. Genome Biol, 2007. **8**(11): p. R243.
323. Grutzner, F., et al., *The monotreme genome: a patchwork of reptile, mammal and unique features?* Comp Biochem Physiol A Mol Integr Physiol, 2003. **136**(4): p. 867-81.
324. Grutzner, F. and J.A. Graves, *A platypus' eye view of the mammalian genome*. Curr Opin Genet Dev, 2004. **14**(6): p. 642-9.
325. Grutzner, F., et al., *In the platypus a meiotic chain of ten sex chromosomes shares genes with the bird Z and mammal X chromosomes*. Nature, 2004. **432**(7019): p. 913-7.
326. Veyrunes, F., et al., *Bird-like sex chromosomes of platypus imply recent origin of mammal sex chromosomes*. Genome Res, 2008. **18**(6): p. 965-73.
327. Rens, W., et al., *Epigenetic modifications on X chromosomes in marsupial and monotreme mammals and implications for evolution of dosage compensation*. Proc Natl Acad Sci U S A, 2010. **107**(41): p. 17657-62.
328. Deakin, J.E., et al., *Unravelling the evolutionary origins of X chromosome inactivation in mammals: insights from marsupials and monotremes*. Chromosome Res, 2009. **17**(5): p. 671-85.

References

329. Deakin, J.E., et al., *The status of dosage compensation in the multiple X chromosomes of the platypus*. PLoS Genet, 2008. **4**(7): p. e1000140.
330. Mahadevaiah, S.K., et al., *Key features of the X inactivation process are conserved between marsupials and eutherians*. Curr Biol, 2009. **19**(17): p. 1478-84.
331. Cooper, D.W., *Directed genetic change model for X chromosome inactivation in eutherian mammals*. Nature, 1971. **230**(5292): p. 292-4.
332. Sharman, G.B., *Late DNA replication in the paternally derived X chromosome of female kangaroos*. Nature, 1971. **230**(5291): p. 231-2.
333. Richardson, B.J., A.B. Czuppon, and G.B. Sharman, *Inheritance of glucose-6-phosphate dehydrogenase variation in kangaroos*. Nat New Biol, 1971. **230**(13): p. 154-5.
334. Cooper, D.W., et al., *Phosphoglycerate kinase polymorphism in kangaroos provides further evidence for paternal X inactivation*. Nat New Biol, 1971. **230**(13): p. 155-7.
335. Kaslow, D.C. and B.R. Migeon, *DNA methylation stabilizes X chromosome inactivation in eutherians but not in marsupials: evidence for multistep maintenance of mammalian X dosage compensation*. Proc Natl Acad Sci U S A, 1987. **84**(17): p. 6210-4.
336. Graves, J.A., *DNA synthesis in chromosomes of cultured leucocytes from two marsupial species*. Exp Cell Res, 1967. **46**(1): p. 37-57.
337. Koina, E., et al., *Specific patterns of histone marks accompany X chromosome inactivation in a marsupial*. Chromosome Res, 2009. **17**(1): p. 115-26.
338. Wakefield, M.J., et al., *Histone underacetylation is an ancient component of mammalian X chromosome inactivation*. Proc Natl Acad Sci U S A, 1997. **94**(18): p. 9665-8.
339. Chaumeil, J., et al., *Evolution from XIST-independent to XIST-controlled X-chromosome inactivation: epigenetic modifications in distantly related mammals*. PLoS One, 2011. **6**(4): p. e19040.
340. Piper, A.A., et al., *Isolation of a clone partially encoding hill kangaroo X-linked hypoxanthine phosphoribosyltransferase: sex differences in methylation in the body of the gene*. Somat Cell Mol Genet, 1993. **19**(2): p. 141-59.
341. Loebel, D.A. and P.G. Johnston, *Methylation analysis of a marsupial X-linked CpG island by bisulfite genomic sequencing*. Genome Res, 1996. **6**(2): p. 114-23.
342. Duret, L., et al., *The Xist RNA gene evolved in eutherians by pseudogenization of a protein-coding gene*. Science, 2006. **312**(5780): p. 1653-5.
343. Hore, T.A., et al., *The region homologous to the X-chromosome inactivation centre has been disrupted in marsupial and monotreme mammals*. Chromosome Res, 2007. **15**(2): p. 147-61.
344. Grant, J., et al., *Rsx is a metatherian RNA with Xist-like properties in X-chromosome inactivation*. Nature, 2012.
345. Kolesnikov, N.N. and E.A. Elisafenko, *[Exon-intron structure of the Xist gene in elephant, armadillo, and the ancestor of placental mammals]*. Genetika, 2010. **46**(10): p. 1379-85.
346. Nishihara, H., S. Maruyama, and N. Okada, *Retroposon analysis and recent geological data suggest near-simultaneous divergence of the three superorders of mammals*. Proc Natl Acad Sci U S A, 2009. **106**(13): p. 5235-40.
347. Gribnau, J. and J.A. Grootegeed, *Origin and evolution of X chromosome inactivation*. Curr Opin Cell Biol, 2012.
348. Shevchenko, A.I., et al., *Genes flanking Xist in mouse and human are separated on the X chromosome in American marsupials*. Chromosome Res, 2007. **15**(2): p. 127-36.
349. Davidow, L.S., et al., *The search for a marsupial XIC reveals a break with vertebrate synten*. Chromosome Res, 2007. **15**(2): p. 137-46.
350. Flynn, M., O. Saha, and P. Young, *Molecular evolution of the LNX gene family*. BMC Evol Biol, 2011. **11**: p. 235.
351. Caparros, M.L., et al., *Functional analysis of the highly conserved exon IV of XIST RNA*. Cytogenet Genome Res, 2002. **99**(1-4): p. 99-105.
352. Horvath, J.E., et al., *Comparative analysis of the primate X-inactivation center region and reconstruction of the ancestral primate XIST locus*. Genome Res, 2011. **21**(6): p. 850-62.
353. Michalak, P., *Coexpression, coregulation, and cofunctionality of neighboring genes in eukaryotic genomes*. Genomics, 2008. **91**(3): p. 243-8.
354. Sado, T., et al., *Regulation of imprinted X-chromosome inactivation in mice by Tsix*. Development, 2001. **128**(8): p. 1275-86.
355. Looijenga, L.H., et al., *X inactivation in human testicular tumors. XIST expression and androgen receptor methylation status*. Am J Pathol, 1997. **151**(2): p. 581-90.

References

356. Migeon, B.R., et al., *Identification of TSIX, encoding an RNA antisense to human XIST, reveals differences from its murine counterpart: implications for X inactivation*. *Am J Hum Genet*, 2001. **69**(5): p. 951-60.
357. Chow, J.C., et al., *Ectopic XIST transcripts in human somatic cells show variable expression and localization*. *Cytogenet Genome Res*, 2002. **99**(1-4): p. 92-8.
358. Hall, L.L., et al., *An ectopic human XIST gene can induce chromosome inactivation in postdifferentiation human HT-1080 cells*. *Proc Natl Acad Sci U S A*, 2002. **99**(13): p. 8677-82.
359. Hall, L.L., et al., *X-inactivation reveals epigenetic anomalies in most hESC but identifies sublines that initiate as expected*. *J Cell Physiol*, 2008. **216**(2): p. 445-52.
360. Shen, Y., et al., *X-inactivation in female human embryonic stem cells is in a nonrandom pattern and prone to epigenetic alterations*. *Proc Natl Acad Sci U S A*, 2008. **105**(12): p. 4709-14.
361. Silva, S.S., et al., *X-chromosome inactivation and epigenetic fluidity in human embryonic stem cells*. *Proc Natl Acad Sci U S A*, 2008. **105**(12): p. 4820-5.
362. Enver, T., et al., *Cellular differentiation hierarchies in normal and culture-adapted human embryonic stem cells*. *Hum Mol Genet*, 2005. **14**(21): p. 3129-40.
363. Hoffman, L.M., et al., *X-inactivation status varies in human embryonic stem cell lines*. *Stem Cells*, 2005. **23**(10): p. 1468-78.
364. Dhara, S.K. and N. Benvenisty, *Gene trap as a tool for genome annotation and analysis of X chromosome inactivation in human embryonic stem cells*. *Nucleic Acids Res*, 2004. **32**(13): p. 3995-4002.
365. Takagi, N., *Imprinted X-chromosome inactivation: enlightenment from embryos in vivo*. *Semin Cell Dev Biol*, 2003. **14**(6): p. 319-29.
366. West, J.D., et al., *Preferential expression of the maternally derived X chromosome in the mouse yolk sac*. *Cell*, 1977. **12**(4): p. 873-82.
367. Silva, J., et al., *Nanog is the gateway to the pluripotent ground state*. *Cell*, 2009. **138**(4): p. 722-37.
368. van den Berg, I.M., et al., *X chromosome inactivation is initiated in human preimplantation embryos*. *Am J Hum Genet*, 2009. **84**(6): p. 771-9.
369. Looijenga, L.H., et al., *Heterogeneous X inactivation in trophoblastic cells of human full-term female placentas*. *Am J Hum Genet*, 1999. **64**(5): p. 1445-52.
370. Ropers, H.H., G. Wolff, and H.W. Hitzeroth, *Preferential X inactivation in human placenta membranes: is the paternal X inactive in early embryonic development of female mammals?* *Hum Genet*, 1978. **43**(3): p. 265-73.
371. Daniels, R., et al., *XIST expression in human oocytes and preimplantation embryos*. *Am J Hum Genet*, 1997. **61**(1): p. 33-9.
372. Ray, P.F., R.M. Winston, and A.H. Handyside, *XIST expression from the maternal X chromosome in human male preimplantation embryos at the blastocyst stage*. *Hum Mol Genet*, 1997. **6**(8): p. 1323-7.
373. Bamforth, F., G. Machin, and M. Innes, *X-chromosome inactivation is mostly random in placental tissues of female monozygotic twins and triplets*. *Am J Med Genet*, 1996. **61**(3): p. 209-15.
374. Migeon, B.R., et al., *Incomplete X chromosome dosage compensation in chorionic villi of human placenta*. *Proc Natl Acad Sci U S A*, 1985. **82**(10): p. 3390-4.
375. Migeon, B.R., J. Axelman, and P. Jeppesen, *Differential X reactivation in human placental cells: implications for reversal of X inactivation*. *Am J Hum Genet*, 2005. **77**(3): p. 355-64.
376. Goto, T., E. Wright, and M. Monk, *Paternal X-chromosome inactivation in human trophoblastic cells*. *Mol Hum Reprod*, 1997. **3**(1): p. 77-80.
377. Harrison, K.B., *X-chromosome inactivation in the human cytotrophoblast*. *Cytogenet Cell Genet*, 1989. **52**(1-2): p. 37-41.
378. Harrison, K.B. and D. Warburton, *Preferential X-chromosome activity in human female placental tissues*. *Cytogenet Cell Genet*, 1986. **41**(3): p. 163-8.
379. Okamoto, I., et al., *Eutherian mammals use diverse strategies to initiate X-chromosome inactivation during development*. *Nature*, 2011. **472**(7343): p. 370-4.
380. Liu, W. and X. Sun, *Skewed X chromosome inactivation in diploid and triploid female human embryonic stem cells*. *Hum Reprod*, 2009. **24**(8): p. 1834-43.
381. Sato, N., et al., *Molecular signature of human embryonic stem cells and its comparison with the mouse*. *Dev Biol*, 2003. **260**(2): p. 404-13.
382. Ginis, I., et al., *Differences between human and mouse embryonic stem cells*. *Dev Biol*, 2004. **269**(2): p. 360-80.

References

383. Xu, R.H., et al., *Basic FGF and suppression of BMP signaling sustain undifferentiated proliferation of human ES cells*. Nat Methods, 2005. **2**(3): p. 185-90.
384. Ying, Q.L., et al., *BMP induction of Id proteins suppresses differentiation and sustains embryonic stem cell self-renewal in collaboration with STAT3*. Cell, 2003. **115**(3): p. 281-92.
385. Boyer, L.A., et al., *Core transcriptional regulatory circuitry in human embryonic stem cells*. Cell, 2005. **122**(6): p. 947-56.
386. Brons, I.G., et al., *Derivation of pluripotent epiblast stem cells from mammalian embryos*. Nature, 2007. **448**(7150): p. 191-5.
387. Tesar, P.J., et al., *New cell lines from mouse epiblast share defining features with human embryonic stem cells*. Nature, 2007. **448**(7150): p. 196-9.
388. Rossant, J., *Stem cells and early lineage development*. Cell, 2008. **132**(4): p. 527-31.
389. Lovell-Badge, R., *Many ways to pluripotency*. Nat Biotechnol, 2007. **25**(10): p. 1114-6.
390. Xu, R.H., et al., *BMP4 initiates human embryonic stem cell differentiation to trophoblast*. Nat Biotechnol, 2002. **20**(12): p. 1261-4.
391. Guo, G., et al., *Klf4 reverts developmentally programmed restriction of ground state pluripotency*. Development, 2009. **136**(7): p. 1063-9.
392. Bao, S., et al., *Epigenetic reversion of post-implantation epiblast to pluripotent embryonic stem cells*. Nature, 2009. **461**(7268): p. 1292-5.
393. Hanna, J., et al., *Metastable pluripotent states in NOD-mouse-derived ESCs*. Cell Stem Cell, 2009. **4**(6): p. 513-24.
394. Nichols, J. and A. Smith, *Naive and primed pluripotent states*. Cell Stem Cell, 2009. **4**(6): p. 487-92.
395. Zvetkova, I., et al., *Global hypomethylation of the genome in XX embryonic stem cells*. Nat Genet, 2005. **37**(11): p. 1274-9.
396. Lagarkova, M.A., et al., *Diverse epigenetic profile of novel human embryonic stem cell lines*. Cell Cycle, 2006. **5**(4): p. 416-20.
397. Rugg-Gunn, P.J., A.C. Ferguson-Smith, and R.A. Pedersen, *Epigenetic status of human embryonic stem cells*. Nat Genet, 2005. **37**(6): p. 585-7.
398. Allegrucci, C., et al., *Restriction landmark genome scanning identifies culture-induced DNA methylation instability in the human embryonic stem cell epigenome*. Hum Mol Genet, 2007. **16**(10): p. 1253-68.
399. Boklage, C.E., *The epigenetic environment: secondary sex ratio depends on differential survival in embryogenesis*. Hum Reprod, 2005. **20**(3): p. 583-7.
400. Han, T.L., et al., *Detection of X- and Y-bearing human spermatozoa after motile sperm isolation by swim-up*. Fertil Steril, 1993. **60**(6): p. 1046-51.
401. Hassold, T., S.D. Quillen, and J.A. Yamane, *Sex ratio in spontaneous abortions*. Ann Hum Genet, 1983. **47**(Pt 1): p. 39-47.
402. Hassold, T., et al., *Cytogenetic and molecular studies of trisomy 13*. J Med Genet, 1987. **24**(12): p. 725-32.
403. Jacobs, P.A., et al., *Trisomy 13 ascertained in a survey of spontaneous abortions*. J Med Genet, 1987. **24**(12): p. 721-4.
404. Rasmussen, S.A., et al., *Population-based analyses of mortality in trisomy 13 and trisomy 18*. Pediatrics, 2003. **111**(4 Pt 1): p. 777-84.
405. Miller, J.F., et al., *Fetal loss after implantation. A prospective study*. Lancet, 1980. **2**(8194): p. 554-6.
406. Ellish, N.J., et al., *A prospective study of early pregnancy loss*. Hum Reprod, 1996. **11**(2): p. 406-12.
407. Wilcox, A.J., et al., *Incidence of early loss of pregnancy*. N Engl J Med, 1988. **319**(4): p. 189-94.
408. Jacobs, P.A. and T.J. Hassold, *The origin of numerical chromosome abnormalities*. Adv Genet, 1995. **33**: p. 101-33.
409. CBS. C.b.v.d.S. 21 April 2012; Available from: www.CBS.nl.
410. Childs, B., *GENETIC ORIGIN OF SOME SEX DIFFERENCES AMONG HUMAN BEINGS*. Pediatrics, 1965. **35**: p. 798-812.
411. Verbrugge, L.M., *Sex differentials in health*. Public Health Rep, 1982. **97**(5): p. 417-37.
412. Crimmins, E.M., J.K. Kim, and A. Sole-Auro, *Gender differences in health: results from SHARE, ELSA and HRS*. Eur J Public Health, 2011. **21**(1): p. 81-91.
413. Gissler, M., et al., *Boys have more health problems in childhood than girls: follow-up of the 1987 Finnish birth cohort*. Acta Paediatr, 1999. **88**(3): p. 310-4.
414. Cui, W., et al., *Sex differences in birth defects: a study of opposite-sex twins*. Birth Defects Res A Clin Mol Teratol, 2005. **73**(11): p. 876-80.

References

415. Lary, J.M. and L.J. Paulozzi, *Sex differences in the prevalence of human birth defects: a population-based study*. Teratology, 2001. **64**(5): p. 237-51.
416. Cartwright, R.A., K.A. Gurney, and A.V. Moorman, *Sex ratios and the risks of haematological malignancies*. Br J Haematol, 2002. **118**(4): p. 1071-7.
417. Horner, J.F., *Die Erbllichkeit des Daltonismus. Amtlicher Bericht ueber die Verwaltung des Medizinahwesens des Kantons Zurich vom Jahr 1876*. 1876, Zurich: Druck der Genossenschaftsbuchdruckerei.
418. Otto, J.C., *An account of an haemorrhagic disposition existing in certain families*. Medical Repository, 1803. **6**: p. 1-4.
419. Jaeger, W., *Horner's law. The first step in the history of the understanding of X-linked disorders*. Ophthalmic Paediatr Genet, 1992. **13**(2): p. 49-56.
420. Dobyns, W.B., *The pattern of inheritance of X-linked traits is not dominant or recessive, just X-linked*. Acta Paediatr Suppl, 2006. **95**(451): p. 11-5.
421. Morgan, T., *Sex-linked inheritance in Drosophila*. Science, 1910. **32**: p. 120-122.
422. Morgan, T., et al., *The mechanism of Mendelian heredity*. 1915, New York: Henry Holt & Company.
423. Morgan, T., et al., *The mechanism of Mendelian heredity*. 1922, New York: Henry Holt & Company.
424. Strachan, T. and A. Read, *Human Molecular Genetics* 3. 3rd ed. 2004, New York: Garland Science.
425. Lyon, M.F., *X-chromosome inactivation and human genetic disease*. Acta Paediatr Suppl, 2002. **91**(439): p. 107-12.
426. Chabas, A. and R. Happle, *Understanding the biology of X-linked diseases*. Acta Paediatr Suppl, 2006. **95**(451): p. 9-10.
427. Migeon, B.R., *Why females are mosaics. X-chromosome inactivation, and sex differences in disease*. Gend Med, 2007. **4**(2): p. 97-105.
428. Dobyns, W.B., et al., *Inheritance of most X-linked traits is not dominant or recessive, just X-linked*. Am J Med Genet A, 2004. **129A**(2): p. 136-43.
429. Migeon, B.R., *Selection and cell communication as determinants of female phenotype*. Basic Life Sci, 1978. **12**: p. 417-32.
430. Fratantoni, J.C., C.W. Hall, and E.F. Neufeld, *Hurler and Hunter syndromes: mutual correction of the defect in cultured fibroblasts*. Science, 1968. **162**(3853): p. 570-2.
431. Dewey, M.J., et al., *Mosaic mice with teratocarcinoma-derived mutant cells deficient in hypoxanthine phosphoribosyltransferase*. Proc Natl Acad Sci U S A, 1977. **74**(12): p. 5564-8.
432. Migeon, B.R., *Studies of skin fibroblasts from 10 families with HGPRT deficiency, with reference in X-chromosomal inactivation*. Am J Hum Genet, 1971. **23**(2): p. 199-210.
433. Filosa, S., et al., *Somatic-cell selection is a major determinant of the blood-cell phenotype in heterozygotes for glucose-6-phosphate dehydrogenase mutations causing severe enzyme deficiency*. Am J Hum Genet, 1996. **59**(4): p. 887-95.
434. Migeon, B.R., et al., *Adrenoleukodystrophy: evidence for X linkage, inactivation, and selection favoring the mutant allele in heterozygous cells*. Proc Natl Acad Sci U S A, 1981. **78**(8): p. 5066-70.
435. Maier, E.M., et al., *Symptoms in carriers of adrenoleukodystrophy relate to skewed X inactivation*. Ann Neurol, 2002. **52**(5): p. 683-8.
436. Wieland, I., et al., *Mutations of the ephrin-B1 gene cause craniofrontonasal syndrome*. Am J Hum Genet, 2004. **74**(6): p. 1209-15.
437. Twigg, S.R., et al., *Mutations of ephrin-B1 (EFNB1), a marker of tissue boundary formation, cause craniofrontonasal syndrome*. Proc Natl Acad Sci U S A, 2004. **101**(23): p. 8652-7.
438. Makarov, R., et al., *The impact of CFNS-causing EFNB1 mutations on ephrin-B1 function*. BMC Med Genet, 2010. **11**: p. 98.
439. Davy, A., J.O. Bush, and P. Soriano, *Inhibition of gap junction communication at ectopic Eph/ephrin boundaries underlies craniofrontonasal syndrome*. PLoS Biol, 2006. **4**(10): p. e315.
440. Amos-Landgraf, J.M., et al., *X chromosome-inactivation patterns of 1,005 phenotypically unaffected females*. Am J Hum Genet, 2006. **79**(3): p. 493-9.
441. Sharp, A., D. Robinson, and P. Jacobs, *Age- and tissue-specific variation of X chromosome inactivation ratios in normal women*. Hum Genet, 2000. **107**(4): p. 343-9.
442. Puck, J.M., C.C. Stewart, and R.L. Nussbaum, *Maximum-likelihood analysis of human T-cell X chromosome inactivation patterns: normal women versus carriers of X-linked severe combined immunodeficiency*. Am J Hum Genet, 1992. **50**(4): p. 742-8.
443. Monteiro, J., et al., *Commitment to X inactivation precedes the twinning event in monozygotic MZ twins*. Am J Hum Genet, 1998. **63**(2): p. 339-46.

References

444. Belmont, J.W., *Genetic control of X inactivation and processes leading to X-inactivation skewing*. Am J Hum Genet, 1996. **58**(6): p. 1101-8.
445. Cattanaach, B.M. and J.H. Isaacson, *Controlling elements in the mouse X chromosome*. Genetics, 1967. **57**(2): p. 331-46.
446. Cattanaach, B.M. and C.E. Williams, *Evidence of non-random X chromosome activity in the mouse*. Genet Res, 1972. **19**(3): p. 229-40.
447. Cattanaach, B.M., *Control of chromosome inactivation*. Annu Rev Genet, 1975. **9**: p. 1-18.
448. Simmler, M.C., et al., *Mapping the murine Xce locus with (CA)_n repeats*. Mamm Genome, 1993. **4**(9): p. 523-30.
449. Chadwick, L.H., et al., *Genetic control of X chromosome inactivation in mice: definition of the Xce candidate interval*. Genetics, 2006. **173**(4): p. 2103-10.
450. Avner, P., et al., *Molecular correlates of the murine Xce locus*. Genet Res, 1998. **72**(3): p. 217-24.
451. Plenge, R.M., et al., *A promoter mutation in the XIST gene in two unrelated families with skewed X-chromosome inactivation*. Nat Genet, 1997. **17**(3): p. 353-6.
452. Pugacheva, E.M., et al., *Familial cases of point mutations in the XIST promoter reveal a correlation between CTCF binding and pre-emptive choices of X chromosome inactivation*. Hum Mol Genet, 2005. **14**(7): p. 953-65.
453. Naumova, A.K., et al., *Heritability of X chromosome--inactivation phenotype in a large family*. Am J Hum Genet, 1996. **58**(6): p. 1111-9.
454. Busque, L., et al., *Nonrandom X-inactivation patterns in normal females: lyonization ratios vary with age*. Blood, 1996. **88**(1): p. 59-65.
455. Naumova, A.K., et al., *Genetic mapping of X-linked loci involved in skewing of X chromosome inactivation in the human*. Eur J Hum Genet, 1998. **6**(6): p. 552-62.
456. Ropers, H.H., et al., *Evidence for preferential X-chromosome inactivation in a family with Fabry disease*. Am J Hum Genet, 1977. **29**(4): p. 361-70.
457. Migeon, B.R. and C. Haisley-Royster, *Familial skewed X inactivation and X-linked mutations: unbalanced X inactivation is a powerful means to ascertain X-linked genes that affect cell proliferation*. Am J Hum Genet, 1998. **62**(6): p. 1555-7; author reply 1557-8.
458. Marcus, S., et al., *Mutation analysis and prenatal diagnosis in a Lesch-Nyhan family showing non-random X-inactivation interfering with carrier detection tests*. Hum Genet, 1992. **89**(4): p. 395-400.
459. Ruttum, M.S., M.F. Lewandowski, and J.B. Bateman, *Affected females in X-linked congenital stationary night blindness*. Ophthalmology, 1992. **99**(5): p. 747-52.
460. Orstavik, K.H., et al., *Inheritance of skewed X chromosome inactivation in a large family with an X-linked recessive deafness syndrome*. Am J Med Genet, 1996. **64**(1): p. 31-4.
461. Bolduc, V., et al., *No evidence that skewing of X chromosome inactivation patterns is transmitted to offspring in humans*. J Clin Invest, 2008. **118**(1): p. 333-41.
462. Lau, A.W., et al., *Skewed X-chromosome inactivation is common in fetuses or newborns associated with confined placental mosaicism*. Am J Hum Genet, 1997. **61**(6): p. 1353-61.
463. Goodship, J., J. Carter, and J. Burn, *X-inactivation patterns in monozygotic and dizygotic female twins*. Am J Med Genet, 1996. **61**(3): p. 205-8.
464. Richards, C.S., et al., *Skewed X inactivation in a female MZ twin results in Duchenne muscular dystrophy*. Am J Hum Genet, 1990. **46**(4): p. 672-81.
465. Lupski, J.R., et al., *Discordance of muscular dystrophy in monozygotic female twins: evidence supporting asymmetric splitting of the inner cell mass in a manifesting carrier of Duchenne dystrophy*. Am J Med Genet, 1991. **40**(3): p. 354-64.
466. Abbadi, N., et al., *Additional case of female monozygotic twins discordant for the clinical manifestations of Duchenne muscular dystrophy due to opposite X-chromosome inactivation*. Am J Med Genet, 1994. **52**(2): p. 198-206.
467. Winchester, B., et al., *Female twin with Hunter disease due to nonrandom inactivation of the X-chromosome: a consequence of twinning*. Am J Med Genet, 1992. **44**(6): p. 834-8.
468. Tiberio, G., *MZ female twins discordant for X-linked diseases: a review*. Acta Genet Med Gemellol (Roma), 1994. **43**(3-4): p. 207-14.
469. Redonnet-Vernhet, I., et al., *Uneven X inactivation in a female monozygotic twin pair with Fabry disease and discordant expression of a novel mutation in the alpha-galactosidase A gene*. J Med Genet, 1996. **33**(8): p. 682-8.
470. Bennett, C.M., E. Boye, and E.J. Neufeld, *Female monozygotic twins discordant for hemophilia A due to nonrandom X-chromosome inactivation*. Am J Hematol, 2008. **83**(10): p. 778-80.

References

471. Costa, T., et al., *Monozygotic twins discordant for Aicardi syndrome*. *J Med Genet*, 1997. **34**(8): p. 688-91.
472. Bum, J., et al., *Duchenne muscular dystrophy in one of monozygotic twin girls*. *J Med Genet*, 1986. **23**(6): p. 494-500.
473. Marguery, M.C., et al., *Fabry's disease: heterozygous form of different expression in two monozygous twin sisters*. *Dermatology*, 1993. **187**(1): p. 9-15.
474. Nance, W.E., *Do twin Lyons have larger spots?* *Am J Hum Genet*, 1990. **46**(4): p. 646-8.
475. Lubinsky, M.S. and J.G. Hall, *Genomic imprinting, monozygous twinning, and X inactivation*. *Lancet*, 1991. **337**(8752): p. 1288.
476. Chitnis, S., et al., *X chromosome-inactivation patterns confirm the late timing of monoamniotic-MZ twinning*. *Am J Hum Genet*, 1999. **65**(2): p. 570-1.
477. Puck, J.M., *The timing of twinning: more insights from X inactivation*. *Am J Hum Genet*, 1998. **63**(2): p. 327-8.
478. Fisk, N.M., et al., *X-chromosome inactivation patterns do not implicate asymmetric splitting of the inner cell mass in the aetiology of twin-twin transfusion syndrome*. *Mol Hum Reprod*, 1999. **5**(1): p. 52-6.
479. Kristiansen, M., et al., *Twin study of genetic and aging effects on X chromosome inactivation*. *Eur J Hum Genet*, 2005. **13**(5): p. 599-606.
480. Allen, R.C., et al., *Application of carrier testing to genetic counseling for X-linked agammaglobulinemia*. *Am J Hum Genet*, 1994. **54**(1): p. 25-35.
481. Li, S.L., et al., *Carrier identification in X-linked immunodeficiency diseases*. *J Paediatr Child Health*, 1998. **34**(3): p. 273-9.
482. Moschese, V., et al., *X-chromosome inactivation and mutation pattern in the Bruton's tyrosine kinase gene in patients with X-linked agammaglobulinemia*. *Italian XLA Collaborative Group*. *Mol Med*, 2000. **6**(2): p. 104-13.
483. Hendriks, R.W., et al., *Inactivation of Btk by insertion of lacZ reveals defects in B cell development only past the pre-B cell stage*. *Embo J*, 1996. **15**(18): p. 4862-72.
484. Hendriks, R.W., M.E. Kraakman, and R.K. Schuurman, *Patterns of X chromosome inactivation in haematopoietic cells of female carriers of X linked severe combined immunodeficiency*. *Immunodeficiency*, 1993. **4**(1-4): p. 263-5.
485. Hendriks, R.W., M.E. Kraakman, and R.K. Schuurman, *X chromosome inactivation patterns in haematopoietic cells of female carriers of X-linked severe combined immunodeficiency determined by methylation analysis at the hypervariable DXS255 locus*. *Clin Genet*, 1992. **42**(3): p. 114-21.
486. Wengler, G., et al., *Nonrandom inactivation of the X chromosome in early lineage hematopoietic cells in carriers of Wiskott-Aldrich syndrome*. *Blood*, 1995. **85**(9): p. 2471-7.
487. Gealy, W.J., J.M. Dwyer, and J.B. Harley, *Allelic exclusion of glucose-6-phosphate dehydrogenase in platelets and T lymphocytes from a Wiskott-Aldrich syndrome carrier*. *Lancet*, 1980. **1**(8159): p. 63-5.
488. Prechal, J.T., et al., *Wiskott-Aldrich syndrome: cellular impairments and their implication for carrier detection*. *Blood*, 1980. **56**(6): p. 1048-54.
489. Goodship, J., et al., *Carrier detection in Wiskott-Aldrich syndrome: combined use of M27 beta for X-inactivation studies and as a linked probe*. *Blood*, 1991. **77**(12): p. 2677-81.
490. Fearon, E.R., et al., *Carrier detection in the Wiskott Aldrich syndrome*. *Blood*, 1988. **72**(5): p. 1735-9.
491. Mantuano, E., et al., *Analysis of X-chromosome inactivation in bone marrow precursors from carriers of Wiskott-Aldrich syndrome and X-linked severe combined immunodeficiency: evidence that the Wiskott-Aldrich gene is expressed prior to granulocyte-macrophage colony-forming-unit*. *Immunodeficiency*, 1993. **4**(1-4): p. 271-6.
492. Greer, W.L., et al., *X-chromosome inactivation in the Wiskott-Aldrich syndrome: a marker for detection of the carrier state and identification of cell lineages expressing the gene defect*. *Genomics*, 1989. **4**(1): p. 60-7.
493. Puck, J.M., R.L. Nussbaum, and M.E. Conley, *Carrier detection in X-linked severe combined immunodeficiency based on patterns of X chromosome inactivation*. *J Clin Invest*, 1987. **79**(5): p. 1395-400.
494. Goodship, J., et al., *Use of X chromosome inactivation analysis to establish carrier status for X-linked severe combined immunodeficiency*. *Lancet*, 1988. **1**(8588): p. 729-32.
495. Winkelstein, J.A. and E. Fearon, *Carrier detection of the X-linked primary immunodeficiency diseases using X-chromosome inactivation analysis*. *J Allergy Clin Immunol*, 1990. **85**(6): p. 1090-7.

References

496. Wong, C.C., et al., *A longitudinal twin study of skewed X chromosome-inactivation*. PLoS Onc, 2011. **6**(3): p. e17873.
497. Christensen, K., et al., *X-linked genetic factors regulate hematopoietic stem-cell kinetics in females*. Blood, 2000. **95**(7): p. 2449-51.
498. Hatakeyama, C., et al., *The dynamics of X-inactivation skewing as women age*. Clin Genet, 2004. **66**(4): p. 327-32.
499. Sandovici, I., et al., *A longitudinal study of X-inactivation ratio in human females*. Hum Genet, 2004. **115**(5): p. 387-92.
500. Knudsen, G.P., et al., *Increased skewing of X chromosome inactivation with age in both blood and buccal cells*. Cytogenet Genome Res, 2007. **116**(1-2): p. 24-8.
501. Fey, M.F., et al., *Clonality and X-inactivation patterns in hematopoietic cell populations detected by the highly informative M27 beta DNA probe*. Blood, 1994. **83**(4): p. 931-8.
502. Cotter, P.D., et al., *Late-onset X-linked sideroblastic anemia. Missense mutations in the erythroid delta-aminolevulinate synthase (ALAS2) gene in two pyridoxine-responsive patients initially diagnosed with acquired refractory anemia and ringed sideroblasts*. J Clin Invest, 1995. **96**(4): p. 2090-6.
503. Cazzola, M., et al., *Familial-skewed X-chromosome inactivation as a predisposing factor for late-onset X-linked sideroblastic anemia in carrier females*. Blood, 2000. **96**(13): p. 4363-5.
504. Manco, L., et al., *Chronic hemolytic anemia is associated with a new glucose-6-phosphate dehydrogenase in-frame deletion in an older woman*. Blood Cells Mol Dis, 2011. **46**(4): p. 288-93.
505. Au, W.Y., et al., *Glucose 6-phosphate dehydrogenase (G6PD) deficiency in elderly Chinese women heterozygous for G6PD variants*. Am J Med Genet A, 2004. **129A**(2): p. 208-11.
506. Au, W.Y., et al., *Glucose-6-phosphate dehydrogenase deficiency in female octogenarians, nanogenarians, and centenarians*. J Gerontol A Biol Sci Med Sci, 2006. **61**(10): p. 1086-9.
507. Gale, R.E., et al., *Acquired skewing of X-chromosome inactivation patterns in myeloid cells of the elderly suggests stochastic clonal loss with age*. Br J Haematol, 1997. **98**(3): p. 512-9.
508. Mengel-From, J., et al., *Skewed X inactivation and survival: a 13-year follow-up study of elderly twins and singletons*. Eur J Hum Genet, 2012. **20**(3): p. 361-4.
509. Vickers, M.A., et al., *Assessment of mechanism of acquired skewed X inactivation by analysis of twins*. Blood, 2001. **97**(5): p. 1274-81.
510. Abkowitz, J.L., et al., *An X chromosome gene regulates hematopoietic stem cell kinetics*. Proc Natl Acad Sci U S A, 1998. **95**(7): p. 3862-6.
511. Coleman, R., et al., *Interaction of incontinentia pigmenti and factor VIII mutations in a female with biased X inactivation, resulting in haemophilia*. J Med Genet, 1993. **30**(6): p. 497-500.
512. Gibbons, R.J., et al., *X-linked alpha-thalassemia/mental retardation (ATR-X) syndrome: localization to Xq12-q21.31 by X inactivation and linkage analysis*. Am J Hum Genet, 1992. **51**(5): p. 1136-49.
513. Wada, T., et al., *Non-skewed X-inactivation may cause mental retardation in a female carrier of X-linked alpha-thalassemia/mental retardation syndrome (ATR-X): X-inactivation study of nine female carriers of ATR-X*. Am J Med Genet A, 2005. **138**(1): p. 18-20.
514. Orstavik, K.H., et al., *X chromosome inactivation in carriers of Barth syndrome*. Am J Hum Genet, 1998. **63**(5): p. 1457-63.
515. Ferraris, A.M., et al., *Nonrandom X-chromosome inactivation in hemopoietic cells from carriers of dyskeratosis congenita*. Am J Hum Genet, 1997. **61**(2): p. 458-61.
516. Devriendt, K., et al., *Skewed X-chromosome inactivation in female carriers of dyskeratosis congenita*. Am J Hum Genet, 1997. **60**(3): p. 581-7.
517. Vulliamy, T.J., et al., *Skewed X-inactivation in carriers of X-linked dyskeratosis congenita*. Blood, 1997. **90**(6): p. 2213-6.
518. Orstavik, K.H., et al., *Novel splicing mutation in the NEMO (IKK-gamma) gene with severe immunodeficiency and heterogeneity of X-chromosome inactivation*. Am J Med Genet A, 2006. **140**(1): p. 31-9.
519. Van Esch, H., et al., *Duplication of the MECP2 region is a frequent cause of severe mental retardation and progressive neurological symptoms in males*. Am J Hum Genet, 2005. **77**(3): p. 442-53.
520. Jiang, Y.H., et al., *Molecular characterization of co-occurring Duchenne muscular dystrophy and X-linked oculo-facio-cardio-dental syndrome in a girl*. Am J Med Genet A, 2009. **149A**(6): p. 1249-52.
521. Azofeifa, J., et al., *X-chromosome methylation in manifesting and healthy carriers of dystrophinopathies: concordance of activation ratios among first degree female relatives and skewed inactivation as cause of the affected phenotypes*. Hum Genet, 1995. **96**(2): p. 167-76.

References

522. Yoshioka, M., T. Yorifuji, and I. Mituyoshi, *Skewed X inactivation in manifesting carriers of Duchenne muscular dystrophy*. Clin Genet, 1998. **53**(2): p. 102-7.
523. Parolini, O., et al., *X-linked Wiskott-Aldrich syndrome in a girl*. N Engl J Med, 1998. **338**(5): p. 291-5.
524. Puck, J.M. and H.F. Willard, *X inactivation in females with X-linked disease*. N Engl J Med, 1998. **338**(5): p. 325-8.
525. Inoue, H., et al., *X-linked thrombocytopenia in a girl*. Br J Haematol, 2002. **118**(4): p. 1163-5.
526. Andreu, N., et al., *Wiskott-Aldrich syndrome in a female with skewed X-chromosome inactivation*. Blood Cells Mol Dis, 2003. **31**(3): p. 332-7.
527. Conley, M.E., et al., *Atypical Wiskott-Aldrich syndrome in a girl*. Blood, 1992. **80**(5): p. 1264-9.
528. Russell, S.J. and P.D. Nisen, *Random X chromosome inactivation in a female with a variant of Wiskott-Aldrich syndrome*. Br J Haematol, 1995. **90**(1): p. 210-2.
529. Lutskiy, M.I., et al., *Wiskott-Aldrich syndrome in a female*. Blood, 2002. **100**(8): p. 2763-8.
530. Orstavik, K.H., R.E. Orstavik, and M. Schwartz, *Skewed X chromosome inactivation in a female with haemophilia B and in her non-carrier daughter: a genetic influence on X chromosome inactivation?* J Med Genet, 1999. **36**(11): p. 865-6.
531. Knobe, K.E., et al., *Female haemophilia A caused by skewed X inactivation*. Haemophilia, 2008. **14**(4): p. 846-8.
532. Tanner, S.M., et al., *Skewed X-inactivation in a manifesting carrier of X-linked myotubular myopathy and in her non-manifesting carrier mother*. Hum Genet, 1999. **104**(3): p. 249-53.
533. Jungbluth, H., et al., *Early and severe presentation of X-linked myotubular myopathy in a girl with skewed X-inactivation*. Neuromuscul Disord, 2003. **13**(1): p. 55-9.
534. Kristiansen, M., et al., *X-inactivation patterns in carriers of X-linked myotubular myopathy*. Neuromuscul Disord, 2003. **13**(6): p. 468-71.
535. Roos, D., et al., *Molecular basis and enzymatic properties of glucose 6-phosphate dehydrogenase valendam, leading to chronic nonspherocytic anemia, granulocyte dysfunction, and increased susceptibility to infections*. Blood, 1999. **94**(9): p. 2955-62.
536. Badens, C., et al., *ATRX syndrome in a girl with a heterozygous mutation in the ATRX Zn finger domain and a totally skewed X-inactivation pattern*. Am J Med Genet A, 2006. **140**(20): p. 2212-5.
537. Morrone, A., et al., *Fabry disease: molecular studies in Italian patients and X inactivation analysis in manifesting carriers*. J Med Genet, 2003. **40**(8): p. e103.
538. Mattei, M.G., et al., *X-autosome translocations: cytogenetic characteristics and their consequences*. Hum Genet, 1982. **61**(4): p. 295-309.
539. Schmidt, M. and D. Du Sart, *Functional disomies of the X chromosome influence the cell selection and hence the X inactivation pattern in females with balanced X-autosome translocations: a review of 122 cases*. Am J Med Genet, 1992. **42**(2): p. 161-9.
540. Orstavik, K.H., et al., *Severe hypohidrotic ectodermal dysplasia in a girl caused by a de novo 9;X insertion that includes XIST and disrupts the EDA gene*. Am J Med Genet A, 2007. **143A**(13): p. 1510-3.
541. Hodgson, S.V., et al., *A balanced de novo X/autosome translocation in a girl with manifestations of Lowe syndrome*. Am J Med Genet, 1986. **23**(3): p. 837-47.
542. Punnett, H.H., *Simpson-Golabi-Behmel syndrome (SGBS) in a female with an X-autosome translocation*. Am J Med Genet, 1994. **50**(4): p. 391-3.
543. Imai, K., et al., *Female hyper IgM syndrome type 1 with a chromosomal translocation disrupting CD40LG*. Biochim Biophys Acta, 2006. **1762**(3): p. 335-40.
544. Monaco, A.P., et al., *Isolation of candidate cDNAs for portions of the Duchenne muscular dystrophy gene*. Nature, 1986. **323**(6089): p. 646-50.
545. Boyd, Y., et al., *Muscular dystrophy in girls with X;autosome translocations*. J Med Genet, 1986. **23**(6): p. 484-90.
546. Boyd, Y., et al., *Mapping of 12 translocation breakpoints in the Xp21 region with respect to the locus for Duchenne muscular dystrophy*. Cytogenet Cell Genet, 1988. **48**(1): p. 28-34.
547. Jacobs, P.A., et al., *Duchenne muscular dystrophy (DMD) in a female with an X/autosome translocation: further evidence that the DMD locus is at Xp21*. Am J Hum Genet, 1981. **33**(4): p. 513-8.
548. Bodrug, S.E., et al., *Mapping of four translocation breakpoints within the Duchenne muscular dystrophy gene*. Genomics, 1989. **4**(1): p. 101-4.
549. Billuart, P., et al., *Oligophrenin-1 encodes a rhoGAP protein involved in X-linked mental retardation*. Nature, 1998. **392**(6679): p. 923-6.

References

550. Bienvenu, T., et al., *Mapping of the X-breakpoint involved in a balanced X:12 translocation in a female with mild mental retardation*. Eur J Hum Genet, 1997. **5**(2): p. 105-9.
551. Bawle, E., et al., *Aarskog syndrome: full male and female expression associated with an X-autosome translocation*. Am J Med Genet, 1984. **17**(3): p. 595-602.
552. Hodgson, S.V., et al., *Two cases of X/autosome translocation in females with incontinentia pigmenti*. Hum Genet, 1985. **71**(3): p. 231-4.
553. Royer-Pokora, B., et al., *Cloning the gene for an inherited human disorder--chronic granulomatous disease--on the basis of its chromosomal location*. Nature, 1986. **322**(6074): p. 32-8.
554. Baehner, R.L., et al., *DNA linkage analysis of X chromosome-linked chronic granulomatous disease*. Proc Natl Acad Sci U S A, 1986. **83**(10): p. 3398-401.
555. Orstavik, K.H., *X chromosome inactivation in clinical practice*. Hum Genet, 2009. **126**(3): p. 363-73.
556. Franco, B. and A. Ballabio, *X-inactivation and human disease: X-linked dominant male-lethal disorders*. Curr Opin Genet Dev, 2006. **16**(3): p. 254-9.
557. Morleo, M. and B. Franco, *Dosage compensation of the mammalian X chromosome influences the phenotypic variability of X-linked dominant male-lethal disorders*. J Med Genet, 2008. **45**(7): p. 401-8.
558. Bahi-Buisson, N., et al., *Key clinical features to identify girls with CDKL5 mutations*. Brain, 2008. **131**(Pt 10): p. 2647-61.
559. Thauvin-Robinet, C., et al., *Clinical, molecular, and genotype-phenotype correlation studies from 25 cases of oral-facial-digital syndrome type 1: a French and Belgian collaborative study*. J Med Genet, 2006. **43**(1): p. 54-61.
560. Robertson, S.P., et al., *Localized mutations in the gene encoding the cytoskeletal protein filamin A cause diverse malformations in humans*. Nat Genet, 2003. **33**(4): p. 487-91.
561. Parrish, J.E., et al., *Selection against mutant alleles in blood leukocytes is a consistent feature in Incontinentia Pigmenti type 2*. Hum Mol Genet, 1996. **5**(11): p. 1777-83.
562. Shirahama, S., et al., *Skewed X-chromosome inactivation causes intra-familial phenotypic variation of an EBP mutation in a family with X-linked dominant chondrodysplasia punctata*. Hum Genet, 2003. **112**(1): p. 78-83.
563. Hedera, P. and J.L. Gorski, *Oculo-facio-cardio-dental syndrome: skewed X chromosome inactivation in mother and daughter suggest X-linked dominant inheritance*. Am J Med Genet A, 2003. **123A**(3): p. 261-6.
564. Fusco, F., et al., *Molecular analysis of the genetic defect in a large cohort of IP patients and identification of novel NEMO mutations interfering with NF-kappaB activation*. Hum Mol Genet, 2004. **13**(16): p. 1763-73.
565. Kristiansen, M., et al., *Phenotypic variation in Melnick-Needles syndrome is not reflected in X inactivation patterns from blood or buccal smear*. Am J Med Genet, 2002. **108**(2): p. 120-7.
566. Knudsen, G.P., et al., *Increased skewing of X chromosome inactivation in Rett syndrome patients and their mothers*. Eur J Hum Genet, 2006. **14**(11): p. 1189-94.
567. Robertson, S.P., et al., *Frontometaphyseal dysplasia: mutations in FLNA and phenotypic diversity*. Am J Med Genet A, 2006. **140**(16): p. 1726-36.
568. Leoyklang, P., et al., *Three novel mutations in the PORCN gene underlying focal dermal hypoplasia*. Clin Genet, 2008. **73**(4): p. 373-9.
569. Blinkenberg, E.O., et al., *Angioma serpiginosum with oesophageal papillomatosis is an X-linked dominant condition that maps to Xp11.3-Xq12*. Eur J Hum Genet, 2007. **15**(5): p. 543-7.
570. Amir, R.E., et al., *Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2*. Nat Genet, 1999. **23**(2): p. 185-8.
571. Chahrour, M. and H.Y. Zoghbi, *The story of Rett syndrome: from clinic to neurobiology*. Neuron, 2007. **56**(3): p. 422-37.
572. Dayer, A.G., et al., *MECP2 mutant allele in a boy with Rett syndrome and his unaffected heterozygous mother*. Brain Dev, 2007. **29**(1): p. 47-50.
573. Huppke, P., et al., *Very mild cases of Rett syndrome with skewed X inactivation*. J Med Genet, 2006. **43**(10): p. 814-6.
574. Allanson, J. and S. Richter, *Linear skin defects and congenital microphthalmia: a new syndrome at Xp22.2*. J Med Genet, 1991. **28**(2): p. 143-4.
575. Lindsay, E.A., et al., *Microphthalmia with linear skin defects (MLS) syndrome: clinical, cytogenetic, and molecular characterization*. Am J Med Genet, 1994. **49**(2): p. 229-34.

576. Ogata, T., et al., *Microphthalmia with linear skin defects syndrome in a mosaic female infant with monosomy for the Xp22 region: molecular analysis of the Xp22 breakpoint and the X-inactivation pattern*. Hum Genet, 1998. **103**(1): p. 51-6.
577. Carrel, L., et al., *A first-generation X-inactivation profile of the human X chromosome*. Proc Natl Acad Sci U S A, 1999. **96**(25): p. 14440-4.
578. de Conciliis, L., et al., *Characterization of Cxorf5 (71-7A), a novel human cDNA mapping to Xp22 and encoding a protein containing coiled-coil alpha-helical domains*. Genomics, 1998. **51**(2): p. 243-50.
579. Ferrante, M.I., et al., *Identification of the gene for oral-facial-digital type I syndrome*. Am J Hum Genet, 2001. **68**(3): p. 569-76.
580. Ferrante, M.I., et al., *Oral-facial-digital type I protein is required for primary cilia formation and left-right axis specification*. Nat Genet, 2006. **38**(1): p. 112-7.
581. Cai, X.H., et al., *Female hemophilia A heterozygous for a de novo frameshift and a novel missense mutation of factor VIII*. J Thromb Haemost, 2006. **4**(9): p. 1969-74.
582. Windsor, S., et al., *Severe haemophilia A in a female resulting from two de novo factor VIII mutations*. Br J Haematol, 1995. **90**(4): p. 906-9.
583. David, D., et al., *Female haemophiliac homozygous for the factor VIII intron 22 inversion mutation, with transcriptional inactivation of one of the factor VIII alleles*. Haemophilia, 2003. **9**(1): p. 125-30.
584. Vencesla, A., et al., *Severe haemophilia A in a female resulting from an inherited gross deletion and a de novo codon deletion in the F8 gene*. Haemophilia, 2008. **14**(5): p. 1094-8.
585. Radhakrishna, U., et al., *Novel homozygous, heterozygous and hemizygous FRMD7 gene mutations segregated in the same consanguineous family with congenital X-linked nystagmus*. Eur J Hum Genet, 2012.
586. Nagtzaam, I.F., et al., *Clinically manifest X-linked recessive ichthyosis in a female due to a homozygous interstitial 1.6-Mb deletion of Xp22.31*. Br J Dermatol, 2012. **166**(4): p. 905-7.
587. Saito, T., et al., *A patient with hypophosphatemic rickets and ossification of posterior longitudinal ligament caused by a novel homozygous mutation in ENPP1 gene*. Bone, 2011. **49**(4): p. 913-6.
588. Fujii, K., et al., *Homozygous female Becker muscular dystrophy*. Am J Med Genet A, 2009. **149A**(5): p. 1052-5.
589. Israels, M.C., H. Lempert, and E. Gilbertson, *Haemophilia in the female*. Lancet, 1951. **1**(6670): p. 1375-80.
590. Merskey, C., *The occurrence of haemophilia in the human female*. Q J Med, 1951. **20**(79): p. 299-312.
591. Quan, F., et al., *Uniparental disomy of the entire X chromosome in a female with Duchenne muscular dystrophy*. Am J Hum Genet, 1997. **60**(1): p. 160-5.
592. Solomon, I.L. and E.J. Schoen, *Sex-linked ichthyosis in XO gonadal dysgenesis*. Lancet, 1971. **1**(7712): p. 1304-5.
593. Chelly, J., et al., *De novo DNA microdeletion in a girl with Turner syndrome and Duchenne muscular dystrophy*. Hum Genet, 1986. **74**(2): p. 193-6.
594. Gilgenkrantz, S., et al., *A case of female hemophilia with a 46,XXr karyotype studied with X-chromosome DNA probes*. Hum Genet, 1986. **72**(2): p. 157-9.
595. Panarello, C., et al., *Concomitant Turner syndrome and hemophilia A in a female with an idic(X)(p11) heterozygous at locus DXS52*. Cytogenet Cell Genet, 1992. **59**(4): p. 241-2.
596. Gilchrist, G.S., D. Hammond, and J. Melnyk, *Hemophilia A in a phenotypically normal female with XX-XO mosaicism*. N Engl J Med, 1965. **273**(26): p. 1402-6.
597. Chuansumrit, A., et al., *Inversion of intron 22 of the factor VIII gene in a girl with severe hemophilia A and Turner's syndrome*. Thromb Haemost, 1999. **82**(4): p. 1379.
598. Weinspach, S., et al., *Intracranial hemorrhage in a female leading to the diagnosis of severe hemophilia A and Turner syndrome*. Klin Padiatr, 2009. **221**(3): p. 167-71.
599. Katayama, Y., et al., *Co-occurrence of mutations in both dystrophin- and androgen-receptor genes is a novel cause of female Duchenne muscular dystrophy*. Hum Genet, 2006. **119**(5): p. 516-9.
600. Nilsson, I.M., et al., *Haemophilia A in a "girl" with male sex-chromatin pattern*. Lancet, 1959. **2**(7097): p. 264-6.
601. Huisse, M.G., et al., *Hemophilia A in a phenotypic female with normal male karyotype associated with a low factor XII level*. Ann Genet, 1980. **23**(1): p. 31-4.
602. Sitalakshmi, S., et al., *An unusual case of haemophilia--a case report*. Indian J Pathol Microbiol, 2001. **44**(3): p. 367-8.
603. Andrejev, N.J., et al., *Haemophilia A in a patient with testicular feminization*. Thromb Diath Haemorrh, 1975. **33**(2): p. 208-16.

References

604. Witkowski, R., *Germinal "mosaicism"--germline mutation or chimerism?* Hum Genet, 1992. **88**(3): p. 359-60.
605. Malan, V., M. Vekemans, and C. Turleau, *Chimera and other fertilization errors*. Clin Genet, 2006. **70**(5): p. 363-73.
606. Wieacker, P., J. Zimmer, and H.H. Ropers, *X inactivation patterns in two syndromes with probable X-linked dominant, male lethal inheritance*. Clin Genet, 1985. **28**(3): p. 238-42.
607. Neidich, J.A., et al., *Heterogeneity of clinical severity and molecular lesions in Aicardi syndrome*. J Pediatr, 1990. **116**(6): p. 911-7.
608. Hoag, H.M., et al., *Evidence that skewed X inactivation is not needed for the phenotypic expression of Aicardi syndrome*. Hum Genet, 1997. **100**(3-4): p. 459-64.
609. Eble, T.N., et al., *Non-random X chromosome inactivation in Aicardi syndrome*. Hum Genet, 2009. **125**(2): p. 211-6.
610. Canueto, J., et al., *Clinical, molecular and biochemical characterization of nine Spanish families with Conradi-Hunermann-Happle syndrome: new insights into X-linked dominant chondrodysplasia punctata with a comprehensive review of the literature*. Br J Dermatol, 2012. **166**(4): p. 830-8.
611. Konig, A., et al., *A novel missense mutation of NSDHL in an unusual case of CHILD syndrome showing bilateral, almost symmetric involvement*. J Am Acad Dermatol, 2002. **46**(4): p. 594-6.
612. Schluth, C., et al., *Phenotype in X chromosome rearrangements: pitfalls of X inactivation study*. Pathol Biol (Paris), 2007. **55**(1): p. 29-36.
613. Baroncini, A., et al., *Terminal osseous dysplasia with pigmentary defects: clinical description of a new family*. Am J Med Genet A, 2007. **143**(1): p. 51-7.
614. Cutler, J.A., et al., *Germline mosaicism resulting in the transmission of severe hemophilia B from a grandfather with a mild deficiency*. Am J Med Genet A, 2004. **129A**(1): p. 13-5.
615. Leuer, M., et al., *Somatic mosaicism in hemophilia A: a fairly common event*. Am J Hum Genet, 2001. **69**(1): p. 75-87.
616. Youssefian, H. and R.E. Pyeritz, *Mechanisms and consequences of somatic mosaicism in humans*. Nat Rev Genet, 2002. **3**(10): p. 748-58.
617. Kasper, C.K. and C.H. Buzin, *Mosaics and haemophilia*. Haemophilia, 2009. **15**(6): p. 1181-6.
618. Rogaev, E.I., et al., *Genotype analysis identifies the cause of the "royal disease"*. Science, 2009. **326**(5954): p. 817.
619. Stevens, R.F., *The history of haemophilia in the royal families of Europe*. Br J Haematol, 1999. **105**(1): p. 25-32.
620. Orstavik, K.H., et al., *Absence of correlation between X chromosome inactivation pattern and plasma concentration of factor VIII and factor IX in carriers of haemophilia A and B*. Thromb Haemost, 2000. **83**(3): p. 433-7.
621. Song, M.J., et al., *Molecular characterization of female hemophilia A by multiplex ligation-dependent probe amplification analysis and X-chromosome inactivation study*. Blood Coagul Fibrinolysis, 2011. **22**(3): p. 211-4.
622. Luzzatto, L., et al., *Imbalance in X-chromosome expression: evidence for a human X-linked gene affecting growth of hemopoietic cells*. Science, 1979. **205**(4413): p. 1418-20.
623. Usanga, E.A. and L. Luzzatto, *Adaptation of Plasmodium falciparum to glucose 6-phosphate dehydrogenase-deficient host red cells by production of parasite-encoded enzyme*. Nature, 1985. **313**(6005): p. 793-5.
624. Cappadoro, M., et al., *Early phagocytosis of glucose-6-phosphate dehydrogenase (G6PD)-deficient erythrocytes parasitized by Plasmodium falciparum may explain malaria protection in G6PD deficiency*. Blood, 1998. **92**(7): p. 2527-34.
625. Green, A.M. and G.M. Kupfer, *Fanconi anemia*. Hematol Oncol Clin North Am, 2009. **23**(2): p. 193-214.
626. Harigae, H. and K. Furuyama, *Hereditary sideroblastic anemia: pathophysiology and gene mutations*. Int J Hematol, 2010. **92**(3): p. 425-31.
627. Kacena, M.A., et al., *GATA1-Related X-Linked Cytopenia*. 1993.
628. Holden, S.T., et al., *Fanconi anaemia complementation group B presenting as X linked VACTERL with hydrocephalus syndrome*. J Med Genet, 2006. **43**(9): p. 750-4.
629. McCauley, J., et al., *X-linked VACTERL with hydrocephalus syndrome: further delineation of the phenotype caused by FANCB mutations*. Am J Med Genet A, 2011. **155A**(10): p. 2370-80.
630. Aivado, M., N. Gattermann, and S. Bottomley, *X chromosome inactivation ratios in female carriers of X-linked sideroblastic anemia*. Blood, 2001. **97**(12): p. 4000-2.

631. Aivado, M., et al., *X-linked sideroblastic anemia associated with a novel ALAS2 mutation and unfavourable skewed X-chromosome inactivation patterns*. *Blood Cells Mol Dis*, 2006. **37**(1): p. 40-5.
632. Ducamp, S., et al., *Sideroblastic anemia: molecular analysis of the ALAS2 gene in a series of 29 probands and functional studies of 10 missense mutations*. *Hum Mutat*, 2011. **32**(6): p. 590-7.
633. Freson, K., et al., *Different substitutions at residue D218 of the X-linked transcription factor GATA1 lead to altered clinical severity of macrothrombocytopenia and anemia and are associated with variable skewed X inactivation*. *Hum Mol Genet*, 2002. **11**(2): p. 147-52.
634. Fish, E.N., *The X-files in immunity: sex-based differences predispose immune responses*. *Nat Rev Immunol*, 2008. **8**(9): p. 737-44.
635. Thompson, D.J., et al., *Excess risk of staphylococcal infection and disease in newborn males*. *Am J Epidemiol*, 1966. **84**(2): p. 314-28.
636. Green, M.S., *The male predominance in the incidence of infectious diseases in children: a postulated explanation for disparities in the literature*. *Int J Epidemiol*, 1992. **21**(2): p. 381-6.
637. Purtilo, D.T. and J.L. Sullivan, *Immunological bases for superior survival of females*. *Am J Dis Child*, 1979. **133**(12): p. 1251-3.
638. Butterworth, M., B. McClellan, and M. Allansmith, *Influence of sex in immunoglobulin levels*. *Nature*, 1967. **214**(5094): p. 1224-5.
639. Aspinall, R., *Longevity and the immune response*. *Biogerontology*, 2000. **1**(3): p. 273-8.
640. Libert, C., L. Dejager, and I. Pinheiro, *The X chromosome in immune functions: when a chromosome makes the difference*. *Nat Rev Immunol*, 2010. **10**(8): p. 594-604.
641. Morris, J.A. and L.M. Harrison, *Hypothesis: increased male mortality caused by infection is due to a decrease in heterozygous loci as a result of a single X chromosome*. *Med Hypotheses*, 2009. **72**(3): p. 322-4.
642. Spolarics, Z., *The X-files of inflammation: cellular mosaicism of X-linked polymorphic genes and the female advantage in the host response to injury and infection*. *Shock*, 2007. **27**(6): p. 597-604.
643. Selmi, C., et al., *The X chromosome and the sex ratio of autoimmunity*. *Autoimmun Rev*, 2011.
644. Gleicher, N. and D.H. Barad, *Gender as risk factor for autoimmune diseases*. *J Autoimmun*, 2007. **28**(1): p. 1-6.
645. Koker, M.Y., et al., *Skewing of X-chromosome inactivation in three generations of carriers with X-linked chronic granulomatous disease within one family*. *Eur J Clin Invest*, 2006. **36**(4): p. 257-64.
646. Notarangelo, L.D., et al., *Analysis of X-chromosome inactivation in X-linked immunodeficiency with hyper-IgM (HIGM1): evidence for involvement of different hematopoietic cell lineages*. *Hum Genet*, 1991. **88**(2): p. 130-4.
647. Hendriks, R.W., et al., *Evidence that in X-linked immunodeficiency with hyperimmunoglobulinemia M the intrinsic immunoglobulin heavy chain class switch mechanism is intact*. *Eur J Immunol*, 1990. **20**(12): p. 2603-8.
648. Tommasini, A., et al., *X-chromosome inactivation analysis in a female carrier of FOXP3 mutation*. *Clin Exp Immunol*, 2002. **130**(1): p. 127-30.
649. van den Bogaard, R., et al., *Molecular characterisation of 10 Dutch properdin type I deficient families: mutation analysis and X-inactivation studies*. *Eur J Hum Genet*, 2000. **8**(7): p. 513-8.
650. Woon, S.T., et al., *Follicular lymphoma in a X-linked lymphoproliferative syndrome carrier female*. *Scand J Immunol*, 2008. **68**(2): p. 153-8.
651. Marsh, R.A., et al., *A rapid flow cytometric screening test for X-linked lymphoproliferative disease due to XIAP deficiency*. *Cytometry B Clin Cytom*, 2009. **76**(5): p. 334-44.
652. Ozcelik, T., *X chromosome inactivation and female predisposition to autoimmunity*. *Clin Rev Allergy Immunol*, 2008. **34**(3): p. 348-51.
653. Kast, R.E., *Predominance of autoimmune and rheumatic diseases in females*. *J Rheumatol*, 1977. **4**(3): p. 288-92.
654. Stewart, J.J., *The female X-inactivation mosaic in systemic lupus erythematosus*. *Immunol Today*, 1998. **19**(8): p. 352-7.
655. Uz, E., et al., *Increased frequency of extremely skewed X chromosome inactivation in juvenile idiopathic arthritis*. *Arthritis Rheum*, 2009. **60**(11): p. 3410-2.
656. Broen, J.C., et al., *Skewed X-chromosomal inactivation impacts T regulatory cell function in systemic sclerosis*. *Ann Rheum Dis*, 2010. **69**(12): p. 2213-6.
657. Brix, T.H., et al., *High frequency of skewed X-chromosome inactivation in females with autoimmune thyroid disease: a possible explanation for the female predisposition to thyroid autoimmunity*. *J Clin Endocrinol Metab*, 2005. **90**(11): p. 5949-53.

References

658. Ozbalkan, Z., et al., *Skewed X chromosome inactivation in blood cells of women with scleroderma*. *Arthritis Rheum*, 2005. **52**(5): p. 1564-70.
659. Ozcelik, T., et al., *Evidence from autoimmune thyroiditis of skewed X-chromosome inactivation in female predisposition to autoimmunity*. *Eur J Hum Genet*, 2006. **14**(6): p. 791-7.
660. Uz, E., et al., *Skewed X-chromosome inactivation in scleroderma*. *Clin Rev Allergy Immunol*, 2008. **34**(3): p. 352-5.
661. Scofield, R.H., et al., *Klinefelter's syndrome (47,XXY) in male systemic lupus erythematosus patients: support for the notion of a gene-dose effect from the X chromosome*. *Arthritis Rheum*, 2008. **58**(8): p. 2511-7.
662. Knudsen, G.P., et al., *X chromosome inactivation in females with multiple sclerosis*. *Eur J Neurol*, 2007. **14**(12): p. 1392-6.
663. Chitnis, S., et al., *The role of X-chromosome inactivation in female predisposition to autoimmunity*. *Arthritis Res*, 2000. **2**(5): p. 399-406.
664. Lu, Q., et al., *Demethylation of CD40LG on the inactive X in T cells from women with lupus*. *J Immunol*, 2007. **179**(9): p. 6352-8.
665. Forsdyke, D.R., *X chromosome reactivation perturbs intracellular self/not-self discrimination*. *Immunol Cell Biol*, 2009. **87**(7): p. 525-8.
666. Zhou, Y., et al., *T cell CD40LG gene expression and the production of IgG by autologous B cells in systemic lupus erythematosus*. *Clin Immunol*, 2009. **132**(3): p. 362-70.
667. Brooks, W.H., *X chromosome inactivation and autoimmunity*. *Clin Rev Allergy Immunol*, 2010. **39**(1): p. 20-9.
668. Ranke, M.B. and P. Saenger, *Turner's syndrome*. *Lancet*, 2001. **358**(9278): p. 309-14.
669. Elsheikh, M., J.A. Wass, and G.S. Conway, *Autoimmune thyroid syndrome in women with Turner's syndrome--the association with karyotype*. *Clin Endocrinol (Oxf)*, 2001. **55**(2): p. 223-6.
670. Sybert, V.P. and E. McCauley, *Turner's syndrome*. *N Engl J Med*, 2004. **351**(12): p. 1227-38.
671. Invernizzi, P., et al., *X chromosome monosomy: a common mechanism for autoimmune diseases*. *J Immunol*, 2005. **175**(1): p. 575-8.
672. Invernizzi, P., et al., *Frequency of monosomy X in women with primary biliary cirrhosis*. *Lancet*, 2004. **363**(9408): p. 533-5.
673. Invernizzi, P., et al., *X monosomy in female systemic lupus erythematosus*. *Ann N Y Acad Sci*, 2007. **1110**: p. 84-91.
674. Chiurazzi, P., et al., *XLMR genes: update 2007*. *Eur J Hum Genet*, 2008. **16**(4): p. 422-34.
675. Bonnet, C., et al., *Exploring the potential role of disease-causing mutation in a gene desert: duplication of noncoding elements 5' of GRIA3 is associated with GRIA3 silencing and X-linked intellectual disability*. *Hum Mutat*, 2012. **33**(2): p. 355-8.
676. Yonath, H., et al., *X inactivation testing for identifying a non-syndromic X-linked mental retardation gene*. *J Appl Genet*, 2011. **52**(4): p. 437-41.
677. Plenge, R.M., et al., *Skewed X-chromosome inactivation is a common feature of X-linked mental retardation disorders*. *Am J Hum Genet*, 2002. **71**(1): p. 168-73.
678. Bittel, D.C., et al., *Comparison of X-chromosome inactivation patterns in multiple tissues from human females*. *J Med Genet*, 2008. **45**(5): p. 309-13.
679. van de Kamp, J.M., et al., *Clinical features and X-inactivation in females heterozygous for creatine transporter defect*. *Clin Genet*, 2011. **79**(3): p. 264-72.
680. Oostra, B.A. and R. Willemsen, *The X chromosome and fragile X mental retardation*. *Cytogenet Genome Res*, 2002. **99**(1-4): p. 257-64.
681. Kirchgessner, C.U., S.T. Warren, and H.F. Willard, *X inactivation of the FMR1 fragile X mental retardation gene*. *J Med Genet*, 1995. **32**(12): p. 925-9.
682. Fryns, J.P., et al., *Inactivation pattern of the fragile X in heterozygous carriers*. *Hum Genet*, 1984. **65**(4): p. 400-1.
683. Schmidt, M., et al., *Unusual X chromosome inactivation in a mentally retarded girl with an interstitial deletion Xq27: implications for the fragile X syndrome*. *Hum Genet*, 1990. **84**(4): p. 347-52.
684. Martinez, R., et al., *Skewed X inactivation of the normal allele in fully mutated female carriers determines the levels of FMRP in blood and the fragile X phenotype*. *Mol Diagn*, 2005. **9**(3): p. 157-62.
685. Martorell, L., et al., *Four sisters compound heterozygotes for the pre- and full mutation in fragile X syndrome and a complete inactivation of X-functional chromosome: implications for genetic counseling*. *J Hum Genet*, 2011. **56**(1): p. 87-90.

686. Heine-Suner, D., et al., *Fragile-X syndrome and skewed X-chromosome inactivation within a family: a female member with complete inactivation of the functional X chromosome*. Am J Med Genet A, 2003. **122A**(2): p. 108-14.
687. Nolin, S.L., et al., *Mosaicism in fragile X affected males*. Am J Med Genet, 1994. **51**(4): p. 509-12.
688. de Vries, B.B., et al., *Variable FMR1 gene methylation of large expansions leads to variable phenotype in three males from one fragile X family*. J Med Genet, 1996. **33**(12): p. 1007-10.
689. Willemsen, R., et al., *Twin sisters, monozygotic with the fragile X mutation, but with a different phenotype*. J Med Genet, 2000. **37**(8): p. 603-4.
690. Garcia-Alegria, E., et al., *Analysis of FMR1 gene expression in female premutation carriers using robust segmented linear regression models*. RNA, 2007. **13**(5): p. 756-62.
691. Ramocki, M.B., et al., *Autism and other neuropsychiatric symptoms are prevalent in individuals with MECP2 duplication syndrome*. Ann Neurol, 2009. **66**(6): p. 771-82.
692. del Gaudio, D., et al., *Increased MECP2 gene copy number as the result of genomic duplication in neurodevelopmentally delayed males*. Genet Med, 2006. **8**(12): p. 784-92.
693. Meins, M., et al., *Submicroscopic duplication in Xq28 causes increased expression of the MECP2 gene in a boy with severe mental retardation and features of Rett syndrome*. J Med Genet, 2005. **42**(2): p. e12.
694. Ramocki, M.B., Y.J. Tavyev, and S.U. Peters, *The MECP2 duplication syndrome*. Am J Med Genet A, 2010. **152A**(5): p. 1079-88.
695. Kemohan, K.D., et al., *ATRX partners with cohesin and MeCP2 and contributes to developmental silencing of imprinted genes in the brain*. Dev Cell, 2010. **18**(2): p. 191-202.
696. Honda, S., et al., *Concomitant microduplications of MECP2 and ATRX in male patients with severe mental retardation*. J Hum Genet, 2011.
697. De La Fuente, R., C. Baumann, and M.M. Viveiros, *Role of ATRX in chromatin structure and function: implications for chromosome instability and human disease*. Reproduction, 2011. **142**(2): p. 221-34.
698. Grasshoff, U., et al., *De novo MECP2 duplication in two females with random X-inactivation and moderate mental retardation*. Eur J Hum Genet, 2011. **19**(5): p. 507-12.
699. Mayo, S., et al., *De novo interstitial triplication of MECP2 in a girl with neurodevelopmental disorder and random X chromosome inactivation*. Cytogenet Genome Res, 2011. **135**(2): p. 93-101.
700. Pichavant, C., et al., *Current status of pharmaceutical and genetic therapeutic approaches to treat DMD*. Mol Ther, 2011. **19**(5): p. 830-40.
701. Schanzer, A., et al., *Duchenne Muscular Dystrophy in a 4-Year-Old Girl due to Heterozygous Frame Shift Deletion of the Dystrophin Gene and Skewed X-Inactivation*. Klin Padiatr, 2012.
702. Soltanzadeh, P., et al., *Clinical and genetic characterization of manifesting carriers of DMD mutations*. Neuromuscul Disord, 2010. **20**(8): p. 499-504.
703. Lesca, G., et al., *[Symptomatic carriers of dystrophinopathy with chromosome X inactivation bias]*. Rev Neurol (Paris), 2003. **159**(8-9): p. 775-80.
704. Melacini, P., et al., *Cardiac transplantation in a Duchenne muscular dystrophy carrier*. Neuromuscul Disord, 1998. **8**(8): p. 585-90.
705. Yoon, J., et al., *Carrier woman of Duchenne muscular dystrophy mimicking inflammatory myositis*. J Korean Med Sci, 2011. **26**(4): p. 587-91.
706. Seemann, N., et al., *Symptomatic dystrophinopathies in female children*. Neuromuscul Disord, 2011. **21**(3): p. 172-7.
707. Takeda, A., et al., *Eponym: Barth syndrome*. Eur J Pediatr, 2011. **170**(11): p. 1365-7.
708. Grogan, P.M., et al., *Myopathy with skeletal asymmetry and hemidiaphragm elevation is caused by myotubularin mutations*. Neurology, 2005. **64**(9): p. 1638-40.
709. Hedberg, C., et al., *Myopathy in a woman and her daughter associated with a novel splice site MTM1 mutation*. Neuromuscul Disord, 2012. **22**(3): p. 244-51.
710. Sutton, I.J., et al., *Limb girdle and facial weakness in female carriers of X-linked myotubular myopathy mutations*. Neurology, 2001. **57**(5): p. 900-2.
711. Drouot, A., et al., *[Unilateral presentation of X-linked myotubular myopathy (XLMTM) in two out of three female carriers in a family with no affected male]*. Rev Neurol (Paris), 2008. **164**(2): p. 169-76.
712. Finsterer, J., *Perspectives of Kennedy's disease*. J Neurol Sci, 2010. **298**(1-2): p. 1-10.
713. Mariotti, C., et al., *Phenotypic manifestations associated with CAG-repeat expansion in the androgen receptor gene in male patients and heterozygous females: a clinical and molecular study of 30 families*. Neuromuscul Disord, 2000. **10**(6): p. 391-7.

References

714. Sobue, G., et al., *Subclinical phenotypic expressions in heterozygous females of X-linked recessive bulbospinal neuronopathy*. J Neurol Sci, 1993. **117**(1-2): p. 74-8.
715. Chen, R.S., et al., *Random X chromosome methylation patterns in the carriers with clinical phenotypic expressions of X-linked recessive bulbospinal neuronopathy*. Acta Neurol Scand, 1999. **100**(4): p. 249-53.
716. Ishihara, H., et al., *Clinical features and skewed X-chromosome inactivation in female carriers of X-linked recessive spinal and bulbar muscular atrophy*. J Neurol, 2001. **248**(10): p. 856-60.
717. Paradas, C., et al., *Highly skewed inactivation of the wild-type X-chromosome in asymptomatic female carriers of spinal and bulbar muscular atrophy (Kennedy's disease)*. J Neurol, 2008. **255**(6): p. 853-7.
718. Yang, Z. and M. Vatta, *Danon disease as a cause of autophagic vacuolar myopathy*. Congenit Heart Dis, 2007. **2**(6): p. 404-9.
719. Majer, F., et al., *Danon disease: a focus on processing of the novel LAMP2 mutation and comments on the beneficial use of peripheral white blood cells in the diagnosis of LAMP2 deficiency*. Gene, 2012. **498**(2): p. 183-95.
720. Fanin, M., et al., *Generalized lysosome-associated membrane protein-2 defect explains multisystem clinical involvement and allows leukocyte diagnostic screening in Danon disease*. Am J Pathol, 2006. **168**(4): p. 1309-20.
721. Yiu, E.M., et al., *A retrospective review of X-linked Charcot-Marie-Tooth disease in childhood*. Neurology, 2011. **76**(5): p. 461-6.
722. Lin, G.S., et al., *A unique mutation in connexin32 associated with severe, early onset CMTX in a heterozygous female*. Ann N Y Acad Sci, 1999. **883**: p. 481-4.
723. Siskind, C.E., et al., *Phenotype expression in women with CMT1X*. J Peripher Nerv Syst, 2011. **16**(2): p. 102-7.
724. Murphy, S.M., et al., *X inactivation in females with X-linked Charcot-Marie-Tooth disease*. Neuromuscul Disord, 2012.
725. Yoriifuji, T., et al., *X-inactivation pattern in the liver of a manifesting female with ornithine transcarbamylase (OTC) deficiency*. Clin Genet, 1998. **54**(4): p. 349-53.
726. Gale, R.E., et al., *Tissue specificity of X-chromosome inactivation patterns*. Blood, 1994. **83**(10): p. 2899-905.
727. Pinto, L.L., et al., *Expression of the disease on female carriers of X-linked lysosomal disorders: a brief review*. Orphanet J Rare Dis, 2010. **5**: p. 14.
728. Zarate, Y.A. and R.J. Hopkin, *Fabry's disease*. Lancet, 2008. **372**(9647): p. 1427-35.
729. Kaneski, C.R., et al., *Use of lissamine rhodamine ceramide trihexoside as a functional assay for alpha-galactosidase A in intact cells*. J Lipid Res, 2010. **51**(9): p. 2808-17.
730. Wang, R.Y., et al., *Heterozygous Fabry women are not just carriers, but have a significant burden of disease and impaired quality of life*. Genet Med, 2007. **9**(1): p. 34-45.
731. Giacomini, P.S., et al., *Fabry's disease presenting as stroke in a young female*. Can J Neurol Sci, 2004. **31**(1): p. 112-4.
732. Dobrovolny, R., et al., *Relationship between X-inactivation and clinical involvement in Fabry heterozygotes. Eleven novel mutations in the alpha-galactosidase A gene in the Czech and Slovak population*. J Mol Med (Berl), 2005. **83**(8): p. 647-54.
733. Maier, E.M., et al., *Disease manifestations and X inactivation in heterozygous females with Fabry disease*. Acta Paediatr Suppl, 2006. **95**(451): p. 30-8.
734. Elstein, D., et al., *X-inactivation in Fabry disease*. Gene, 2012.
735. Menkes, J.H., et al., *A sex-linked recessive disorder with retardation of growth, peculiar hair, and focal cerebral and cerebellar degeneration*. Pediatrics, 1962. **29**: p. 764-79.
736. Vulpe, C., et al., *Isolation of a candidate gene for Menkes disease and evidence that it encodes a copper-transporting ATPase*. Nat Genet, 1993. **3**(1): p. 7-13.
737. Mercer, J.F., et al., *Isolation of a partial candidate gene for Menkes disease by positional cloning*. Nat Genet, 1993. **3**(1): p. 20-5.
738. Chelly, J., et al., *Isolation of a candidate gene for Menkes disease that encodes a potential heavy metal binding protein*. Nat Genet, 1993. **3**(1): p. 14-9.
739. Desai, V., et al., *Favorably skewed X-inactivation accounts for neurological sparing in female carriers of Menkes disease*. Clin Genet, 2011. **79**(2): p. 176-82.
740. Sirleto, P., et al., *Lyonization effects of the t(X;16) translocation on the phenotypic expression in a rare female with Menkes disease*. Pediatr Res, 2009. **65**(3): p. 347-51.

References

741. Lazoff, S.G., et al., *Skeletal dysplasia, occipital horns, diarrhea and obstructive uropathy- a new hereditary syndrome*. Birth Defects Orig Artic Ser, 1975. **11**(5): p. 71-4.
742. Kaler, S.G., et al., *Occipital horn syndrome and a mild Menkes phenotype associated with splice site mutations at the MNK locus*. Nat Genet, 1994. **8**(2): p. 195-202.
743. Yamaguchi, S., et al., *Mutations and polymorphisms in the human ornithine transcarbamylase (OTC) gene*. Hum Mutat, 2006. **27**(7): p. 626-32.
744. Klein, O.D., et al., *Acute fatal presentation of ornithine transcarbamylase deficiency in a previously healthy male*. Hepatol Int, 2008. **2**(3): p. 390-4.
745. Trivedi, M., et al., *Ornithine transcarbamylase deficiency unmasked because of gastrointestinal bleeding*. J Clin Gastroenterol, 2001. **32**(4): p. 340-3.
746. Schimanski, U., et al., *A novel two-nucleotide deletion in the ornithine transcarbamylase gene causing fatal hyperammonia in early pregnancy*. Hepatology, 1996. **24**(6): p. 1413-5.
747. Pridmore, C.L., J.T. Clarke, and S. Blaser, *Ornithine transcarbamylase deficiency in females: an often overlooked cause of treatable encephalopathy*. J Child Neurol, 1995. **10**(5): p. 369-74.
748. Felig, D.M., S.W. Brusilow, and J.L. Boyer, *Hyperammonemic coma due to parenteral nutrition in a woman with heterozygous ornithine transcarbamylase deficiency*. Gastroenterology, 1995. **109**(1): p. 282-4.
749. Migeon, B.R., *X inactivation, female mosaicism, and sex differences in renal diseases*. J Am Soc Nephrol, 2008. **19**(11): p. 2052-9.
750. Rheault, M.N., et al., *X-inactivation modifies disease severity in female carriers of murine X-linked Alport syndrome*. Nephrol Dial Transplant, 2010. **25**(3): p. 764-9.
751. Kashtan, C.E., *Alport syndrome and the X chromosome: implications of a diagnosis of Alport syndrome in females*. Nephrol Dial Transplant, 2007. **22**(6): p. 1499-505.
752. Rheault, M.N., *Women and Alport syndrome*. Pediatr Nephrol, 2012. **27**(1): p. 41-6.
753. Guo, C., et al., *Severe alport phenotype in a woman with two missense mutations in the same COL4A5 gene and preponderant inactivation of the X chromosome carrying the normal allele*. J Clin Invest, 1995. **95**(4): p. 1832-7.
754. Cau, M., et al., *A locus for familial skewed X chromosome inactivation maps to chromosome Xq25 in a family with a female manifesting Lowe syndrome*. J Hum Genet, 2006. **51**(11): p. 1030-6.
755. Demura, M., et al., *Surgical stress-induced transient nephrogenic diabetes insipidus (NDI) associated with decreased Vasopressin receptor2 (AVPR2) expression linked to nonsense-mediated mRNA decay and incomplete skewed X-inactivation in a female patient with a heterozygous AVPR2 mutation (c. 89-90 delAC)*. Clin Endocrinol (Oxf), 2004. **60**(6): p. 773-5.
756. Nomura, Y., et al., *Detection of skewed X-inactivation in two female carriers of vasopressin type 2 receptor gene mutation*. J Clin Endocrinol Metab, 1997. **82**(10): p. 3434-7.
757. van Lieburg, A.F., et al., *Clinical phenotype of nephrogenic diabetes insipidus in females heterozygous for a vasopressin type 2 receptor mutation*. Hum Genet, 1995. **96**(1): p. 70-8.
758. Satoh, M., S. Ogikubo, and A. Yoshizawa-Ogasawara, *Correlation between clinical phenotypes and X-inactivation patterns in six female carriers with heterozygote vasopressin type 2 receptor gene mutations*. Endocr J, 2008. **55**(2): p. 277-84.
759. Oohashi, T., et al., *Clonal overgrowth of esophageal smooth muscle cells in diffuse leiomyomatosis-Alport syndrome caused by partial deletion in COL4A5 and COL4A6 genes*. Matrix Biol, 2011. **30**(1): p. 3-8.
760. Orstavik, K.H., et al., *X chromosome inactivation pattern in female carriers of X linked hypophosphataemic rickets*. J Med Genet, 1996. **33**(8): p. 700-3.
761. Owen, C.J., et al., *Discordance for X-linked hypophosphataemic rickets in identical twin girls*. Horm Res, 2009. **71**(4): p. 237-44.
762. Faerch, M., et al., *Skewed X-chromosome inactivation causing diagnostic misinterpretation in congenital nephrogenic diabetes insipidus*. Scand J Urol Nephrol, 2010. **44**(5): p. 324-30.
763. Decaux, G., et al., *Nephrogenic syndrome of inappropriate antidiuresis in adults: high phenotypic variability in men and women from a large pedigree*. J Am Soc Nephrol, 2007. **18**(2): p. 606-12.
764. De Gregorio, L., et al., *Lesch-Nyhan disease in a female with a clinically normal monozygotic twin*. Mol Genet Metab, 2005. **85**(1): p. 70-7.
765. Rinat, C., et al., *Molecular, biochemical, and genetic characterization of a female patient with Lesch-Nyhan disease*. Mol Genet Metab, 2006. **87**(3): p. 249-52.

References

766. Blaschko, A., *Die Nervenverteilung in der Haut in ihrer Beziehung zu den Erkrankungen der Haut. Beilage zu den Verhandlungen der Deutschen Dermatologischen Gesellschaft*. 1901, Wien Leipzig: Braumüller.
767. Happle, R., *Lyonization and the lines of Blaschko*. *Hum Genet*, 1985. **70**(3): p. 200-6.
768. Happle, R., *X-chromosome inactivation: role in skin disease expression*. *Acta Paediatr Suppl*, 2006. **95**(451): p. 16-23.
769. Lexner, M.O., et al., *X-linked hypohidrotic ectodermal dysplasia. Genetic and dental findings in 67 Danish patients from 19 families*. *Clin Genet*, 2008. **74**(3): p. 252-9.
770. Balmer, R., et al., *Enamel defects and Lyonization in focal dermal hypoplasia*. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*, 2004. **98**(6): p. 686-91.
771. Witkop, C.J., Jr., *Partial expression of sex-linked recessive amelogenesis imperfecta in females compatible with the Lyon hypothesis*. *Oral Surg Oral Med Oral Pathol*, 1967. **23**(2): p. 174-82.
772. Rott, H.D., *Extracutaneous analogies of Blaschko lines*. *Am J Med Genet*, 1999. **85**(4): p. 338-41.
773. Thomas, G.A., D. Williams, and E.D. Williams, *The demonstration of tissue clonality by X-linked enzyme histochemistry*. *J Pathol*, 1988. **155**(2): p. 101-8.
774. Tan, S.S. and S. Breen, *Radial mosaicism and tangential cell dispersion both contribute to mouse neocortical development*. *Nature*, 1993. **362**(6421): p. 638-40.
775. König, A., et al., *Mutations in the NSDHL gene, encoding a 3beta-hydroxysteroid dehydrogenase, cause CHLD syndrome*. *Am J Med Genet*, 2000. **90**(4): p. 339-46.
776. Jurkiewicz, D., et al., *Four novel RSK2 mutations in females with Coffin-Lowry syndrome*. *Eur J Med Genet*, 2010. **53**(5): p. 268-73.
777. Wang, Y., et al., *A novel RSK2 (RPS6KA3) gene mutation associated with abnormal brain MRI findings in a family with Coffin-Lowry syndrome*. *Am J Med Genet A*, 2006. **140**(12): p. 1274-9.
778. Simensen, R.J., et al., *Cognitive function in Coffin-Lowry syndrome*. *Clin Genet*, 2002. **61**(4): p. 299-304.
779. Aten, E., et al., *Keratosis Follicularis Spinulosa Decalvans is caused by mutations in MBTPS2*. *Hum Mutat*, 2010. **31**(10): p. 1125-33.
780. Graham, J.M., Jr., et al., *Behavior of 10 patients with FG syndrome (Opitz-Kaveggia syndrome) and the p.R961W mutation in the MED12 gene*. *Am J Med Genet A*, 2008. **146A**(23): p. 3011-7.
781. Yano, S., et al., *Familial Simpson-Golabi-Behmel syndrome: studies of X-chromosome inactivation and clinical phenotypes in two female individuals with GPC3 mutations*. *Clin Genet*, 2011. **80**(5): p. 466-71.
782. Hernandez-Martin, A., R. Gonzalez-Sarmiento, and P. De Unamuno, *X-linked ichthyosis: an update*. *Br J Dermatol*, 1999. **141**(4): p. 617-27.
783. Happle, R., *Cutaneous manifestation of X-linked genes escaping inactivation*. *Clin Exp Dermatol*, 1992. **17**(1): p. 69.
784. Simunovic, M.P., *Colour vision deficiency*. *Eye (Lond)*, 2010. **24**(5): p. 747-55.
785. Martinez-Garcia, M., et al., *Identification of a novel deletion in the OAI gene: report of the first Spanish family with X-linked ocular albinism*. *Clin Experiment Ophthalmol*, 2010. **38**(5): p. 489-95.
786. Kondo, H., et al., *Novel mutations in Norrie disease gene in Japanese patients with Norrie disease and familial exudative vitreoretinopathy*. *Invest Ophthalmol Vis Sci*, 2007. **48**(3): p. 1276-82.
787. Sato, M., et al., *Three novel mutations in the X-linked juvenile retinoschisis (XLRSL) gene in 6 Japanese patients, 1 of whom had Turner's syndrome*. *Ophthalmic Res*, 2003. **35**(5): p. 295-300.
788. Mendoza-Londono, R., et al., *A Colombian family with X-linked juvenile retinoschisis with three affected females finding of a frameshift mutation*. *Ophthalmic Genet*, 1999. **20**(1): p. 37-43.
789. Rodriguez, F.J., et al., *X-linked retinoschisis in three females from the same family: a phenotype-genotype correlation*. *Retina*, 2005. **25**(1): p. 69-74.
790. Watkins, R.J., et al., *The Role of FRMD7 in Idiopathic Infantile Nystagmus*. *J Ophthalmol*, 2012. **2012**: p. 460956.
791. Zhang, Q., et al., *FRMD7 mutations in Chinese families with X-linked congenital motor nystagmus*. *Mol Vis*, 2007. **13**: p. 1375-8.
792. Kaplan, Y., et al., *Skewed X inactivation in an X linked nystagmus family resulted from a novel, p.R229G, missense mutation in the FRMD7 gene*. *Br J Ophthalmol*, 2008. **92**(1): p. 135-41.
793. Self, J.E., et al., *Allelic variation of the FRMD7 gene in congenital idiopathic nystagmus*. *Arch Ophthalmol*, 2007. **125**(9): p. 1255-63.
794. Teraï, N. and F. Raiskup, *[Bilateral visual loss in a young male patient]*. *Ophthalmologe*, 2012. **109**(5): p. 487-90.

References

795. Callea, M., et al., *Infantile bilateral glaucoma in a child with ectodermal dysplasia*. *Ophthalmic Genet*, 2012.
796. ACOG practice bulletin. *Management of recurrent pregnancy loss*. Number 24, February 2001. (Replaces Technical Bulletin Number 212, September 1995). *American College of Obstetricians and Gynecologists*. *Int J Gynaecol Obstet*, 2002. **78**(2): p. 179-90.
797. Lanasa, M.C., et al., *Highly skewed X-chromosome inactivation is associated with idiopathic recurrent spontaneous abortion*. *Am J Hum Genet*, 1999. **65**(1): p. 252-4.
798. Sangha, K.K., et al., *Extremely skewed X-chromosome inactivation is increased in women with recurrent spontaneous abortion*. *Am J Hum Genet*, 1999. **65**(3): p. 913-7.
799. Robinson, W.P., et al., *Skewed X inactivation and recurrent spontaneous abortion*. *Semin Reprod Med*, 2001. **19**(2): p. 175-81.
800. Uehara, S., et al., *Preferential X-chromosome inactivation in women with idiopathic recurrent pregnancy loss*. *Fertil Steril*, 2001. **76**(5): p. 908-14.
801. Su, M.T., S.H. Lin, and Y.C. Chen, *Association of sex hormone receptor gene polymorphisms with recurrent pregnancy loss: a systematic review and meta-analysis*. *Fertil Steril*, 2011. **96**(6): p. 1435-1444 e1.
802. Bagislar, S., et al., *Extremely skewed X-chromosome inactivation patterns in women with recurrent spontaneous abortion*. *Aust N Z J Obstet Gynaecol*, 2006. **46**(5): p. 384-7.
803. Kuo, P.L., et al., *Association of extremely skewed X-chromosome inactivation with Taiwanese women presenting with recurrent pregnancy loss*. *J Formos Med Assoc*, 2008. **107**(4): p. 340-3.
804. Kim, J.W., et al., *X-chromosome inactivation patterns in Korean women with idiopathic recurrent spontaneous abortion*. *J Korean Med Sci*, 2004. **19**(2): p. 258-62.
805. Aruna, M., et al., *Role of androgen receptor CAG repeat polymorphism and X-inactivation in the manifestation of recurrent spontaneous abortions in Indian women*. *PLoS One*, 2011. **6**(3): p. e17718.
806. Dasoula, A., et al., *Skewed X-chromosome inactivation in Greek women with idiopathic recurrent miscarriage*. *Fetal Diagn Ther*, 2008. **23**(3): p. 198-203.
807. Beever, C.L., et al., *Skewed X-chromosome inactivation is associated with trisomy in women ascertained on the basis of recurrent spontaneous abortion or chromosomally abnormal pregnancies*. *Am J Hum Genet*, 2003. **72**(2): p. 399-407.
808. Bretherick, K., J. Gair, and W.P. Robinson, *The association of skewed X chromosome inactivation with aneuploidy in humans*. *Cytogenet Genome Res*, 2005. **111**(3-4): p. 260-5.
809. Warburton, D., et al., *Skewed X chromosome inactivation and trisomic spontaneous abortion: no association*. *Am J Hum Genet*, 2009. **85**(2): p. 179-93.
810. Toniolo, D. and F. Rizzolio, *X chromosome and ovarian failure*. *Semin Reprod Med*, 2007. **25**(4): p. 264-71.
811. Ferreira, S.I., et al., *X-chromosome terminal deletion in a female with premature ovarian failure: Haploinsufficiency of X-linked genes as a possible explanation*. *Mol Cytogenet*, 2010. **3**: p. 14.
812. Bertini, V., et al., *Molecular cytogenetic definition of a translocation t(X;15) associated with premature ovarian failure*. *Fertil Steril*, 2010. **94**(3): p. 1097 e5-8.
813. Layman, L.C., *Editorial: BMP15--the first true ovarian determinant gene on the X-chromosome?* *J Clin Endocrinol Metab*, 2006. **91**(5): p. 1673-6.
814. Di Pasquale, E., P. Beck-Peccoz, and L. Persani, *Hypergonadotropic ovarian failure associated with an inherited mutation of human bone morphogenetic protein-15 (BMP15) gene*. *Am J Hum Genet*, 2004. **75**(1): p. 106-11.
815. Allingham-Hawkins, D.J., et al., *Fragile X premutation is a significant risk factor for premature ovarian failure: the International Collaborative POF in Fragile X study--preliminary data*. *Am J Med Genet*, 1999. **83**(4): p. 322-5.
816. Sato, K., et al., *Genetic significance of skewed X-chromosome inactivation in premature ovarian failure*. *Am J Med Genet A*, 2004. **130A**(3): p. 240-4.
817. Bretherick, K.L., et al., *Skewed X-chromosome inactivation is associated with primary but not secondary ovarian failure*. *Am J Med Genet A*, 2007. **143A**(9): p. 945-51.
818. Rodriguez-Revenga, L., et al., *Premature ovarian failure and fragile X female premutation carriers: no evidence for a skewed X-chromosome inactivation pattern*. *Menopause*, 2009. **16**(5): p. 944-9.
819. Tejada, M.I., et al., *Analysis of the molecular parameters that could predict the risk of manifesting premature ovarian failure in female premutation carriers of fragile X syndrome*. *Menopause*, 2008. **15**(5): p. 945-9.

References

820. Yoon, S.H., et al., *X chromosome inactivation patterns in patients with idiopathic premature ovarian failure*. Hum Reprod, 2008. **23**(3): p. 688-92.
821. Spath, M.A., et al., *X chromosome inactivation does not define the development of premature ovarian failure in fragile X premutation carriers*. Am J Med Genet A, 2010. **152A**(2): p. 387-93.
822. Pu, D., J. Wu, and J. Liu, *Skewed X chromosome inactivation may be not associated with premature ovarian failure*. Gynecol Endocrinol, 2010. **26**(6): p. 423-8.
823. Medema, R.H. and B.M. Burgering, *The X factor: skewing X inactivation towards cancer*. Cell, 2007. **129**(7): p. 1253-4.
824. Kido, T., J.H. Ou, and Y.F. Lau, *The X-linked tumor suppressor TSPX interacts and promotes degradation of the hepatitis B viral protein HBx via the proteasome pathway*. PLoS One, 2011. **6**(7): p. e22979.
825. Liu, R., M. Kain, and L. Wang, *Inactivation of X-linked tumor suppressor genes in human cancer*. Future Oncol, 2012. **8**(4): p. 463-81.
826. Rivera, M.N., et al., *An X chromosome gene, WTX, is commonly inactivated in Wilms tumor*. Science, 2007. **315**(5812): p. 642-5.
827. Zuo, T., et al., *FOXP3 is an X-linked breast cancer suppressor gene and an important repressor of the HER-2/ErbB2 oncogene*. Cell, 2007. **129**(7): p. 1275-86.
828. Wang, L., et al., *FOXP3 as an X-linked tumor suppressor*. Discov Med, 2010. **10**(53): p. 322-8.
829. Knudson, A.G., Jr., *Mutation and cancer: statistical study of retinoblastoma*. Proc Natl Acad Sci U S A, 1971. **68**(4): p. 820-3.
830. Weakley, S.M., et al., *Expression and function of a large non-coding RNA gene XIST in human cancer*. World J Surg, 2011. **35**(8): p. 1751-6.
831. Kawakami, T., et al., *The roles of supernumerical X chromosomes and XIST expression in testicular germ cell tumors*. J Urol, 2003. **169**(4): p. 1546-52.
832. Lind, G.E., R.I. Skotheim, and R.A. Lothe, *The epigenome of testicular germ cell tumors*. APMIS, 2007. **115**(10): p. 1147-60.
833. Sirchia, S.M., et al., *Misbehaviour of XIST RNA in breast cancer cells*. PLoS One, 2009. **4**(5): p. e5559.
834. Huang, K.C., et al., *Relationship of XIST expression and responses of ovarian cancer to chemotherapy*. Mol Cancer Ther, 2002. **1**(10): p. 769-76.
835. Kawakami, T., et al., *Characterization of loss-of-inactive X in Klinefelter syndrome and female-derived cancer cells*. Oncogene, 2004. **23**(36): p. 6163-9.
836. McDonald, H.L., et al., *Involvement of the X chromosome in non-Hodgkin lymphoma*. Genes Chromosomes Cancer, 2000. **28**(3): p. 246-57.
837. Sirchia, S.M., et al., *Loss of the inactive X chromosome and replication of the active X in BRCA1-defective and wild-type breast cancer cells*. Cancer Res, 2005. **65**(6): p. 2139-46.
838. Wang, N., et al., *Two identical active X chromosomes in human mammary carcinoma cells*. Cancer Genet Cytogenet, 1990. **46**(2): p. 271-80.
839. van Leenders, G.J., et al., *Polycomb-group oncogenes EZH2, BMI1, and RING1 are overexpressed in prostate cancer with adverse pathologic and clinical features*. Eur Urol, 2007. **52**(2): p. 455-63.
840. Simon, J.A. and C.A. Lange, *Roles of the EZH2 histone methyltransferase in cancer epigenetics*. Mutat Res, 2008. **647**(1-2): p. 21-9.
841. Laird, P.W. and R. Jaenisch, *The role of DNA methylation in cancer genetic and epigenetics*. Annu Rev Genet, 1996. **30**: p. 441-64.
842. Iacobuzio-Donahue, C.A., *Epigenetic changes in cancer*. Annu Rev Pathol, 2009. **4**: p. 229-49.
843. Pageau, G.J., L.L. Hall, and J.B. Lawrence, *BRCA1 does not paint the inactive X to localize XIST RNA but may contribute to broad changes in cancer that impact XIST and Xi heterochromatin*. J Cell Biochem, 2007. **100**(4): p. 835-50.
844. Klinefelter, H.F., E.C. Reifenstein, and F. Albright, *Syndrome characterized by gynecomastia, aspermatogenesis without Leydigism, increased excretion of follicle stimulating hormone*. J Clin Endocrinol, 1942. **2**: p. 615-627.
845. Jacobs, P.A. and J.A. Strong, *A case of human intersexuality having a possible XXY sex-determining mechanism*. Nature, 1959. **183**(4657): p. 302-3.
846. Tuttelmann, F. and J. Gromoll, *Novel genetic aspects of Klinefelter's syndrome*. Mol Hum Reprod, 2010. **16**(6): p. 386-95.
847. Morris, J.K., et al., *Is the prevalence of Klinefelter syndrome increasing?* Eur J Hum Genet, 2008. **16**(2): p. 163-70.

848. Iitsuka, Y., et al., *Evidence of skewed X-chromosome inactivation in 47,XXY and 48,XXYY Klinefelter patients*. *Am J Med Genet*, 2001. **98**(1): p. 25-31.
849. Zitzmann, M., et al., *X-chromosome inactivation patterns and androgen receptor functionality influence phenotype and social characteristics as well as pharmacogenetics of testosterone therapy in Klinefelter patients*. *J Clin Endocrinol Metab*, 2004. **89**(12): p. 6208-17.
850. Bojesen, A., J.M. Hertz, and C.H. Gravholt, *Genotype and phenotype in Klinefelter syndrome - impact of androgen receptor polymorphism and skewed X inactivation*. *Int J Androl*, 2011. **34**(6 Pt 2): p. e642-8.
851. Kanaka-Gantenbein, C., et al., *Tall stature, insulin resistance, and disturbed behavior in a girl with the triple X syndrome harboring three SHOX genes: offspring of a father with mosaic Klinefelter syndrome but with two maternal X chromosomes*. *Horm Res*, 2004. **61**(5): p. 205-10.
852. Prendiville, J.S., et al., *Incontinentia pigmenti in a male infant with Klinefelter syndrome*. *J Am Acad Dermatol*, 1989. **20**(5 Pt 2): p. 937-40.
853. Ormerod, A.D., et al., *Incontinentia pigmenti in a boy with Klinefelter's syndrome*. *J Med Genet*, 1987. **24**(7): p. 439-41.
854. Buinauskaite, E., et al., *Incontinentia pigmenti in a male infant with Klinefelter syndrome: a case report and review of the literature*. *Pediatr Dermatol*, 2010. **27**(5): p. 492-5.
855. Kenwick, S., et al., *Survival of male patients with incontinentia pigmenti carrying a lethal mutation can be explained by somatic mosaicism or Klinefelter syndrome*. *Am J Hum Genet*, 2001. **69**(6): p. 1210-7.
856. Cho, S.Y., C.K. Lee, and B.K. Drummond, *Surviving male with incontinentia pigmenti: a case report*. *Int J Paediatr Dent*, 2004. **14**(1): p. 69-72.
857. Turner, H.H., *A syndrome of infantilism, congenital webbed neck and cubitus valgus*. *Endocrinology*, 1938. **23**: p. 566-574.
858. Ford, C.E., et al., *A sex-chromosome anomaly in a case of gonadal dysgenesis (Turner's syndrome)*. *Lancet*, 1959. **1**(7075): p. 711-3.
859. Zinn, A.R., D.C. Page, and E.M. Fisher, *Turner syndrome: the case of the missing sex chromosome*. *Trends Genet*, 1993. **9**(3): p. 90-3.
860. Rao, E., et al., *Pseudoautosomal deletions encompassing a novel homeobox gene cause growth failure in idiopathic short stature and Turner syndrome*. *Nat Genet*, 1997. **16**(1): p. 54-63.
861. Shears, D.J., et al., *Mutation and deletion of the pseudoautosomal gene SHOX cause Leri-Weill dyschondrosteosis*. *Nat Genet*, 1998. **19**(1): p. 70-3.
862. Speed, R.M., *The possible role of meiotic pairing anomalies in the atresia of human fetal oocytes*. *Hum Genet*, 1988. **78**(3): p. 260-6.
863. Cohen, M.M., et al., *Autoradiographic investigations of centric fragments and rings in patients with stigmata of gonadal dysgenesis*. *Cytogenetics*, 1967. **6**(3): p. 254-67.
864. Kushnick, T., et al., *45X/46X,r(X) with syndactyly and severe mental retardation*. *Am J Med Genet*, 1987. **28**(3): p. 567-74.
865. Lindgren, V., et al., *Cytogenetic and molecular characterization of marker chromosomes in patients with mosaic 45,X karyotypes*. *Hum Genet*, 1992. **88**(4): p. 393-8.
866. Van Dyke, D.L., et al., *Mental retardation in Turner syndrome*. *J Pediatr*, 1991. **118**(3): p. 415-7.
867. Van Dyke, D.L., et al., *Ulrich-Turner syndrome with a small ring X chromosome and presence of mental retardation*. *Am J Med Genet*, 1992. **43**(6): p. 996-1005.
868. Dennis, N.R., et al., *Three patients with ring (X) chromosomes and a severe phenotype*. *J Med Genet*, 1993. **30**(6): p. 482-6.
869. Grompe, M., et al., *45,X/46,X,+r(X) can have a distinct phenotype different from Ulrich-Turner syndrome*. *Am J Med Genet*, 1992. **42**(1): p. 39-43.
870. El Abd, S., et al., *Social, communicational, and behavioral deficits associated with ring X turner syndrome*. *Am J Med Genet*, 1999. **88**(5): p. 510-6.
871. Kubota, T., et al., *The proportion of cells with functional X disomy is associated with the severity of mental retardation in mosaic ring X Turner syndrome females*. *Cytogenet Genome Res*, 2002. **99**(1-4): p. 276-84.
872. Migeon, B.R., et al., *The severe phenotype of females with tiny ring X chromosomes is associated with inability of these chromosomes to undergo X inactivation*. *Am J Hum Genet*, 1994. **55**(3): p. 497-504.
873. Migeon, B.R., et al., *Deficient transcription of XIST from tiny ring X chromosomes in females with severe phenotypes*. *Proc Natl Acad Sci U S A*, 1993. **90**(24): p. 12025-9.

References

874. Shchelochkov, O.A., et al., *Mosaicism for r(X) and der(X)del(X)(p11.23)dup(X)(p11.21p11.22) provides insight into the possible mechanism of rearrangement.* Mol Cytogenet, 2008. 1: p. 16.
875. Wolff, D.J., et al., *Small marker X chromosomes lack the X inactivation center: implications for karyotype/phenotype correlations.* Am J Hum Genet, 1994. 55(1): p. 87-95.
876. Cole, H., et al., *Mental retardation and Ullrich-Turner syndrome in cases with 45,X/46X,+mar: additional support for the loss of the X-inactivation center hypothesis.* Am J Med Genet, 1994. 52(2): p. 136-45.
877. Stavropoulou, C., et al., *Severe phenotype resulting from an active ring X chromosome in a female with a complex karyotype: characterisation and replication study.* J Med Genet, 1998. 35(11): p. 932-8.
878. Otter, M., C.T. Schrander-Stumpel, and L.M. Curfs, *Triple X syndrome: a review of the literature.* Eur J Hum Genet, 2010. 18(3): p. 265-71.
879. Tartaglia, N.R., et al., *A review of trisomy X (47,XXX).* Orphanet J Rare Dis, 2010. 5: p. 8.
880. Ratcliffe, S.G., H. Pan, and M. McKie, *The growth of XXX females: population-based studies.* Ann Hum Biol, 1994. 21(1): p. 57-66.
881. Alvesalo, L., E. Tammisalo, and E. Therman, *47,XXX females, sex chromosomes, and tooth crown structure.* Hum Genet, 1987. 77(4): p. 345-8.
882. Gropman, A.L., et al., *Clinical variability and novel neurodevelopmental findings in 49, XXXXY syndrome.* Am J Med Genet A, 2010. 152A(6): p. 1523-30.
883. Monheit, A., et al., *The penta-X syndrome.* J Med Genet, 1980. 17(5): p. 392-6.
884. Linden, M.G., B.G. Bender, and A. Robinson, *Sex chromosome tetrasomy and pentasomy.* Pediatrics, 1995. 96(4 Pt 1): p. 672-82.
885. Visootsak, J., et al., *Behavioral phenotype of sex chromosome aneuploidies: 48,XXYY, 48,XXXY, and 49,XXXXY.* Am J Med Genet A, 2007. 143A(11): p. 1198-203.
886. Therman, E., et al., *X chromosome constitution and the human female phenotype.* Hum Genet, 1980. 54(2): p. 133-43.
887. Sarto, G.E., et al., *What causes the abnormal phenotype in a 49,XXXXY male?* Hum Genet, 1987. 76(1): p. 1-4.
888. Moraes, L.M., et al., *Detailed analysis of X chromosome inactivation in a 49,XXXXX pentasomy.* Mol Cytogenet, 2009. 2: p. 20.
889. Stochholm, K., et al., *Criminality in men with Klinefelter's syndrome and XYY syndrome: a cohort study.* BMJ Open, 2012. 2(1): p. e000650.
890. Freyne, A. and A. O'Connor, *XYY genotype and crime: 2 cases.* Med Sci Law, 1992. 32(3): p. 261-3.
891. Ross, J.L., et al., *Behavioral and social phenotypes in boys with 47,XYY syndrome or 47,XXY Klinefelter syndrome.* Pediatrics, 2012. 129(4): p. 769-78.
892. Wutz, A. and J. Gribnau, *X inactivation Xplained.* Curr Opin Genet Dev, 2007. 17(5): p. 387-93.
893. Bach, I., et al., *RLIM inhibits functional activity of LIM homeodomain transcription factors via recruitment of the histone deacetylase complex.* Nat Genet, 1999. 22(4): p. 394-9.
894. Her, Y.R. and I.K. Chung, *Ubiquitin Ligase RLIM Modulates Telomere Length Homeostasis through a Proteolysis of TRF1.* J Biol Chem, 2009. 284(13): p. 8557-66.
895. Johnsen, S.A., et al., *Regulation of estrogen-dependent transcription by the LIM cofactors CLIM and RLIM in breast cancer.* Cancer Res, 2009. 69(1): p. 128-36.
896. Kalantry, S., et al., *Evidence of Xist RNA-independent initiation of mouse imprinted X-chromosome inactivation.* Nature, 2009. 460(7255): p. 647-51.
897. Becker, T., et al., *Comparing protein stabilities during zebrafish embryogenesis.* Methods Cell Sci, 2003. 25(1-2): p. 85-9.
898. Sado, T., Y. Hoki, and H. Sasaki, *Tsix defective in splicing is competent to establish Xist silencing.* Development, 2006. 133(24): p. 4925-31.
899. Cline, T.W. and B.J. Meyer, *Vive la difference: males vs females in flies vs worms.* Annu Rev Genet, 1996. 30: p. 637-702.
900. Lee, E.C., et al., *A highly efficient Escherichia coli-based chromosome engineering system adapted for recombinogenic targeting and subcloning of BAC DNA.* Genomics, 2001. 73(1): p. 56-65.
901. Capecchi, M.R., *Altering the genome by homologous recombination.* Science, 1989. 244(4910): p. 1288-92.
902. Deng, C. and M.R. Capecchi, *Reexamination of gene targeting frequency as a function of the extent of homology between the targeting vector and the target locus.* Mol Cell Biol, 1992. 12(8): p. 3365-71.
903. Hasty, P., et al., *Target frequency and integration pattern for insertion and replacement vectors in embryonic stem cells.* Mol Cell Biol, 1991. 11(9): p. 4509-17.

References

904. te Riele, H., E.R. Maandag, and A. Berns, *Highly efficient gene targeting in embryonic stem cells through homologous recombination with isogenic DNA constructs*. Proc Natl Acad Sci U S A, 1992. **89**(11): p. 5128-32.
905. Valenzuela, D.M., et al., *High-throughput engineering of the mouse genome coupled with high-resolution expression analysis*. Nat Biotechnol, 2003. **21**(6): p. 652-9.
906. Yang, Y. and B. Seed, *Site-specific gene targeting in mouse embryonic stem cells with intact bacterial artificial chromosomes*. Nat Biotechnol, 2003. **21**(4): p. 447-51.
907. Song, H., S.K. Chung, and Y. Xu, *Modeling disease in human ESCs using an efficient BAC-based homologous recombination system*. Cell Stem Cell, 2010. **6**(1): p. 80-9.
908. Osoegawa, K., et al., *Bacterial artificial chromosome libraries for mouse sequencing and functional analysis*. Genome Res, 2000. **10**(1): p. 116-28.
909. Frazer, K.A., et al., *A sequence-based variation map of 8.27 million SNPs in inbred mouse strains*. Nature, 2007. **448**(7157): p. 1050-3.
910. Barakat, T.S., et al., *X-changing information on X inactivation*. Exp Cell Res, 2010. **316**(5): p. 679-87.
911. Barakat, T.S., et al., *RNF12 activates Xist and is essential for X chromosome inactivation*. PLoS Genet, 2011. **7**(1): p. e1002001.
912. Eggan, K., et al., *Hybrid vigor, fetal overgrowth, and viability of mice derived by nuclear cloning and tetraploid embryo complementation*. Proc Natl Acad Sci U S A, 2001. **98**(11): p. 6209-14.
913. Bradley, A., et al., *Formation of germ-line chimaeras from embryo-derived teratocarcinoma cell lines*. Nature, 1984. **309**(5965): p. 255-6.
914. Thomas, K.R. and M.R. Capecchi, *Site-directed mutagenesis by gene targeting in mouse embryo-derived stem cells*. Cell, 1987. **51**(3): p. 503-12.
915. Mansour, S.L., K.R. Thomas, and M.R. Capecchi, *Disruption of the proto-oncogene int-2 in mouse embryo-derived stem cells: a general strategy for targeting mutations to non-selectable genes*. Nature, 1988. **336**(6197): p. 348-52.
916. Smithies, O., et al., *Insertion of DNA sequences into the human chromosomal beta-globin locus by homologous recombination*. Nature, 1985. **317**(6034): p. 230-4.
917. Doetschman, T., et al., *Targeted correction of a mutant HPRT gene in mouse embryonic stem cells*. Nature, 1987. **330**(6148): p. 576-8.
918. Koller, B.H., et al., *Normal development of mice deficient in beta 2M, MHC class I proteins, and CD8+ T cells*. Science, 1990. **248**(4960): p. 1227-30.
919. Schwartzberg, P.L., S.P. Goff, and E.J. Robertson, *Germ-line transmission of a c-abl mutation produced by targeted gene disruption in ES cells*. Science, 1989. **246**(4931): p. 799-803.
920. Zijlstra, M., et al., *Germ-line transmission of a disrupted beta 2-microglobulin gene produced by homologous recombination in embryonic stem cells*. Nature, 1989. **342**(6248): p. 435-8.
921. International Mouse Knockout, C., et al., *A mouse for all reasons*. Cell, 2007. **128**(1): p. 9-13.
922. Austin, C.P., et al., *The knockout mouse project*. Nat Genet, 2004. **36**(9): p. 921-4.
923. Auwerx, J., et al., *The European dimension for the mouse genome mutagenesis program*. Nat Genet, 2004. **36**(9): p. 925-7.
924. Pardo, M., et al., *An expanded Oct4 interaction network: implications for stem cell biology, development, and disease*. Cell Stem Cell, 2010. **6**(4): p. 382-95.
925. Ciotta, G., et al., *Recombineering BAC transgenes for protein tagging*. Methods, 2011. **53**(2): p. 113-9.
926. Zhang, P., M.Z. Li, and S.J. Elledge, *Towards genetic genome projects: genomic library screening and gene-targeting vector construction in a single step*. Nat Genet, 2002. **30**(1): p. 31-9.
927. Testa, G., et al., *Engineering the mouse genome with bacterial artificial chromosomes to create multipurpose alleles*. Nat Biotechnol, 2003. **21**(4): p. 443-7.
928. Cotta-de-Almeida, V., et al., *A new method for rapidly generating gene-targeting vectors by engineering BACs through homologous recombination in bacteria*. Genome Res, 2003. **13**(9): p. 2190-4.
929. Testa, G., et al., *BAC engineering for the generation of ES cell-targeting constructs and mouse transgenes*. Methods Mol Biol, 2004. **256**: p. 123-39.
930. Shizuya, H., et al., *Cloning and stable maintenance of 300-kilobase-pair fragments of human DNA in Escherichia coli using an F-factor-based vector*. Proc Natl Acad Sci U S A, 1992. **89**(18): p. 8794-7.
931. Ioannou, P.A., et al., *A new bacteriophage P1-derived vector for the propagation of large human DNA fragments*. Nat Genet, 1994. **6**(1): p. 84-9.
932. Marra, M.A., et al., *High throughput fingerprint analysis of large-insert clones*. Genome Res, 1997. **7**(11): p. 1072-84.

References

933. Kelley, J.M., et al., *High throughput direct end sequencing of BAC clones*. Nucleic Acids Res, 1999. **27**(6): p. 1539-46.
934. Osoegawa, K. and P.J. de Jong, *BAC library construction*. Methods Mol Biol, 2004. **255**: p. 1-46.
935. Mozo, T., et al., *A complete BAC-based physical map of the Arabidopsis thaliana genome*. Nat Genet, 1999. **22**(3): p. 271-5.
936. Hoskins, R.A., et al., *A BAC-based physical map of the major autosomes of Drosophila melanogaster*. Science, 2000. **287**(5461): p. 2271-4.
937. McPherson, J.D., et al., *A physical map of the human genome*. Nature, 2001. **409**(6822): p. 934-41.
938. Kim, U.J., et al., *Construction and characterization of a human bacterial artificial chromosome library*. Genomics, 1996. **34**(2): p. 213-8.
939. Frengen, E., et al., *A modular, positive selection bacterial artificial chromosome vector with multiple cloning sites*. Genomics, 1999. **58**(3): p. 250-3.
940. Frengen, E., et al., *Modular bacterial artificial chromosome vectors for transfer of large inserts into mammalian cells*. Genomics, 2000. **68**(2): p. 118-26.
941. Hejna, J.A., et al., *Functional complementation by electroporation of human BACs into mammalian fibroblast cells*. Nucleic Acids Res, 1998. **26**(4): p. 1124-5.
942. Jessen, J.R., et al., *Modification of bacterial artificial chromosomes through chi-stimulated homologous recombination and its application in zebrafish transgenesis*. Proc Natl Acad Sci U S A, 1998. **95**(9): p. 5121-6.
943. Muyrers, J.P., et al., *ET recombination: DNA engineering using homologous recombination in E. coli*. Methods Mol Biol, 2004. **256**: p. 107-21.
944. Muyrers, J.P., et al., *Rapid modification of bacterial artificial chromosomes by ET-recombination*. Nucleic Acids Res, 1999. **27**(6): p. 1555-7.
945. Narayanan, K., et al., *Efficient and precise engineering of a 200 kb beta-globin human/bacterial artificial chromosome in E. coli DH10B using an inducible homologous recombination system*. Gene Ther, 1999. **6**(3): p. 442-7.
946. Chan, W., et al., *A recombineering based approach for high-throughput conditional knockout targeting vector construction*. Nucleic Acids Res, 2007. **35**(8): p. e64.
947. Adams, D.J., et al., *Mutagenic insertion and chromosome engineering resource (MICER)*. Nat Genet, 2004. **36**(8): p. 867-71.
948. Yang, X.W., P. Model, and N. Heintz, *Homologous recombination based modification in Escherichia coli and germline transmission in transgenic mice of a bacterial artificial chromosome*. Nat Biotechnol, 1997. **15**(9): p. 859-65.
949. Lalioti, M. and J. Heath, *A new method for generating point mutations in bacterial artificial chromosomes by homologous recombination in Escherichia coli*. Nucleic Acids Res, 2001. **29**(3): p. E14.
950. Zhang, Y., et al., *A new logic for DNA engineering using recombination in Escherichia coli*. Nat Genet, 1998. **20**(2): p. 123-8.
951. Murphy, K.C., *Use of bacteriophage lambda recombination functions to promote gene replacement in Escherichia coli*. J Bacteriol, 1998. **180**(8): p. 2063-71.
952. Sharan, S.K., et al., *Recombineering: a homologous recombination-based method of genetic engineering*. Nat Protoc, 2009. **4**(2): p. 206-23.
953. Copeland, N.G., N.A. Jenkins, and D.L. Court, *Recombineering: a powerful new tool for mouse functional genomics*. Nat Rev Genet, 2001. **2**(10): p. 769-79.
954. Wu, S., et al., *A protocol for constructing gene targeting vectors: generating knockout mice for the cadherin family and beyond*. Nat Protoc, 2008. **3**(6): p. 1056-76.
955. Gong, S., L. Kus, and N. Heintz, *Rapid bacterial artificial chromosome modification for large-scale mouse transgenesis*. Nat Protoc, 2010. **5**(10): p. 1678-96.
956. Hofemeister, H., et al., *Recombineering, transfection, Western, IP and ChIP methods for protein tagging via gene targeting or BAC transgenesis*. Methods, 2011. **53**(4): p. 437-52.
957. Narayanan, K. and Q. Chen, *Bacterial artificial chromosome mutagenesis using recombineering*. J Biomed Biotechnol, 2011. **2011**: p. 971296.
958. Barakat, T.S., et al., *Precise BAC targeting of genetically polymorphic mouse ES cells*. Nucleic Acids Res, 2011 Oct;39(18):e121.
959. Beck, J.A., et al., *Genealogies of mouse inbred strains*. Nat Genet, 2000. **24**(1): p. 23-5.
960. Abremski, K. and R. Hoess, *Bacteriophage P1 site-specific recombination. Purification and properties of the Cre recombinase protein*. J Biol Chem, 1984. **259**(3): p. 1509-14.

References

961. Abremski, K., R. Hoess, and N. Sternberg, *Studies on the properties of P1 site-specific recombination: evidence for topologically unlinked products following recombination*. Cell, 1983. **32**(4): p. 1301-11.
962. Zhang, Z. and B. Lutz, *Cre recombinase-mediated inversion using lox66 and lox71: method to introduce conditional point mutations into the CREB-binding protein*. Nucleic Acids Res, 2002. **30**(17): p. e90.
963. Oberdoerfler, P., et al., *Unidirectional Cre-mediated genetic inversion in mice using the mutant loxP pair lox66/lox71*. Nucleic Acids Res, 2003. **31**(22): p. e140.
964. Hogan B, B.R., Costantini F, Lacy E, ed. *Manipulating the Mouse Embryo*. 1994, Cold Spring Harbor Laboratory Press: Cold Spring Harbor, New York.
965. Eakin, G.S. and A.K. Hadjantonakis, *Production of chimeras by aggregation of embryonic stem cells with diploid or tetraploid mouse embryos*. Nat Protoc, 2006. **1**(3): p. 1145-53.
966. Eggan, K., et al., *Male and female mice derived from the same embryonic stem cell clone by tetraploid embryo complementation*. Nat Biotechnol, 2002. **20**(5): p. 455-9.
967. Zhao, X.-Y., et al., *Production of mice using iPS cells and tetraploid complementation*. Nat Protoc, 2010. **5**(5): p. 963-71.
968. Sambrook J, *Molecular Cloning: A Laboratory Manual* third edition ed. 2000, Cold Spring Harbor, New York: Cold Spring Harbor Laboratory Press
969. Lin, S. and P. Talbot, *Methods for culturing mouse and human embryonic stem cells*. Methods Mol Biol, 2011. **690**: p. 31-56.
970. Meissner, A., S. Emlinli, and R. Jaenisch, *Derivation and manipulation of murine embryonic stem cells*. Methods Mol Biol, 2009. **482**: p. 3-19.
971. Nichols, J. and Q.L. Ying, *Derivation and propagation of embryonic stem cells in serum- and feeder-free culture*. Methods Mol Biol, 2006. **329**: p. 91-8.
972. Laird, P.W., et al., *Simplified mammalian DNA isolation procedure*. Nucleic Acids Res, 1991. **19**(15): p. 4293.
973. Buecker, C., et al., *A murine ESC-like state facilitates transgenesis and homologous recombination in human pluripotent stem cells*. Cell Stem Cell, 2010. **6**(6): p. 535-46.
974. Lyon, M.F., et al., *A Mouse Translocation Suppressing Sex-Linked Variegation*. Cytogenetics, 1964. **15**: p. 306-23.
975. Chow, J. and E. Heard, *X inactivation and the complexities of silencing a sex chromosome*. Curr Opin Cell Biol, 2009. **21**(3): p. 359-66.
976. Lee, J.T., *Regulation of X-chromosome counting by Tsix and Xite sequences*. Science, 2005. **309**(5735): p. 768-71.
977. Xu, M. and P.R. Cook, *The role of specialized transcription factories in chromosome pairing*. Biochim Biophys Acta, 2008. **1783**(11): p. 2155-60.
978. Haileselasse Sene, K., et al., *Gene function in early mouse embryonic stem cell differentiation*. BMC Genomics, 2007. **8**: p. 85.
979. Burgoyne, P.S., et al., *The genetic basis of XX-XY differences present before gonadal sex differentiation in the mouse*. Philos Trans R Soc Lond B Biol Sci, 1995. **350**(1333): p. 253-60 discussion 260-1.
980. Lee, J.T., *Functional intergenic transcription: a case study of the X-inactivation centre*. Philos Trans R Soc Lond B Biol Sci, 2003. **358**(1436): p. 1417-23; discussion 1423.
981. Shin, J., et al., *Maternal Rnf12/RLIM is required for imprinted X-chromosome inactivation in mice*. Nature, 2010. **467**(7318): p. 977-81.
982. Ahn, J.Y. and J.T. Lee, *Retinoic acid accelerates downregulation of the Xist repressor, Oct4, and increases the likelihood of Xist activation when Tsix is deficient*. BMC Dev Biol, 2010. **10**: p. 90.
983. Kim, J., et al., *An extended transcriptional network for pluripotency of embryonic stem cells*. Cell, 2008. **132**(6): p. 1049-61.
984. Ostendorff, H.P., et al., *Functional characterization of the gene encoding RLIM, the corepressor of LIM homeodomain factors*. Genomics, 2000. **69**(1): p. 129-30.
985. Takagi, N., *Differentiation of X chromosomes in early female mouse embryos*. Exp Cell Res, 1974. **86**(1): p. 127-35.
986. Okamoto, I., et al., *Evidence for de novo imprinted X-chromosome inactivation independent of meiotic inactivation in mice*. Nature, 2005. **438**(7066): p. 369-73.
987. Barakat, T.S. and J. Gribnau, *X chromosome inactivation in the cycle of life*. Development, 2012. **139**(12): p. 2085-9.

References

988. Lee, J.T., *Disruption of imprinted X inactivation by parent-of-origin effects at Tsix*. Cell, 2000. **103**(1): p. 17-27.
989. Navarro, P., et al., *Molecular coupling of Tsix regulation and pluripotency*. Nature, 2010. **468**(7322): p. 457-60.
990. Navarro, P., et al., *The X-inactivation trans-activator Rnf12 is negatively regulated by pluripotency factors in embryonic stem cells*. Hum Genet, 2011. **130**(2): p. 255-64.
991. Gillich, A., et al., *Epiblast stem cell-based system reveals reprogramming synergy of germline factors*. Cell Stem Cell, 2012. **10**(4): p. 425-39.
992. Tsai, C.L., et al., *Higher order chromatin structure at the X-inactivation center via looping DNA*. Dev Biol, 2008. **319**(2): p. 416-25.
993. Navarro, P. and P. Avner, *When X-inactivation meets pluripotency: an intimate rendezvous*. FEBS Lett, 2009. **583**(11): p. 1721-7.
994. Nesterova, T.B., et al., *Pluripotency factor binding and Tsix expression act synergistically to repress Xist in undifferentiated embryonic stem cells*. Epigenetics Chromatin, 2011. **4**(1): p. 17.
995. Cecchi, C. and P. Avner, *Genomic organization of the mottled gene, the mouse homologue of the human Menkes disease gene*. Genomics, 1996. **37**(1): p. 96-104.
996. Hosler, B.A., et al., *Expression of REX-1, a gene containing zinc finger motifs, is rapidly reduced by retinoic acid in F9 teratocarcinoma cells*. Mol Cell Biol, 1989. **9**(12): p. 5623-9.
997. Scotland, K.B., et al., *Analysis of Rex1 (zfp42) function in embryonic stem cell differentiation*. Dev Dyn, 2009. **238**(8): p. 1863-77.
998. Kim, J.D., et al., *Rex1/Zfp42 as an epigenetic regulator for genomic imprinting*. Hum Mol Genet, 2011. **20**(7): p. 1353-62.
999. Masui, S., et al., *Rex1/Zfp42 is dispensable for pluripotency in mouse ES cells*. BMC Dev Biol, 2008. **8**: p. 45.
1000. Kim, J.D., C. Faulk, and J. Kim, *Retroposition and evolution of the DNA-binding motifs of YY1, YY2 and REX1*. Nucleic Acids Res, 2007. **35**(10): p. 3442-52.
1001. van den Berg, D.L., et al., *An Oct4-centered protein interaction network in embryonic stem cells*. Cell Stem Cell, 2010. **6**(4): p. 369-81.
1002. Soler, E., et al., *A systems approach to analyze transcription factors in mammalian cells*. Methods, 2011. **53**(2): p. 151-62.
1003. Dignam, J.D., R.M. Lebovitz, and R.G. Roeder, *Accurate transcription initiation by RNA polymerase II in a soluble extract from isolated mammalian nuclei*. Nucleic Acids Res, 1983. **11**(5): p. 1475-89.
1004. Nora, E.P., et al., *Spatial partitioning of the regulatory landscape of the X-inactivation centre*. Nature, 2012. **485**(7398): p. 381-5.
1005. Anguera, M.C., et al., *Tsx produces a long noncoding RNA and has general functions in the germline, stem cells, and brain*. PLoS Genet, 2011. **7**(9): p. e1002248.
1006. Tian, D., S. Sun, and J.T. Lee, *The long noncoding RNA, Jpx, is a molecular switch for X chromosome inactivation*. Cell, 2011. **143**(3): p. 390-403.
1007. Chureau, C., et al., *Ftx is a non-coding RNA which affects Xist expression and chromatin structure within the X-inactivation center region*. Hum Mol Genet, 2011. **20**(4): p. 705-18.
1008. Spencer, R.J., et al., *A boundary element between Tsix and Xist binds the chromatin insulator Ctf and contributes to initiation of X-chromosome inactivation*. Genetics, 2011. **189**(2): p. 441-54.
1009. Gontan, C., et al., *RNF12 initiates X-chromosome inactivation by targeting REX1 for degradation*. Nature, 2012. **485**(7398): p. 386-90.
1010. Schoenfelder, S., et al., *Preferential associations between co-regulated genes reveal a transcriptional interactome in erythroid cells*. Nat Genet, 2010. **42**(1): p. 53-61.
1011. Augui, S., E.P. Nora, and E. Heard, *Regulation of X-chromosome inactivation by the X-inactivation centre*. Nat Rev Genet, 2011. **12**(6): p. 429-42.
1012. Dixon, J.R., et al., *Topological domains in mammalian genomes identified by analysis of chromatin interactions*. Nature, 2012. **485**(7398): p. 376-80.
1013. Nesterova, T.B., et al., *Loss of Xist imprinting in diploid parthenogenetic preimplantation embryos*. Dev Biol, 2001. **235**(2): p. 343-50.
1014. Wang, Z. and R. Jaenisch, *At most three ES cells contribute to the somatic lineages of chimeric mice and of mice produced by ES-tetraploid complementation*. Dev Biol, 2004. **275**(1): p. 192-201.
1015. Dvash, T., N. Lavon, and G. Fan, *Variations of X chromosome inactivation occur in early passages of female human embryonic stem cells*. PLoS One, 2010. **5**(6): p. e11330.

References

1016. Lengner, C.J., et al., *Derivation of pre-X inactivation human embryonic stem cells under physiological oxygen concentrations*. *Cell*, 2010. **141**(5): p. 872-83.
1017. Hanna, J., et al., *Human embryonic stem cells with biological and epigenetic characteristics similar to those of mouse ESCs*. *Proc Natl Acad Sci U S A*, 2010. **107**(20): p. 9222-7.
1018. Diaz Perez, S.V., et al., *Derivation of new human embryonic stem cell lines reveals rapid epigenetic progression in vitro that can be prevented by chemical modification of chromatin*. *Hum Mol Genet*, 2012. **21**(4): p. 751-64.
1019. Tchieu, J., et al., *Female human iPSCs retain an inactive X chromosome*. *Cell Stem Cell*, 2010. **7**(3): p. 329-42.
1020. Ananiev, G., et al., *Isogenic pairs of wild type and mutant induced pluripotent stem cell (iPSC) lines from Rett syndrome patients as in vitro disease model*. *PLoS One*, 2011. **6**(9): p. e25255.
1021. Cheung, A.Y., et al., *Isolation of MECP2-null Rett Syndrome patient hiPS cells and isogenic controls through X-chromosome inactivation*. *Hum Mol Genet*, 2011. **20**(11): p. 2103-15.
1022. Amenduni, M., et al., *iPS cells to model CDKL5-related disorders*. *Eur J Hum Genet*, 2011. **19**(12): p. 1246-55.
1023. Pomp, O., et al., *Unexpected X chromosome skewing during culture and reprogramming of human somatic cells can be alleviated by exogenous telomerase*. *Cell Stem Cell*, 2011. **9**(2): p. 156-65.
1024. Marchetto, M.C., et al., *A model for neural development and treatment of Rett syndrome using human induced pluripotent stem cells*. *Cell*, 2010. **143**(4): p. 527-39.
1025. Kim, K.Y., E. Hysolli, and I.H. Park, *Neuronal maturation defect in induced pluripotent stem cells from patients with Rett syndrome*. *Proc Natl Acad Sci U S A*, 2011. **108**(34): p. 14169-74.
1026. Bruck, T. and N. Benvenisty, *Meta-analysis of the heterogeneity of X chromosome inactivation in human pluripotent stem cells*. *Stem Cell Res*, 2011. **6**(2): p. 187-93.
1027. Sperger, J.M., et al., *Gene expression patterns in human embryonic stem cells and human pluripotent germ cell tumors*. *Proc Natl Acad Sci U S A*, 2003. **100**(23): p. 13350-5.
1028. Mitjavila-Garcia, M.T., et al., *Partial reversal of the methylation pattern of the X-linked gene HUMARA during hematopoietic differentiation of human embryonic stem cells*. *J Mol Cell Biol*, 2010. **2**(5): p. 291-8.
1029. Park, I.H., et al., *Disease-specific induced pluripotent stem cells*. *Cell*, 2008. **134**(5): p. 877-86.
1030. Huangfu, D., et al., *Induction of pluripotent stem cells from primary human fibroblasts with only Oct4 and Sox2*. *Nat Biotechnol*, 2008. **26**(11): p. 1269-75.
1031. Lyssiotis, C.A., et al., *Reprogramming of murine fibroblasts to induced pluripotent stem cells with chemical complementation of Klf4*. *Proc Natl Acad Sci U S A*, 2009. **106**(22): p. 8912-7.
1032. Warlich, E., et al., *Lentiviral vector design and imaging approaches to visualize the early stages of cellular reprogramming*. *Mol Ther*, 2011. **19**(4): p. 782-9.
1033. Mekhoubad, S., et al., *Erosion of dosage compensation impacts human iPSC disease modeling*. *Cell Stem Cell*, 2012. **10**(5): p. 595-609.
1034. Anguera, M.C., et al., *Molecular signatures of human induced pluripotent stem cells highlight sex differences and cancer genes*. *Cell Stem Cell*, 2012. **11**(1): p. 75-90.
1035. Nazor, K.L., et al., *Recurrent variations in DNA methylation in human pluripotent stem cells and their differentiated derivatives*. *Cell Stem Cell*, 2012. **10**(5): p. 620-34.
1036. Tomoda, K., et al., *Derivation conditions impact x-inactivation status in female human induced pluripotent stem cells*. *Cell Stem Cell*, 2012. **11**(1): p. 91-9.
1037. Najm, F.J., et al., *Isolation of epiblast stem cells from preimplantation mouse embryos*. *Cell Stem Cell*, 2011. **8**(3): p. 318-25.
1038. O'Leary, T., et al., *Tracking the progression of the human inner cell mass during embryonic stem cell derivation*. *Nat Biotechnol*, 2012. **30**(3): p. 278-82.
1039. Ostendorff, H.P., et al., *Ubiquitination-dependent cofactor exchange on LIM homeodomain transcription factors*. *Nature*, 2002. **416**(6876): p. 99-103.
1040. Hiratani, I., et al., *Selective degradation of excess Ldb1 by Rnf12/RLIM confers proper Ldb1 expression levels and Xlim-1/Ldb1 stoichiometry in Xenopus organizer functions*. *Development*, 2003. **130**(17): p. 4161-75.
1041. Ostendorff, H.P., et al., *Dynamic expression of LIM cofactors in the developing mouse neural tube*. *Dev Dyn*, 2006. **235**(3): p. 786-91.
1042. Scanlan, M.J., et al., *Antigens recognized by autologous antibody in patients with renal-cell carcinoma*. *Int J Cancer*, 1999. **83**(4): p. 456-64.

References

1043. Gaspar, C., et al., *Cross-species comparison of human and mouse intestinal polyps reveals conserved mechanisms in adenomatous polyposis coli (APC)-driven tumorigenesis*. *Am J Pathol*, 2008. **172**(5): p. 1363-80.
1044. Zhang, L., et al., *RNF12 Controls Embryonic Stem Cell Fate and Morphogenesis in Zebrafish Embryos by Targeting Smad7 for Degradation*. *Mol Cell*, 2012. **46**(5): p. 650-61.
1045. Macdonald, D.H., et al., *Cloning and characterization of RNF6, a novel RING finger gene mapping to 13q12*. *Genomics*, 1999. **58**(1): p. 94-7.
1046. Lo, H.S., et al., *Identification of somatic mutations of the RNF6 gene in human esophageal squamous cell carcinoma*. *Cancer Res*, 2002. **62**(15): p. 4191-3.
1047. Lopez, P., et al., *Gene control in germinal differentiation: RNF6, a transcription regulatory protein in the mouse sertoli cell*. *Mol Cell Biol*, 2002. **22**(10): p. 3488-96.
1048. Jaenisch, R. and R. Young, *Stem cells, the molecular circuitry of pluripotency and nuclear reprogramming*. *Cell*, 2008. **132**(4): p. 567-82.
1049. Jani, M.M., et al., *Molecular characterization of tiny ring X chromosomes from females with functional X chromosome disomy and lack of cis X inactivation*. *Genomics*, 1995. **27**(1): p. 182-8.
1050. Tomkins, D.J., et al., *Lack of expression of XIST from a small ring X chromosome containing the XIST locus in a girl with short stature, facial dysmorphism and developmental delay*. *Eur J Hum Genet*, 2002. **10**(1): p. 44-51.
1051. Namekawa, S.H., et al., *Postmeiotic sex chromatin in the male germline of mice*. *Curr Biol*, 2006. **16**(7): p. 660-7.
1052. Williams, L.H., et al., *Transcription precedes loss of Xist coating and depletion of H3K27me3 during X-chromosome reprogramming in the mouse inner cell mass*. *Development*, 2011. **138**(10): p. 2049-57.
1053. Chuvp de Sousa Lopes, S.M., et al., *X chromosome activity in mouse XX primordial germ cells*. *PLoS Genet*, 2008. **4**(2): p. e30.
1054. de Napoles, M., T. Nesterova, and N. Brockdorff, *Early loss of Xist RNA expression and inactive X chromosome associated chromatin modification in developing primordial germ cells*. *PLoS One*, 2007. **2**(9): p. e860.
1055. Sugimoto, M. and K. Abe, *X chromosome reactivation initiates in nascent primordial germ cells in mice*. *PLoS Genet*, 2007. **3**(7): p. e116.
1056. Mise, N., et al., *Differences and similarities in the developmental status of embryo-derived stem cells and primordial germ cells revealed by global expression profiling*. *Genes Cells*, 2008. **13**(8): p. 863-77.
1057. Takagi, N., et al., *Reversal of X-inactivation in female mouse somatic cells hybridized with murine teratocarcinoma stem cells in vitro*. *Cell*, 1983. **34**(3): p. 1053-62.
1058. Do, J.T., et al., *Erasure of cellular memory by fusion with pluripotent cells*. *Stem Cells*, 2007. **25**(4): p. 1013-20.
1059. Guo, G. and A. Smith, *A genome-wide screen in EpiSCs identifies Nr5a nuclear receptors as potent inducers of ground state pluripotency*. *Development*, 2010. **137**(19): p. 3185-92.
1060. Eggan, K., et al., *X-Chromosome inactivation in cloned mouse embryos*. *Science*, 2000. **290**(5496): p. 1578-81.
1061. Inoue, K., et al., *Impeding Xist expression from the active X chromosome improves mouse somatic cell nuclear transfer*. *Science*, 2010. **330**(6003): p. 496-9.
1062. Maherali, N. and K. Hochedlinger, *Tgfbeta signal inhibition cooperates in the induction of iPSCs and replaces Sox2 and cMyc*. *Curr Biol*, 2009. **19**(20): p. 1718-23.
1063. Takagi, N. and K. Abe, *Detrimental effects of two active X chromosomes on early mouse development*. *Development*, 1990. **109**(1): p. 189-201.
1064. Guy, J., et al., *Reversal of neurological defects in a mouse model of Rett syndrome*. *Science*, 2007. **315**(5815): p. 1143-7.
1065. Gadalla, K.K., M.E. Bailey, and S.R. Cobb, *MeCP2 and Rett syndrome: reversibility and potential avenues for therapy*. *Biochem J*, 2011. **439**(1): p. 1-14.
1066. Michalon, A., et al., *Chronic pharmacological mGlu5 inhibition corrects fragile X in adult mice*. *Neuron*, 2012. **74**(1): p. 49-56.

Appendix

- Abbreviations
- Summary
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Abbreviations

BAC: Bacterial Artificial Chromosome
BF: Blocking Factor
CF: Competence Factor
(h)ES cell: (human) Embryonic Stem cell
EpiSC: Epiblast derived Stem Cell
FISH: Fluorescent *In Situ* Hybridization
ICM: Inner Cell Mass
(h)iPS cell: (human) induced Pluripotent Stem cell
iXCI: imprinted X Chromosome Inactivation
Mb: Mega base
MSCI: Meiotic Sex Chromosome Inactivation
MYA: Million Years Ago
OMIM: Online Mendelian Inheritance in Man database
PAR: Pseudoautosomal Region
PGCs: Primordial Germ Cells
rXCI: random X Chromosome Inactivation
TBP: TATA Binding Protein
Xa: active X chromosome
XAR: X Added Region
Xce: X controlling element
XCI: X Chromosome Inactivation
XCR: X Conserved Region
Xi: inactive X chromosome
Xic: X inactivation center
XicHR: X inactivation center Homologous Region
Xist: X inactive specific transcript
Xm: maternally derived X chromosome
Xp: paternally derived X chromosome, or short art of the human X chromosome
Xpr: X pairing region
Xq: long arm of the human X chromosome
YAC: Yeast Artificial Chromosome

Summary

The evolution of placental mammals has resulted in a genetic mechanism of sex determination with X and Y sex chromosomes, where females of most species have two X chromosomes and the males have a single X chromosome and a small Y chromosome. These sex chromosomes originated from a non-sex-specific pair of autosomes. In males the deterioration of the Y chromosome resulted in a loss of genes, compared to the X chromosome. To balance this loss, the single X chromosome in males became transcriptionally up-regulated, resulting in a dosage compensation mechanism between genes located on the sex chromosomes and genes on autosomes. In females, this up-regulation of X-linked genes resulted in a subsequent dosage problem, as two copies of X chromosomes are present. Therefore, in females, an additional dosage compensation mechanism was required, which results in the transcriptional silencing of a single X chromosome in females. This silencing of an X chromosome in females, leaving one active X chromosome per diploid genome, is called X chromosome inactivation (XCI), and is the central topic in this thesis work.

XCI occurs during early female embryonic development, and results in transcriptional silencing of one X chromosome in females, thereby equalizing the dosage of X-linked genes between both sexes and between the sex chromosomes and autosomes. This is needed for proper biochemical and physiological functioning of cells in the body, as gene products derived from the X chromosome need to interact with gene products from autosomal genes. Aberrations of the crucial XCI process are believed to be lethal, as no healthy females are observed having more than one active X chromosome, and females in which parts of both X chromosomes are fully active suffer from severe phenotypes. The XCI process itself is random regarding the choice of the X chromosome that will be inactivated, resulting in silencing of either the X chromosome derived from the father or the X chromosome derived from the mother, in the cells forming the embryo. Therefore, growing and adult females consist of a mosaic of two different cell populations, which can have profound impacts on health and disease manifestations in females, as described in the Introduction in Chapter 1. Many genes located on the X chromosome can cause diseases when mutated. In general, females suffer less from X-chromosomal diseases, due to the presence of the mosaic, in which some cells will express the mutant gene, whereas others will express the non-mutated backup copy from the healthy X chromosome. Such a backup mechanism is not possible in males carrying a single X chromosome, who therefore always show expression of an X-linked mutation. Besides this advantage for females, of having two X chromosomes, there is also a disadvantage. The XCI process seems to be difficult to establish properly in all female cells, which might explain higher female lethality during peri-implantation development. Furthermore, the presence of mosaicism can sometimes cause a disease, when the choice of inactivation is not random between the X chromosomes present. Therefore, insight in the mechanisms resulting in epigenetic silencing of a single X chromosome in females is needed, and XCI is an important subject to study.

Following the initial discovery of the XCI process in 1961 by Mary Lyon, many investigations have tried to unravel the mechanisms leading to the shut-down of one X chromosome in females. Most of these studies have been performed in mice, by making use of either genetic mouse models or embryonic stem (ES) cells. ES cells are pluripotent cells derived from early embryos, and can be used to study several aspects of the XCI process in a culture dish, as undifferentiated female ES cells have two active X chromosomes, and upon differentiation to more specialized cells these cells engage in XCI, similar to what happens in early embryonic development. By making use of these models, studies have shown that a small region of the X chromosome, the X inactivation center (Xic), is crucial for XCI to occur, as this process is only initiated in cells having two copies of this regulatory region on two distinct X chromosomes. The Xic harbors several conserved genes, of which the *X inactive transcript (Xist)* is crucial for XCI to occur. *Xist* is the only gene which is specifically expressed from the inactive X chromosome, and transcription of this gene results in a non-coding RNA, which spreads along the X chromosome and induces silencing. Another non-coding gene, *Tsix*, has been found in the Xic, and this gene is transcribed in the opposite direction to *Xist* and functions as a negative regulator of *Xist*.

During female embryonic development, or upon ES cell differentiation, *Xist* is specifically up-regulated on a single X chromosome, thereby starting the XCI process. Even 20 years after the discovery of *Xist*, a longstanding question has remained how this gene is specifically up-regulated on a single X chromosome in cells having more than one X chromosome. Several models have tried to explain this female specific initiation of XCI, and in the past, we have proposed that sex-specific initiation of XCI is regulated by a stochastic process. In this model, autosomally-encoded XCI-inhibitors prevent XCI in undifferentiated cells or cells in early embryos, by directly inhibiting *Xist* or stimulating *Tsix*. These XCI-inhibitors are encoded by genes located on autosomes, in both sexes, so that an identical level of XCI repression will be present. In contrast, X-encoded activators of XCI will stimulate the XCI process. These activators are encoded by X-linked genes, which implies that in females a two-fold higher dosage of XCI-activators will be present compared to males. These activators are most likely developmentally regulated, with low or absent expression in undifferentiated cells. Therefore, in undifferentiated cells, XCI will be absent, and will only be initiated during differentiation, as XCI activators will become up-regulated. This up-regulation will most likely occur in both sexes, but only in females the level of activation will be high enough to reach a threshold which is set by the combined action of the XCI-inhibitors. In this model, XCI-activators work in *trans*, meaning that they can activate XCI on all X chromosomes in a cell nucleus, independent from which X chromosome they are derived, resulting in a stochastic choice which *Xist* allele will become up-regulated. Once up-regulation has occurred, silencing of the XCI-activator gene in *cis*, thus on the same X chromosome where *Xist* will spread, will also silence the XCI-activator gene. Hence, the level of XCI-activator will drop, to a level which is now similar to that of a male cell harboring a single X chromosome. This simple mechanism ensures that XCI initiation on the other X chromosome will be prevented. *Tsix* is also silenced in *cis*, so that XCI can be maintained on the X chromosome which has started inactivation despite the lower level of the XCI-activators. Therefore, in this model, initiation of XCI is regulated by a sex-specific dosage difference in XCI-activators.

Although this model could principally explain all features of the XCI initiation mechanism, the identity of the XCI-activator genes remained illusive. We therefore set out to identify genes encoding such activators. By making use of a Bacterial Artificial Chromosome (BAC) transgenesis screen in ES cells, described in Chapter 2, we identified X-encoded RNF12 as the first known XCI activator. When *Rnf12* was over-expressed in male ES cells, this resulted in XCI initiation on the single X chromosome, and over-expression in female ES cells resulted in XCI on both X chromosomes in a significant portion of cells. During differentiation of ES cells, endogenous *Rnf12* becomes up-regulated, and a two-fold higher dose of RNF12 has been found in female compared to male ES cells prior to and during XCI initiation. After XCI has occurred, expression levels are equal between male and female cells. This two-fold higher dosage of RNF12 explains female specific initiation of XCI, as only in female cells a threshold will be reached upon differentiation which is needed to overcome repression of XCI by XCI-inhibitors, resulting in *Xist* accumulation. These XCI-inhibitors are mainly pluripotency associated factors, which either repress *Xist* directly, or indirectly, via stimulation of *Tsix* expression, or they might even work via repression of XCI-activators. In line with this, over-expression of *Rnf12* even leads to initiation of XCI in undifferentiated cells, indicating that absence of XCI-activator activity is important in preventing XCI in undifferentiated cells. More evidence for an important role of RNF12 in the initiation of XCI was obtained when one of the two copies of *Rnf12* was removed from female ES cells, as this resulted in delayed XCI initiation in these cells, favoring inactivation of the X chromosome harboring the *Rnf12* deletion. To generate this and other genetic modifications of ES cells described throughout this thesis work, we developed a new BAC targeting strategy described in Chapter 3, which makes use of polymorphic ES cells. Chapter 4 discusses the identification of RNF12 as an XCI-activator, and describes this discovery in more detail in the context of the stochastic model of XCI initiation. We next asked the question whether RNF12 is essential for XCI to occur, by generation and analysis of *Rnf12* knockout ES cells, described in Chapter 5. These cells were found to be devoid of XCI, providing evidence that XCI is not occurring in the absence of this crucial activator. Additional genetic experiments performed indicated that the XCI-activating function of RNF12 works mainly on *Xist*, as *Xist* transgenes were higher expressed when *Rnf12* was over-expressed, whereas the expression of *Tsix* transgenes was not affected. As RNF12 is an E3 ubiquitin ligase, it seems likely that such *Xist* activation should involve an indirect mechanism, in which RNF12 would target a repressor of *Xist* for proteasomal degradation. Indeed, studies aiming to identify the interaction partners of RNF12, described in Chapter 6, indicated that RNF12 targets the pluripotency associated transcription factor REX1 for degradation. REX1 was found to bind the *Xist* promoter and distal region, thereby repressing *Xist* expression in undifferentiated cells. Upon differentiation, *Rnf12* will become up-regulated, leading to a dose-dependent catalysis of the breakdown of REX1, which will result in loss of repression of *Xist*, and hence will lead to initiation of XCI. The *Rnf12* gene is located in the near vicinity of *Xist*, which ensures rapid silencing of *Rnf12* transcription in *cis* shortly after initiation of XCI. Chapter 7 addresses the question whether other genes located between *Xist* and *Rnf12* are also functional in the regulation of the XCI process. We found that *Jpx*, *Ftx* and the *Xpr* region have a *cis* regulatory role, in which these regions likely create an active environment, which is needed to enable activation of *Xist* by the *trans* action of

RNF12. We propose to divide the Xic in a *cis*-Xic, composed of all genes and elements involved in the regulation of *Xist* and *Tsix* expression in *cis*, and a *trans*-Xic, which is required for *trans* regulation of the *cis*-Xic, and hence for counting the number of X chromosomes present. We also provide evidence which suggest that direct interaction between the two X chromosomes in a female nucleus, referred to as X-pairing, is not functionally required for XCI to occur. Rather, more evidence was obtained supporting an indispensable role for the *trans* action of RNF12, even for the maintenance of XCI initiation.

We next proceeded to test the role of RNF12 *in vivo*, by generating an *Rnf12* knockout mouse model, described in Chapter 8. As expected from the observations showing loss of XCI in *Rnf12* homozygous knockout ES cells, homozygous *Rnf12* knockout females are not born, suggesting that RNF12 has an important role in XCI initiation during *in vivo* development. Females heterozygous for the *Rnf12* mutation also have an XCI phenotype, with transcriptional activity from two X chromosomes in some cells from adult tissues representing loss of XCI. This surprising and remarkable phenotype might make it necessary to change the dogma that living with two active X chromosomes is not possible. This awaits detailed investigation, but promises to have important implications for the understanding and future therapies of X-linked diseases in humans. Finally, in Chapter 9, we proceeded to translate the newly obtained knowledge on the regulation of XCI initiation from mouse to human. We studied XCI in female human induced pluripotent stem cells, and provide evidence that the X chromosome that is inactivated in a founder human fibroblast becomes reactivated upon somatic reprogramming. This might allow future generation of novel model systems to study XCI in humans. The General Discussion, Chapter 10, aims to integrate our current views on the initiation of XCI with previously published observations, and provides an outlook for future investigations.

In conclusion, the work described in this thesis has allowed us to obtain novel insights in the mechanisms controlling the initiation of XCI, including the role of RNF12, and provides a framework for future studies investigating the possibility to live with two active X chromosomes.

Samenvatting

De evolutie van zoogdieren heeft geresulteerd in een genetisch mechanisme van geslachtsdeterminatie waarbij vrouwen van de meeste soorten twee X geslachtschromosomen hebben, in tegenstelling tot mannen met een enkel X chromosoom en daarnaast een Y geslachtschromosoom. De X en Y geslachtschromosomen zijn ontstaan uit een niet-geslachtsgebonden chromosomenpaar. Het snelle verlies van genen van het Y chromosoom, in tegenstelling tot een geleidelijke toename van het aantal genen op het X chromosoom, heeft geresulteerd in een situatie waarbij het X chromosoom verreweg de meeste genen bevat. Het Y-chromosomale verlies van genen werd gecompenseerd door een verhoging van de gen-transcriptie van het enkele X chromosoom in mannen, zodat een vergelijkbare dosis van geslachtsgebonden en niet-geslachtsgebonden genen behouden werd. Aangezien vrouwen twee X chromosomen hebben, zou een dergelijke verhoogde transcriptie van X-chromosomale genen tot een nieuw gendosis-probleem in vrouwen leiden. Dit werd voorkomen door een mechanisme, waarbij één van beide X chromosomen van de vrouw wordt uitgeschakeld. Dit proces, genaamd X-chromosoom-inactivatie (XCI), resulteert in niet meer dan één actief X chromosoom per vrouwelijk diploid genoom, en is het centrale onderwerp van dit proefschrift.

XCI vindt plaats tijdens de vroege embryonale ontwikkeling van vrouwen, en resulteert in het uitschakelen van één van beide X chromosomen, waardoor de juiste dosis van X-gebonden genen tussen de geslachten, en tussen de geslachts- en de niet-geslachtsgebonden chromosomen behouden blijft. Dit is noodzakelijk om belangrijke processen in cellen goed te kunnen laten verlopen, aangezien veel genproducten van het X chromosoom in de juiste verhouding moeten kunnen samenwerken met genproducten van andere chromosomen. Afwijkingen in het cruciale XCI proces zijn dan ook embryonaal lethaal. Er zijn geen gezonde vrouwen waarbij volledige activiteit van beide X chromosomen werd vastgesteld, en zelfs de dubbele activiteit van kleine delen van het X chromosoom kan leiden tot aanzienlijke ziekteprocessen. Het XCI proces zelf kent een aspect van toeval, waarbij in cellen van het vrouwelijke embryo het X chromosoom verkregen van moeder of het X chromosoom verkregen van vader wordt uitgeschakeld. Door dit willekeurige proces zijn vrouwen een mozaïek van twee verschillende celpopulaties, hetgeen een aanzienlijk effect kan hebben op de manifestatie van ziekten, zoals beschreven in de Inleiding in Hoofdstuk 1. Vele genen gelokaliseerd op het X chromosoom kunnen ziekten veroorzaken indien zij afwijkend zijn, maar over het algemeen zijn ziekteverschijnselen bij vrouwen dankzij dit mozaïek milder vergeleken met mannen, aangezien veel cellen in vrouwen het intacte gen tot expressie zullen brengen. In mannen is een dergelijke back-up mechanisme niet mogelijk, en daarom zullen afwijkingen op het enkele X chromosoom altijd tot uitdrukking komen. Afgezien van deze voordelen voor vrouwen, zijn er ook nadelen verbonden aan het bezit van twee X chromosomen. Er zijn aanwijzingen dat het moeilijk is om het XCI proces goed te laten verlopen in alle vrouwelijke cellen tijdens de embryonale ontwikkeling, hetgeen de hogere sterfte van vrouwelijke embryo's tijdens de vroege embryonale ontwikkeling voorafgaand aan en rondom de implantatie fase zou kunnen verklaren. Tevens kan de aanwezigheid van een mozaïek soms ook problemen veroorzaken, in het bijzonder als de keus tussen het inactiveren van één van de X chromosomen niet willekeurig is. Daardoor is het van belang

om meer over dit belangrijke proces, dat kan worden gekenmerkt als een epigenetisch proces, te weten te komen.

Sinds de ontdekking van het XCI proces door Mary Lyon in 1961, zijn vele studies gewijd aan het ontrafelen van de mechanismen die er toe leiden dat in vrouwen één X chromosoom wordt uitgeschakeld. Het merendeel van deze studies heeft gebruik gemaakt van muismodellen of muis embryonale stamcellen (ES cellen). ES cellen zijn pluripotente cellen, nog niet gedifferentieerd, verkregen uit embryo's, die gebruikt kunnen worden om het XCI proces in een kweekschaal te bestuderen. Ongedifferentieerde vrouwelijke ES cellen hebben twee actieve X chromosomen, maar ondergaan het XCI proces als deze cellen naar andere, meer gespecialiseerde celtypen worden gedifferentieerd, waarbij deze cellen een eerste differentiatie stap tijdens de vroege embryonale ontwikkeling simuleren. Door gebruik te maken van deze modellen hebben studies laten zien dat een klein gedeelte van het X chromosoom, het zogenaamde X inactivatie centrum (Xic), vereist is om XCI plaats te laten vinden. Meerdere, in verschillende soorten geconserveerde, genen zijn gelokaliseerd in dit Xic, en het meest belangrijke is het *X inactieve transcript (Xist)* gen. *Xist* is het enige gen dat specifiek actief is op het inactieve X chromosoom, en transcriptie van dit gen resulteert in een niet-coderend RNA, hetgeen over het X chromosoom kan spreiden en daardoor het X chromosoom kan uitschakelen. Een ander gen, *Tsix*, is tevens gelokaliseerd in de buurt van *Xist*. *Tsix* wordt in de tegenovergestelde richting afgeschreven in vergelijking met *Xist*, en is een negatieve regulator van het *Xist* gen.

Tijdens de vroege embryonale ontwikkeling van vrouwen of tijdens ES cel-differentiatie wordt het XCI proces gestart met de specifieke verhoging van transcriptie van het *Xist* gen. Ondanks dat het reeds 20 jaar geleden is dat *Xist* ontdekt werd, blijft een belangrijke vraag hoe het komt dat dit gen alleen wordt geactiveerd als meer dan één X chromosoom aanwezig is in de celkern. Verschillende modellen hebben geprobeerd om dit te verklaren, en wij hebben een stochastisch model voorgesteld. In dit model wordt XCI in ongedifferentieerde ES cellen of vroege embryo's voorkomen door de werking van XCI-remmers, die gecodeerd worden door niet-geslachtsgebonden chromosomen. Deze remmen *Xist* direct, of stimuleren de negatieve regulator *Tsix*. Aangezien in dit model de XCI-remmers afkomstig zijn van genen op niet-geslachtsgebonden chromosomen, zal de mate van repressie in vrouwelijke en mannelijke cellen gelijk zijn. In tegenstelling tot deze XCI-remmers, leiden XCI-activatoren tot activatie van het XCI proces. Indien deze XCI-activatoren gecodeerd worden door genen gelokaliseerd op het X chromosoom, zal in ongedifferentieerde vrouwelijke cellen met nog twee actieve X chromosomen een tweevoudige dosis van activatoren aanwezig zijn vergeleken met mannelijke cellen die maar één X chromosoom hebben. Naar verwachting zijn XCI-activatoren ontwikkelingsbiologisch gereguleerd, zodat ze tijdens ongedifferentieerde celstadia laag, of zelfs geheel niet, tot expressie komen. Hierdoor zal het proces van XCI in ongedifferentieerde cellen niet op gang komen. Echter tijdens differentiatie en verdere ontwikkeling van de cellen zal de expressie van deze XCI-activator genen toenemen, in beide geslachten. De twee actieve X chromosomen in vrouwelijke cellen zorgen voor een hoge dosis XCI-activatoren, voldoende om de remming van het XCI proces door de XCI-remmers te overkomen. De XCI-activatoren werken in *trans*, hetgeen betekend dat ze zowel XCI op het X chromosoom van waar zij geproduceerd worden als ook op alle andere X chromosomen in de celkern kunnen activeren. Dit heeft tot gevolg dat het een

willekeurige keuze zal zijn, welke kopie van *Xist* geactiveerd wordt. Eenmaal geactiveerd, zal *Xist* RNA over het X chromosoom spreiden, in *cis*, en X chromosomale genen op datzelfde X chromosoom uitschakelen. Hierdoor worden ook vrij snel de XCI-activator genen uitgeschakeld, hetgeen er voor zal zorgen dat de concentratie van XCI-activatoren in de celkern weer zal afnemen, zodat XCI niet plaats kan vinden op het tweede X chromosoom in dezelfde celkern. Aangezien de negatieve regulator *Tsix* ook wordt uitgeschakeld, zal de lagere dosis XCI-activator, die nu vergelijkbaar is met de dosis in mannelijke cellen, voldoende zijn om initiatie van XCI te behouden. Samengevat, in dit model wordt initiatie van het XCI proces dus gereguleerd door een geslachtsspecifiek verschil in de dosis van XCI-activatoren.

Hoewel dit model in principe alle eigenschappen van het XCI proces zou kunnen beschrijven, was de identiteit van de XCI-activator nog onbekend. We hebben daarom dit onderzoek gestart met een zoek-strategie om deze XCI-activatoren te identificeren, waarbij we gebruik maakten van bacteriële artificiële chromosomen (BACs) die we tot hoge expressie brachten in ES cellen. Door deze strategie, die beschreven staat in Hoofdstuk 2, ontdekten wij dat het door het X-chromosoom gecodeerde RNF12 eiwit de functie van XCI-activator kan uitoefenen. Wanneer het *Rnf12* gen tot hoge expressie werd gebracht in mannelijke ES cellen, resulteerde dit in XCI initiatie van het enkele X chromosoom in deze mannelijke cellen, en hoge expressie in vrouwelijke ES cellen resulteerde in XCI van beide X chromosomen in een hoog percentage van de cellen. Tijdens differentiatie van ES cellen komt het normaal aanwezige X-gebonden *Rnf12* gen ook hoger tot expressie, en een tweevoudige dosis van RNF12 werd gevonden in vrouwelijke cellen, vergeleken met mannelijke cellen, voor en tijdens XCI initiatie. Nadat de initiatie van het XCI proces is afgerond, is de hoeveelheid RNF12 gelijk tussen cellen van beide geslachten. Met dit dosis-afhankelijke mechanisme kan worden verklaard dat initiatie van XCI plaatsvindt in vrouwelijke cellen. De XCI-remmers zijn met name betrokken bij het in stand houden van de pluripotente staat van ongedifferentieerde cellen, en remmen *Xist* direct, of indirect via stimulatie van *Tsix* expressie. Tevens zouden zij kunnen werken via remming van de XCI-activatoren. Dit laatste wordt ondersteund door de vinding dat hoge expressie van RNF12 reeds tot activatie van XCI leidt in ongedifferentieerde cellen, hetgeen erop wijst dat remming van XCI-activatoren een belangrijk mechanisme zou kunnen zijn om XCI in ongedifferentieerde cellen te voorkomen. Meer bewijs voor een belangrijke rol van RNF12 bij de initiatie van XCI werd verkregen door het genetisch verwijderen van één van de twee X-gebonden *Rnf12* genen in vrouwelijke ES cellen. Dit resulteerde in vertraagde initiatie van XCI in deze cellen, die bij voorkeur het X chromosoom waarop het gemuteerde *Rnf12* gen was gelokaliseerd uitzetten. Om deze en andere genetische modificaties van ES cellen te bewerkstelligen, ontwikkelden wij een nieuwe BAC targeting strategie, die beschreven staat in Hoofdstuk 3 van dit proefschrift, en die gebruik maakt van polymorfe ES cellen verkregen door kruisingen van verschillende muizenstammen. In Hoofdstuk 4 bespreken we meer uitvoerig de bevindingen verkregen uit de *Rnf12* proeven, en zetten we deze uiteen in de context van het stochastische model van XCI. In het vervolg vroegen wij ons af of RNF12 een essentiële rol heeft in het XCI proces. Om dit te beantwoorden genereerden wij *Rnf12* knockout ES cellen, zoals beschreven in Hoofdstuk 5. In deze cellen bleek XCI niet op te treden tijdens differentiatie, hetgeen bevestigd dat RNF12 een cruciale rol bij het tot stand brengen van dit XCI proces speelt. Andere experimenten lieten zien dat de XCI-

activerende functie van RNF12 voornamelijk via *Xist* activatie werkt, aangezien *Xist* transgenen hoger tot expressie kwamen wanneer *Rnf12* tot hoge expressie werd gebracht, terwijl expressie van *Tsix* transgenen niet werd beïnvloed. RNF12 is een E3 ubiquitine ligase, een enzym dat betrokken is bij proteasomale eiwit-afbraak, en het is daarom aannemelijk dat deze *Xist* activatie verloopt via een indirect mechanisme, waarbij RNF12 een remmer van *Xist* zou markeren voor afbraak. Inderdaad lieten studies beschreven in Hoofdstuk 6 zien dat RNF12 de pluripotentie-geassocieerde transcriptiefactor REX1 target voor deze vorm van afbraak. We vonden dat REX1 bindt aan de *Xist* promotor, waardoor het de expressie van *Xist* remt in ongedifferentieerde cellen. Tijdens differentiatie komt RNF12 hoger tot expressie, hetgeen zal leiden tot een dosis-afhankelijke afbraak van REX1, waardoor de rem op *Xist* expressie verdwijnt en het XCI proces op gang zal komen. Op het X chromosoom is *Rnf12* dicht bij *Xist* gelokaliseerd. Dit zorgt ervoor dat na XCI initiatie *Rnf12* spoedig wordt uitgeschakeld in *cis*. In Hoofdstuk 7 behandelen wij de vraag of ook andere genen in de buurt van *Xist* en *Rnf12* betrokken zijn bij het XCI proces. Hier vonden wij dat *Jpx*, *Ftx* en de *Xpr* regio een rol spelen in de *Xist* regulatie in *cis*, hetgeen nodig zal zijn om RNF12 in staat te stellen om *Xist* in *trans* te kunnen activeren. Daarom stellen wij een nieuwe indeling van het Xic voor, waarbij we spreken van een *cis*-Xic, een regio die alle genen en elementen bevat die noodzakelijk zijn voor de *cis* regulatie van *Xist* en *Tsix*, en een *trans*-Xic, die nodig is voor de *trans* regulatie van de *cis*-Xic, met andere woorden voor het tellen van de hoeveelheid X chromosomen die aanwezig zijn. Verder vonden wij in experimenten beschreven in hetzelfde Hoofdstuk dat directe interactie tussen de beide X chromosomen, aangeduid met *X-pairing*, niet noodzakelijk is voor normale regulatie van XCI. In tegenstelling daarvan brachten deze experimenten verder inzicht in de belangrijke rol van RNF12 ook tijdens het in stand houden van de XCI. Om vast te stellen of RNF12 inderdaad een belangrijke rol *in vivo* speelt hebben wij een *Rnf12* deficiënt muis-model ontwikkeld, hetgeen beschreven staat in Hoofdstuk 8. Zoals reeds verwacht aan de hand van de *Rnf12* knockout ES cellen die geen XCI meer initiëren, bleken vrouwelijke *Rnf12* knockout muizen niet levensvatbaar, hetgeen suggereert dat RNF12 ook een belangrijke rol speelt tijdens de embryonale ontwikkeling *in vivo*. Heterozygote *Rnf12*^{+/-} vrouwtjes hebben ook een XCI fenotype, waarbij in volwassen muizen cellen aanwezig zijn die twee actieve X chromosomen hebben. Deze verrassende vinding dient verder te worden onderzocht, en zou belangrijke nieuwe inzichten kunnen verschaffen over onze kennis en mogelijk nieuwe behandelingen van X-chromosomale ziekten. Tevens kan aan de hand van dit muismodel wellicht het dogma worden bijgesteld, waarbij ervan wordt uitgegaan dat de aanwezigheid van twee actieve X chromosomen niet levensvatbaar is. Tenslotte, in Hoofdstuk 9 proberen we onze nieuwe inzichten in de XCI regulatie in muizen te vertalen naar de mens, waarbij we XCI hebben bestudeerd in vrouwelijke humane geïnduceerde pluripotente stamcellen. Hier vonden wij dat het geïnactiveerde X chromosoom in humane fibroblasten tijdens het reprogrammeren gereactiveerd wordt. Dit kan mogelijk helpen om toekomstige modelsystemen te genereren om humane XCI te bestuderen. De Algemene Discussie in Hoofdstuk 10 vat de gevonden data samen, integreert deze met eerder gepubliceerde studies en biedt een vooruitzicht op toekomstige studies. Concluderend heeft het werk beschreven in dit proefschrift bijgedragen aan nieuwe inzichten in de regulatie van XCI en de rol van RNF12 hierin, en biedt het een basis voor toekomstige studies die zullen onderzoeken in hoeverre het mogelijk is om met twee actieve X chromosomen te leven.

Curriculum Vitae

Name: Tahsin Stefan Barakat

Born: 25 May 1984

City of Birth: Meerbusch, Germany

Place of Living: Wissenkerke, The Netherlands

Educational Background:

2008-2012	PhD student Erasmus MC, University Medical Center, Rotterdam Department of Reproduction and Development
2004-2008	Master of Science in Molecular Medicine Erasmus MC, University Medical Center, Rotterdam
2007-2008	Department of Reproduction and Development Master thesis " <i>Searching for an X-linked activator of X chromosome inactivation</i> ", supervisor Prof.dr. Joost Gribnau
2006-2007	Department of Experimental Pathology Internship, " <i>Characterization of tumor-microenvironment derived effects on Wnt/β-catenin signalling and invasiveness of colorectal cancer cells</i> ", supervisor Prof.dr. Riccardo Fodde
2006	Short lab rotations Department of Cell Biology, supervisor Prof.dr. Elaine Dzierzak Department of Experimental Pathology, supervisor Dr. Ron Smits
2003-2007	Medicine study, Erasmus MC, University Medical Center, Rotterdam
2004-2007	Doctoral exam (<i>cum laude</i>)
2003-2004	Propaedeutic exam
1997-2003	Gymnasium, Pontes Scholengemeenschap, Locatie Het Goese Lyceum, Goes
2003	Voorbereidend Wetenschappelijk onderwijs (VWO) exam (<i>cum laude</i>)
1990-1997	Primary education in Germany and The Netherlands

Curriculum Vitae

Tahsin Stefan Barakat was born on the 25th May of 1984 in Meerbusch, West-Germany. After primary school in Germany and The Netherlands, he graduated *cum laude* for his *Voorbereidend Wetenschappelijk Onderwijs (VWO)* exam in 2003 at the *Pontes Scholengemeenschap, Locatie Het Goese Lyceum*, Goes. In 2003 he started his medical study at the Erasmus MC / Erasmus University, Rotterdam. In 2004 he graduated for his Propaedeutic exam, and was elected for the Master of Science in Molecular Medicine programme at the Erasmus MC. After following several courses belonging to the programme, he performed in 2006 short laboratory rotations at the Departments of Cell Biology (Prof.dr. Elaine Dzierzak) and Experimental Pathology (Dr. Ron Smits). In 2007 he performed a six month research project at the Department of Experimental Pathology under supervision of Prof.dr. Riccardo Fodde, entitled *Characterization of tumor-microenvironment derived effects on Wnt/ β -catenin signalling and invasiveness of colorectal cancer cells*. In 2007 he graduated for his Medical Doctoral exam (*cum laude*). From September 2007 until summer 2008 he was at the Department of Reproduction and Development under supervision of Dr. Joost Gribnau, where he performed the studies described in his Master Thesis, entitled *Searching for an X-linked activator of X chromosome inactivation*. From 2008 onwards, he continued his research in the same laboratory as a PhD student, performing the studies described in this thesis, with Prof.dr. J. Anton Grootegeed and Prof.dr. Joost Gribnau as promotor. In July 2012, he visited the laboratory of Prof.dr. Edith Heard in Paris, to perform FISH experiments on post-implantation embryos. After his public thesis defence in September 2012, he will continue his medical study with the clinical internships to obtain the degree of Medical Doctor, to be followed by a further scientific research career as a post-doctoral fellow.

List of publications

1. Iris Jonkers*, **Tahsin Stefan Barakat***, Eskeatnaf Mulugeta Achame, Kim Monkhorst, Annegien Kenter, Eveline Rentmeester, Frank Grosveld, J. Anton Grootegoed and Joost Gribnau (2009) "RNF12 is an X-Encoded dose-dependent activator of X chromosome inactivation" *Cell* 139(5):999-1011
* both authors contributed equally
2. **Tahsin Stefan Barakat**, Iris Jonkers, Kim Monkhorst and Joost Gribnau (2010) "X-changing information on X inactivation" *Exp Cell Res* 10;316(5):679-87
3. Christa Buecker, Hsu-Hsin Chen, Jose Maria Polo, Laurence Daheron, Lei Bu, **Tahsin Stefan Barakat**, Patricia Okwieka, Andrew Porter, Joost Gribnau, Konrad Hochedlinger, and Niels Geijsen (2010) "A murine ESC-like state facilitates transgenesis and homologous recombination in human pluripotent stem cells" *Cell Stem Cell* 6:535-46.
4. **Tahsin Stefan Barakat** and Joost Gribnau (2010) "X chromosome inactivation and embryonic stem cells" *Adv Exp Med Biol* 695: 132-154.
5. **Tahsin Stefan Barakat**, Nilhan Gunhanlar, Cristina Gontan Pardo, Eskeatnaf Mulugeta Achame, Mehrnaz Ghazvini, Ruben Boers, Annegien Kenter, Eveline Rentmeester, J. Anton Grootegoed and Joost Gribnau (2011) "RNF12 activates *Xist* and is essential for X chromosome inactivation" *PLoS Genet.* 2011;7(1):e1002001
6. **Tahsin Stefan Barakat**, Eveline Rentmeester, Frank Sleutels, J. Anton Grootegoed and Joost Gribnau (2011) "Precise BAC targeting of genetically polymorphic mouse ES cells" *Nucleic Acids Res* 2011 Oct;39(18):e121
7. Long Zhang, Huizhe Huang, Fang Fang Zhou, Joost Schimmel, Cristina Gontan Pardo, Tingting Zhang, **Tahsin Stefan Barakat**, Kelly-Ann Sheppard, Craig Mickanin, Jeff A. Porter, Alfred C.O. Vertegaal, Hans van Dam, Joost Gribnau, Chris X. Lu, and Peter ten Dijke (2012) "RNF12 Controls Embryonic Stem Cell Fate and Morphogenesis in Zebrafish Embryos by Targeting SMAD7 for Degradation" *Mol Cell* 46:650-61.
8. Cristina Gontan, Eskeatnaf Mulugeta Achame, Jeroen Demmers, **Tahsin Stefan Barakat**, Eveline Rentmeester, Wilfred van IJcken, J. Anton Grootegoed and Joost Gribnau (2012) "RNF12 initiates X chromosome inactivation by targeting REX1 for degradation" *Nature* 2012 **485**(7398): p. 386-90
9. **Tahsin Stefan Barakat** and Joost Gribnau (2012) "X chromosome inactivation in the cycle of life" *Development* 139 (12): 2085-2089
10. **Tahsin Stefan Barakat** and Joost Gribnau (2012) "A Restriction Fragment Length Polymorphism based Bacterial Artificial Chromosome targeting strategy for efficient and fast generation of knockout alleles in polymorphic mouse Embryonic Stem cells" (*Manuscript in preparation*)
11. Erwin Brosens, Elisabeth M. de Jong, **Tahsin Stefan Barakat**, Bert H Eussen, Barbara D'haene, Elfride de Baere, Pino P Poddighe, Robert-Jan Galjaard, Joost Gribnau, Alice S Brooks, Dick Tibboel, and Annelies de Klein (2012) "Congenital malformations of the gastro-intestinal tract and Triple X syndrome" (*Submitted*)

12. **Tahsin Stefan Barakat**, Selma van Staveren, Friedemann Loos, Elvira Myronova, Mehrnaz Ghazvini, J. Anton Grootegoed and Joost Gribnau (2012) “Initiation of X inactivation is regulated by *trans*-acting activators and *cis*-acting elements: no evidence for a functional involvement of X-pairing” (*Submitted*)

PhD portfolio

General Academic Skills:

From Development to Disease course, 2009

Article 9 Animal Experimentation course, 2010

Presentations:

2009

BSIK Stem Cells in Development and Disease Conference, Rotterdam

7th Winter School of the International Graduiertenkolleg, Kleinwalsertal, Austria

2nd Dutch Stem Cell Meeting, Rotterdam

1st Erasmus Stem Cell Institute (ESI) meeting, Rotterdam

16th MGC PhD Student Workshop, Brugge, Belgium (Best Oral Presentation)

3rd SCDD International Symposium “Stem Cells, Development and Regulation”
Amsterdam

2010

8th Winter School of the International Graduiertenkolleg, Kleinwalsertal, Austria

2011

Winter Symposium, “Epigenetics in Development and Disease”, Miami, United States of America

4th Dutch Stem Cell Meeting, Leiden

18th MGC PhD Student Workshop, Maastricht

3rd X inactivation Conference, “50 years of X inactivation research”, EMBO-workshop,
Oxford, United Kingdom

2012

2nd Winter School of the Collaborative Research Center (TRR 81), “Chromatin Changes in Differentiation and Malignancies”, Kleinwalsertal, Austria

Paris-Rotterdam X chromosome inactivation meeting, Spetses, Greece

10th Annual meeting of the International Society for Stem Cell Research (ISSCR),
Yokohama, Japan

Awards:

Travel grant of the International Society for Stem Cell Research (ISSCR) for the 10th
Annual meeting in Yokohama, Japan, 2012

International seminars, workshops and conferences:

2008

Stem Cell Symposium, “Stem Cell, Development and Regulation”, Amsterdam

2009

European Molecular Biology Organisation (EMBO) meeting, Amsterdam

MGC day 2009, Rotterdam

Stem Cell Symposium, “Stem Cell, Development and Regulation”, Amsterdam

JNI Symposium, “The stem of cancer”, Rotterdam

2010

Epigenome Network of Excellence meeting: “Imprinting and X inactivation”, Emmetten, Switzerland

3rd Dutch Stem Cell meeting, Utrecht

Erasmus Stem Cell Institute Opening Symposium, Rotterdam (poster contribution)

Stem Cell Symposium, “Stem Cell, Development and Regulation”, Amsterdam

8th Annual Meeting of the International Society for Stem Cell Research (ISSCR), San Francisco, United States of America

2011

6th International meeting, Stem Cell Network North-Rhine-Westphalia, Essen, Germany (Poster contribution)

International Symposium “Chromatin Changes in Differentiation and Malignancies”, Giessen, Germany (Poster contribution)

Teaching activities:

Junior Science Program:

Supervision lab rotation Junior Science program, October 2009

Supervision Profielwerkstuk Junior Science student 2010-2011

Teaching Medical Curriculum Erasmus MC:

Course “Development of internal and external genitalia”, November 2008

Course “Development of internal and external genitalia”, December 2009

Course “Early embryos, cloning and transgenesis” November 2010

Course Embryology 2nd year Medicine/Master of Science programme, January 2010

Master of Science in Molecular Medicine Program:

Supervision lab rotation MSc Mol Med student, February 2009

Supervision lab rotation MSc Mol Med student, October 2009

Supervision lab rotation MSc Mol Med student, January 2010

Supervision MSc Thesis Selma van Staveren, “Initiation of X chromosome Inactivation” 2010-2011

Supervision MSc Thesis Elvira Myronova, 2011-2012

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Finally, after a long journey, I'm arriving at the finish. For sure it has been a long journey, in which I travelled approximately 516780 km or 12.9 times around the earth; that's the distance you travel if you live in Zeeland and study and work in Rotterdam, during 5 years of study and 4 years of PhD. If I just look back at the last 4 years, many things have happened during that period, here just a small selection of the "really important" things:

The layout of the faculty changed completely and we also changed a bit the view on X chromosome inactivation; unnecessary wars have unfortunately been characterizing the politics in the world, we arrived in a economical crisis, the 7 billions human being has been born, a mad person with colored hair tried to come up for the rights of *Henk* and *Ingrid* in Dutch politics, but instead yet another round of new elections is upcoming. A nuclear catastrophe was necessary to finally result in a different view on nuclear energy. And two Olympic Games and several football tournaments passed by, with the only constant being that in every important football match I look at, my favorite team does not win.

So a lot has happened and changed in the world, and for sure also I changed due to the many positive but sometimes also negative events which happened during this journey. I came across many people during that exciting trip, and to all of them I would like to say thank you!

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Tahsin Stefan Barakat, August 2012