Ten-year neonatal hepatitis B vaccination program, the Netherlands, 1982–1992: protective efficacy and long-term immunogenicity


From 1982 to 1989, 705 infants born to HBsAg-positive mothers entered the Dutch neonatal hepatitis B vaccination program and received passive-active hepatitis B immunization in three randomized controlled trials testing variations in time of starting active vaccination, dose and type of vaccine, and number of hepatitis B immunoglobulin (HBIg) injections. A meta-analysis of individual patient data of the three randomized trials was performed to determine which independent host and vaccination related factors influence protective efficacy and long-term immunogenicity, and to assess whether hepatitis B vaccination concomitant with standard DKTP vaccination provides optimal protection. Statistical methodology included multivariate logistic regression analysis. Eight infants (1.1%), all born to HBeAg-positive mothers, became HBsAg carriers within the first year of life. The protective efficacy rate (PER) of passive-active immunization at 12 months follow-up was 92% for the total group of children from 114 HBeAg-positive mothers with no significant differences between children starting active immunization at birth or at 3 months of age, between infants starting at 3 months of age receiving one or two doses of HBIg or between those receiving plasma derived or recombinant vaccine. The only factor that affected the PER significantly was the level of maternal HBV DNA; PER was 100% if maternal HBV DNA was < 15 pg ml\(^{-1}\) and 68% for HBV DNA levels > 15 pg ml\(^{-1}\). After 5 years of follow-up, the group that started active immunization at birth had significantly more infants with loss of seroprotection (anti-HBs levels < 10 IU l\(^{-1}\), 15%) than the corresponding group starting at 3 months of age (anti-HBs < 10 IU l\(^{-1}\), 2%). One of 35 children with loss of seroprotection at 2 years became a HBsAg carrier in the fifth year of follow-up. This meta-analysis shows that the protective efficacy of passive-active hepatitis B vaccination is mainly influenced by maternal HBV DNA levels, and independent of the time of starting active vaccination at birth or at 3 months of age; long-term immunity was enhanced by starting active vaccination concomitant with DKTP vaccination. These findings allow incorporation of hepatitis B vaccine into the standard infant immunization programs for countries with a passive-active immunization strategy for the control of hepatitis B. Additional measures are needed to protect neonates of highly viremic women. © 1997 Elsevier Science Ltd.

Keywords: hepatitis B, vaccine plasma, hepatitis B immunoglobulin, recombinant, neonates, meta-analysis, randomized controlled trials.

*Department of Internal Medicine II, University Hospital Dijkzigt, Rotterdam, Netherlands, and the Dutch study group Prevention Neonatal Hepatitis B. †General Practice, Borne, Netherlands. ‡Department of Virology, Erasmus University, Rotterdam, Netherlands. §Department of Epidemiology and Biostatistics, Erasmus University, Rotterdam, Netherlands. ||Wilhelmina Children's Hospital, University Hospital Utrecht, Utrecht, Netherlands. ¶Department of Hematology, University Hospital Utrecht, Utrecht, Netherlands. ‡‡Department of Pediatrics, St. Sophia Hospital, Zwolle, Netherlands. *Department of Pediatrics, Ruwaard van Putten Hospital, Spijkenisse, Netherlands. **Department of Hepatogastroenterology (section Hepatology), University Hospital Dijkzigt, Dr. Molewaterplein 40, 3015 GD Rotterdam, Netherlands. Tel.: 31-10-4635942; fax: 31-10-4365961. (Received 24 July 1996; revised version received 12 February 1997; accepted 4 March 1997)
When hepatitis B vaccine was licensed in the Netherlands in 1982, a program was started to screen mothers for HBsAg positivity and to immunize their offspring. In the first randomized trial a passive–active immunization schedule starting at birth was compared with a schedule with active immunization starting at 3 months of age, concomitant with diphtheria–tetanus–pertussis and poliomyelitis (DTPP) vaccination. The authors hypothesized that efficacy of delayed active immunization would be similar, but long-term immunogenicity superior to immunization starting at birth. Such a result would allow incorporation of active hepatitis B immunization in the standard infant immunization program, which was thought to be advantageous with respect to logistics and costs. Our preliminary results supported this hypothesis.

In 1984 a second randomized trial was started to compare the short- and long-term immunogenicity of the standard vaccine dose with a pediatric dose. With the introduction of recombinant vaccine in 1987, a third randomized trial was performed to confirm the efficacy of delayed vaccination with the recombinant vaccine; this trial also allowed testing the need for supplementary hepatitis B immunoglobulins (HBIG) at the start of delayed active immunization.

Our recent findings, also observed by others, that vaccination failure is also related to high maternal HBV DNA levels made it necessary to re-evaluate the effects of the components of the various schedules: time of starting active immunization, types and doses of vaccine and HBIG used, in relation to maternal HBV DNA levels.

This final report of the Dutch program for prevention of perinatal hepatitis B describes the protective efficacy and long-term immunogenicity of passive–active hepatitis B immunization over a period of 10 years.

SUBJECTS AND METHODS

Hepatitis B screening

The study was started in July 1982 in three large city hospitals in Rotterdam and Utrecht and one large rural area, Twente-Gelderse Achterhoek. Blood samples obtained from all pregnant women at their first visit to the prenatal clinic of the participating centers were tested for the presence of HBsAg. If HBsAg positivity was confirmed, randomization to one of the immunization schedules took place after informed consent was obtained from the mother. In the two participating hospitals in Rotterdam, the HBsAg status of expectant mothers was checked soon after arrival in the delivery room. Whenever prenatal HBsAg test results were missing, blood was obtained and tested the next morning with a rapid hemagglutination HBsAg test. If the rapid HBsAg test was positive, the mother was asked for informed consent and the baby was randomized and included in the immunization trial. All pregnant HBsAg-positive women were also tested for the presence of HBeAg. In December 1992, maternal HBV DNA levels were quantified retrospectively in the available stored serum samples positive for HBeAg.

Subjects and immunization schedules

From July 1982 to March 1984, 238 eligible babies were randomly allocated to one of the two initial plasma vaccine immunization schedules (Table I, groups I–II); from April 1984 to December 1987, 257 babies entered the plasma vaccine immunogenicity study (Table I, groups III–IV); from January 1988 to October 1989, another 210 eligible babies were allocated to one of the two recombinant vaccine immunization schedules (Table I, groups V–VI).

All infants received HBIG (200–300 IU, Central Laboratory of the Netherlands Red Cross Blood Transfusion Service, Amsterdam) i.m. within 2 h of birth by the physician or midwife in charge of the delivery. For active immunization, infants were referred to a pediatrician. Infants received plasma vaccine (10 and 5 μg HBvax, Merck Sharp & Dohme, West Point, USA) or recombinant vaccine (20 μg Engerix-B, SmithKline Beecham, Rixensart, Belgium). Nine infants (one in group II, three in group III and five in group IV) with an anti-HBs level ≤10 IU/l at 12 months of age and a negative test for HBsAg received an additional course of plasma vaccine or recombinant vaccine in their second year of life.

Serological assays and laboratory methods

Blood samples were taken from the infants at birth (cord blood) and in groups I and II at months 3, 6, 11, 12 and then yearly until 9 years of age; in groups III and IV at months 3, 6, 12 and then yearly until 5 years of age; in groups V and VI at months 3, 4, 6, 11, 12 and at 2 years of age. All serum samples were tested for anti-HBs and anti-HBe; HBsAg was assayed in all samples with anti-HBs below 100 IU/l.

Table 1 Immunization schedules of study groups

<table>
<thead>
<tr>
<th>Group</th>
<th>Entry period</th>
<th>Total (mother HBeAg +)</th>
<th>Evaluated for immunogenicity</th>
<th>Evaluated for efficacy (12 months)</th>
<th>HBlg</th>
<th>Vaccine</th>
<th>Schedule</th>
<th>Months after birth</th>
<th>Type/dose</th>
<th>Months after birth</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>1982–1984</td>
<td>117 (38)</td>
<td>110 (35)</td>
<td>103 (37)</td>
<td>0/200</td>
<td>plasma 10 μg</td>
<td>0, 1, 2, 11</td>
<td>3.4.5.6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>II</td>
<td>1982–1984</td>
<td>121 (42)</td>
<td>109 (37)</td>
<td>105 (41)</td>
<td>0/200</td>
<td>plasma 10 μg</td>
<td>0, 1.6</td>
<td>3.4.5.6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>III</td>
<td>1984–1987</td>
<td>133 (29)</td>
<td>128 (2)</td>
<td>127 (3)</td>
<td>0/200</td>
<td>plasma 10 μg</td>
<td>0, 1.6</td>
<td>3.4.5.6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IV</td>
<td>1984–1987</td>
<td>124 (4)</td>
<td>122 (4)</td>
<td>115 (2)</td>
<td>0/200</td>
<td>plasma 5 μg</td>
<td>0, 1, 6</td>
<td>3.4.5.6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>V</td>
<td>1988–1989</td>
<td>112 (14)</td>
<td>102 (13)</td>
<td>98 (14)</td>
<td>0/200</td>
<td>recombinant 20 μg</td>
<td>3.4.5.6</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>VI</td>
<td>1988–1989</td>
<td>98 (17)</td>
<td>93 (17)</td>
<td>83 (17)</td>
<td>0/200</td>
<td>recombinant 20 μg</td>
<td>3.4.5.6</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>1982–1989</td>
<td>705 (118)</td>
<td>664 (108)</td>
<td>631 (114)</td>
<td>0/200</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Hepatitis B immunoglobulin, Central Laboratory of the Dutch Red Cross Blood Transfusion Service, The Netherlands; †infants of HBeAg positive mothers entered in group I and II until December 1987; ‡between parentheses number of infants with HBeAg positive mothers; HBvax, Merck Sharp & Dohme, Westport, USA; Engerix-B, SmithKline Beecham Biologicals, Rixensart, Belgium;
HBsAg, HBeAg, anti-HBc and anti-HBs were assessed using a commercial radioimmunoassay kit (Abbott Laboratories, Chicago, IL, USA). HBV DNA was measured quantitatively by a solution hybridization assay (HBV DNA, Abbott Laboratories).

**Definition of HBV infection**

A HBV carrier state was defined as being HBsAg positive for more than 6 months. Transient HBV infection was characterized by the presence of HBsAg in serum for less than 6 months. Inapparent HBV infection was defined as anti-HBc positivity without HBsAg on two or more occasions after 12 months.

**Statistical analysis**

Available data were analysed according to the intention-to-treat principle. Separate per-protocol analysis, i.e. analysis of data of children who received vaccinations according to protocol, was also performed. The results of these analyses did not differ significantly. Therefore, the authors report on the protective efficacy using outcomes of the intention-to-treat analysis; the results on the immunogenicity are reported using the outcomes of the per-protocol analysis.

For analysis of factors affecting the protective efficacy rate, logistic regression analysis was applied. Confidence limits for odds ratio were calculated using the statistical software package ‘STATXACT’. Differences in percentages were analysed by the two-sample Wilcoxon rank-sum test. The limit for significance was set to 0.05 (two-sided). In case of evaluations at various timepoints, the limit of significance was set according to Bonferroni’s principle to allow for the multiplicity of statistical tests. Geometric mean titers (GMT) were calculated only for those infants who had anti-HBs ≥10 IU l⁻¹.

**Ethics**

The study was approved by the local Medical Ethics Committee of the participating centres.

**RESULTS**

**Follow-up**

From July 1982 until October 1989, 705 infants of HBsAg-positive mothers were randomized into three controlled trials. Sixteen infants were withdrawn after informed consent but before vaccination, 12 infants received at least one vaccination, but no serum sample was available after month 0. In total, 677 infants received full courses of passive–active immunization according to six schedules (Table 1). Thirteen infants received vaccinations but not according to protocol (one in group I, four in group II, one in group III, one in group IV, three in group V and three in group VI). Thus, 664 infants received passive–active immunization according to protocol. Serum samples were available from 590 children at 1 year of follow-up, 546 after 2 years, 262 after 5 years and 126 after 8 years. Incomplete data from these children were primarily due to secondary refusal of the parents, related to the frequency of blood sampling (up to ten samples) during the study, and migration.

**Protective efficacy of perinatal HBV immunization**

At month 12, HBsAg positivity was found in eight of 590 infants. Of 33 infants no serum sample was available at month 12 but they were HBsAg negative thereafter, so the assumption was made that there was no HBsAg positivity at month 12. From eight of the 13 infants who received a vaccination schedule not according to protocol, serum samples were available and found to be HBsAg negative at month 12 or thereafter. Of 631 (i.e. 590+33+8) infants analysed, eight (1.3%) were found to be HBsAg-positive during the first year of life. All HBsAg-positive children were born to HBeAg-positive mothers.

One infant of a HBeAg-negative mother became HBsAg positive at the age of 5 years (Figure 1); it had a peak anti-HBs level between 10 and 100 IU l⁻¹ after vaccination, but was anti-HBs negative from 2 years onwards. This child and all infants found to be HBsAg-positive in the first year became hepatitis B carriers.

Inapparent HBV infection (anti-HBc positivity with negative HBsAg tests on two or more occasions after month 12) was observed in eight infants, all born to HBeAg-positive mothers.

At month 12, the protective efficacy rate (PER) of passive–active immunization for the 114 infants of HBsAg- and HBeAg-negative mothers was 90% (Table 2). No significant differences were found between the PER for infants starting active vaccination at birth and those starting at 3 months; between infants starting at 3 months and receiving one or two doses of HBlg and between infants receiving plasma vaccine or recombinant vaccine.

In 72 of the 114 HBeAg-positive mothers, residual frozen serum was available for quantitative HBV DNA assessment in 1992. Table 3 shows the relation of maternal HBV DNA levels and the number of infants who became HBV carriers. The PER at month 12 for the two groups with HBV DNA levels <150 pg ml⁻¹ were 100% and significantly higher than the PER (68%) for the group with maternal HBV DNA levels ≥150 pg ml⁻¹ (P = 0.009).

[Figure 1] Chronic HBV infection after 5 years of follow-up in one infant, born to an HBeAg-negative mother, despite a response to neonatal passive–active hepatitis B immunization (group II)
Table 2 Protective efficacy of passive–active hepatitis B immunization, at month 12 in infants of HBeAg-positive mothers, according to different vaccination schedules

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of infants</th>
<th>HBsAg positive (%)</th>
<th>HBsAg positivity difference in % (95% CI)</th>
<th>PER12 months (%)</th>
<th>Expected % of infants with HBV infections without prophylaxis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Vaccine starting month 0</td>
<td>42</td>
<td>3 (7.1)</td>
<td>0.2 (--9.5, +9.9)</td>
<td>92</td>
<td>89</td>
</tr>
<tr>
<td>Vaccine starting month 3</td>
<td>72</td>
<td>5 (6.9)</td>
<td></td>
<td>92</td>
<td>89</td>
</tr>
<tr>
<td>HBlg month 0</td>
<td>56</td>
<td>4 (7.1)</td>
<td>0.2 (--9.2, +9.2)</td>
<td>92</td>
<td>89</td>
</tr>
<tr>
<td>HBlg months 0.3</td>
<td>58</td>
<td>4 (6.9)</td>
<td></td>
<td>92</td>
<td>89</td>
</tr>
<tr>
<td>Plasma vaccine</td>
<td>83</td>
<td>6 (7.2)</td>
<td>0.7 (--9.6, +11.0)</td>
<td>92</td>
<td>89</td>
</tr>
<tr>
<td>Recombinant vaccine</td>
<td>31</td>
<td>2 (6.5)</td>
<td></td>
<td>92</td>
<td>90</td>
</tr>
<tr>
<td>Total</td>
<td>114</td>
<td>8 (7.0)</td>
<td></td>
<td>92</td>
<td>90</td>
</tr>
</tbody>
</table>

*PER = (expected number of HBV infections without immunoprophylaxis minus measured number of HBV infections in immunization group) divided by the expected number of HBV infections without immunoprophylaxis x 100%; the expected number of HBV infections without immunoprophylaxis for infants from HBeAg positive mothers is usually estimated at 90%22–23, however, PER was also calculated for 67% expected HBV infections without immunoprophylaxis, in view of the finding that one-third of infants of HBeAg-positive mothers had HBV DNA < 5 pg ml−1 and are therefore unlikely to transmit hepatitis B to their infants6.

Table 3 Protective efficacy rate (PER) of passive–active hepatitis B immunization at month 12 according to maternal HBV DNA levels

<table>
<thead>
<tr>
<th>HBV DNA (pg ml−1)</th>
<th>No. of infants</th>
<th>HBsAg positive (%)</th>
<th>PER12 months (%)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 6</td>
<td>24</td>
<td>0 (0)</td>
<td>n.a.</td>
<td>0.000*</td>
</tr>
<tr>
<td>7–150</td>
<td>24</td>
<td>0 (0)</td>
<td>100</td>
<td></td>
</tr>
<tr>
<td>&gt; 150</td>
<td>24</td>
<td>7 (29)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Fisher’s exact test between number of HBsAg-positive infants in group with HBV DNA > 150 pg ml−1 and groups with HBV DNA of 7–150 and < 6 pg ml−1; owing to small numbers of infants, the exact 95% CI of the ratio of the odds of HBsAg positivity in the group of infants with HBV DNA > 150 pg ml−1 (odds 0/48 versus the corresponding odds 7/24 in the group with HBV DNA > 150 pg ml−1 was calculated (this 95% CI ranged from 0 to 0.22); not applicable, in view of the finding that infants of HBeAg-positive mothers with HBV DNA < 5 pg ml−1 are unlikely to transmit hepatitis B to their infants6.

DISCUSSION.

In this meta-analysis of individual patient data of three randomized controlled trials, the PER against hepatitis B infection for infants of HBeAg-positive carrier mothers was 90% at 12 months of age; for infants of HBeAg-negative mothers it was 100%. These rates are comparable to those found in other passive–active immunization studies, with either plasma-derived or recombinant vaccine4,5. During follow-up beyond 1 year, the PER remained above 90%, with the protective efficacy rate decreasing to 89% at 3 years and 83% at 5 years. These results suggest that hepatitis B vaccine administered from birth onwards provides long-term protection against HBV infection in infants of HBeAg-positive mothers.

Figure 2 shows the maternal HBV DNA levels for HBV infected infants and non-infected infants. The median maternal HBV DNA level of the HBV carrier infants and that of the inapparently infected infants were ten times higher (~350 pg ml−1) than the median maternal HBV DNA level (31 pg ml−1) of the infants without HBV infection (P = 0.001 and 0.03, respectively).

When data on time of starting active vaccination, number of doses of HBIG and type of vaccine were analysed by multivariate logistic regression, no significant differences were found (odds ratio confidence intervals were from 0.001 to > 10). When the analysis was expanded with HBV DNA data, both the factor HBV DNA as well as HBV DNA > 150 pg ml−1 were significantly associated with the PER.

Long-term immunogenicity

Assuming a minimal risk for hepatitis B infection in vaccinees with anti-HBs levels > 10 IU l−1, and a potential risk with anti-HBs levels < 10 IU l−1, the authors calculated the percentages of infants with anti-HBs < 10 IU l−1 in the different immunization groups (Figure 3). At the age of 5 years, the group that started at 3 months of age with plasma vaccine (group II) had a significantly lower percentage (2%, 95% CI 0–6%) of children with anti-HBs < 10 IU l−1 than group I (14%, 95% CI 6–23%) that started immunization at birth (Fisher’s exact test, P = 0.02). The percentage of infants with anti-HBs < 10 IU l−1 in group II never exceeded 5% during the 5 year follow-up, whereas the corresponding percentage in the other immunization groups increased to > 15% during follow-up.

Table 4 shows the GMT of anti-HBs (anti-HBs ≥ 10 IU l−1) in the different immunization groups during follow-up. GMT of group II (10 µg plasma vaccine administration from 3 months of age onwards) was approximately double that of group I (starting at birth with 10 µg plasma vaccine). In the groups with different dosages of vaccine, the GMT at month 12 in group III with three doses of 10 µg plasma vaccine administered from birth onwards, was approximately double that of group IV, which received the same schedule but only 5 µg of plasma vaccine. However, this difference was not significant after 36 months of follow-up. Comparison of GMT values of groups given plasma vaccine or recombinant vaccine from 3 months onwards showed that the group receiving plasma vaccine (group II) had an approximately one and half times higher GMT than the corresponding group receiving recombinant vaccine (group VI) (P < 0.001 at all times measured).

Table 3 Protective efficacy rate (PER) of passive–active hepatitis B immunization at month 12 according to maternal HBV DNA levels

**Vaccine 1997 Volume 15 Number 15 1627**
year, extending to 9 years, one vaccinated infant of an HBeAg-negative mother became positive for HBsAg.

The timing of the initial vaccine injection, the type of the vaccine and the number of doses of HBIg had no effect on the PERs. These results confirm earlier findings. Beasley et al. reported that, with HBIg, the median HBV DNA levels of seven chronic HBV infected infants, four inapparent HBV infected infants and of 61 non-infected infants. Cut-off level of HBV DNA assay is 1.7 pg ml −1.

Figure 2 Maternal HBV DNA levels of seven chronic HBV infected infants, four inapparent HBV infected infants and of 61 non-infected infants. Cut-off level of HBV DNA assay is 1.7 pg ml −1.

Figure 3 Percentages of infants with anti-HBs > 10 IU l −1 in the six immunization groups. Bars indicate standard errors. Results at 2 years are similar for groups starting vaccination at birth or at 3 months of age. Long-term follow-up suggests excellent persistence of protective humoral immunity for infants starting vaccination at 3 months of age.

Table 4 GMT anti-HBs (anti-HBs ≥ 10 IU l −1) in the six immunization groups during follow-up. Values in parentheses are the 95% CI.

<table>
<thead>
<tr>
<th>Group</th>
<th>Month 12</th>
<th>Month 24</th>
<th>Month 36</th>
<th>Month 48</th>
<th>Month 60</th>
</tr>
</thead>
<tbody>
<tr>
<td>I (0,1,2,11; 10 μg pl.)</td>
<td>8730(6238–12217)</td>
<td>737(533–1019)</td>
<td>353(248–502)</td>
<td>207(145–296)</td>
<td>137(92–203)</td>
</tr>
<tr>
<td>II (3,4,5,11; 10 μg pl.)</td>
<td>15739(11738–21104)</td>
<td>1728(1284–2325)</td>
<td>820(596–1129)</td>
<td>484(356–231)</td>
<td>331(231–473)</td>
</tr>
<tr>
<td>III (0,1,6; 10 pg pl.)</td>
<td>1142(849–1537)</td>
<td>331(245–447)</td>
<td>202(151–271)</td>
<td>138(100–190)</td>
<td>100(70–143)</td>
</tr>
<tr>
<td>IV (0,1,6; 5 pg pl.)</td>
<td>608(438–846)</td>
<td>203(146–283)</td>
<td>163(116–228)</td>
<td>114(87–166)</td>
<td>75(52–109)</td>
</tr>
</tbody>
</table>

*Between groups I and II; **between groups III and IV; †between groups V and VI.
in a low-endemic region like northwestern Europe, but the extent of the problem appears limited.

Long-term immunogenicity was significantly higher in the group receiving late active immunization than in the group starting directly after birth. At the age of 5 years, the group with delayed active immunization had a significantly lower percentage (2%) of children with anti-HBs <10 IU l−1 than groups in which immunization was started at birth (15−25%). This finding is in agreement with others who found an enhancement of the immune response if the infant was older at the time of the initial injection, probably related to a more mature immune system 3,4,9. The implications of these findings are at present unclear. If protection against hepatitis B infection depends on the degree of immunologic priming reflected by the persistence of antibody, then a strong argument could be made for adoption of schedules that maximize anti-HBs levels.

In the present study, there was a total follow-up of 186 person-years in 71 infants with anti-HBs <10 IU l−1. One infant, born to a HBsAg-negative mother, with an initial response between 10 and 100 IU l−1 became HBsAg carrier after 4 years of follow-up without detectable anti-HBs. In other studies, no HBsAg positivity after 5−9 years of follow-up was found for infants with initial anti-HBs ≥10 IU l−1, whether the infants lost their anti-HBs or not 3,4,9,10. In long-term follow-up studies of immunized adults, no persistent HBsAg positivity was detected in persons with an initial anti-HBs response >10 SRI, but transient HBsAg positivity and/or anti-HBc positivity was detected in this group, <1% in 100 person-years exposed 9. At present, until more long-term follow-up studies are available, it seems advisable to aim for an initial anti-HBs response of 100 IU l−1 or more for the prevention of clinically important forms of hepatitis B.

The confirmation of an old finding that there was no effect of the time of starting active immunization on PER facilitates the incorporation of hepatitis B vaccine into the existing Expanded Programme on Immunization (EPI) by allowing active hepatitis B vaccination concomitant with DTP immunization. The number of visits can then be reduced as well as the number of injections if a multivalent vaccine becomes available.

Acknowledgements

The following participants of the Dutch study group Prevention Neonatal Hepatitis B are acknowledged for their contribution: J.W. Wladimiroff, Department of Obstetrics, Academic Hospital Dijkzigt, Rotterdam; A.S.M. Nuijten, State School of Training for Obstetricians, Rotterdam; J.R.J. Bänffer, Regional Public Health Laboratory, Rotterdam; M.J. Botman, Regional Laboratory of Pathology and Microbiology, Enschede; G.C.M.L. Christiaens, Department of Gynecology and Obstetrics, Academic Hospital, Utrecht; W. Baerts, Department of Pediatrics, St. Sophia Hospital, Zwolle. The study group is grateful to the Steering Committee formed by Professor Dr. J. Huismans, Professor Dr J.W. Stoop, Dr H. Bijkerk and the late Professor S. Kragman. The authors are grateful to M. Bakker-Bendik, Th. van der Laar-Kokx, H. Ribbers-Smeenk, H.C. van Dijk and M. Voogd-Schotanus for their outstanding help in this study for several years. Special thanks go to Y.A.M. Weber, J. Kruining and H.G.M. Niesters for performing the laboratory tests, J. Boot for technical assistance; P.N. Lelie for making available the maternal HBV DNA assays of the Hong Kong neonatal hepatitis B vaccination program; and B.E. Hansen for performing multivariate analysis. This study was supported by a grant from the Pravecintefonds, Netherlands (no. 28-760-2).

References

Efficacy and immunogenicity neonatal hepatitis B vaccination: R. del Canho et al.


