Multiple Infectious Complications in a Severely Injured Patient with Single Nucleotide Polymorphisms in Important Innate Immune Response Genes

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Abstract: Trauma is a major public health problem worldwide. Infectious complications, sepsis, and multiple organ dysfunction syndrome (MODS) remain important causes for morbidity and mortality in patients who survive the initial trauma. There is increasing evidence for the role of genetic variation in the innate immune system on infectious complications in severe trauma patients. We describe a trauma patient with multiple infectious complications caused by multiple micro-organisms leading to prolonged hospital stay with numerous treatments. This patient had multiple single nucleotide polymorphisms (SNPs) in the *MBL2, MASP2, FCN2* and *TLR2* genes, most likely contributing to increased susceptibility and severity of infectious disease.

Keywords: Complications, genetic variation, infection, multiple organ dysfunction syndrome, single nucleotide polymorphism, systemic inflammatory response syndrome, trauma.

INTRODUCTION

Trauma is a major public health problem worldwide, ranking as the fourth leading cause of death. In 2010, there were 5.1 million deaths from injuries and the total number of deaths from injuries was greater than the number of deaths from HIV/AIDS, tuberculosis, and malaria combined (3.8 million) [1, 2]. Infectious complications, sepsis and multiple organ dysfunction syndrome (MODS) remain important causes for morbidity and mortality in patients who survive the initial trauma [3, 4]. These complications increase the burden of cost to society.

There is increasing evidence for the role of genetic variation in the innate immune system on infectious complications in sepsis and trauma [5-10]. We describe a trauma patient with multiple infectious complications caused by multiple micro-organisms leading to prolonged hospital stay with numerous treatments. This patient had multiple single nucleotide polymorphisms (SNPs) in the innate immune system, most likely contributing to increased susceptibility and severity of infectious disease.

SINGLE NUCLEOTIDE POLYMORPHISMS

Humans have 23 pairs of chromosomes and, on average, all humans are 99.9% similar to any other human in terms of DNA sequence. The remaining 0.1% account for all the

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differences between humans These differences are known as 'polymorphisms'. The coding regions of DNA contain the approximately 20,000 human protein-coding genes. The coding regions take up less than 2% of all DNA. More than 98% of the human genome is composed of non-coding DNA of which the function is partly unknown. Human diploid cells contain around six billion base pairs. These base pairs are pairs of nucleotides, the building blocks of DNA (A, C, T, and G).

Single Nucleotide Polymorphisms (SNPs; pronounced '*snip*', plural '*snips*') are variations of only one nucleotide in the sequence of DNA. Around 90% of all DNA variation is caused by SNPs making them the most common type of sequence variation. To date more than 60 million SNPs have been discovered in the human genome.

SNPs in coding regions of DNA may have the potential to alter the amino acid sequence in a protein but as a result of degeneracy this is not always the case; some proteins are coded by more than one codon. SNPs in coding regions are called synonymous if they do not affect the amino acid sequence and non-synonymous if they do influence the amino acid sequence of a protein. The non-synonymous SNPs can be divided into missense SNPs and nonsense SNPs. Missense SNPs result in the transcription of a different amino-acid, changing the functionality of the resulting protein as is the case in Factor V Leiden thrombophilia [11] and sickle cell disease [12]. A nonsense SNP results in the formation of a premature stopcodon leading to a truncated, incomplete protein as is the case in β thalassemia in Sardinia [13] and some forms of cystic fibrosis [14].

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CASE REPORT

Patient A, a 57 year old mechanic with a medical history of occasional use of cocaine and of gradually worsening vision in the last five months as a result of optic nerve atrophy tripped over a low brick wall at work and fell about one meter on the back of his head. He was immediately found to be tetraplegic by the paramedics and was transferred to a level 1 trauma center. Clinically, the complete cord lesion was found to be on the level of C3 and C4. Computed tomography showed a congenital narrowing of the spinal canal at the level of C3 and C4 as well as an old fracture of the third thoracic vertebra (Fig. 1). Magnetic resonance scanning showed hemorrhage in the myelum at the level of C3 and C4 as the reason for the tetraplegia (Fig. 2).



Fig. (1). Computed tomography of discussed patient showing the congenital narrowing of the spinal canal at the level of C3 and C4. Also, a preexisting injury at the level of T2 and T3 can be seen.

He was transferred to the Intensive Care Unit for ventilator support. On day 3 he developed acute respiratory distress syndrome (ARDS) and pneumonia in the right lower lobe, possibly as a result of aspiration, from which purulent sputum was removed repeatedly. The sputum grew Haemolytic Streptococcus group C, Streptococcus pneumoniae, Haemophilus influenzae and Enterobacter cloacae for which he was treated with piperacillin/tazobactam. On day 10 a percutaneous tracheostomy was used and weaning was possible. On day 20 patient was no longer dependent on ventilator support. He spent a total of 32 days on the Intensive Care Unit and 76 days in the hospital before being discharged to a rehabilitation center. After three months he was admitted again, this time to the department of Internal Medicine, for fever, diarrhea and productive cough. Radiology was suspect for pulmonary tuberculosis and

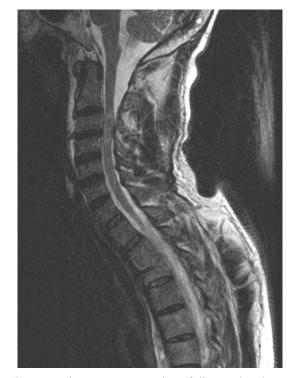


Fig. (2). Magnetic resonance scanning of discussed patient shows bleeding in the myelum at the level of C3 and C4 as the cause for his complete cord syndrome.

Mycobacterium tuberculosis was eventually found in gastric contents. He was placed on tuberculostatic triple therapy. His diarrhea was explained by pseudomembranous colitis caused by *Clostridium* toxins. He was given metronidazol and was free of diarrhea after ten days. He developed deep venous thrombosis in the right subclavian vein ultimately leading to erysipelas and ulceration on the fingers from which *Escherichia coli, Klebsiella pneumoniae, Pseudomonas aeruginosa, Morganella morganii* and *Proteus mirabilis* were cultured. The urine was positive for *Klebsiella pneumoniae* and *Enterococcus faecalis*. The ulcers were treated surgically.

Because this patient participated in a scientific trial studying genetic variation in trauma patients, his genome was sequenced for several single nucleotide polymorphisms (SNPs). Several SNPs were found: heterozygosity in *MBL2* exon 1 (*i.e.*, AC genotype), heterozygosity in *MBL2* promoter region Y-221X, (YX genotype) homozygosity for the minor allele in *MASP2* Y371D (DD genotype), heterozygosity in *FCN2* T236M (TM genotype), heterozygosity in *FCN2* A258S (AS genotype), homozygosity for the minor allele in *TLR2* T-16934A (AA genotype), and heterozygosity in *CD14* C-159T (CD genotype).

DISCUSSION

Mortality as a result of sepsis in the third peak of Trunkey [15] has not changed in recent times despite the improvements in treatments in the Intensive Care Unit leading to reduction in the incidence of sepsis itself [4]. The onset of sepsis in trauma patients is of course multifactorial, but genetic variation at the level of the innate immune

Gene	SNP	OMIM	Cytogenic Location	dbSNP ID	Genotype of Our Patient
MBL2 exon 1	codon 57 [C allele]	154545	10q21	rs1800451	AC
MBL2 promoter	Y-221X	154545	10q21	rs7096206	YX
MASP2	Y371D	605102	1p36	rs12711521	DD
FCN2	A258S	601624	9q34	rs7851696	AS
TLR2	T-16934A	603028	4q31	rs4696480	AA

Table 1. Genotypes found in presented patient.

SNP: single nucleotide polymorphism. **OMIM**: Online Mendelian Inheritance in Man, an online catalog of human genes and genetic disorders. **dbSNP ID**: Single Nucleotide Polymorphism Database, a free public archive for genetic variation hosted by the National Center for Biotechnology Information [NCBI].

system is certainly one important contributing factor. SNPs in genes coding for important proteins in the innate immune system, such as the complement system, may produce low serum levels of these proteins or they may leave these proteins dysfunctional. Hence, the immune response and cytokine response to trauma and infection is reduced leading to increased susceptibility and severity of infectious complications. These complications cause prolonged hospital stay and increase the use of antibiotics, the number of complications, and the cost of care to society.

The above presented patient was proband in a prospective study in severely injured trauma patients focusing on SNPs in the innate immune system and the influence on infectious complications. A number of SNPs were found in this patient (see Table 1).

In the lectin pathway of complement activation three important genes were studied: MBL2, MASP2, and FCN2. The MBL2 gene encodes for mannose-binding lectin (MBL), a protein that is secreted by the liver as part of the acutephase response and is involved in innate immune defense. The ligands for MBL are expressed by a wide variety of microorganisms, and binding of the protein leads to opsonisation of the pathogen as well as activation of the complement system. Genetic variation in this gene leads to a dramatic decrease in circulating serum MBL. Heterozygosity for variants in exon 1 (i.e., an A0 genotype) conferred an increased risk for wound colonization and infection in severely injured patients [6]. This had previously only been demonstrated in a murine model of burns [16]. Also, the YX promoter genotype increased the risk of fungal colonization and infection in trauma patients [6]. Presented patient carried an AC genotype in exon 1 and a YX genotype in the promoter region. MBL activates the complement pathway through mannan-binding lectin serine protease 2 (MASP2). MASP2 Y371D DD homozygosity significantly increased the risk for SIRS and septic shock in trauma patients [6]. Moreover, a trend was noted for an increased risk of Grampositive infections in patients with MASP2 Y371D DD genotype. Above presented patient carried the MASP2 Y371D DD genotype. FCN2 encodes for Ficolin-2, previously termed L-ficolin, a protein which is mainly produced in the liver and has been shown to have carbohydrate binding and opsonic properties in the innate immune system. The homozygous FCN2 A258S AS genotype increased the risk for developing septic shock in

trauma patients [6]. Also, wound colonization and infection were significantly increased. A trend was noted for Gramnegative infections. In this article presented patient carried the *FCN2* A258S AS genotype.

Deficiencies in the lectin pathway have been linked to susceptibility of various pathogens, for example MASP deficient mice are highly susceptible to Streptococcus pneumoniae [17] and also children [18] and adults [19] have previously been shown to be highly susceptible to S. pneumoniae with deficiencies in MBL and MASP2. In children lectin pathway deficiencies may play a role in Haemophilus influenzae [20] but this was not found in a cohort of adults with community acquired pneumonia [21]. The effect of MBL genotype on the susceptibility to Mycobacterium tuberculosis is controversial [22, 23]. In burn injury patients [16] and cystic fibrosis patients [24] MBL deficiency plays an important role on susceptibility to Pseudomonas aeruginosa infection and colonization. The FCN2 A258S polymorphism was previously shown to influence susceptibility to leprosy [25], influence colonization with Pseudomonas aeruginosa in cystic fibrosis patients [24] and influence renal transplant outcome [26].

As a membrane surface receptor, TLR-2 recognizes many bacterial, fungal, viral, and certain endogenous substances. The *TLR2* T-16934A polymorphism was previously linked to spontaneous bacterial peritonitis in liver cirrhosis patients [27], atopic dermatitis, asthma and wheezing [28-30] and sarcoidosis [31]. The *TLR2* T-16934A genotype was studied in a trauma population by one author [7] who found that the *TLR2* T-16934A TA genotype increased the risk of a Grampositive infection and SIRS. The *TLR2* T-16934A AA genotype seemed to protect against urinary infection, oddly. However, patient A carried the *TLR2* T-16934A AA genotype but developed positive urine cultures.

Infectious complications are multifactorial in origin. SNPs in the innate immune system contribute to susceptibility and severity of these infections and lead to prolonged hospital stay and increased cost. The presented patient demonstrates the clinical course of such complications that will be recognized by all surgeons. In the future we expect that initial genotyping will become routine workup in all trauma patients to quantify the individual risk for developing infections. Patients identified to be at risk for developing infectious complications can be prophylactically treated with antibiotics in an early stage or can be supplemented with plasma from mixed donors containing the deficient proteins. Substitution therapy with purified or recombinant proteins has also produced clinical results, for example in the case of MBL-deficiency [32-36]. Further studies are needed in order to determine which genes affect this risk and to quantify their effect.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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REFERENCES

- Norton R, Kobusingye O. Injuries. N Engl J Med 2013; 368(18): 1723-30.
- [2] Lozano R, Naghavi M, Foreman K, et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet 2012; 380(9859): 2095-128.
- [3] Mann EA, Baun MM, Meininger JC, Wade CE. Comparison of mortality associated with sepsis in the burn, trauma, and general intensive care unit patient: a systematic review of the literature. Shock 2012; 37(1): 4-16.
- [4] Wafaisade A, Lefering R, Bouillon B, et al. Epidemiology and risk factors of sepsis after multiple trauma: an analysis of 29,829 patients from the Trauma Registry of the German Society for Trauma Surgery. Crit Care Med 2011; 39(4): 621-8.
- [5] Pothlichet J, Quintana-Murci L. The genetics of innate immunity sensors and human disease. Int Rev Immunol 2013; 32(2): 157-208.
- [6] Bronkhorst MWGA, Lomax MAZ, Vossen RHAM, Bakker J, Patka P, van Lieshout EMM. Risk of infection and sepsis in severely injured patients related to single nucleotide polymorphisms in the lectin pathway. Br J Surg 2013; 100(13): 1818-26.
- [7] Bronkhorst MWGA, Boye ND, et al. Single-nucleotide polymorphisms in the Toll-like receptor pathway increase susceptibility to infections in severely injured trauma patients. J Trauma Acute Care Surg 2013; 74(3): 862-70.
- [8] Zeng L, Zhang AQ, Gu W, et al. Clinical relevance of single nucleotide polymorphisms of the high mobility group box 1 protein gene in patients with major trauma in southwest China. Surgery 2012; 151(3): 427-36.
- [9] Manson J, Thiemermann C, Brohi K. Trauma alarmins as activators of damage-induced inflammation. The British journal of surgery. 2012; 99 (Suppl 1): 12-20.
- [10] Hildebrand F, Mommsen P, Frink M, van Griensven M, Krettek C. Genetic predisposition for development of complications in multiple trauma patients. Shock 2011; 35(5): 440-8.
- [11] Bertina RM, Koeleman BP, Koster T, et al. Mutation in blood coagulation factor V associated with resistance to activated protein C. Nature 1994; 369(6475): 64-7.
- [12] Sebastiani P, Solovieff N, Hartley SW, et al. Genetic modifiers of the severity of sickle cell anemia identified through a genome-wide association study. Am J Hematol 2010; 85(1): 29-35.
- [13] Cao A, Rosatelli C, Pirastu M, Galanello R. Thalassemias in Sardinia: molecular pathology, phenotype-genotype correlation, and prevention. Am J Pediatr Hematol Oncol 1991; 13(2): 179-88.
- [14] Rolfini R, Cabrini G. Nonsense mutation R1162X of the cystic fibrosis transmembrane conductance regulator gene does not reduce messenger RNA expression in nasal epithelial tissue. J Clin Invest 1993; 92(6): 2683-7.

- [15] Trunkey DD. Trauma. Accidental and intentional injuries account for more years of life lost in the U.S. than cancer and heart disease. Among the prescribed remedies are improved preventive efforts, speedier surgery and further research. Sci Am 1983; 249(2): 28-35.
- [16] Moller-Kristensen M, Ip WK, Shi L, et al. Deficiency of mannosebinding lectin greatly increases susceptibility to postburn infection with Pseudomonas aeruginosa. J Immunol 2006; 176(3): 1769-75.
- [17] Ali YM, Lynch NJ, Haleem KS, *et al.* The lectin pathway of complement activation is a critical component of the innate immune response to pneumococcal infection. PLoS pathogens 2012; 8(7): e1002793.
- [18] Munoz-Almagro C, Bautista C, Arias MT, et al. High prevalence of genetically-determined mannose binding lectin deficiency in young children with invasive pneumococcal disease. Clin Microbiol Infect 2014; 20(10): O745-52.
- [19] Eisen DP. Mannose-binding lectin deficiency and respiratory tract infection. J Innate Immun 2010; 2(2): 114-22.
- [20] Vuononvirta J, Toivonen L, Grondahl-Yli-Hannuksela K, et al. Nasopharyngeal bacterial colonization and gene polymorphisms of mannose-binding lectin and toll-like receptors 2 and 4 in infants. PLoS One 2011; 6(10): e26198.
- [21] Endeman H, Herpers BL, de Jong BA, et al. Mannose-binding lectin genotypes in susceptibility to community-acquired pneumonia. Chest 2008; 134(6): 1135-40.
- [22] Singla N, Gupta D, Joshi A, Batra N, Singh J, Birbian N. Association of mannose-binding lectin gene polymorphism with tuberculosis susceptibility and sputum conversion time. Int J Immunogenet 2012; 39(1): 10-4.
- [23] You HL, Lin TM, Wang JC, et al. Mannose-binding lectin gene polymorphisms and mycobacterial lymphadenitis in young patients. Pediatr Infect Dis J 2013; 32(9): 1005-9.
- [24] Haerynck F, Van Steen K, Cattaert T, *et al.* Polymorphisms in the lectin pathway genes as a possible cause of early chronic Pseudomonas aeruginosa colonization in cystic fibrosis patients. Hum Immunol 2012; 73(11): 1175-83.
- [25] Zhang DF, Huang XQ, Wang D, Li YY, Yao YG. Genetic variants of complement genes ficolin-2, mannose-binding lectin and complement factor H are associated with leprosy in Han Chinese from Southwest China. Human Genetics 2013; 132(6): 629-40.
- [26] Eikmans M, de Canck I, van der Pol P, et al. The functional polymorphism Ala258Ser in the innate receptor gene ficolin-2 in the donor predicts improved renal transplant outcome. Transplantation 2012; 94(5): 478-85.
- [27] Nischalke HD, Berger C, Aldenhoff K, et al. Toll-like receptor (TLR) 2 promoter and intron 2 polymorphisms are associated with increased risk for spontaneous bacterial peritonitis in liver cirrhosis. J Hepatol 2011; 55(5): 1010-6.
- [28] Custovic A, Rothers J, Stern D, et al. Effect of day care attendance on sensitization and atopic wheezing differs by Toll-like receptor 2 genotype in 2 population-based birth cohort studies. J Allergy Clin Immunol 2011; 127(2): 390-7 e1-9.
- [29] Oh DY, Schumann RR, Hamann L, Neumann K, Worm M, Heine G. Association of the toll-like receptor 2 A-16934T promoter polymorphism with severe atopic dermatitis. Allergy 2009; 64(11): 1608-15.
- [30] Eder W, Klimecki W, Yu L, *et al.* Toll-like receptor 2 as a major gene for asthma in children of European farmers. J Allergy Clin Immunol 2004; 113(3): 482-8.
- [31] Veltkamp M, Wijnen PA, van Moorsel CH, et al. Linkage between Toll-like receptor (TLR) 2 promotor and intron polymorphisms: functional effects and relevance to sarcoidosis. Clin Exp Immunol 2007; 149(3): 453-62.
- [32] Brouwer N, Frakking FN, van de Wetering MD, et al. Mannosebinding lectin (MBL) substitution: recovery of opsonic function in vivo lags behind MBL serum levels. J Immunol 2009; 183(5): 3496-504.
- [33] Frakking FN, Brouwer N, van de Wetering MD, et al. Safety and pharmacokinetics of plasma-derived mannose-binding lectin (MBL) substitution in children with chemotherapy-induced neutropaenia. Eur J Cancer 2009; 45(4): 505-12.

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- [34] Bang P, Laursen I, Thornberg K, et al. The pharmacokinetic profile of plasma-derived mannan-binding lectin in healthy adult volunteers and patients with Staphylococcus aureus septicaemia. Scand J Infect Dis 2008; 40(1): 44-8.
- [35] Vorup-Jensen T. Challenges and opportunities in fractionation of recombinant human mannan-binding lectin. Methods Mol Biol 2014; 1100: 109-21.

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[36] Valdimarsson H, Vikingsdottir T, Bang P, *et al.* Human plasmaderived mannose-binding lectin: a phase I safety and pharmacokinetic study. Scand J Immunol 2004; 59(1): 97-102.

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