

AN *IN VITRO* MODEL OF UROTHELIAL REGENERATION: EFFECTS OF GROWTH FACTORS AND EXTRACELLULAR MATRIX PROTEINS

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SUMMARY

Although the cellular turnover of resting urothelium is very low, its regenerative capacity is known to be outstanding. In organotypic mouse urothelial cultures closely mimicking the differentiation and multilayering of normal urothelium, we examined the cell biological mechanisms underlying urothelial regeneration and the specific role of growth factors and several extracellular matrix (ECM) components. Exposure to epidermal growth factor (EGF) and acidic fibroblast growth factor (aFGF) and culture on laminin resulted in enhanced expansion of the urothelium. Microscopy and assessment of proliferative activity revealed that enhanced urothelial expansion due to EGF could be attributed to increased proliferative activity and an increase in cell numbers, whereas aFGF-stimulated expansion must be considered the consequence of increased cellularity and migration. Laminin-enhanced urothelial expansion was shown to be the result of spreading of the entire urothelial organotypic culture. This was associated with a considerable decrease in the number of cell layers. A synergistic effect of growth factors and laminin was not found. This organotypic urothelial cell culture model seems to be very useful in studying strategies to improve urothelial regeneration.

KEY WORDS—Regeneration, bladder explant, growth factors, extracellular matrix components, migration, proliferation, cell spreading.

INTRODUCTION

Human bladder cancer is a common cancer in Western society, with considerable morbidity due to its high recurrence rate after local resection of superficial transitional cell carcinomas.¹ Clinical and experimental data indicate that a substantial proportion of tumour recurrence is the consequence of seeding of tumour cells on traumatized and denuded bladder mucosa.² It can be envisaged that variations in the regeneration rate of traumatized urothelium can influence the incidence of tumour recurrence. The regenerative potential of urothelium after denudation of the bladder mucosa has been studied in several species, including man and mouse, and is known to be outstanding.^{3,4} within 48 h of the

urothelium being stripped, the whole mouse bladder is again covered with urothelium.

In vitro studies on keratinocytes, thyrocytes, and endothelial cells have suggested that growth factors such as epidermal growth factor (EGF), fibroblast growth factor (FGF), and transforming growth factor β (TGF β) can influence the migration of these cell types.^{5–9} For the NBT II rat bladder carcinoma cell line, it has been shown that exposure to acid FGF (aFGF) leads to increased migration.¹⁰

Basement membranes of epithelium, including urothelium, are composed of several extracellular matrix (ECM) components such as laminin, fibronectin (FN), and collagen type IV (COL IV).^{11,12} ECM–cytoskeletal interactions are also reported to be crucial in the regulation of

regeneration and wound healing. For example, FN appears to enhance the migration of corneal epithelial cells following wounding¹³ and laminin stimulates the migration of olfactory neuroepithelial cells.¹⁴ Since data on growth factor and ECM modulation of urothelial regeneration are scarce, we studied these aspects in an *in vitro* model using organotypic cultures of murine urothelium. For that purpose, explant cultures of adult mouse bladder mucosa on transparent porous membranes, uncoated or coated with ECM, were used as an *in vitro* model for regenerating urothelium. The culture on transparent membranes allowed daily measurements of the circumference of the explant outgrowths. Previous studies from our laboratory on this culture system had revealed that the outgrowths of the primary murine bladder explants closely mimicked the *in vivo* situation, since multilayering and maturation to umbrella cells occurred.^{15,16} Our approach contrasts with most previous *in vitro* studies in which the migration or cell spreading of single cells or cell lines was investigated.

MATERIALS AND METHODS

Bladder explant culture on porous membranes

Six- to 8-week-old female C3H/HE mice were killed and the urinary bladder was removed aseptically. The mucosa was stripped from the underlying muscle layer, cut into two pieces, and each half was placed on a transparent porous membrane (Falcon cell-culture insert, cyclopore membrane with a diameter of 25 mm; pore size 0.45 μ m). The lamina propria of the explant was always in contact with the membrane, as described previously.¹⁵ The medium of the porous membrane cultures was replaced every other day with standard medium [a 1:1 mixture of Dulbecco's modified Eagle's medium (DMEM) and Ham's F10 medium with 10 per cent heat-inactivated fetal calf serum (FCS) and supplement as described previously¹⁵]. In some experiments, growth factors were added to the standard medium beginning at day 4 (as indicated in the text). The growth factors used were aFGF, 20 ng/ml, obtained from Boehringer Mannheim (Almere, The Netherlands); and EGF, 20 ng/ml, and TGF β , 1 ng/ml, obtained from Sigma (St. Louis, U.S.A.). This growth factor-supplemented medium was changed every other day.

The circumference of the bladder outgrowth was monitored daily through a light microscope. From

these overviews the expansion of the outgrowth was determined. The surface area as determined in mm² was related to the total surface of the porous membrane. When the outgrowth covered the whole membrane, 100 per cent outgrowth was reached. Only explant cultures exceeding an outgrowth of 8.5 per cent on day 4 were included in the study in order to obtain only actively growing cultures.

Two hours before termination of the culture, 40 μ g/ml bromodeoxyuridine (BrdU) was added.

Culture substrates

FN and laminin (Sigma, St. Louis, U.S.A.) were diluted in phosphate-buffered saline (PBS) to a concentration of 25 μ g/ml. COL IV (kindly donated by Dr J. P. M. Cleutjens of the Department of Pathology, University of Limburg, The Netherlands) and collagen type I (COL I) (Vitro-nogen 100; ICN Laboratories, Amsterdam, The Netherlands) were diluted in Ham's F10 to a concentration of 25 μ g/ml. Porous membranes were covered with 0.5 ml of a solution containing one of the ECM proteins and incubated overnight at 37°C in a humidified atmosphere. Coated membranes were then incubated with 2.5 mg of bovine serum albumin (BSA) for 30 min at 37°C to block non-specific binding sites on the membrane. Explants on BSA-coated membranes were regarded as negative controls, since BSA is not an adhesion molecule. Membranes were washed with H₂O and dried overnight. The next day, the bladder explants were placed on the membrane.

[³H]Thymidine incorporation

When the primary bladder culture covered an area of 60–65 per cent, the standard medium was replaced by serum-free DMEM with supplements containing 20 μ Ci of [³H]thymidine/ml (1.25 ml/membrane), in which the urothelium was cultured for another 2 h. Subsequently, the culture was rinsed with PBS with an excess of non-labelled thymidine. The incorporated radioactivity was counted in Ultima Gold (Packard, Groningen, The Netherlands) using an α,β -scintillation analyser (as described previously¹⁶). Before incorporated [³H]thymidine was counted, the membranes were immunohistochemically stained and counterstained with haematoxylin. To preserve an adequate morphology, trichloroacetic acid (TCA) treatment of the membranes was omitted. Control

experiments revealed no differences in the [^3H]thymidine incorporation values between membranes incubated and washed with 15 per cent TCA and the corresponding untreated membranes.

Reagents for immunohistochemistry

Monoclonal antibody RGE 53 (specific for cytokeratin 18) was obtained from Eurodiagnostica (Apeldoorn, The Netherlands). RGE 53 was employed to visualize the more mature urothelial cells. With this staining, cells with a typical morphology of umbrella cells can be distinguished more easily. The monoclonal antibody against BrdU was obtained from Dr Ramaekers (University of Maastricht, The Netherlands). For BrdU staining, the ethanol-fixed membranes were pre-treated with 1 M HCl following the procedure described in ref. 17. The membranes were incubated with the primary antibody overnight at 4°C. Subsequently, an indirect conjugated peroxidase method was applied. A goat anti-mouse immunoglobulin (Ig)-peroxidase conjugate obtained from DAKO (Glostrup, Denmark) was used. Diaminobenzidine (DAB) was used as a chromogen. The DAB solution was prepared by dissolving 75 mg of DAB tetrahydrochloride (Fluka, Oud-Beyerland, The Netherlands) in 100 ml of PBS, and 50 μl of 30 per cent hydrogen peroxidase was added as a substrate. For visualization of bound antibody, the membranes were incubated for 7 min with DAB solution and the sections were washed in distilled water. All membranes were counterstained with Mayer's haematoxylin.

To evaluate the number of immunostained urothelial cell layers more accurately, strokes of the immunostained porous membranes covered with urothelial cells were dipped in 5 per cent gelatin, dried overnight in air, and embedded in paraffin. Cross-sections (5 μm) perpendicular to the surface were made. The remaining parts of the membranes were dehydrated in alcohol and embedded in Euparal (Chroma-Gesellschaft Stuttgart, Germany).

Quantification of cell density and cell spreading

To quantify the cell density of the urothelium, the number of nuclei was counted in 14 random areas of 0.15 mm² in the outgrowth explant cultures used for [^3H]thymidine incorporation. At this time, the bladder outgrowth covered about 60–65 per cent of the membrane. The average \pm SEM of

three independent experiments with two samples each was taken.

In cultures covering approximately 100 per cent of the supportive membrane, the degree of cell spreading was evaluated by counting the number of basal nuclei on cross-sections over a distance of 5 mm. In each membrane, eight random positions at the peripheral part of the culture were chosen for evaluation.

RESULTS

Modulation of expansion of the primary bladder outgrowth

In explant cultures kept in standard medium containing 10 per cent FCS (control outgrowth), the urothelium immediately began to expand as a sheet of epithelial cells after attachment of the explants to the cyclopore membrane. The edges of the outgrowth of primary urothelium were well demarcated throughout the experimental period. This permitted easy measurement of the surface covered by the expanding urothelium. On day 4, the urothelial outgrowth covered more than 8.5 per cent of the membrane in 80 per cent of the cases. The whole membrane was covered in approximately 22 days (100 per cent urothelial outgrowth). Addition of EGF and aFGF to the standard medium containing 10 per cent FCS led to an increase in the expansion rate from day 7 onwards as compared with the controls ($P \leq 0.01$) (Fig. 1). Addition of TGF β resulted in significant inhibition of the expansion from day 7 onwards.

The expansion rate of primary urothelium was also examined on membranes coated with different ECM proteins. No growth factors were added to the standard medium. In 95 per cent of the cultures grown on ECM-coated membranes, the outgrowth was over 8.5 per cent on day 4. The attachment percentage (40 per cent) and the expansion rate of explants on BSA-coated membranes were significantly smaller than on uncoated membranes (Fig. 2). No significant difference in the expansion rate was observed between urothelial cultures grown on COL IV-, FN-, or COL I-coated supportive membranes and uncoated membranes (Fig. 2). It is probable that constituents of FCS provide an adhesive substrate allowing the expansion of urothelial cells on uncoated membranes. However, the proportion of successfully attached explants was higher

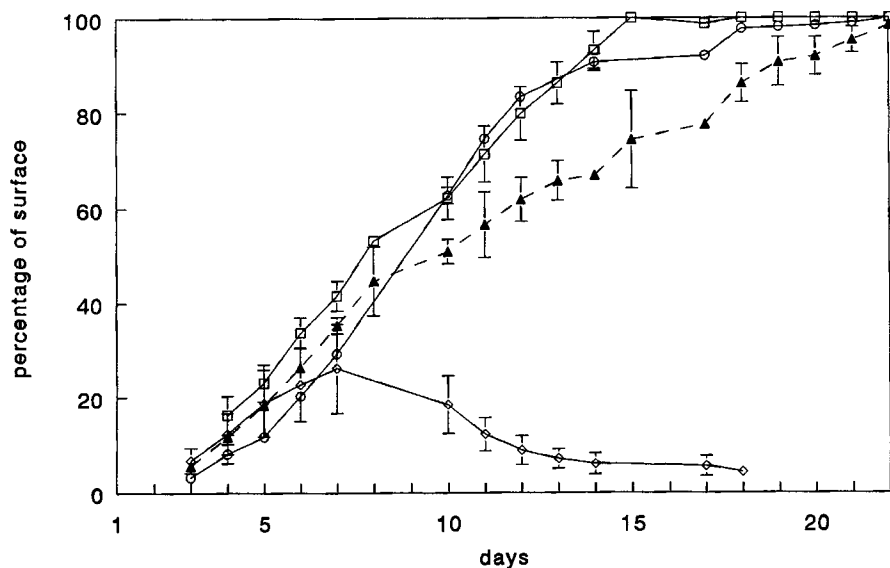


Fig. 1—Influence of growth factors on the expansion rate of explant cultures. ○, EGF; □, aFGF; ◇, TGFβ; ▲, control. Data are expressed as the mean \pm SEM of three independent experiments with three samples each

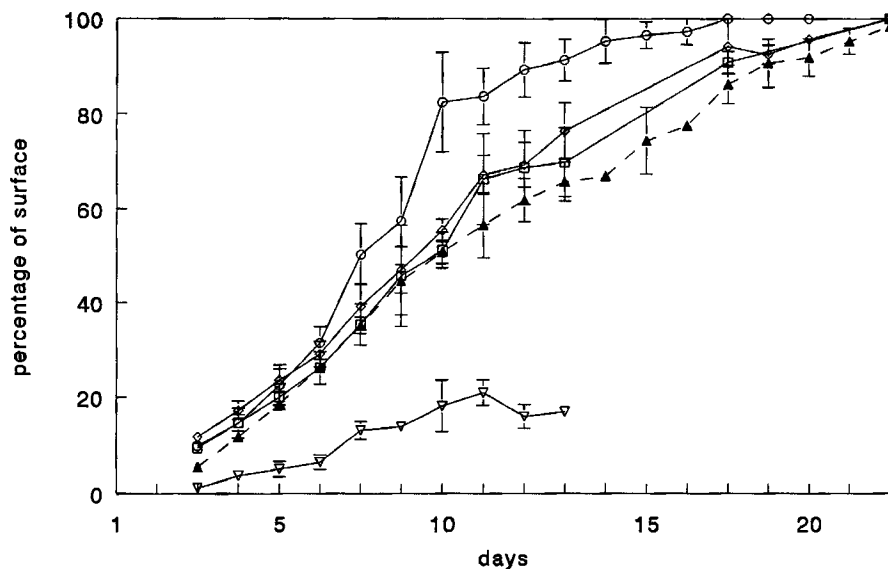


Fig. 2—Influence of ECM on the expansion rate of explant cultures. ◇, FN; ○, laminin; ▲, control; ▽, BSA; □, COL IV. Mean \pm SEM was calculated as explained in the legend of Fig. 1

on the ECM-coated membranes than on the uncoated membranes (i.e., 95 per cent vs. 80 per cent). The expansion of the urothelium cultured on laminin increased from day 7 onwards compared with the other cultures ($P \leq 0.01$).

Figure 3 illustrates the outgrowth of urothelial cultures grown on FN-, laminin-, COL IV-, or COL I-coated membranes in the presence or absence of EGF. Addition of EGF led to an increased expansion rate of urothelium on FN,

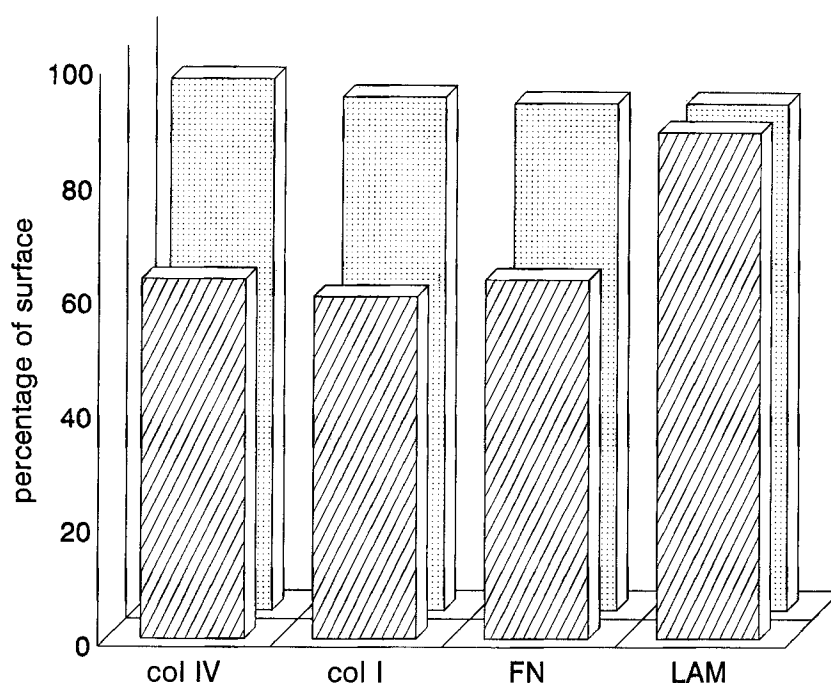


Fig. 3—Influence of EGF on the expansion rate of explant cultures grown on ECM. Cultured for 11 days. Hatched bars represent explants cultured in the absence of EGF and dotted bars represent explants cultured in the presence of EGF

COL IV, and COL I, but did not influence the expansion rate of urothelium grown on laminin.

Modulation of multilayering and cellularity in the primary bladder outgrowth

After termination of the cultures, immunohistochemistry with antibody RGE 53 was performed in order to visualize maturation to umbrella cells. Cells with the typical morphology of umbrella cells were identified in control explant cultures, in cultures exposed to aFGF and TGF β , and in urothelial outgrowths on all coated membranes without addition of growth factors. No RGE 53-positive cells with the typical morphology of umbrella cells were found in EGF-treated cultures. Furthermore, the exposure of urothelium on ECM-coated membranes to EGF was associated with the disappearance of RGE 53-positive cells with the morphology of umbrella cells (Figs 4a–4c).

Cross-sections of immunostained membranes (Figs 5a–5d) revealed that at near-confluence, EGF-treated cultures had an increased thickness of 3–5 cell layers, as opposed to 2–3 cell layers in

untreated cultures. The cellularity increased from 179 ± 35 cells/ 0.15 mm^2 in untreated cultures to 414 ± 14 cells/ 0.15 mm^2 in EGF-treated cultures. In aFGF-treated cultures, the cellularity also increased to 280 ± 52 cells and the cultures consisted of 2–3 cell layers with an upper layer of flattened RGE 53-positive umbrella cells. The TGF β -treated culture consisted of one single cell layer of very large RGE 53-positive cells. Its cellularity decreased to 67 ± 1 cell/ 0.15 mm^2 .

In cross-sections of the urothelial outgrowths on COL I-, COL IV-, or FN-coated membranes (Figs 5e–5h), 2–3 cell layers were observed. The top layer of these cultures consisted of flattened RGE 53-positive cells with the morphology of umbrella cells. Urothelium on laminin-coated membranes consisted of one cell layer of large flattened RGE 53-positive cells with a remarkably low nuclear density in the basal cell layer (Table I).

Cross-sections of immunostained coated membranes showed that in the presence of EGF the number of cell layers and the nuclear density of the basal cell layer (Table I) were increased in all cultures irrespective of the coating (Figs 5i and 5j).

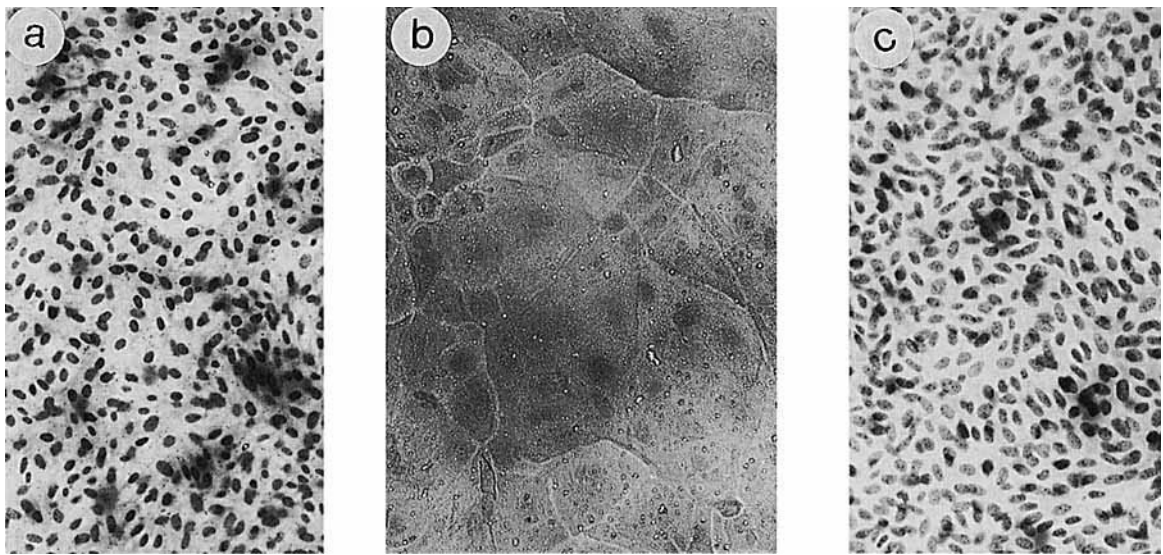


Fig. 4—Overviews of the bladder explant after 20 days of culture. All cultures were immunohistologically stained with RGE 53, an antibody against cytokeratin 18, and counterstained with haematoxylin. (a) Overview of an EGF-treated culture. Note the absence of RGE 53-positive cells. (b) Overview of a bladder explant cultured on COL IV. The cells have the typical morphological appearance of umbrella cells. (c) A bladder explant cultured on COL IV with addition of EGF. Note the absence of RGE 53-positive cells

Modulation of the proliferation of the primary bladder outgrowth

Table I shows that an increase in proliferative activity, as measured by [^3H]thymidine incorporation at the time that the culture reached 60–65 per cent outgrowth, was observed only in cultures exposed to EGF. Direct visualization of prolifer-

ating cells by immunostaining with anti-BrdU antibody revealed that these cells were evenly dispersed over the membrane. In contrast, 50 per cent of the aFGF-exposed cultures lacked these BrdU-positive cells in the periphery of the culture (data not shown).

Cultures on ECM-coated membranes generally had a lower proliferative activity than the control.

Table I—Proliferation, spreading of basal cells, and number of cell layers in organotypic urothelial cell cultures

	[^3H]thymidine incorporation, DPM \pm SD	No. of nuclei in the basal cell layer \pm SEM	No. of cell layers
Control	75 893 \pm 10 550	7.7 \pm 0.2	2–3
EGF	311 047 \pm 3790	15.3 \pm 0.6	3–5
aFGF	72 154 \pm 9495	9.3 \pm 0.8	2–3
COL IV	35 344 \pm 18 602	9.3 \pm 0.7	2–3
COL I	39 448 \pm 15 102	7.9 \pm 0.8	2–3
FN	32 577 \pm 14 187	7.3 \pm 0.6	2–3
Laminin	20 007 \pm 11 776	3.7 \pm 0.8	1
COL IV+EGF	179 624 \pm 36 791	13.1 \pm 2.8	3–4
COL I+EGF	139 756 \pm 3788	13.4 \pm 2.6	3–4
FN+EGF	103 313 \pm 21 266	13.8 \pm 3.1	3–4
Laminin+EGF	130 460 \pm 24 255	15.1 \pm 2.4	3–4

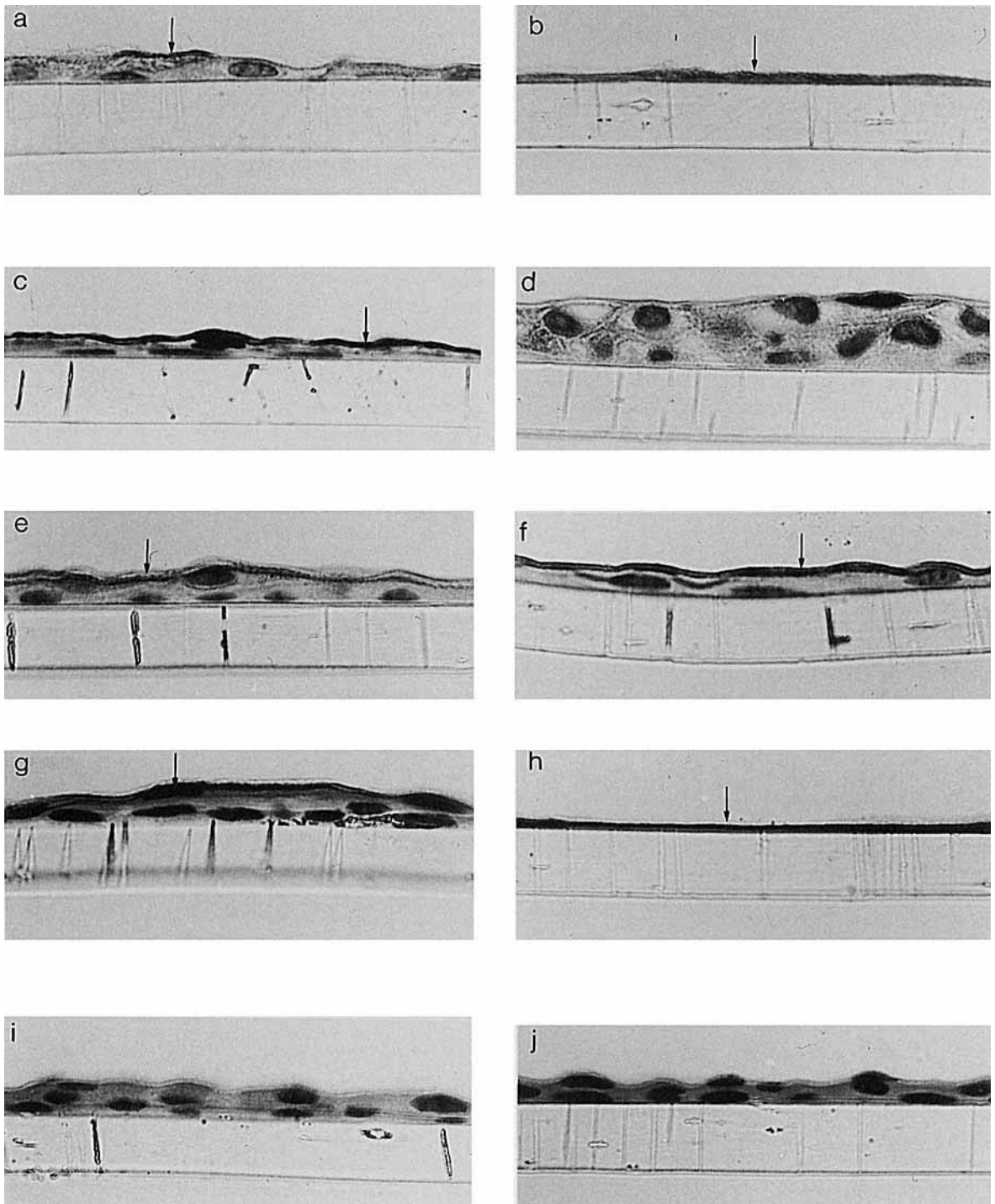


Fig. 5—Cross-sections of different urothelial cultures ($\times 1000$). Supportive membranes are immediately below the urothelium. Arrows indicate the RGE 53-positive cells. (a) Control; (b) $TGF\beta$ culture; (c) aFGF culture; (d) EGF culture; (e) grown on COL I; (f) grown on FN; (g) grown on COL IV; (h) grown on laminin; (i) grown on COL IV with addition of EGF; (j) grown on laminin with addition of EGF

No difference in [^3H]thymidine incorporation was found between the cultures when grown on different ECM-coated membranes. Direct visualization of BrdU-containing urothelial cells demonstrated proliferating cells in a peripheral concentric zone in 75 per cent of the cultures.

Cultures grown on ECM components in the presence of EGF demonstrated an increase in [^3H]thymidine incorporation compared with the primary urothelium grown on the ECM-coated membranes in the absence of EGF (Table I).

DISCUSSION

The high regenerative capacity of the bladder urothelium after acute damage is remarkable in view of the slow turnover of the normal transitional epithelium. Several mechanisms may account for the rapid re-epithelialization of the bladder surface. These could include extension of the surfaces of residual cells, migration of cells out of the urethra and ureteric orifices, and an increase in the proliferation of the residual urothelial cells.¹⁸

In this study we present evidence that each of these mechanisms was operating in our organotypic regeneration model, either separately or in combination. In this organotypic culture system we studied the effects of growth factors or ECM components on the expansion of a cohesive sheet of primary urothelial cells, instead of the effect of these factors on individual cells, as most studies do. We demonstrated an increase in the expansion rate of mouse bladder explants cultured in the presence of either EGF or aFGF, and when grown on laminin. Strikingly, the observed increase in expansion of the urothelium was accompanied by different morphological and functional features, dependent on the culture conditions applied.

EGF stimulation of the urothelial expansion rate was associated with an increased number of cell layers and a higher cellular density, as well as a loss of maturation into umbrella cells. The effect of EGF on expansion may thus be attributed to the observed strong increase in proliferative capacity. Previous studies on mouse and human epithelium also demonstrated an EGF-mediated stimulation of proliferation.¹⁹⁻²¹ Similarly, Schultz *et al.*²² showed that regeneration of corneal epithelial cells *in vivo* is regulated by EGF.

In urothelial cultures grown on laminin-coated membranes, the proliferative activity was decreased and at the peripheral parts of the culture

the urothelium consisted of only one flattened cell layer, with low nuclear density. The latter feature indicates enhanced cell spreading of this cohesive sheet of cells. Laminin-induced spreading of dispersed individual cells has been described for other cell types,²³⁻²⁵ but as yet not for organotypic cultures. It is most likely that the laminin-induced increase in urothelial expansion is due to this spreading of the urothelial cells.

To explain the mechanism by which aFGF induced an increased expansion rate is more complex. Addition of aFGF increased the cellularity of the urothelial culture, but at 60 per cent of confluence the proliferation rate of the aFGF-stimulated cultures was similar to controls. Furthermore, at the peripheral edge of the aFGF-stimulated cultures, proliferating (BrdU-positive) cells were largely absent. This suggests that proliferation does not greatly contribute to the aFGF-induced urothelial expansion. More likely, migration rather than stretching or proliferation of the urothelial cells contributes to the observed aFGF-enhanced expansion. This hypothesis is in line with the work of Valles *et al.*²⁶ in which the authors showed that aFGF can serve as a migration factor in a rat bladder carcinoma cell line. Similarly, migration of urothelial cells may be involved in the expansion of explants cultured on COL I, COL IV, or FN. In these cultures, proliferative activity is decreased, whereas the number of cell layers and the expansion rate remain equal to that of the cultures on uncoated membranes. Migration of epithelial cells on COL IV, COL I, and FN has also been found in other studies,^{27,28} but as yet not in organotypic urothelial cultures. The enhanced migration of ECM-coated membranes of the urothelial cells is probably due to an interaction of the ECM with ECM-specific integrins.²⁹ Unfortunately, we were not able to detect $\alpha 3$, $\alpha 5$, $\alpha 6$, $\beta 1$, or $\beta 4$ subunits using immunohistochemistry.

The negative effect of TGF β on the expansion rate of the urothelium is probably due to the induction of maturation and apoptosis in urothelial cells.^{30,31}

Another question addressed by this study was whether growth factors can act synergistically with the ECM in order to enhance expansion, which was tested by culturing the urothelium on COL I, COL IV, FN, or laminin and adding EGF to the culture. Exposure to EGF led to greater urothelial expansion of COL I-, COL IV-, and FN-coated membranes, but not on laminin-coated membranes. Apparently, the mechanism of

(laminin-mediated) cell spreading is not synergistic with the expanding effect of EGF-mediated proliferation. Indeed, the high nuclear density of basal cells in EGF-treated cultures on laminin-coated membranes demonstrated that the laminin-mediated cell spreading is inhibited by exposure to EGF. EGF may have changed the integrin expression pattern in such a way that the laminin-mediated cellular spreading is diminished. On the other hand, ECM coating of membranes reduced the EGF-associated increase in the number of cell layers, but not the increased level of [³H]thymidine incorporation.

In conclusion, this *in vitro* mouse urothelium regeneration model has permitted the identification of separate factors which selectively mediate the different mechanisms underlying the regeneration of urothelium *in vivo*. We failed to show a synergism between the different mechanisms of regeneration. Future studies should indicate whether these factors may also operate *in vivo*, during the regeneration of normal urothelium after therapeutic denudation of the urothelium.

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