Intranasal Cold Dry Air Is Superior to Histamine Challenge in Determining the Presence and Degree of Nasal Hyperreactivity in Nonallergic Noninfectious Perennial Rhinitis

JOSEPH P. M. BRAAT, PAUL G. MULDER, WYTSKE J. FOKKENS, ROY GERTH van WIJK, and EVERT RIJNTJES

Departments of Ear, Nose and Throat Surgery, Biostatistics, and Allergology, Erasmus University Rotterdam and University Hospital ‘Dijkzigt,’ Rotterdam; and Department of Ear, Nose and Throat Surgery, Leyenburg Hospital, The Hague, The Netherlands

The objective of the study was to compare cold dry air (CDA) and histamine in differentiating patients with nonallergic noninfectious perennial rhinitis (NANIPER) from control subjects. Nasal reactivity (nasal patency, mucus production, and sneezing) in 16 symptomatic nonsmoking patients with NANIPER and seven nonsmoking control subjects was measured with standardized CDA and histamine provocation series in a randomized crossover study. Intranasal CDA resulted in increased mucus production and nasal blockage in a dose-dependent manner in patients with NANIPER but not in control subjects. Sneezing did not occur. The reproducibility of CDA for patency and mucus production was good. Sensitivity for CDA was 87% compared with 100% for histamine. However, specificity was 71% for CDA and 0% for histamine. It is concluded that the new standardized intranasal CDA provocation method uses a recognizable natural nonspecific stimulus and seems to be more suitable than histamine for characterizing and assessing the presence and degree of nasal reactivity in NANIPER. Braat JPM, Mulder PG, Fokkens WJ, van Wijk RG, Rijntjes E. Intranasal cold dry air is superior to histamine challenge in determining the presence and degree of nasal hyperreactivity in nonallergic noninfectious perennial rhinitis.


Nonallergic noninfectious perennial rhinitis (NANIPER) (also referred to as vasomotor rhinitis) was first described by Wihl and colleagues (1). It is a chronic ongoing type of rhinitis characterized by nasal hyperreactivity that results in nasal blockage, mucus production, and sneezing (2, 3). The diagnosis is established by exclusion. When patients with noninfectious rhinitis respond negatively to a skin prick test and when symptoms persist throughout the year and no anatomic or medical disorders of the nose are present, the diagnosis points to NANIPER. No single diagnostic test, symptom, or clinical feature exists to demonstrate the presence of this disorder. Nor have distinct changes been found in cellular patterns in the epithelium and lamina propria of the nasal mucosa (4).

Nasal hyperreactivity is an increased sensitivity of the nasal mucosa to nonspecific stimuli and was first described by van Lier (5). Cold dry air (CDA) provocation has been proven by the Baltimore group to be an effective tool in quantifying the secretory response of hyperreactivity in persons susceptible to CDA (6, 7). In this population, significant changes were found in cell mediators and neurogenic reflex mechanisms. It was, however, not clear from which type of rhinitis these subjects suffered or what happened to other nasal parameters such as nasal patency or sneezing, important parameters when studying blockers, runners, and sneezers. Pulmonary CDA provocation of patients with asthma results in bronchial obstruction, and it is found to be a suitable method for assessing bronchial hyperreactivity (8).

Histamine and methacholine are the most widely used provocation methods for measuring the degree of nasal hyperreactivity today. These tests have not been validated in patients with NANIPER. The objective of the study was to validate and standardize a new intranasal CDA challenge method and compare it with that of histamine. Nasal patency, mucus production, and sneezing were studied after CDA and histamine challenge in patients with NANIPER and in control subjects. The dose dependency, sensitivity, specificity, and reproducibility of both challenge methods were compared.

METHODS

Patients and Control Subjects

Thirty-eight patients and seven control subjects were selected on the basis of inclusion and exclusion criteria (Table 1). These patients had experienced nasal complaints such as nasal obstruction, sneezing, and/or rhinorrhea for more than 1 yr. These symptoms could not be attributed to an atopic rhinitis, nasal or paranasal sinus infection, anatomic disorders affecting nasal function, pregnancy or lactation, and/or systemic disorders. Symptoms 2 wk prior to entry varied from moderate...
to severe (daily average of 2, on a scale of 0 to 4). Local and systemic corticosteroids or antihistamine treatment had been stopped 4 wk prior to the study, and xylometazoline had been stopped 6 hr prior to each visit. There was no history of xylometazoline abuse.

A subgroup of 16 (five male, 11 female) of the 38 patients and all seven control subjects (three male, four female) met additional requirements: they were nonsmokers and avoided physical or chemical irritants during the study (Group A). These additional restrictions were not imposed on a second group of 16 patients (Group B), randomly selected from the 22 remaining patients. The study was approved by the local medical ethical committee.

Study Design

The study had a randomized crossover controlled design. Patient Group A and the seven control subjects visited the hospital on 16 occasions. Participants were randomly assigned to start either with a CDA challenge period of 4 wk followed by a 4-wk histamine challenge period or vice versa. These periods consisted of three complete provocation series at weekly intervals. Subsequently, five daily repeat challenges followed in order to test for possible increased responsiveness after repeat challenges (see Table 2).

Patient Group B visited the hospital twice. After the run-in period, patients were subjected to one CDA challenge in order to assess whether patency, mucus production, and/or sneezing data of random patients were subjected to one CDA challenge in order to assess responsiveness after repeat challenges (see Table 2).

Table 2

<table>
<thead>
<tr>
<th>Run-in</th>
<th>CDA or Histamine</th>
<th></th>
<th>Histamine or CDA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Visits:</td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Days:</td>
<td>1</td>
<td>14</td>
<td>21</td>
</tr>
</tbody>
</table>

*Underlines = complete provocation series; boldface = provocation at predefined arbitrary threshold: average of the three previous threshold doses resulting in 0.5 g mucus production and 30% patency reduction.

Cold Dry Air

Subjects were allowed to acclimatize to room temperature (20°C) for 15 min. Patients then blew their noses gently to remove remaining secretions. The nose was then rinsed with N saline. A respiratory heat exchanger (Jäger GmbH, Würzburg, Germany) was modified slightly: Before cooling, the air was dried by passing it through a column with silicon granules. The temperature of the air leaving the system was filtered and entered the nose at −10°C with a relative humidity of < 10%. A purpose-designed nose cap (Respricare, The Hague, The Netherlands) ensured a tight fit. The dosage was increased in steps by analogy with the histamine series as follows: 12.5, 25, 50, 100, 200, and 400 L. This involved CDA provocation steps of 1, 1, 2, 4, 8, and 16 min with a flow of 12.5 for the first step and 25 L/min for the following steps. Instructions were given to inhale through the nose and exhale through the mouth during the challenge and to maintain normal breathing frequency as at rest. The provocation was stopped in case of hyperventilation. A posterior rhinomanometry (Rhinoscreen; Jäger) was repeated until no further decrease in patency was measured. Secretions were measured by weighing the amount of mucus produced per provocation step in preweighed paper tissues on a precision scale (Mettler, Würzburg, Germany) with an accuracy of 0.01 g. Sneeze were counted. Maximal effects per dose step were used for calculations. Repeat measurements were performed every time the dose was increased. Patency reduction and mucus production were determined and sneezes were counted in the following 5 min (see below). At the 400 L CDA step, patients were allowed to blow their nose after 8 min in order to correct for drying of nasal secretions during the 16-min provocation with CDA.
Histamine

Acclimatization and prerinsing with NaCl saline were the same as with the CDA challenges. Histamine was used in the concentrations 0.125, 0.25, 0.5, 1, 2, and 4 mg/ml (9-11). Measurements started 1 min after provocation and continued for 4 min, allowing adequate contact between histamine and the nasal epithelium.

Statistical Analysis

Thresholds of outcome parameters were calculated as the intersection of the dose-response curve with the effect cutoff point. Reproducibility and dose dependency were assessed for the outcome parameters “nasal patency” and “mucus production” separately by calculating reliability indices (RIs) (12) at several thresholds. The RI was defined as the ratio of intersubject variance to total variance. Sensitivity and specificity were determined at the “mean 100 dose” and the “mean 400 dose” (see Table 3) and compared for CDA and histamine using Fisher’s exact test.

Specificity was also calculated for maximal values and regression lines of the outcome parameters. Regression line analysis was achieved by pooling three consecutive provocation series and subsequent ln transformation per patient using Biomedical Package Software (BMDP-5V; BMDP Statistical Software, Inc., Los Angeles, CA). They represented the function: ln(y/y0) = -ax. A 2 regression lines started at the predefined value of 100%; a, the slope coefficient of this regression line, represented a decline or increase per doubling of the dose step. Differences between patients with NA-NIPER and control subjects and between provocation types were analyzed using the Mann-Whitney U test. Confidence analysis was performed using SPSS PC software (SPSS Inc., Chicago, IL).

A random coefficient model with an intersubject grouping factor in BMDP 5V software was used for (1) optimal curve estimation for outcome parameters, (2) determination of a possible change in nasal responsiveness after repeat provocations, and (3) calculations of correlation coefficients between clinical features and outcome parameters. The Mann-Whitney U test was used to compare carryover and treatment effects. Intrapatient and control totals for the two provocation periods were used to test the carryover effect. The differences between intrapatient and control totals for two provocation periods were used to test the treatment effect. We assumed that there was no significant carryover or treatment effect if the means of these totals did not significantly differ between the two provocation series. Medians of daily record chart data were analyzed with the Mann-Whitney U test in three ways: (1) symptom score on the same day, (2) on the previous day, and (3) as an average for the preceding week. Visual Analogue Scales for (1) patency reduction, (2) mucus production, and (3) sneezing were analyzed separately and as a total using the Mann-Whitney U test.

Table 3

<table>
<thead>
<tr>
<th>CDA dose</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>400 dose</td>
<td>The dose resulting in 40% reduction of nasal patency and/or 0.5 g mucus production (cutoff lines) in one CDA/histamine provocation series.</td>
</tr>
<tr>
<td>Mean 400 dose</td>
<td>The average of three repeat doses of which at least one resulted in a &gt; 400 dose after three repeat histamine or CDA challenge series performed weekly. A positive response demonstrates the degree of nasal CDA reactivity in patients with NA-NIPER. It is reproducible and sensitive but less specific.</td>
</tr>
<tr>
<td>Mean 100 dose</td>
<td>The dose &lt; 100 L CDA or 1 mg/ml histamine resulting in 40% reduction of nasal patency and/or 0.5 g mucus production (cutoff lines) in a CDA/histamine provocation series.</td>
</tr>
</tbody>
</table>

Figure 1. Optimal curve fits for observed patency results in percentages demonstrating intranasal CDA specificity and the lack of specificity after intranasal histamine challenge: Curves (a) and (b) versus (c), statistically significant different slope coefficients; curves (d) and (e) not statistically significant. External reproducibility of intranasal CDA challenge is shown: Curves (a) and (b), no statistically significant difference.
RESULTS

Dose Dependency

CDA challenges were tolerated well throughout the study. CDA provocation did not lead to sneezing of any importance. Histamine challenge provoked sneezing, but not in a reproducible or dose-dependent manner (data not shown). Nasal patency and mucus production were dose-dependent outcome parameters. To demonstrate and illustrate specificity, maximal effects and regression lines were also assessed (see Figures 1 and 2).

Threshold Doses

Reproducibility and positivity for nasal CDA challenge. Cutoff lines of 10, 20, 30, 40, and 50% patency reduction or 0.2, 0.3, 0.4, 0.5, 0.75, or 1.0 g mucus production were assessed. In order to achieve an accurate calculation of reproducibility, the first intersection of the dose-response line with the cutoff line was calculated and used to determine the reproducibility of the threshold dose. The test-retest reproducibility of one single CDA provocation was inadequate.

In order to increase reproducibility for intranasal CDA provocation, a series of three provocation series was performed. Threshold doses were calculated for three repeated provocations at intervals of 1 wk within a 2-wk period (mean 400 dose and mean 100 dose; see Table 3). The mean 400 dose represented the mean of three threshold doses given at intervals of 1 wk, which resulted in 40% nasal patency reduction and/or 0.5 g mucus production after one challenge series. This dose was used to evaluate intrapatient and interpatient differences. The mean 100 dose is similar to the mean 400 dose, with the added restriction that this dose is reached at or before 10 mL CDA or 1 mg/ml histamine in one of the three CDA or histamine challenge series. Reproducibility was calculated by determining the reliability index (RI) (12). Mean 400 dose and mean 100 dose thresholds for CDA provocations in patients with NANIPER resulted in a satisfactory RI of 76% for nasal patency and a RI of 86% for mucus production. The RIs of histamine challenge in patients with NANIPER for these thresholds were 65 and 92%, respectively. For control subjects, these RIs for patency and mucus production thresholds were, respectively, 32 and 65% after CDA and 69 and 77% after histamine challenge. Thresholds at the other cutoff points mentioned above resulted in lower RIs for CDA reactivity. A rule, histamine reactivity also resulted in lower RIs (results not shown). When patients reached the mean 100 dose they were considered to be CDA reactive. For statistical reasons, patients who did not reach the 400 dose were considered to have a threshold of 800 L CDA or 8 mg/ml histamine. When reaching the 400 dose or the 100 dose after N saline, 6.25 L CDA (or 0.0625 mg/ml histamine) was adopted as the basis for calculations.

Figure 2. 95% confidence intervals pooled over all dose steps demonstrating for mucus production and nasal patency a dose-response relationship. Less overlap between NANIPER Group A and the Control Group results for CDA- versus histamine-provoked mucus production and patency decrease is clearly visible.
Sensitivity, specificity, and degree of nasal CDA challenge. The first dose step resulting in the 400 dose was considered to be a positive reaction to intranasal CDA (Table 4). Sensitivity was calculated as the number of patients with NANIPIER responding positively to intranasal CDA divided by the total number of patients with NANIPIER x 100%. Specificity was calculated as the number of control subjects with one or more positive reactions (false positives) x 100%. The mean 400 dose resulted in a comparable sensitivity for CDA and histamine, 100% and 94%, respectively, whereas specificity was better for CDA at 42% as opposed to 0%. This difference in specificity was even more evident for the mean 100 dose (Table 5). The specificity of CDA compared with that of histamine was 71% as opposed to 0%. Sensitivity for the mean 100 dose was a little less when compared with the mean 400 dose at 87% as opposed to 94% (Table 5). For other histamine thresholds (for patency and mucus production separately, or for a combination of patency and mucus production) a combination of less sensitivity and specificity was found (results not shown).

The intrapatient standard deviations (SDs) for the doubled-dose provocation steps were determined in order to assess the degree of nasal reactivity to the mean 400 dose. Data for 15 of the 16 patients were available. For nasal patency, respectively, mucus production after CDA SDs were 1.1 and 0.9. For histamine provocation these SDs were 0.8 versus 1.3. The same SDs were found for the mean 100 dose.

In short, the presence of CDA reactivity in NANIPIER was best described by the mean 100 dose. This was a dose-dependent, reproducible, sensitive, and specific threshold dose. Specificity was in favor of CDA rather than of histamine (p < 0.05). Sensitivity was the same for both provocation methods (p = 0.35). The degree of nasal CDA reactivity in NANIPIER was best described by the mean 400 dose. This dose was reproducible and sensitive but less specific than the mean 100 dose.

Maximal Effects Over All Dose Steps and Regression Lines

The CDA challenge was superior to histamine in discriminating between patients with NANIPIER and control subjects for data of maximal effects on patency (CDA: p = 0.003; histamine: p = 0.052) and for the same data of mucus production (CDA: p = 0.001; histamine: p = 0.023) (see Table 6).

A significant difference was found between the patency slope coefficients of Group A and those of Group B patients with NANIPIER compared with control subjects (NANIPIER Group A versus control subjects: p = 0.04). No statistical significance was found between these curve slopes for histamine (p = 0.30). In the case of mucus production, the differences between patients with NANIPIER and control subjects were even more apparent than for nasal patency (CDA: p = 0.0021; histamine: p = 0.16) (see Table 6).

In short, CDA provocation—expressed as maximal effects and regression lines—resulted in better discrimination between patients with NANIPIER and healthy control subjects than did histamine provocation in terms of both nasal patency and nasal mucus production. This is shown by the confidence intervals for nontransformed data (Figure 2).

Variability. Variability after histamine provocation was greater in patients with NANIPIER than after CDA challenge (see Figure 3). In control subjects, however, histamine challenge resulted in lower variability than did CDA challenge.

External reproducibility. A second group of 16 patients with NANIPIER (Group B) was subjected to one CDA provocation session in order to assess whether the patency and mucus production data of other patients with NANIPIER (those who did not meet additional restrictions) corresponded to those of Group A (external reproducibility). After plotting the mucus production and patency data of patients from Group A, a best-fit function was found to describe the patency data only: ln(y/y_0) = -βx or y = y_0 e^{-βx}, where y = nasal patency (ml/s); y_0 = preprovoction patency value; x = dose step (preprovoction x = 0; 400 L CDA or 4 mg/ml histamine x = 6); β = slope coefficient.

Patency data after CDA provocation resulted in mean slope coefficients β for NANIPIER Groups A and B of 0.110 and 0.077, respectively. The difference between the groups (0.033) was not statistically significant (p = 0.14; 95% confidence interval: 0.022 to 0.045), thus demonstrating external reproducibility. For control subjects, β = 0.026. It was not possible to find an adequate function to describe mucus production.

Correlation between clinical parameters and nasal reactivity. (1) Prerinsing the nose with N saline, (2) patient’s perception

### Table 4

<table>
<thead>
<tr>
<th>Subjects</th>
<th>Age (yr)</th>
<th>Baseline Patency</th>
<th>Baseline Patency</th>
<th>CDA Dose in Doubling Dose Steps</th>
<th>Total IgE (E/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NANIPIER Group A, n = 16</td>
<td>44 ± 10.9</td>
<td>527.6 ± 163.5</td>
<td>518.2 ± 185.2</td>
<td>4.25 ± 1.78</td>
<td>3.93 ± 2.46</td>
</tr>
<tr>
<td>NANIPIER Group B, n = 16</td>
<td>37 ± 12</td>
<td>856.7 ± 139.5</td>
<td>ND</td>
<td>4.56 ± 1.63</td>
<td>ND</td>
</tr>
<tr>
<td>Control group, n = 7</td>
<td>30 ± 8.2</td>
<td>775.1 ± 205.5</td>
<td>792.8 ± 176.5</td>
<td>5.86 ± 2.26</td>
<td>6.57 ± 0.78</td>
</tr>
</tbody>
</table>

* Values are mean ± SD. Baseline patency and threshold doses are normally distributed within population groups in this study.
† Flow in ml/s at 150 Pa.
‡ 1 = 0.12 mg/ml histamine or 12.5 L CDA; 6 = 4 mg/ml histamine or 400 L CDA.

### Table 5

<table>
<thead>
<tr>
<th>Cold Dry Air</th>
<th>Histamine</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sensitivity</td>
<td>Specificity</td>
</tr>
<tr>
<td>Mean 100 dose</td>
<td>14/16 (87%)</td>
</tr>
<tr>
<td>Mean 400 dose</td>
<td>15/16 (94%)</td>
</tr>
</tbody>
</table>

* The sensitivity of the two challenge types is more or less equal. The specificity of histamine challenge, on the other hand, is far less than that of CDA provocation. Only combined patency and/or mucus production data were assessed. For patency and mucus production data taken separately, similar percentages were found (data not shown).
of the nasal symptoms as expressed by a daily record chart or visual analogue scale, and (3) the color of the nasal mucosa were not statistically significant correlated with the mean 400 dose or the mean 100 dose.

Tachyphylaxis: increased responsiveness, carryover, or treatment effect? The responses to 100 dose and 400 dose (see Table 3) of CDA/histamine provocation series on Day 5 were compared with the responses to the mean 100 dose and mean 400 dose administered during the previous 2 wk. No statistically significant change was noted in the response to the 100 dose and the 400 dose during the CDA provocation period. Histamine provocation, however, gave an increased responsiveness effect for the 100 dose and the 400 dose ($p < 0.025$). A carryover effect was not found ($p = 0.13$). Nor was any treatment effect of histamine after CDA or vice versa found ($p = 0.27$).

### DISCUSSION

This study has shown that, unlike histamine, intranasal CDA provocation is a reliable method for measuring nasal reactivity symptoms in patients with NANIPER in a dose-dependent and, for patients with NANIPER, recognizable way. CDA challenge is a natural stimulus, although exposure conditions are artificial. CDA is thought to provoke neurogenic reflex mechanisms in the nasal mucosa (2, 3). In this study, the assessment of intranasal CDA reactivity is based on nasal patency and mucus production as the two main outcome parameters, making the method very suitable for studying nasal hyperreactivity in patients with NANIPER (blockers and runners). Although time-consuming, it has been found to be a well tolerated, safe, and easy to dose provocation method. Its relation to histamine and the separate assessment of patency

### TABLE 6

RESULTS FOR THE ALTERNATIVE OUTCOME PARAMETERS: MAXIMUM EFFECT AND REGRESSION LINE ANALYSIS*

<table>
<thead>
<tr>
<th>Maximum Effect</th>
<th>Regression Line</th>
<th>95% Confidence</th>
<th>95% Confidence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patency</td>
<td>Mucus Production</td>
<td>Patency</td>
<td>Mucus Production</td>
</tr>
<tr>
<td>CDA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Patient Group A</td>
<td>0.51</td>
<td>0.72</td>
<td>-8.13</td>
</tr>
<tr>
<td>Control group</td>
<td>0.26</td>
<td>0.27</td>
<td>-2.70</td>
</tr>
<tr>
<td>p Values</td>
<td>p = 0.003</td>
<td>p = 0.001</td>
<td>p = 0.04</td>
</tr>
<tr>
<td>Histamine</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Patient Group A</td>
<td>0.67</td>
<td>1.53</td>
<td>-9.17</td>
</tr>
<tr>
<td>Control group</td>
<td>0.42</td>
<td>0.94</td>
<td>-6.30</td>
</tr>
<tr>
<td>p Values</td>
<td>p = 0.052</td>
<td>p = 0.023</td>
<td>p = 0.30</td>
</tr>
</tbody>
</table>

* The specificity of CDA provocation is shown by the significant differences between patients with NANIPER and control subjects. Histamine challenge failed to make this distinction. Note that a 0.071 g increase of mucus production per dose step after CDA challenge indicates NANIPER.

![Figure 3](image)

**Figure 3.** Box plot diagrams demonstrating a large variability of patency results of patients with NANIPER after histamine provocation compared with CDA provocation. In control subjects no significant differences were found between CDA provocation doses and prechallenge values, whereas histamine showed a significant decrease. *$p < 0.05$, **$p < 0.01$ (paired $t$ test).
Patients with NA\textsc{iper} are an interesting group to study because no single test has been developed until now for monitoring changes in this condition. A common and characteristic feature of patients with NA\textsc{iper} is nasal hyperreactivity. Until now, the most common diagnostic test for measuring nasal hyperreactivity was intranasal histamine provocation. Histamine provocation, however, fails to differentiate between patients with NA\textsc{iper} and control subjects. CDA is able to make this distinction. We therefore find that CDA is superior to histamine challenge for diagnosing NA\textsc{iper}. CDA challenge could be an alternative to histamine in characterizing nonspecific nasal hyperreactivity in other subgroups of rhinitis, examples being allergic (9), nonallergic (10, 15), and infectious (14) rhinitis.

A cigarette smoke (16) and ozone (17) have been proven to induce airway hyperreactivity, we tried to eliminate these factors in the first group of patients with NA\textsc{iper} (Group A), as mentioned in Table 1. We were able to demonstrate corresponding results for nasal patency in a second NA\textsc{iper} group without these additional restrictions (Group B), indicating that in this study these environmental factors were not of major importance.

A reproducible measure for intranasal CDA provocation was difficult to establish. Sneezing, one of the three outcome parameters, was not useful. Several criteria for mucus production and patency such as maximal effects, regression line coefficients, and threshold doses at several cutoff points were assessed. Single measurements did not result in a reproducible measure. In order to achieve acceptable reproducibility and sensitivity in both histamine and CDA challenges, requiring as little time as possible, repeated measurements expressed as mean 100 dose and mean 400 dose (see Table 3) were needed. However, specificity was acceptable only in the case of CDA challenge. It was not possible to achieve comparable specificity for histamine challenge using other threshold doses.

In this study, three complete provocation series were performed. This study has demonstrated that the 100 dose is indicative of the presence of CDA reactivity. A soon as the 100 dose is reached, the subsequent dose steps in the series are unnecessary. The degree of CDA reactivity will be indicated by the mean 400 dose. It will be possible to note changes in CDA reactivity, bearing in mind intrasubject variability.

Repeated daily CDA or histamine provocations for 5 d did not lead to increased responsiveness for CDA, but it did so for histamine. Some studies (18, 19) report unchanged nasal responsiveness to histamine after repeated histamine challenges, but these studies are difficult to compare to ours. Gronburg and colleagues (18) rechallenged nonrhinitic subjects after 1 h and 1 d, whereas Ohm and Juto (19) rechallenged nonallergic rhinitis subjects on three to seven occasions, sometimes at intervals of several weeks. A possible explanation for our finding that repeated histamine challenges lead to increased responsiveness could be that we used five consecutive daily provocations and that these resulted in increased inflammation. Söderberg and colleagues (20) found that bronchial histamine challenges lead to an increase in inflammatory cells in bronchoalveolar fluid 24 h after the inhalation of histamine in normal subjects.

Surprisingly, several clinical parameters such as symptom scores and visual analogue scales correlated poorly with CDA and histamine reactivity in patients with NA\textsc{iper}. In patients with perennial allergic rhinitis, however, De Graaf and colleagues (21) found significant correlations between nasal responsiveness to histamine and symptom scores for the preceding 2 wk. Hallen and Juto (22) found reasonable correlations between symptom scores for the preceding 2 wk and histamine reactivity of nasal mucosa in nonallergic nasal-hyperreactive patients. The reason for the conflict with our findings could be that the thriceweekly CDA/histamine provocations interfered with the natural symptomatology of NA\textsc{iper}.

Earlier biopsy studies in patients with NA\textsc{iper} did not support the hypothesis of cellular inflammation (4). This emphasizes the need for further investigation of neurogenic structures and mediators such as neuropeptides in hyperreactive nasal mucosa. Further studies evaluating the effects of CDA/histamine challenge on these parameters are in progress. This new standardized intranasal CDA provocation method uses a recognizable natural nonspecific stimulus and seems to be more suitable than histamine for characterizing and assessing the presence and degree of nasal reactivity in NA\textsc{iper}.

Acknowledgment: The writers wish to thank Prof. C. D. A. Verwoerd for valuable comments, and Miss E. van Schalk for assisting with patient matters.

References


