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# Dynamic and structural properties of interphase chromatin mapped in vivo with fluorescence correlation spectroscopy and quantitative modelling

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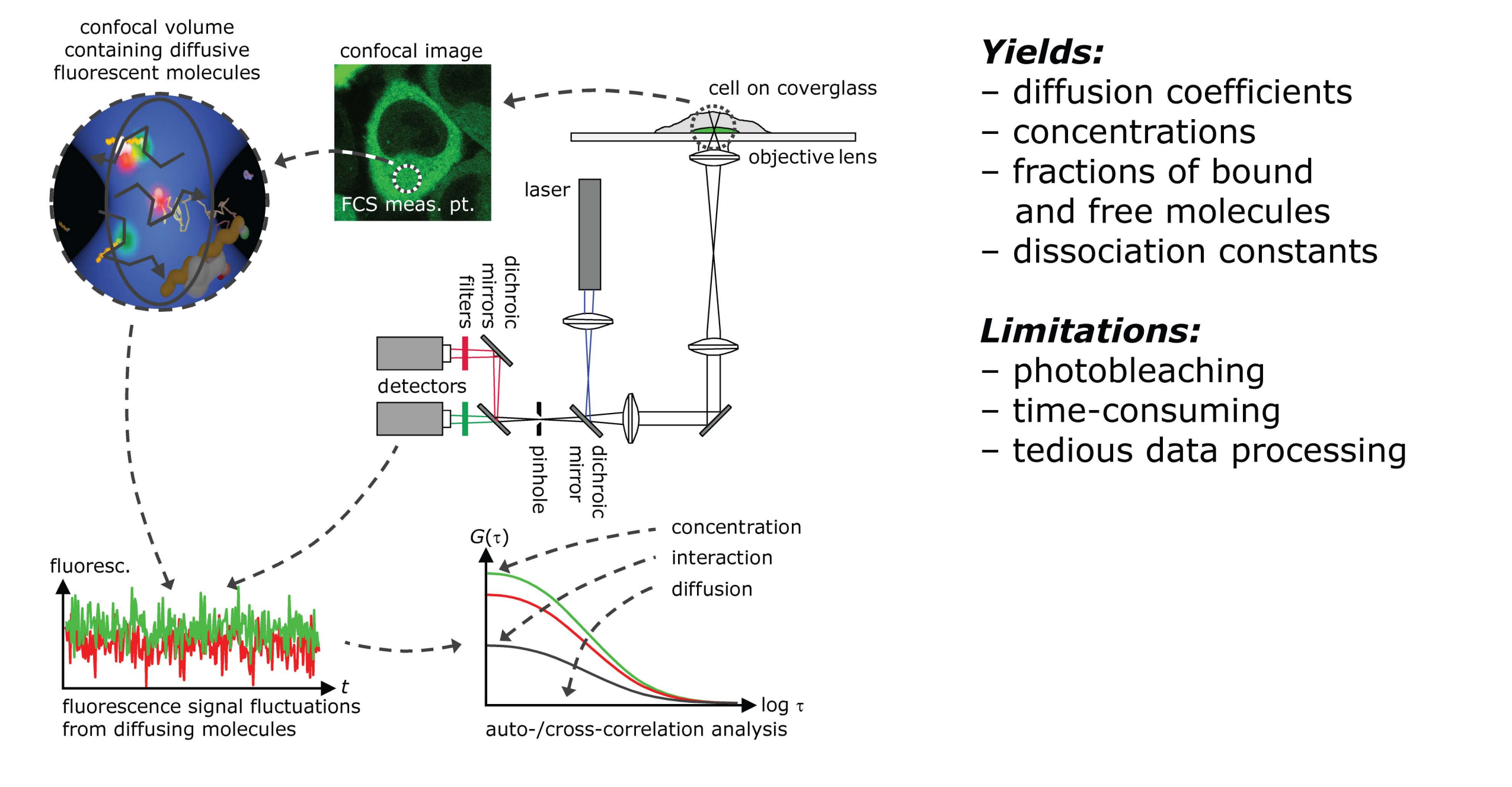
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## Abstract

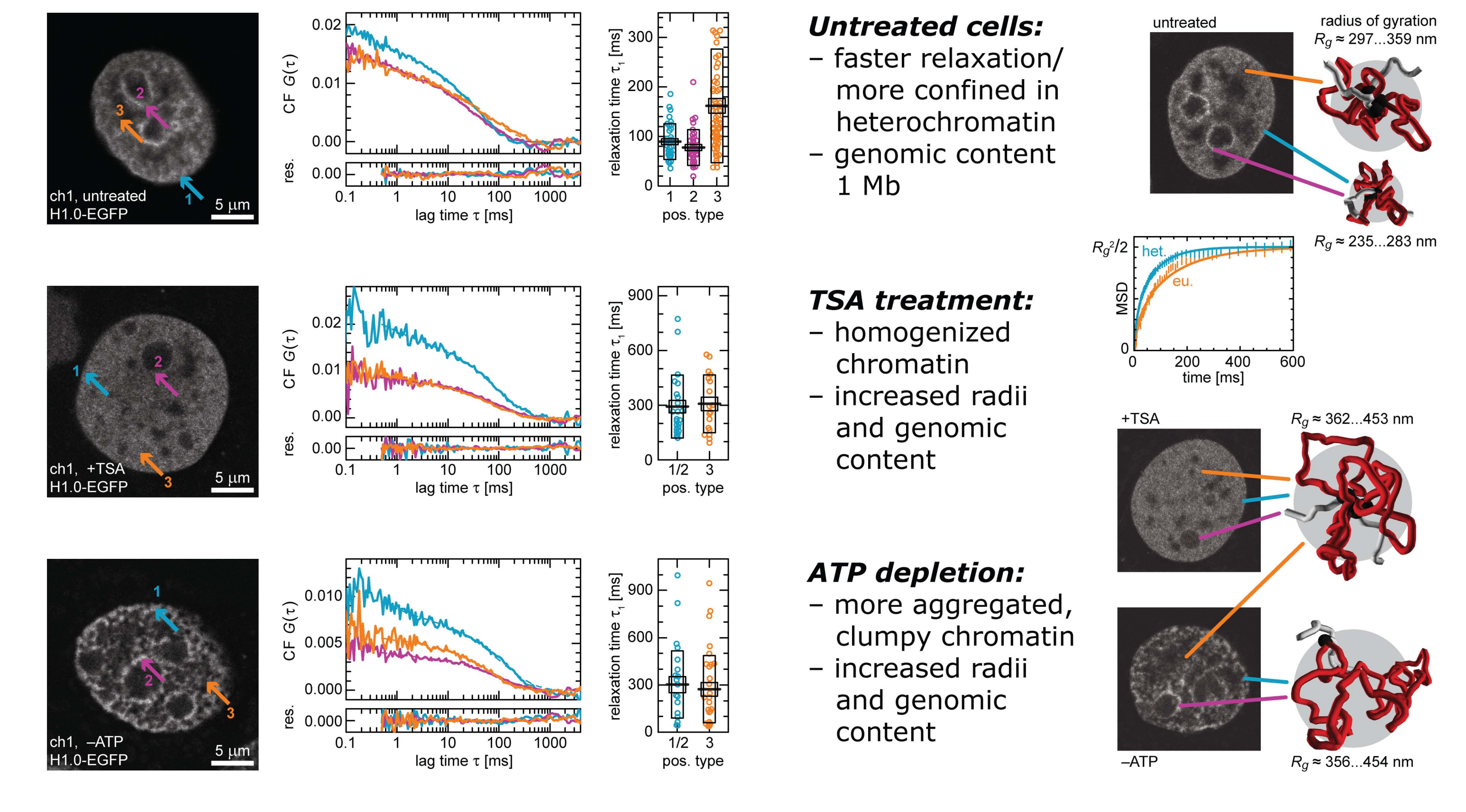
The three-dimensional organization of chromosomes of eukaryotic interphase cells is emerging as an important parameter for the regulation of storage, replication and expression of the genome. While a range of techniques like electron microscopy (EM), fluorescence in situ hybridization (FISH), or chromosome conformation capture techniques (3C, 4C, 5C, HiC, or the novel targeted chromatin capture, T2C) have been very successfully used to study chromosomal architecture, many details of structural and especially of dynamic properties remain unresolved.

Here, we present a novel approach to dissect intramolecular polymer dynamics from fluorescence intensity fluctuations measured with fluorescence correlation spectroscopy (FCS) to investigate the higher order chromatin dynamics in living cells. Using fluorescently tagged linker histone H1 and core histones H2A and H2B as tracer molecules, we find distinct chromatin relaxation times of  $\sim 160$  ms for open and  $\sim 90$  ms for dense chromatin areas, corresponding to radii of gyration of 240 and 300 nm for the topologically independent chromatin units. According to their genomic content of  $\sim 1$  Mb, these units correspond to the distinct topologically associating domains (TADs) found recently by 3C-derived techniques and to sub-chromosomal domains seen earlier by FISH and in vivo chromatin labelling. We have also obtained light-sheet microscopy-based FCS maps of chromatin domain dynamics. Based on these results, we have developed a quantitative analytical and numerical model of chromatin dynamics that provides access to mass density, persistence length and topological information of chromatin. It allows to extract these parameters from dynamics and 5C/HiC results, to predict chromatin conformation and distance data, and to identify complex looping as crucial for domain formation. Data and model suggest the existence of several connected loops of  $\sim 100$  kb each per domain. Especially in combination with the recently developed highly selective T2C method (see abstract T.A. Knoch & M. Wachsmuth, "Determination of the three-dimensional organization of chromatin by modelling-supported targeted chromosomal interaction capture (T2C)", which provides very good signal-to-noise ratio at high genomic resolution, we present a comprehensive systematic approach for the understanding of chromatin dynamics and its relation to structure as well as for insight into the impact on gene regulation.

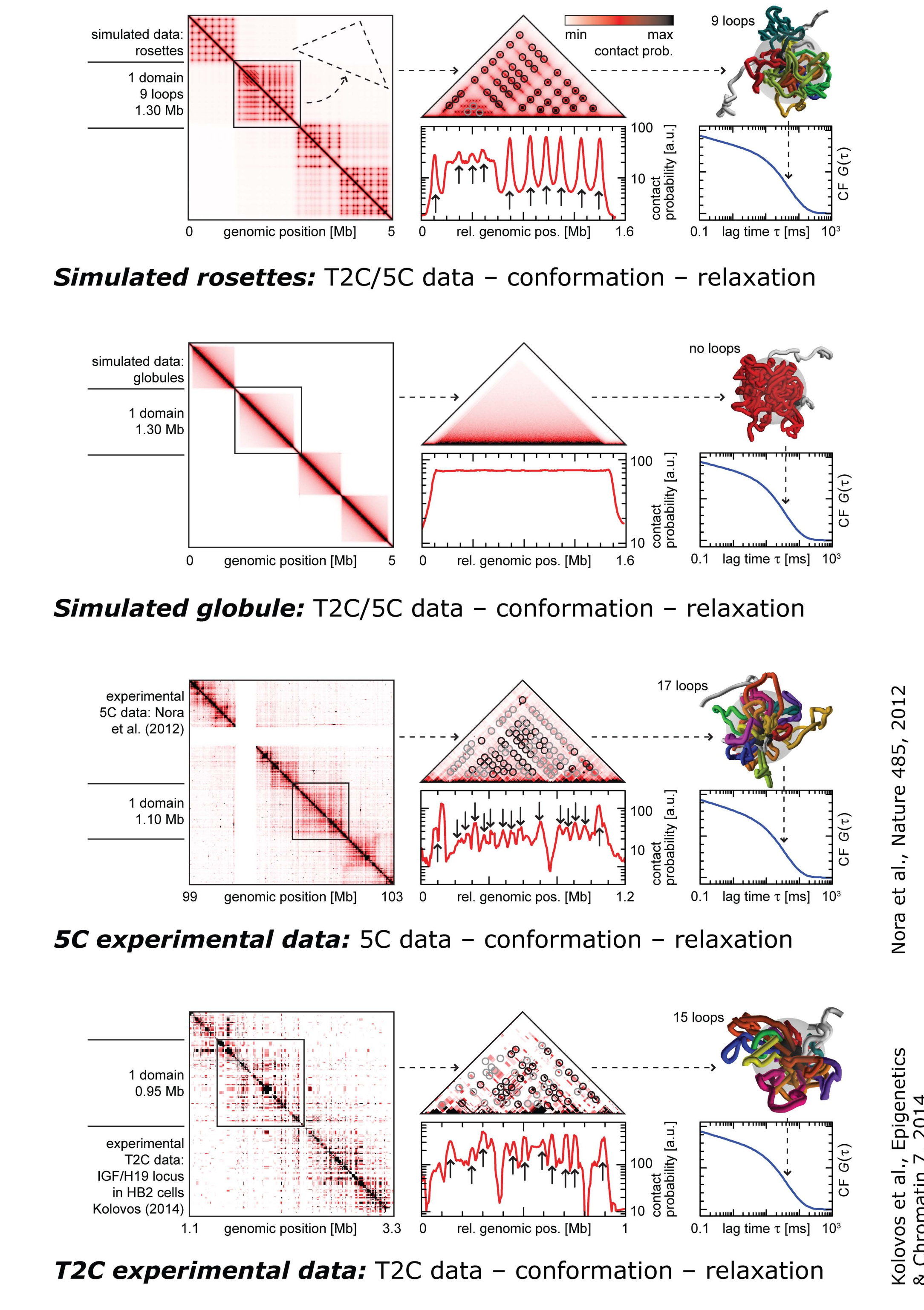
## Confocal fluorescence correlation spectroscopy (FCS)



## Polymer models: 1 Mb content and 240-300 nm size of domains

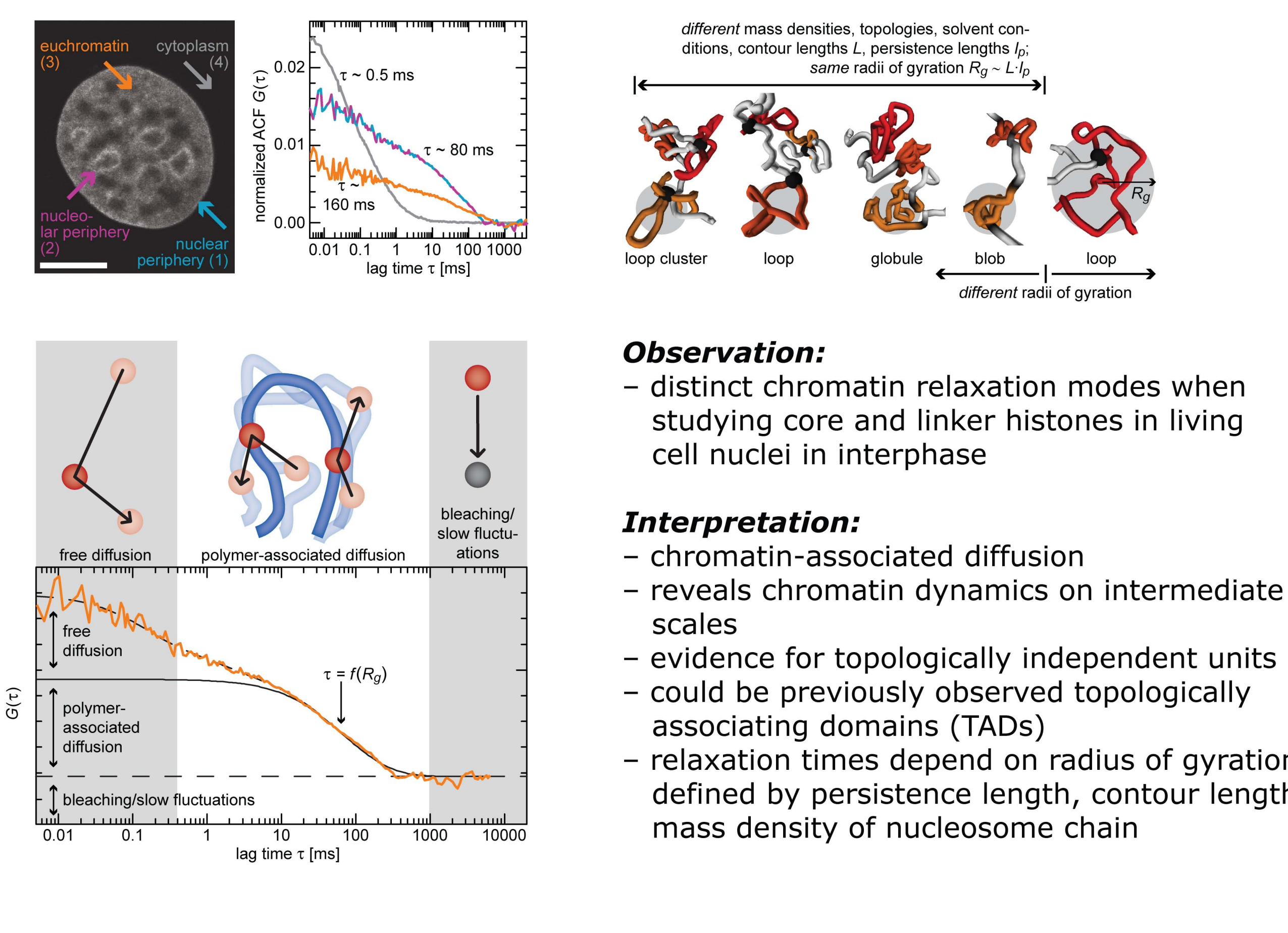


## Rosettes or globules or ...?

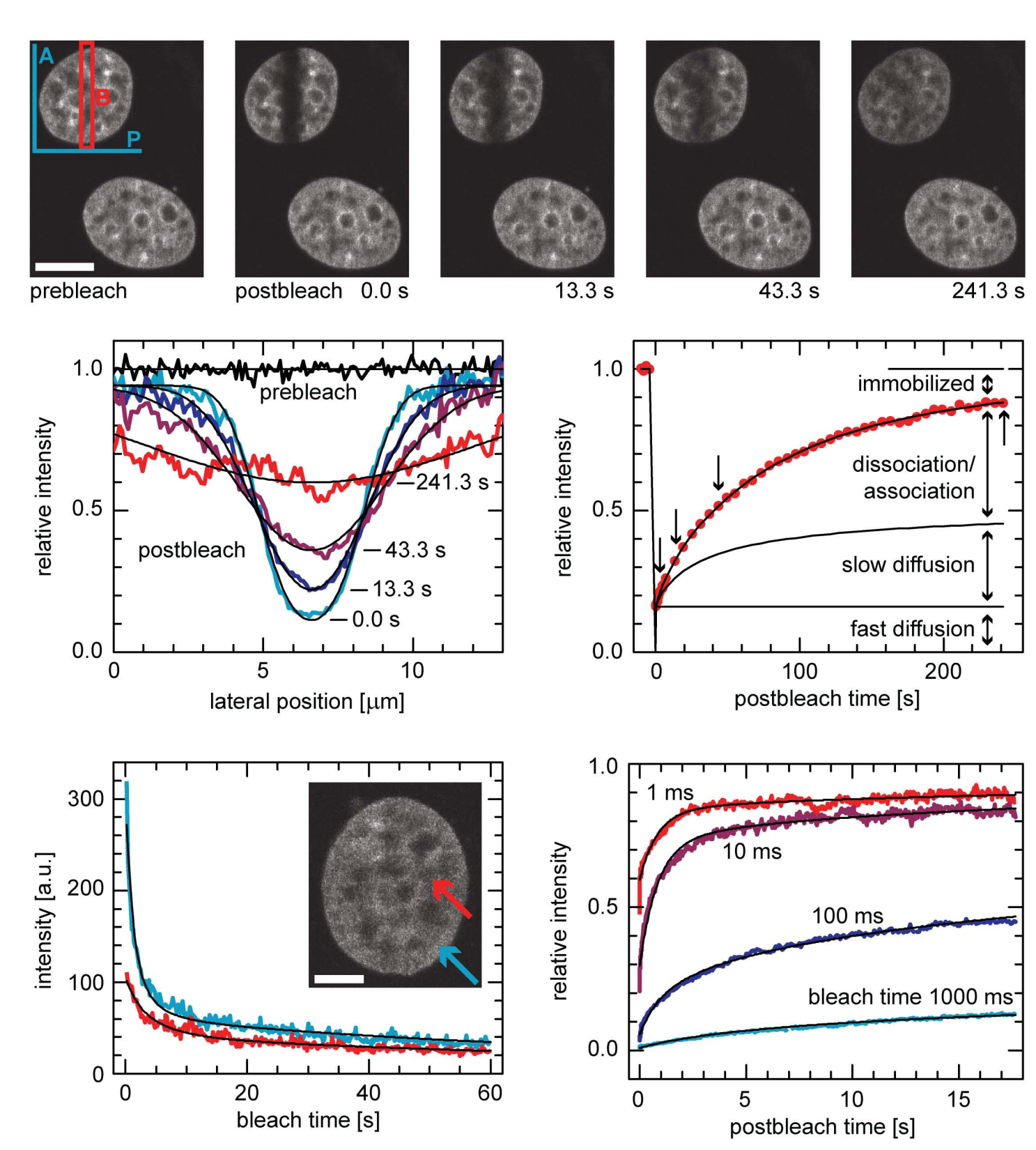


Nora et al., Nature 485, 2012  
Kolovos et al., Epigenetics & Chromatin 7, 2014

## Distinct chromatin relaxation modes reveal indynamically and topologically independent domains



## H1.0 binding is slower than the observed relaxations

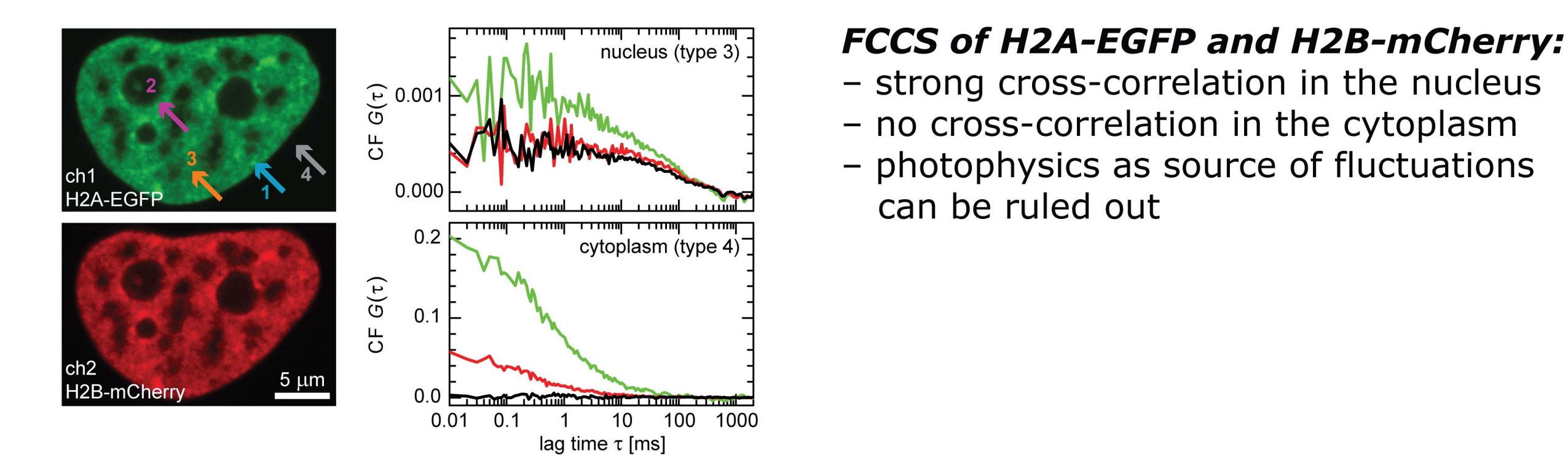


**Fluorescence recovery after/ continuous photobleaching:**

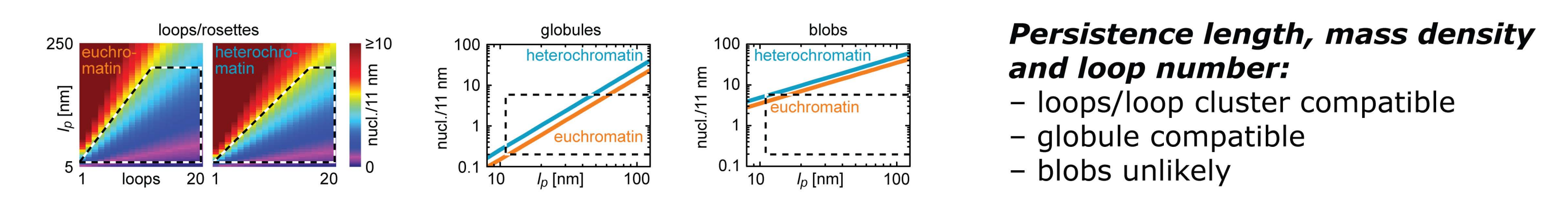
- three-step binding kinetics
- 1st step: mean dwell time  $\sim 1$  s
- 2nd step: switching to longer-lived state
- 3rd step: mean dwell time  $\sim 120$  s

**Interpretation:** overall, association/dissociation is far slower than the observed relaxations

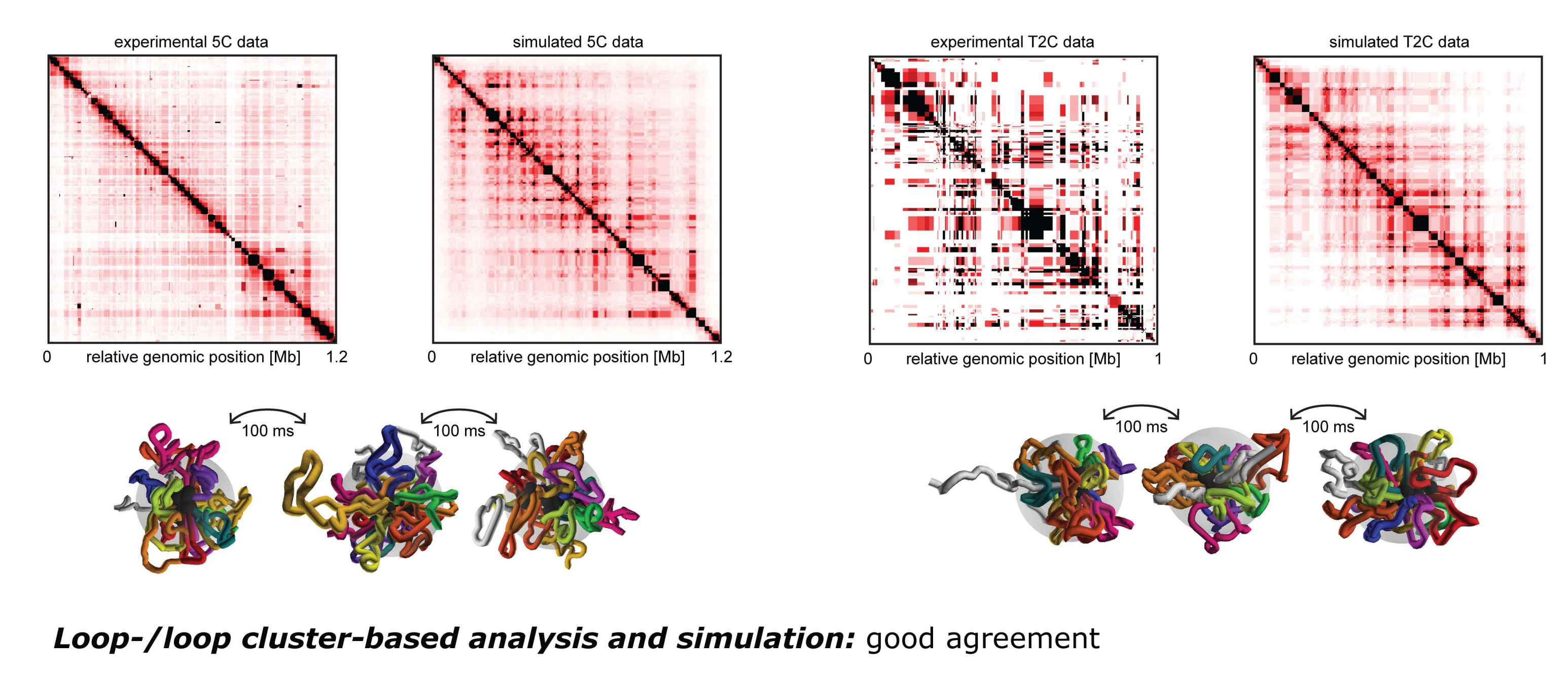
## Cross-correlations of core histones: no photophysics



## Physical parameter space for different conformations



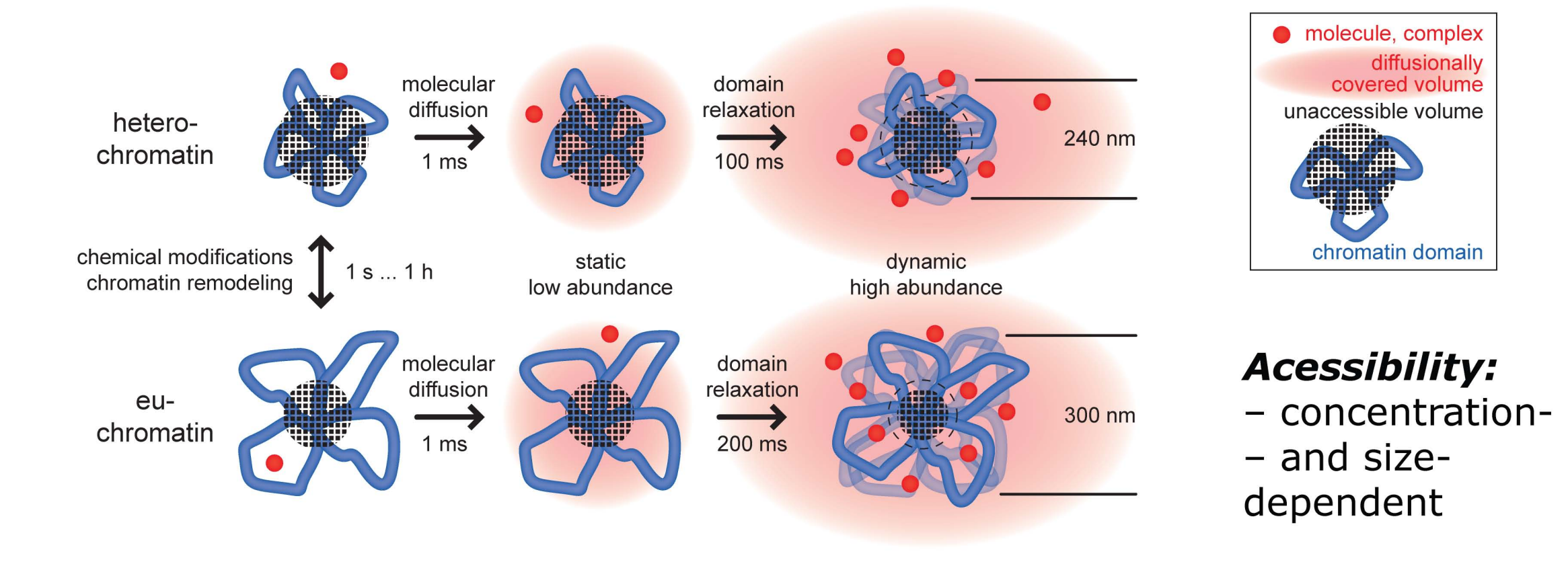
## Analysis and simulations of contact probability maps



## Summary

- The observation of distinct chromatin relaxations indicates the existence of dynamically and topologically independent domains
- Chromatin fiber dynamics provides additional constraints for domain architecture

## Differential regulation of chromatin accessibility



# Dynamic and Structural *in vivo* Properties of Interphase Chromatin

Mapped with

## Fluorescence Correlation Spectroscopy (FCS) and Quantitative Modelling

Wachsmuth, M. & Knoch, T. A.

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6th – 10th December, 2014.*

### *Abstract*

The three-dimensional organization of chromosomes of eukaryotic interphase cells is emerging as an important parameter for the regulation of genes as well as the storage, replication and expression of genetic information in general. Whereas a couple of techniques like electron microscopy (EM), fluorescence *in situ* hybridization, or chromosome capture techniques as 3C, 4C, 5C, HiC, or the novel T2C have been used for researching genomic architecture, its dynamics has remained underexplored. Here, we present a novel approach to dissect intramolecular polymer dynamics from fluorescence intensity fluctuations measured with fluorescence correlation spectroscopy (FCS) to investigate the higher order chromatin dynamics in living cells. Using fluorescently tagged linker histone H1 and core histones H2A and H2B as tracer molecules, we found distinct chromatin relaxation times of ~160 ms for open and ~90 ms for dense chromatin areas, corresponding to radii of gyration of 240 and 300 nm for the topologically independent chromatin units. According to their genomic content of ~1 Mb, these domains correspond to subchromosomal domains, already seen by FISH, and histone GFP labelling beforehand and correspond to so called distinct topologically associating domains (TADs). This is an impressive advancement in the field, since details on the dynamic and structural properties of subchromosomal domains/TADs in living cells are scarce.

Based on these results, quantitative analytical and numerical modelling provided access to mass density, persistence length and topological information of chromatin. It allowed to extract these parameters from 3C/5C/HiC/T2C results, to predict chromatin conformation and distance data, and to identify complex looping as crucial for domain formation. Especially, in combination with T2C, a recently developed selective high-throughput high-resolution chromosomal interaction capture technique (see abstract T.A. Knoch & M. Wachsmuth, Determination of the three-dimensional organization of chromatin by modelling-supported selective chromosomal interaction capture (T2C)), which provides very good signal-to-noise ratio, we present a comprehensive systematic approach for the understanding of chromatin dynamics and structure as well as for insight into their impact on gene regulation. As an outlook, we show light-sheet microscopy-based FCS maps of chromatin domain dynamics.

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### Keywords:

Genome, genomics, genome organization, genome architecture, structural sequencing, architectural sequencing, systems genomics, coevolution, holistic genetics, genome mechanics, genome statistical mechanics, genomic uncertainty principle, multilism genotype-phenotype, genome function, genetics, gene regulation, replication, transcription, repair, homologous recombination, simultaneous co-transfection, cell division, mitosis, metaphase, interphase, cell nucleus, nuclear structure, nuclear organization, chromatin density distribution, nuclear morphology, chromosome territories, subchromosomal domains, chromatin loop aggregates, chromatin rosettes, chromatin loops, chromatin quasi fibre, chromatin density, persistence length, spatial distance measurement, histones, H1.0, H2A, H2B, H3, H4, mH2A1.2, DNA sequence, complete sequenced genomes, molecular transport, obstructed diffusion, anomalous diffusion, percolation, long-range correlations, fractal analysis, scaling analysis, exact yard-stick dimension, box-counting dimension, lacunarity dimension, local nuclear dimension, nuclear diffuseness, parallel super computing, grid computing, volunteer computing, polymer model, analytic mathematical model, Brownian Dynamics, Monte Carlo, fluorescence *in situ* hybridization (FISH), targeted chromatin capture (T2C) confocal laser scanning microscopy, fluorescence correlation spectroscopy, spatial precision distance microscopy, super-resolution microscopy, two dimensional fluorescence correlations spectroscopy (2D-FCS) auto-fluorescent proteins, CFP, GFP, YFP, DsRed, fusion protein, *in vivo* labelling, information browser, visual data base access, holistic viewing system, integrative data management, extreme visualization, three-dimensional virtual environment, virtual paper tool.

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