

Modell sketches and rendered images of simulated chromosome organizations



Tobias A. Knoch

A: RW/GL-modell

B: MLS-modell

C: RW/GL, 5 Mbp loops

D: MLS, 126 kbp loops/linkers

E: RWTL, 126 kbp loops

F: Starting configuration

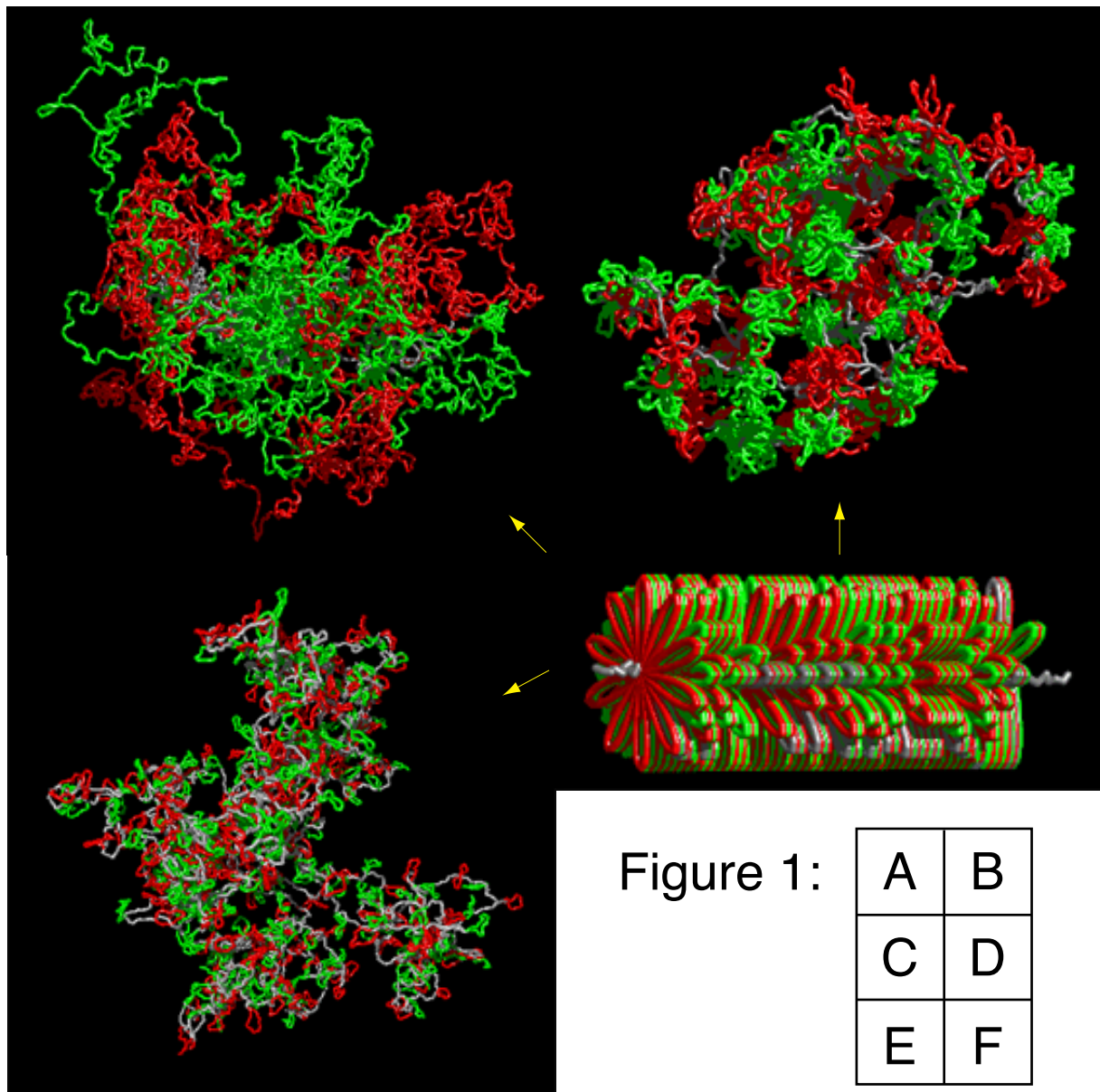
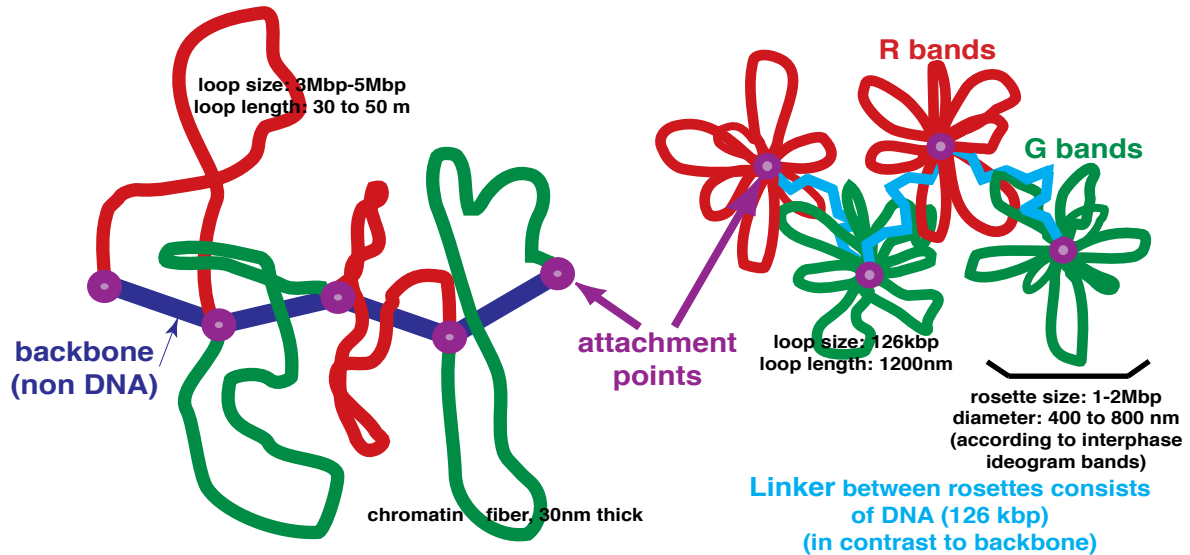
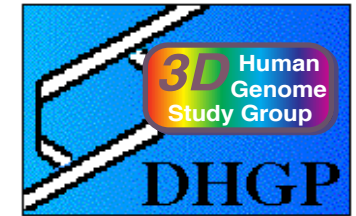


Figure 1:

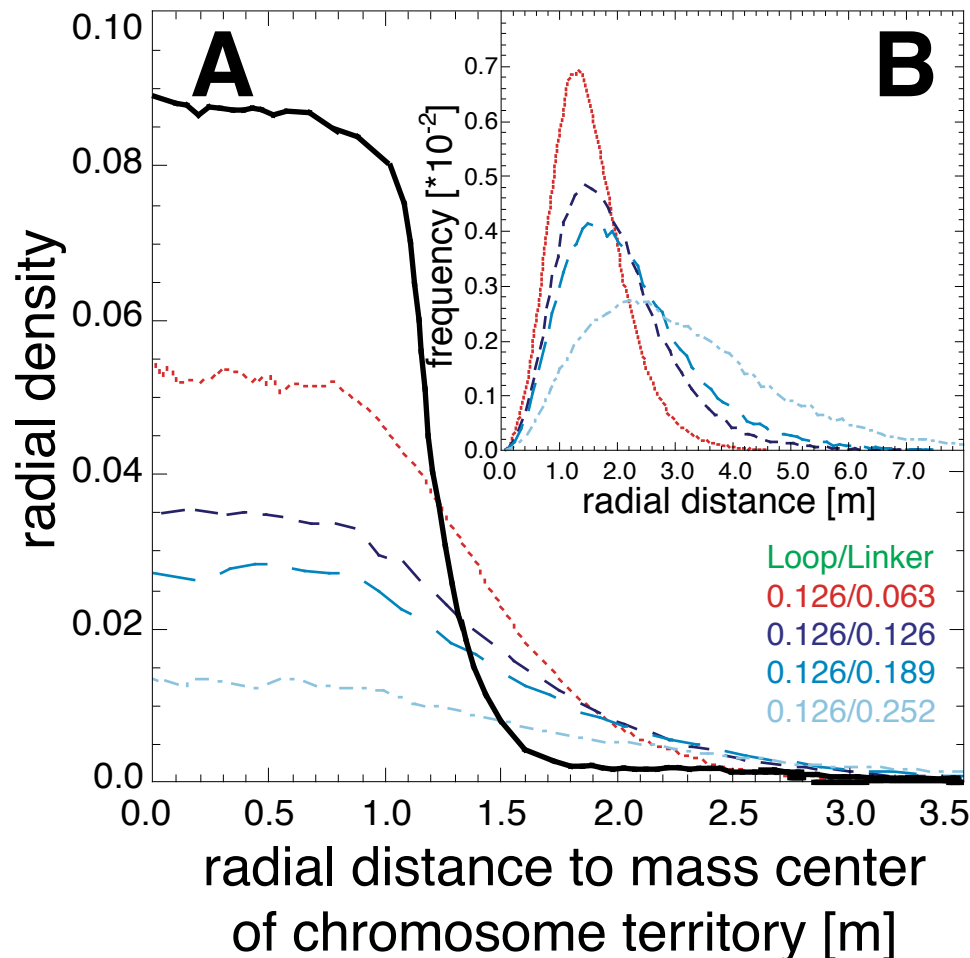
A	B
C	D
E	F

The mean radial mass distribution and radial density of chromosome territories is proportional *only* to the linker size in the MLS model and proportional to the linker *and* loop size in the RW/GL model. In the MLS model the radial density forms a plateau in contrast to the RW/GL model.

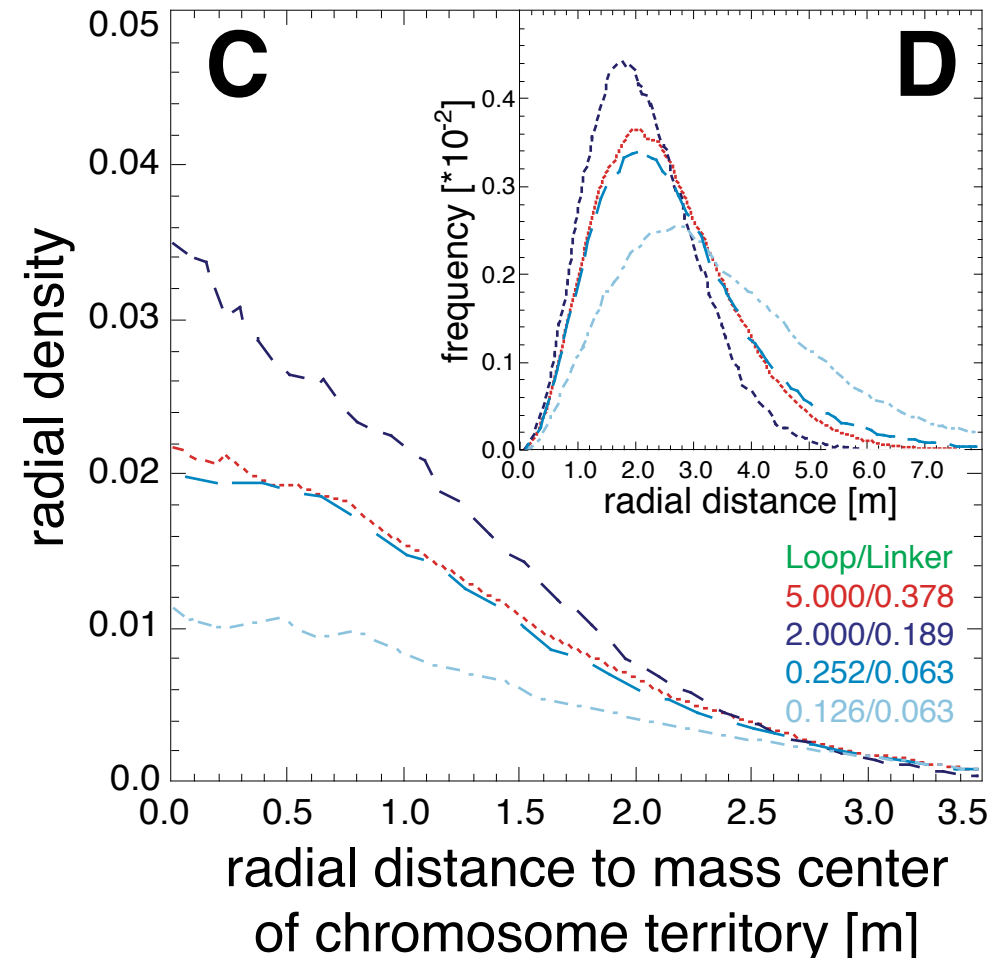


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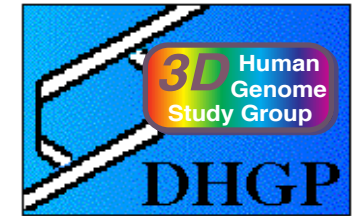
MLS-Model



RW/GL-model

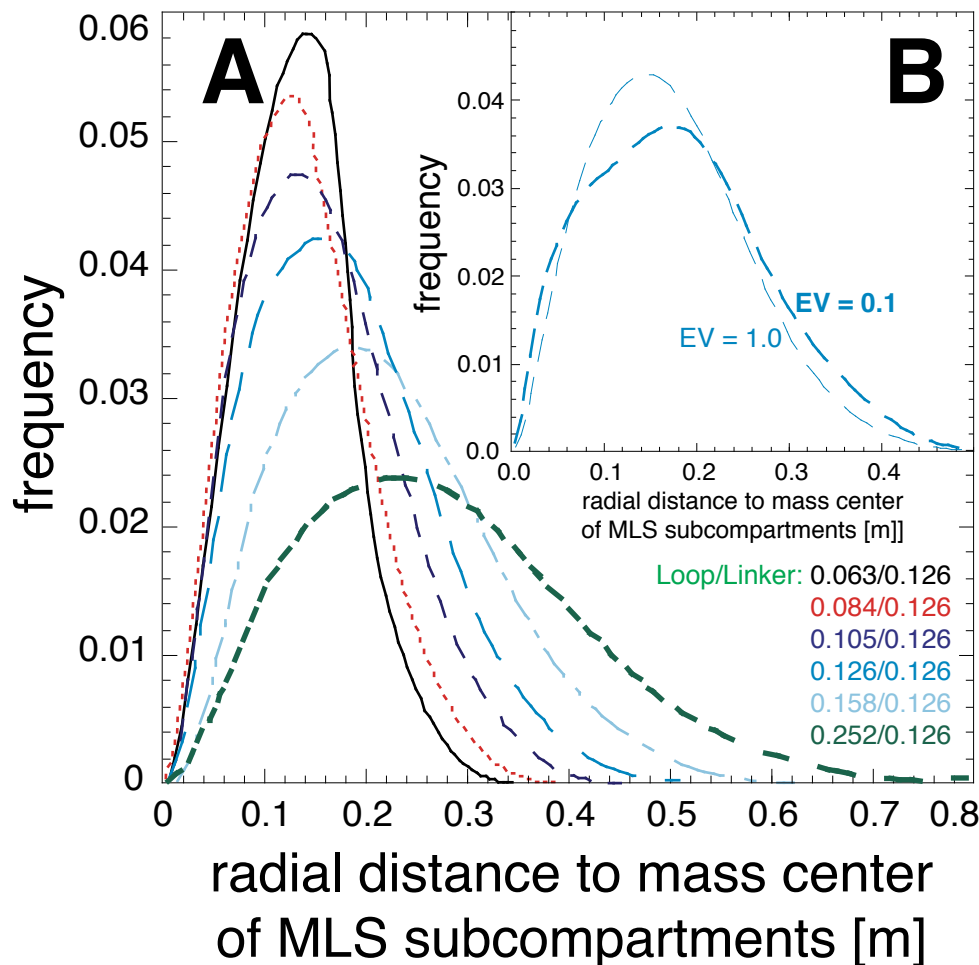


The mean extension of MLS subcompartments is proportional to the loop size (A) and the excluded volume interaction (B). The mean distance between succeeding subcompartments is only proportional to the linker size (C). The nearest neighbour subcompartment distance are proportional to the linker size and the embedding volume (D).

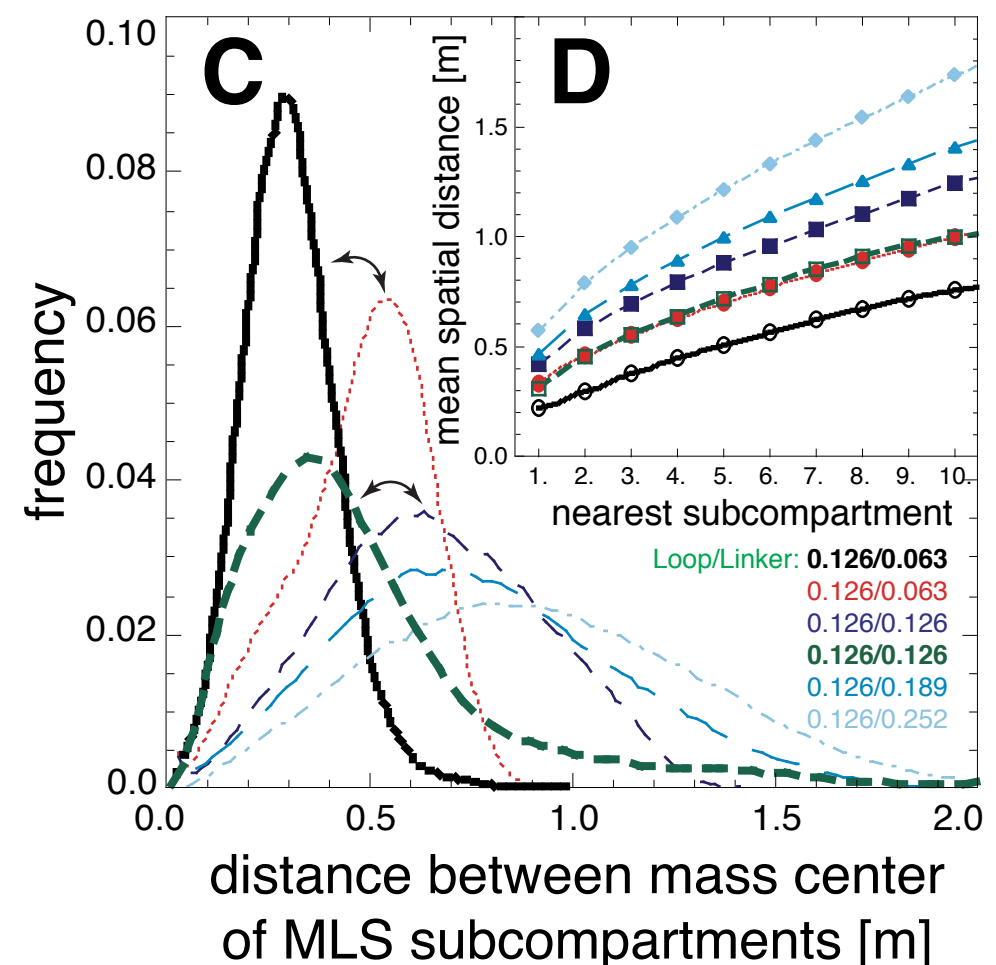


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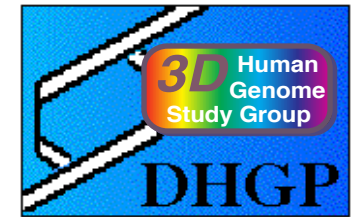
MLS-Model



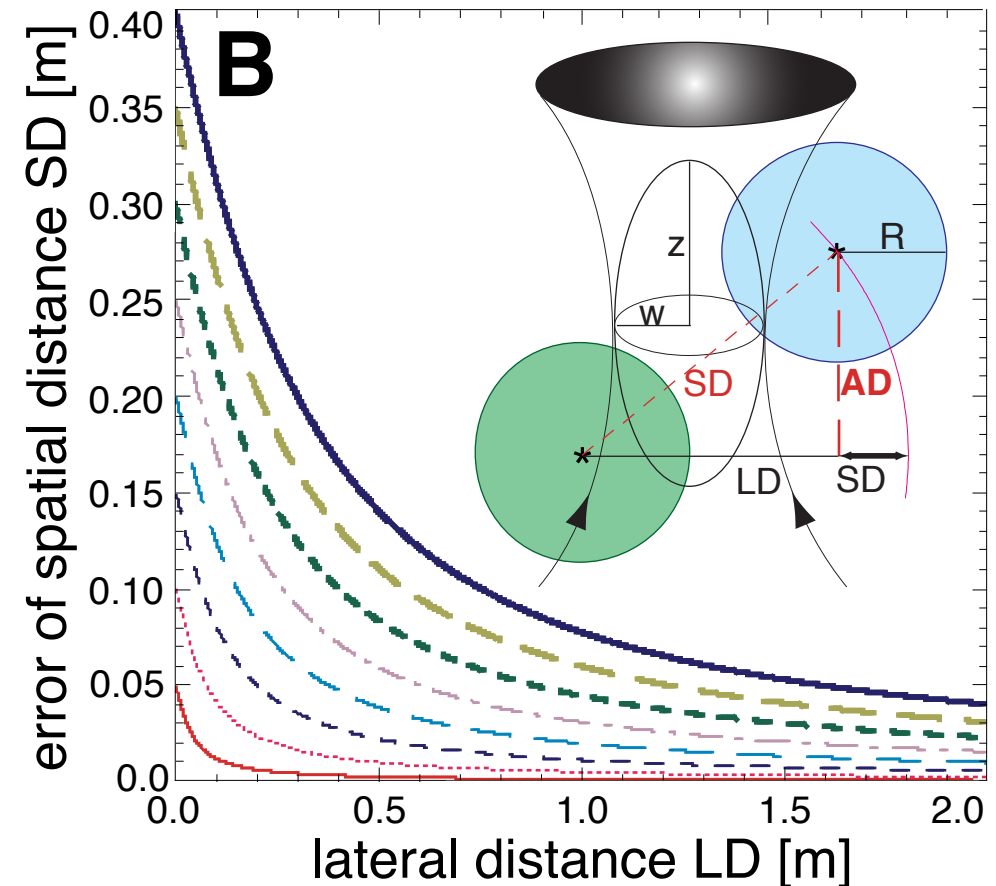
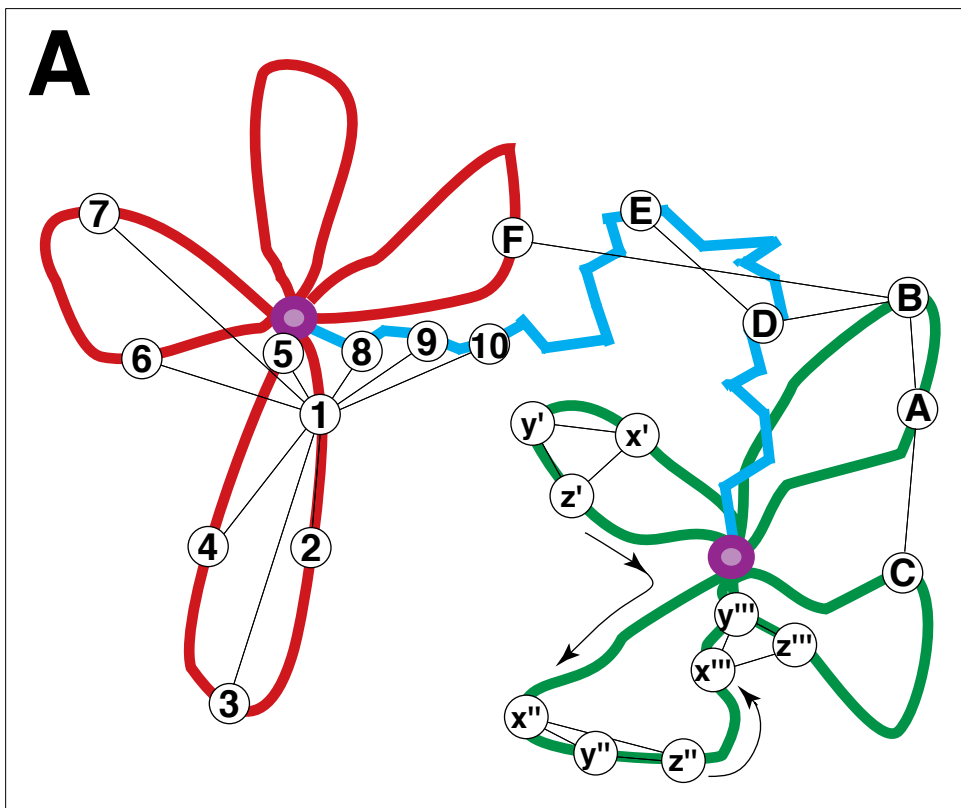
MLS-model



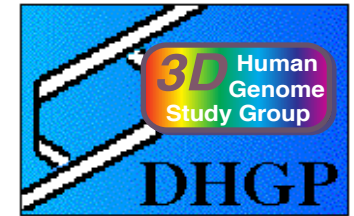
The determination of spatial distances between genomic markers as function of their genomic separation depends on different assumptions about the structure, stability and dynamics of chromosome organizations and the experimental error made depends on the microscopic method.



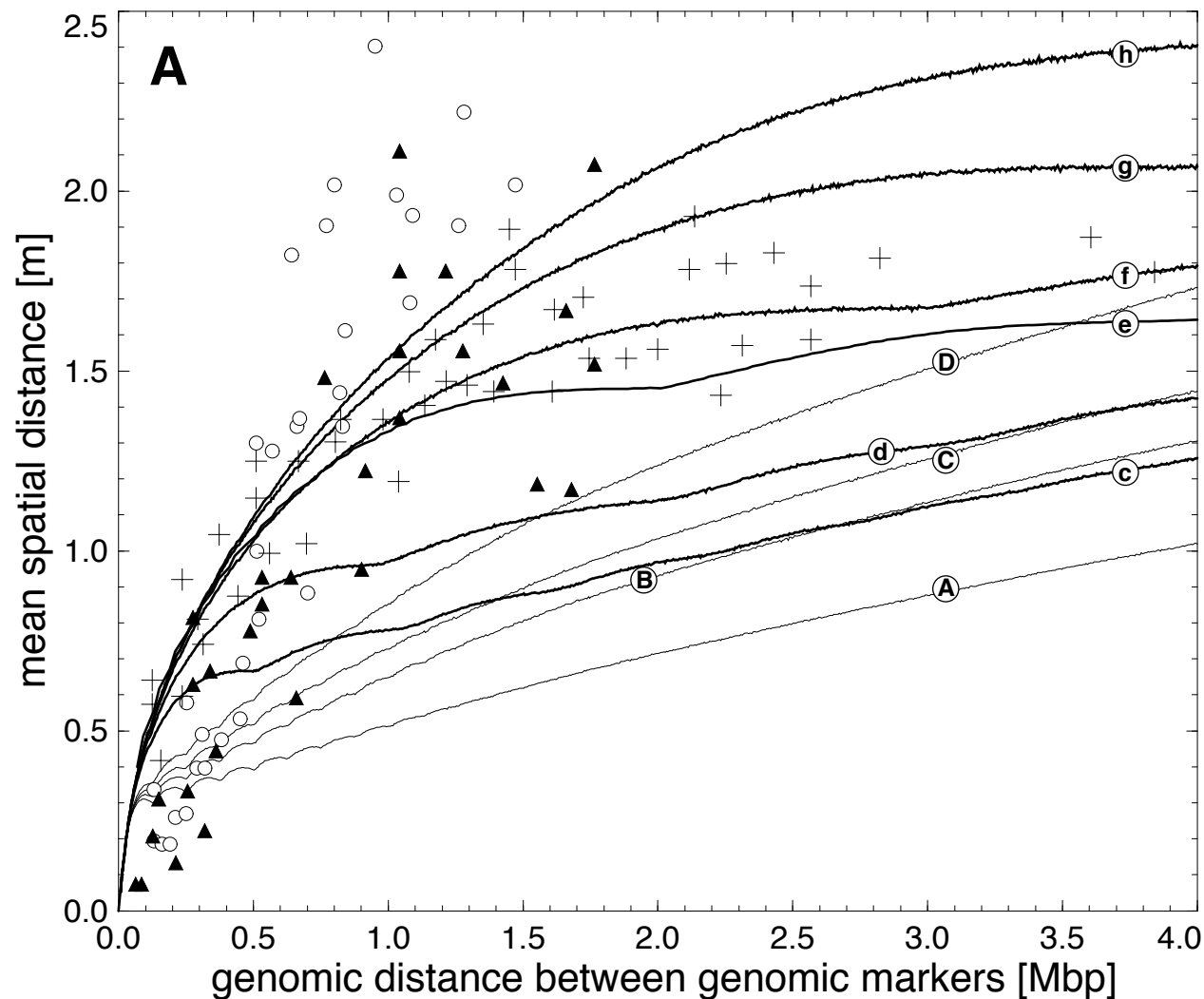
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Comparison between simulated and experimental spatial distance measurements as function of their genomic separation.

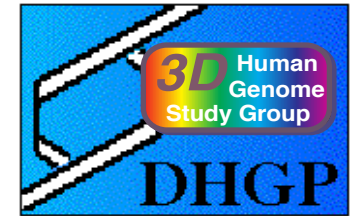


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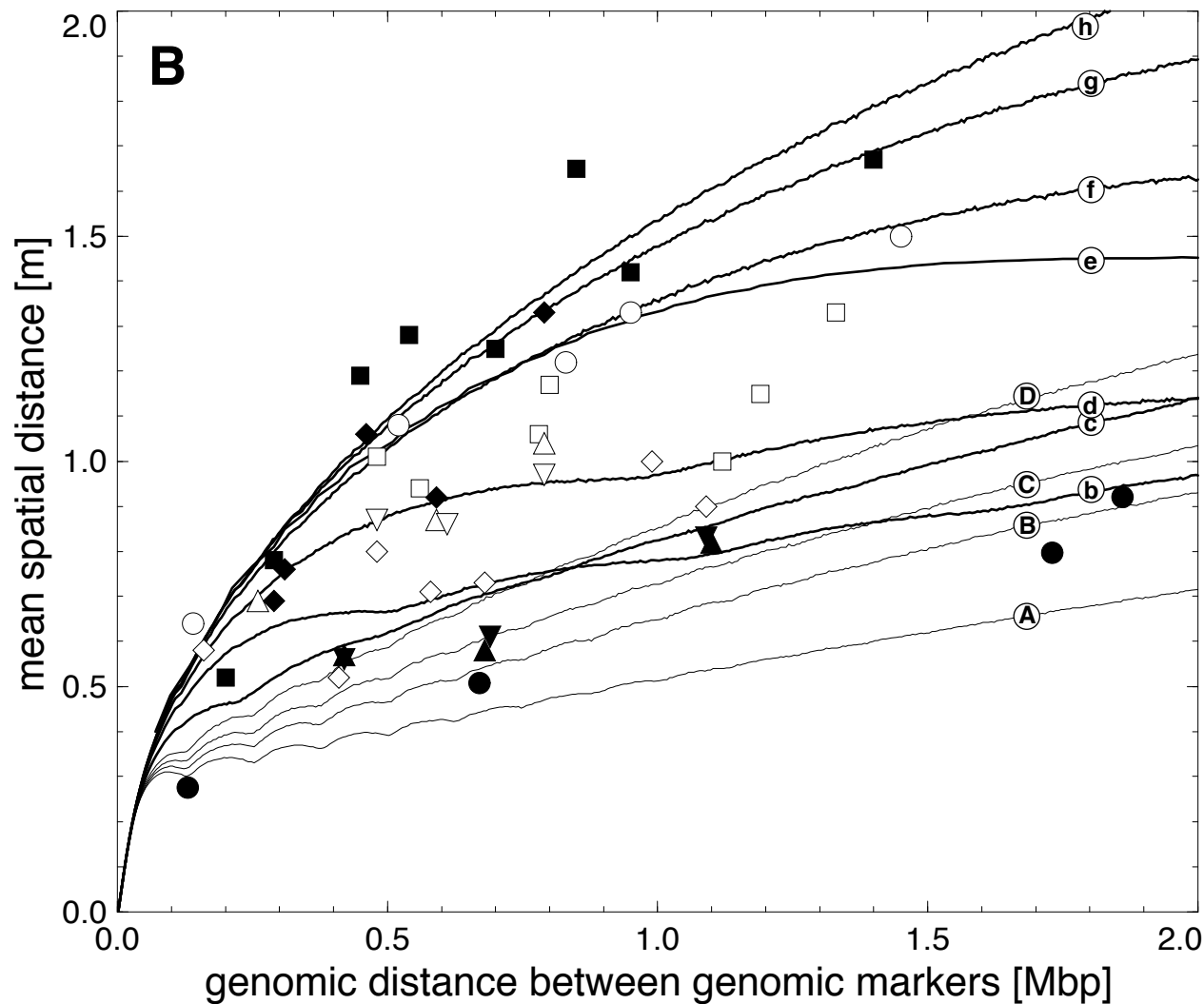


- van den Engh *et al.*, 1992
- ▲ Trask *et al.*, 1993
- + Yokota *et al.*, 1995

Comparison between simulated and experimental spatial distance measurements as function of their genomic separation.



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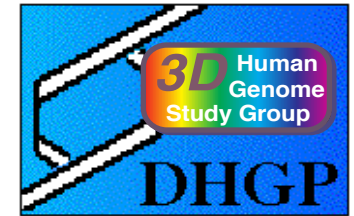


● Yokota *et al.*, 1995

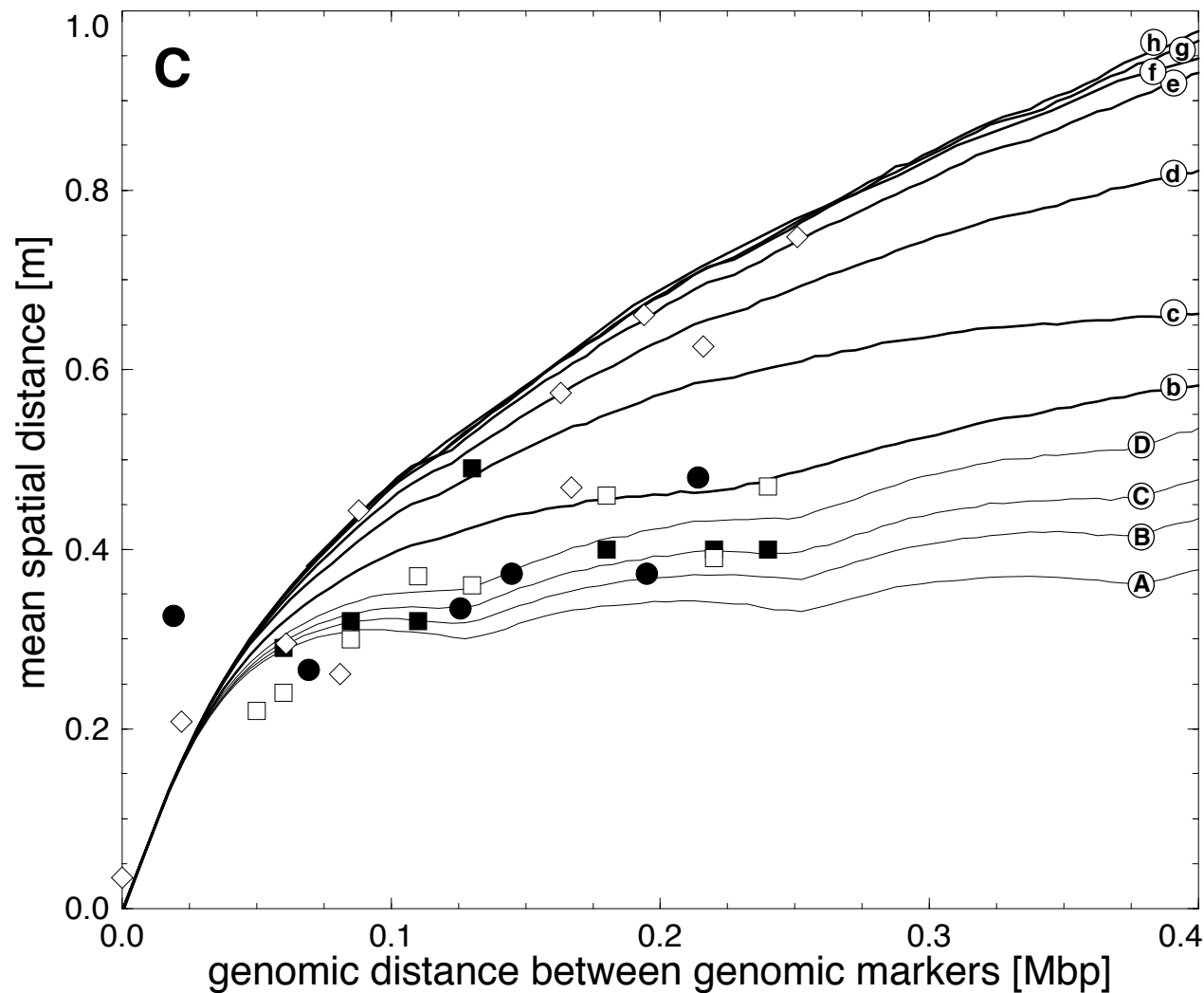
Yokota *et al.*, 1997:

- Fig. 2A 4p16.3
- Fig. 2B 6p21.3
- Fig. 2C 21q22.2
- ◆ Fig. 2D Xq28
- ◇ Fig. 2D Xp21.3
- ▲ Fig. 4B MAA-Xp21.3
- △ Fig. 4B MAA-Xq28
- ▼ Fig. 4A PFA-Xp21.3
- ▽ Fig. 4A PFA-Xq28

Comparison between simulated and experimental spatial distance measurements as function of their genomic separation.

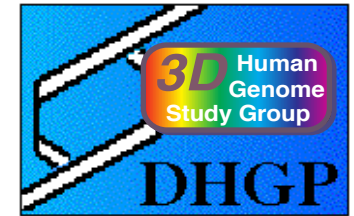


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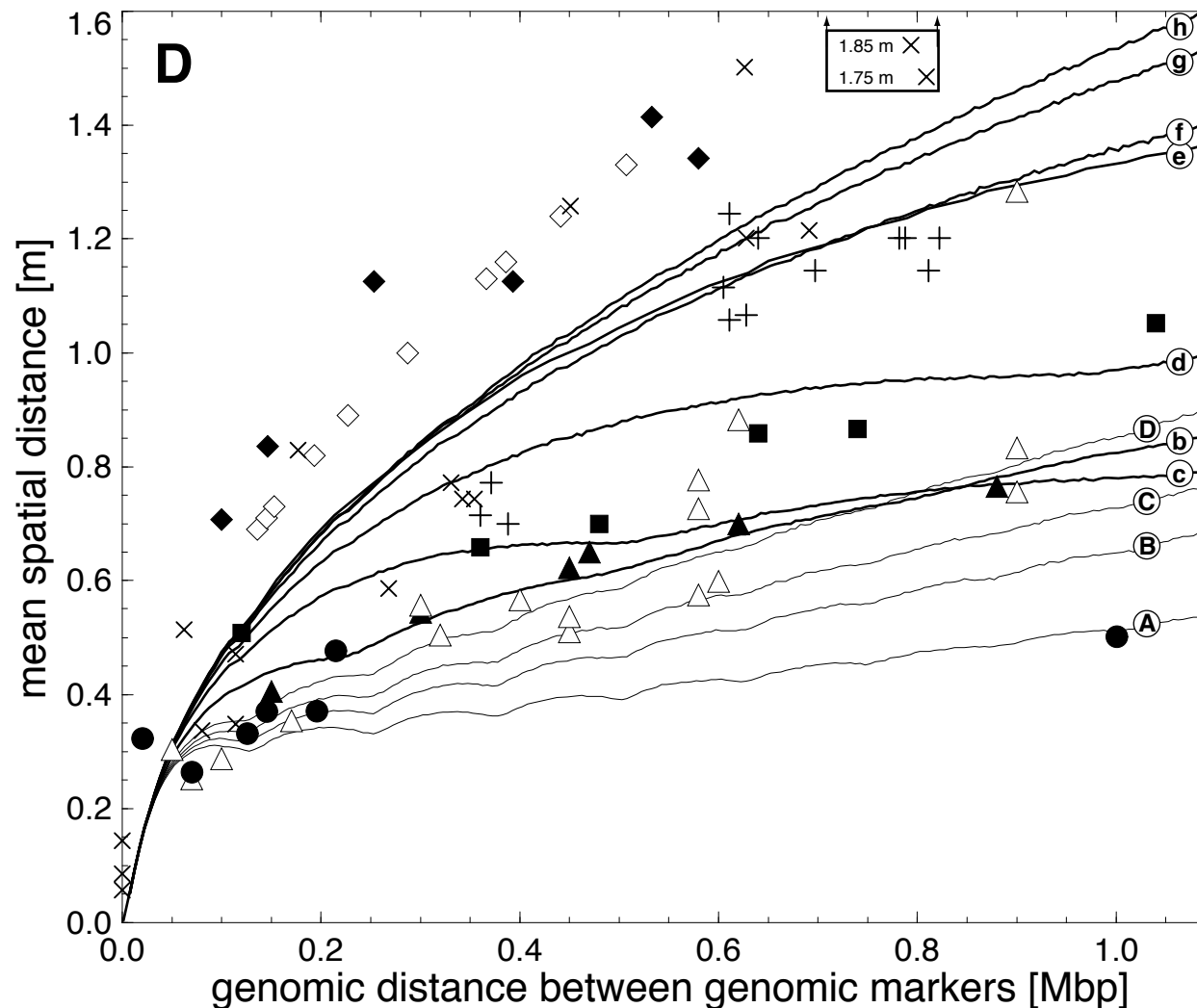


- our data:
J. Rauch, T. A. Knoch
- Monier:
- fibroblasts 11q13
lymphocytes 11q13
- ◇ Trask *et al.*, genomics.

Comparison between simulated and experimental spatial distance measurements as function of their genomic separation.

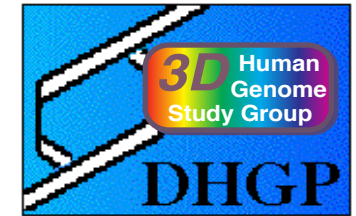


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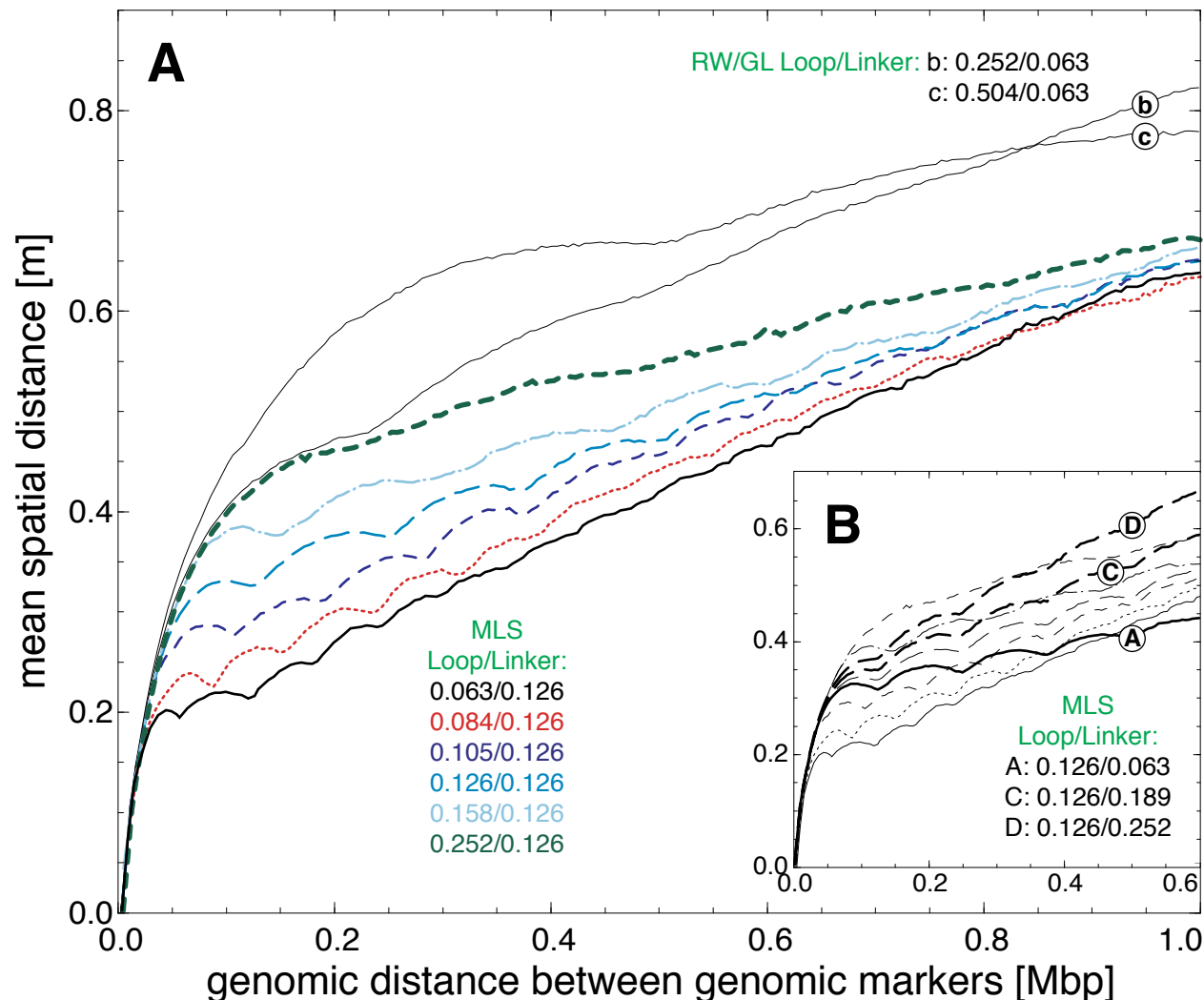


- our data:
J. Rauch, T. A. Knoch
- Lawrence *et al.*, 1990.
- ▲ Senger *et al.*, 1993:
one colour
- △ Warrington *et al.*, 1994:
two colour
- ◆ reference
- ◇ prediction
- Trask *et al.*, 1991:
- + Fig. 4A
- × Fig. 4B.

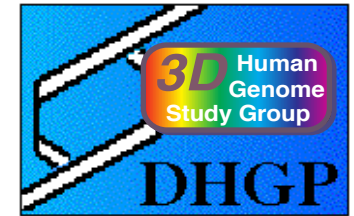
Position independent spatial distances are proportional to the loop size for genomic separations below 800 kbp in the MLS model (A). The loop size and the linker size can complement each other (B). With a comparison between the slope below and above 100 kbp and the point of change, it is possible to determine the loop and linker size separately.



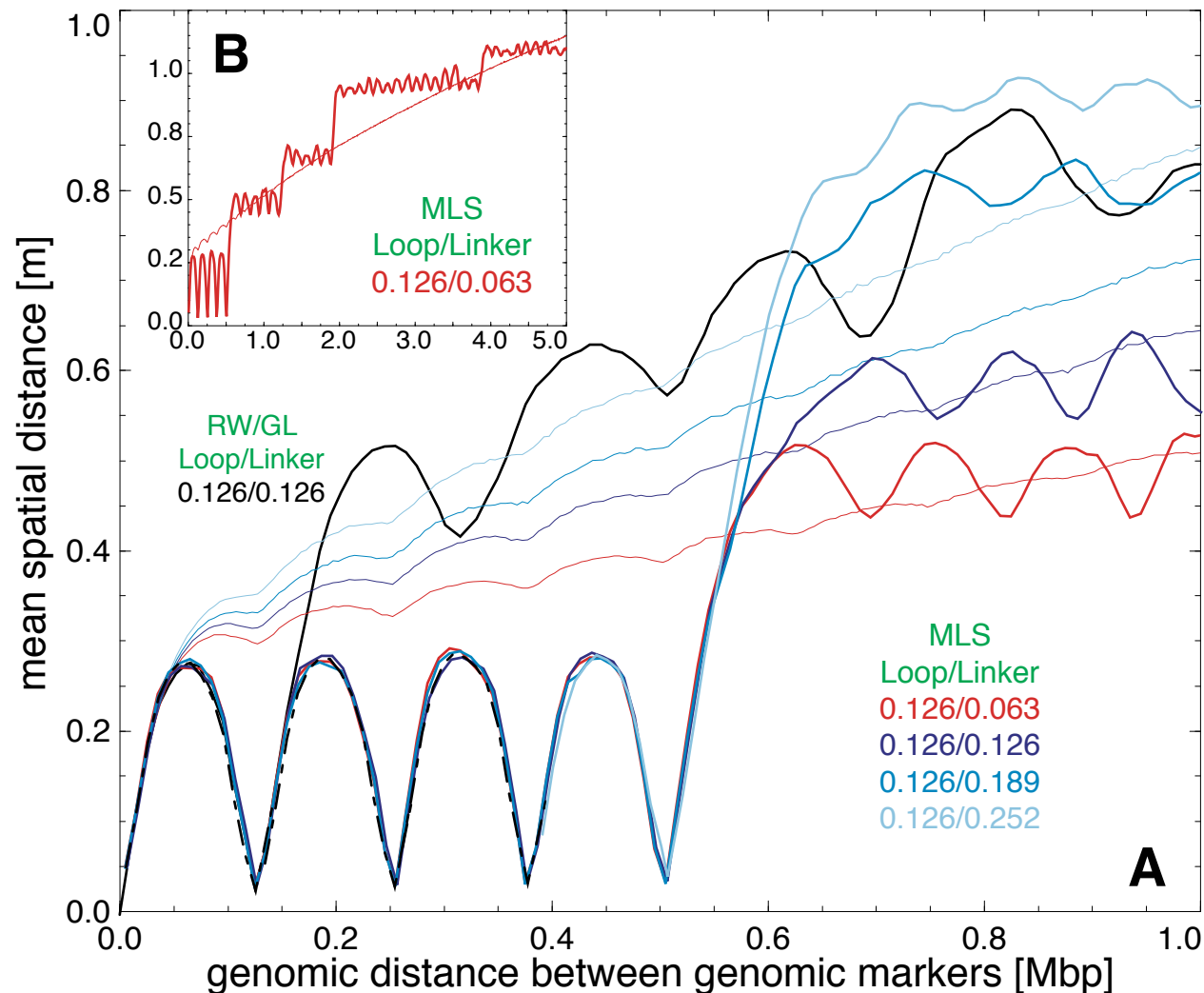
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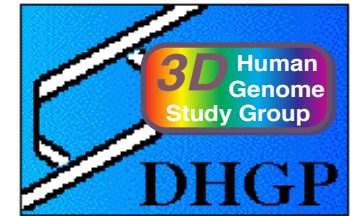
Position dependent spatial distance measurements allow the fine mapping of chromosomes (A & B). Position independent spatial distance measurements are the smeared out or the mean of the position dependent spatial distance measurements.



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Categorizing the available spatial distance measurements between genomic markers as function of their genomic separation leads to a better evaluation of the comparison between simulations and experiments.



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Purpose

Preparation Methods

Imaging Methods

**Order of clones
for sequencing**

**hypotonic swelling
dropping on coverslips
MAA fixation**

**2D
fluorescence microscopy**

**Transition phase
from sequencing to
packing of chromosomes**

**hypotonic swelling
dropping on coverslips
MAA fixation**

**2D
fluorescence microscopy
digital image analysis**

**detailed 3D organization
of the genome**

PFA fixation

**3D/4D CLSM
image reconstruction methods
correction for optical errors**

Simulation of Single Chromosomes, their Properties and Comparison to Experimental Spatial Distance Measurements

Knoch, T. A.

*Biophysics of Macromolecules Seminar, German Cancer Research Centre (DKFZ),
Heidelberg, Germany, April, 2000.*

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Keywords:

Genome, genomics, genome organization, genome architecture, structural sequencing, architectural sequencing, systems genomics, coevolution, holistic genetics, genome mechanics, genome function, genetics, gene regulation, replication, transcription, repair, homologous recombination, simultaneous co-transfection, cell division, mitosis, metaphase, interphase, cell nucleus, nuclear structure, nuclear organization, chromatin density distribution, nuclear morphology, chromosome territories, subchromosomal domains, chromatin loop aggregates, chromatin rosettes, chromatin loops, chromatin fibre, chromatin density, persistence length, spatial distance measurement, histones, H1.0, H2A, H2B, H3, H4, mH2A1.2, DNA sequence, complete sequenced genomes, molecular transport, obstructed diffusion, anomalous diffusion, percolation, long-range correlations, fractal analysis, scaling analysis, exact yard-stick dimension, box-counting dimension, lacunarity dimension, local nuclear dimension, nuclear diffuseness, parallel super computing, grid computing, volunteer computing, Brownian Dynamics, Monte Carlo, fluorescence in situ hybridization, confocal laser scanning microscopy, fluorescence correlation spectroscopy, super resolution microscopy, spatial precision distance microscopy, autofluorescent proteins, CFP, GFP, YFP, DsRed, fusionprotein, in vivo labelling.

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Knoch, T. A., Munkel, C. & Langowski, J. Three-dimensional organization of chromosome territories in the human interphase nucleus. *High Performance Computing in Science and Engineering 1999*, editors Krause, E. & Jäger, W., High-Performance Computing Center (HLRS) Stuttgart, University of Stuttgart, Springer Berlin-Heidelberg-New York, ISBN 3-540-66504-8, 229-238, 2000.

Bestvater, F., **Knoch, T. A.**, Langowski, J. & Spiess, E. GFP-Walking: Artificial construct conversions caused by simultaneous cotransfection. *BioTechniques* 32(4), 844-854, 2002.