Novel Leptin Receptor Mutations Identified in Two Girls with Severe Obesity Are Associated with Increased Bone Mineral Density

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Established Facts

- Leptin receptor (LEPR) deficiency causes hyperphagia and severe early-onset obesity.
- Other features such as hypogonadotropic hypogonadism, reduced growth hormone secretion, hypothalamic hypothyroidism and altered immune function are more variable.
- Leptin has been implicated as an important factor in bone metabolism.

Novel Insights

- Two girls with LEPR deficiency have a high bone mineral density, as assessed by dual-energy X-ray absorptiometry, which may be directly or indirectly related to leptin resistance.
- Bone age is remarkably advanced in the prepubertal girl, suggesting leptin resistance might also favor bone maturation.

Key Words

Leptin · Leptin receptor · Obesity · Hypogonadotropic hypogonadism · Bone mineral density

Abstract

**Background:** Recessive mutations in the leptin receptor (LEPR) are a rare cause of hyperphagia and severe early-onset obesity. To date, the phenotype has only been described in 25 obese children, some of whom also had altered immune function, hypogonadotropic hypogonadism, reduced growth hormone secretion, hypothalamic hypothyroidism or reduced adult height. We provide a detailed description of the phenotype of 2 affected girls to add to this knowledge. **Methods:** Whole-exome sequencing and targeted sequencing were used to detect the LEPR mutations. RNA analysis was performed to assess the effect of splice-site mu-
tions. Results: In 2 unrelated girls with severe obesity, three novel LEPR mutations were detected. Longitudinal growth data show normal childhood growth, and in the older girl, a normal adult height despite hypogonadotropic hypogonadism and the lack of an obvious pubertal growth spurt. Bone age is remarkably advanced in the younger (prepubertal) girl, and bone mineral density (BMD) is high in both girls, which might be directly or indirectly related to leptin resistance. Conclusion: The spectrum of clinical features of LEPR deficiency may be expanded with increased BMD. Future observations in LEPR-deficient subjects should help further unravel the role of leptin in human bone biology.

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Introduction

Leptin is a key regulator of energy homeostasis. It is produced by adipocytes and signals information on the amount of energy stored as body fat to the brain. It binds to the leptin receptor (LEPR), which is expressed in the hypothalamus, where it influences satiety and plays an important role in the regulation of the hypothalamus-pituitary-gonadal axis. LEPR is also expressed in other brain regions such as the cortex, midbrain and hindbrain, where its role is not completely clear yet [1].

Recessive mutations of LEPR are a rare cause of hyperphagia and severe early-onset obesity [2, 3]. To date, the phenotype has only been described in 25 children [2-10]. All were severely obese; in some subjects, additional features were reported, e.g. (clinical features of) altered immune function/T-cell numbers [3, 4, 7, 10], hypogonadotropic hypogonadism [3, 6, 8], reduced growth hormone (GH) secretion [2, 8], hypothalamic hypothyroidism [2, 8], and reduced adult height [3]. No data are available on bone health in these patients, although LEPR is known to be involved in bone metabolism [11, 12].

Here, we describe the clinical and laboratory features of 2 girls with LEPR deficiency to further delineate the phenotype in childhood and adolescence. We show normal childhood growth, normal adult height in the oldest girl, and high bone mineral density (BMD) in both girls.

Case Reports

Patient 1
The girl was born at term with a birth weight of 3,560 g. She suffered from pyelonephritis at the age of 4 months but experienced no other serious infections. She showed normal psychomotor development. From infancy, her appetite seemed insatiable and she gained weight quickly (fig. 1a–d). Despite a restricted diet and exercise program, she remained severely obese. Weight at the age of 9.0 years was 75.7 kg (+3.4 SDS weight-for-height), BMI 33.1 (+4.6 SDS), with a lean body mass of 40.5 kg, a fat mass of 32.5 kg and a fat percentage of 43.7% measured by dual-energy X-ray absorptiometry with the Hologic Discovery A system (Hologic Inc., Waltham, Mass., USA). Growth accelerated between the age of 1 and 4 years, followed by stable growth along +2.3 SDS at the upper border of the target height (TH) range (+0.8 SDS ± 1.6) but close to maternal height SDS (185.2 cm, +2.4 SDS). Pubic hair first appeared at the age of 8.5 years; however, breast development had not started by the age of 9 years.

The family is of Dutch origin, with all grandparents being Dutch. The patient’s father is obese (height 181 cm, weight 123 kg, BMI 37.5 at the age of 49 years) but had normal weight as a child. Her mother (height 186 cm, weight 76 kg, BMI 22.0 at the age of 38 years), sister (height 152 cm, weight 40 kg, BMI –0.1 SDS at the age of 12 years) and brother (height 122 cm, weight 25 kg, BMI +1.2 SDS at the age of 6.5 years) are not overweight.

Laboratory investigations at the age of 9.5 years showed a serum leptin level of 67.2 μg/l, which is above the reference range based on individuals with a normal BMI (3.7–11.1 μg/l) but within the range found in obese children (52.5 ± 20.2 μg/l in 151 children at the age of 12.3 ± 2.4 years and with a BMI z-score +2.7) [13]. Thyroid function, plasma insulin-like growth factor 1 (IGF1; +1.1 SDS), fasting glucose, liver enzymes, total and LDL cholesterol were normal, and serum LH, estradiol, testosterone, FSH and androstenedione were consistent with prepubertal status. Fasting insulin was mildly elevated (23 mU/l), triglycerides were high (1.54 mmol/l, >95th centile) and HDL cholesterol was low (0.79 mmol/l, <5th centile for age).

Bone age (BA) according to Greulich and Pyle [14], however, was remarkably advanced [12 years at a calendar age (CA) of 8.6 years], and dual-energy X-ray absorptiometry showed BMD z-scores of +2.7, +3.1 and +3.4 at the lumbar spine (LS), neck area of the left hip and of the right hip, respectively (reference data for BMD are from NHANES). LS bone mineral apparent density (BMAD) z-scores were +1.9 at the age of 8.6 years and +2.3 at the age of 6.7 years, calculated as described by Ward et al. [15].

Patient 2
The girl was born at term with a birth weight of 3,600 g. Her development was normal, although walking was somewhat delayed. Parents reported a normal appetite and food intake throughout childhood. However, her weight increased dramatically after the age of 1 year, and various diets and exercise programs had no effect. She had no health complaints and grew along her TH SDS (fig. 1e, f). Breast development started around the age of 11 years, but menarche had not yet occurred at the age of 15.2 years (maternal menarcheal age was 14 years). Height was 170.3 cm (+0.5 SDS, close to the TH of 168.4 cm), weight was 110.3 kg, BMI was 38 (+3.8 SDS), and Tanner stage was B5P5A3. Pubic hair was Tanner 5, but remarkably sparse.

The family is of Dutch origin, with all grandparents being Dutch. Her father is overweight (height 180 cm, weight 87 kg, BMI 26.9). Her mother (height 168 cm, weight 57 kg, BMI 20.2) and sister (height 136 cm, weight 32 kg, BMI +0.8 SDS at the age of 9 years) are not overweight.

Fasting glucose was normal with elevated insulin (36.7 mU/l), and an oral glucose tolerance test revealed impaired glucose toler-
Fig. 1. a, b Photographs of patient 1 at the age of 6 years and 8 months, showing severe obesity but no dysmorphic features. c–f Growth charts. c Height for age of patient 1, showing early growth acceleration followed by steady growth at the upper border of the TH range. The shaded area represents −2 to +2 SDS. TH is indicated by ‘TH’ on the right hand side of the graph. Note the advanced BA, which is indicated by a dot connected by a horizontal line to the height data point. d Weight for height of patient 1, showing early-onset obesity. e Height for age of patient 2, showing normal growth but no pubertal growth spurt. f Weight for height of patient 2.
BMAD z-score was +1.7, were +1.3 and +2.0 at the LS and proximal left femur, and the LS cleoSpin were +1.3 and +2.0 at the LS and proximal left femur, and the LS cleoSpin and proximal left femur, and the LS cleoSpin and the LS BMAD z-score was +1.7 [15].

Methods

Genomic DNA was isolated from peripheral blood samples using the AUTOPURE LS Instrument (Gentra Systems, Minneapolis, Minn., United States). Cytogenetic microarray analysis was performed using the Affymetrix CytoScan HD Array according to the manufacturer’s procedures. Copy number was assessed in the proband using ChAS software (Chromosome Analysis Suite; Affymetrix, El Segundo, Calif., USA). Sanger sequencing of the complete coding region including intron-exon boundaries was performed using standard procedures (primer sequences available upon request). Exomes were captured by the NimbleGen SeqCap EZ V2 kit, followed by Illumina paired-end sequencing (2 x 100 bp) with at least x40 mean coverage. Whole-exome sequencing data were generated in the Human Genotyping Facility of the Genetic Laboratory at the Department of Internal Medicine, Erasmus Medical Centre. Downstream analysis was performed with an in-house pipeline [16].

Two peripheral blood cultures for each patient were set up for RNA analysis: one with and one without cycloheximide. RNA was isolated with the RNA-Bee (Bio-connect B.V., Huissen, The Netherlands) method using Nucleospin® RNA II columns (Nucleospin® RNA II kit; Macherey-Nagel, Düren, Germany) according to the manufacturer’s instructions. RT-PCR was performed with home-made primers (sequences available upon request), and Sanger sequencing was performed according to standard procedures. The investigations in patient 1 were approved by the Medical Ethics Committee of the Leiden University Medical Hospital (P06.118), and the investigations in patient 2 were performed in the context of routine patient care. Informed consent was obtained from patients and parents.

Results

In both patients, a monogenic cause of obesity was suspected because of severe early-onset obesity and lean family members. In patient 1, whole-exome sequencing was performed in the index, her parents and two siblings. This revealed two novel LEPR mutations in the index, a splice-site mutation c.1753–1dupG in intron 13 and a missense mutation c.2168C>T p.Ser723Phe in exon 16. RNA analysis of the splice-site mutation r.1753–1dupG showed that the splice-acceptor site shifted one nucleotide, causing the retention of an extra nucleotide in the mRNA and a frameshift with a premature stop codon p.Met585Aspfs*2 (fig. 2, 3). The substituted serine (p.Ser723Phe) is a highly conserved amino acid (conserved in 11 species) in the fibronectin type III domain. The mutation of this residue is predicted to be pathogenic by in silico prediction programs [SIFT: deleterious (score 0.0; median 3.38); PolyPhen: probably damaging; HumDiv: score 1.000; HumVar: score 0.999; Mutation Taster: disease causing a p value of 1.0]. The mother and sister carry the missense mutation, the brother has two wild-type alleles and the father carries the splice mutation.

A CytoScan in patient 2 showed a paternal interstitial duplication on chromosome 2 containing part of the CNTNAP5 gene. Since her father was not severely obese, we concluded that the duplication was likely to be a rare polymorphism without clinical significance. Of interest was a region of homozygosity of 7.5 Mb on chromosome 1 containing the LEPR gene. Sanger sequencing confirmed the presence of a novel homozygous LEPR mutation, c.1604–8A>G, in intron 12. mRNA analysis showed two transcripts, one with retention of 7 nucleotides from intron 12 (r.1604–7_1604–1insTTTCTAG) and one missing exon 13 (r.1604–1752del; fig. 2, 3). Both transcripts result in a frameshift and premature stop codon (p.Lys597Serfs*34 and p.Val596Aspfs*3, respectively). The transcripts were present at very low levels, indicating rapid degradation.

Once the LEPR mutations had been identified, laboratory tests were performed to exclude an immune deficiency. Lymphocyte subsets and T-cell proliferative responses to stimulation by aCD3, IL-2/CD28, PHA/IL2 and tetanus toxoid were normal in both patients.

Discussion

These 2 girls, together carrying three novel LEPR mutations, show the classical phenotype of LEPR deficiency with severe obesity from the first year of life [3]. Patient 2 denies hyperphagia, which has been reported in all other patients, but appetite was not objectively assessed.

In patient 1, pathogenicity of the amino acid substitution variant was not confirmed by functional studies, but based on the evolutionary conservation of this amino acid and the in silico prediction, we consider it highly likely...
that this was a pathogenic mutation. RNA analysis showed that the splice-site mutation gave rise to a frameshift and premature stop codon. RNA analysis in patient 2 showed that the homozygous mutation, due to alternative splicing, resulted in two transcripts that were formed from the same pre-mRNA. Both these mRNA molecules resulted in a frameshift and premature stop codon.

The father of patient 1 is obese (BMI 37.5), as has previously been reported in some heterozygous carriers [7] but not in others [3]. He had normal weight as a child and

Fig. 2. Sanger sequencing results of the RNA of patient 1 (upper panel) and patient 2 (lower panel) compared to the wild-type sequence. Both patients show two transcripts. In patient 1, the wild type and mutant r.1753–1dupG transcript are visible, while in patient 2, the mutant transcript r.1604–7_1604–1insTTTCTAG (exons 12–13) and the mutant transcript with the skip of exon 13 (r.1604_1752del; exons 12–14) are seen. Exon numbers are indicated below the sequences. The vertical box indicates the first nucleotide of the exon. F = forward; R = reverse.
only gained weight after adolescence. The fact that obesity did not develop until adulthood may be interpreted as evidence against a causal role of the obviously congenital LEPR mutation. However, the development of obesity during adulthood was also observed by others in heterozygous subjects [8]. The heterozygous mother and sister have normal weight.

Prepubertal LEPR-deficient children were reported to be tall [5], but to the same extent as equally obese subjects without LEPR mutations [3]. However, two LEPR-deficient sisters with poor growth and low GH secretion have also been described [2]. A lack of pubertal growth spurt with reduced adult height was described by some [3], although others reported normal adult height [6]. Our patient 1 showed growth acceleration during the first 4 years, followed by steady growth within the TH range, and patient 2 did not show an evident pubertal growth spurt but continued to grow along her TH SDS to achieve a normal adult height.

Patient 1 had a remarkably advanced BA (BA/CA ratio 1.4) despite prepubertal levels of estrogens and androgens. BA was more advanced than usually seen in children of similar BMI SDS and age (BA/CA ratio 1.2 ± 0.3) [17]. A very advanced BA was previously reported in prepubertal children with leptin deficiency [11, 18–20], but a 13.8-year-old leptin-deficient girl with arrested puberty and low estradiol had a slightly retarded BA of 13.3 years [21]. Several studies also reported an advanced BA in...
some, but not all, patients with lipodystrophy, who have low or undetectable leptin levels [11, 22]. The fact that advanced BA is reported in prepubertal children with absent/reduced leptin levels or LEPR mutations suggests that absent or decreased leptin signaling favors bone maturation. Elfeteriou et al. [11] interpreted the advanced BA as an indirect sign of premature bone formation caused by the absence of leptin as an anti-osteogenic factor. Interestingly, both our patients had relatively high BMD and BMAD levels. In the first patient, tall stature and advanced BA could partially explain the high BMD. However, the second patient had an average height for age and a retarded BA, and since she was hypogonadal, her BMD was expected to be low. Increased BMD may be related to obesity [23, 24], although other studies failed to confirm these findings [17]. This raises the question whether leptin itself might directly affect bone metabolism, which is possible as LEPR is expressed in (adult) osteoblasts and chondrocytes [25].

Data from animal models provide conflicting data on the effect of leptin on bone. In mouse models, defects in leptin signaling were found to result in an increased bone mass by some [26, 27]. In contrast, others found a reduced bone mass in leptin-deficient mice [28, 29]. Different effects of leptin administration on bone have been observed, possibly due to different routes of administration, with intracerebroventricular infusion being associated with bone loss [27] and peripheral administration with increased bone formation [30]. Age may also be a determinant of the effect of leptin, as a leptin antagonist was found to have a positive effect on bone formation in young mice but no effect in older mice [31].

The role of leptin in human bone metabolism also remains elusive. In obese subjects leptin levels are positively correlated to BMD z-scores [23], although this association may not be independent but mediated through fat mass [32]. Unravelling the link between leptin and bone in obesity is complicated by the finding of a state of leptin resistance in obesity. This was first observed in animal models and is evident from the fact that elevated leptin levels do not result in a return to normal weight [33].

In women with hypoleptinemia due to strenuous exercise treatment with recombinant methionyl human leptin (metreleptin) increased the bone mineral content (BMC) of the LS but not at other sites [34]. However, the increased BMC may be mediated by the increase in estradiol levels and decrease in cortisol levels observed during metreleptin treatment [34]. A study of three leptin-deficient adults reported normal BMC at the LS in 2 females and low BMD (z-score –2.36) in 1 male, who also suffered from hypogonadism [35]. Farooqi et al. [20] described normal BMD in 3 leptin-deficient children, and BMD in these children continued to develop as expected for age and gender during metreleptin treatment, although actual z-scores were not provided. On the other hand, individuals, including children, with hypoleptinemia due to lipodystrophy were found to have an increased BMC SDS of the total body less head, also when adjusted for height, and an increased BMD at the LS and hip [22]. Treatment with recombinant leptin had no effect on BMD in individuals with lipodystrophy [22, 36].

These findings led us to hypothesize that in humans, decreased or absent leptin signaling favors an increase in bone mass. As seen within a group of individuals with lipodystrophy, BMC may not be outside the normal range in all affected subjects despite a clearly higher BMC of the group as a whole [22]. This might also explain why the 3 leptin-deficient children had a normal rather than an elevated BMD, although this remains speculative as no z-scores were provided so that it is unclear whether these children even had a BMD >0 SDS. In older individuals, other factors such as hypogonadism may explain a low BMD despite reduced or lost leptin signaling.

Leptin signaling has been suggested to affect bone metabolism directly through effects on osteoblasts and indirectly through altered activity of the (sympathetic) nervous system, GH, IGF1, parathyroid and thyroid hormones, cortisol and estrogen [12]. The finding that obese subjects with a mutation in MC4R, which acts downstream of the LEPR, also have a high BMD [37] suggests that leptin may act on proopiomelanocortin-producing neurons and then through the melanocortin 4 receptor on paraventricular neurons to influence bone metabolism. The pathway further downstream is not clear. In the patients we describe herein, no abnormalities in the serum levels of IGF1, PTH or thyroid function were observed, although this does not exclude a role of these hormones in the bone phenotype.

Patient 2 had hypogonadotropic hypogonadism, which has previously been reported in several LEPR-deficient subjects [2, 3, 6, 8]. In leptin-deficient patients with hypogonadotropic hypogonadism, a normal response to GnRH has been described by some, which was seen as evidence for the hypothalamic origin of hypogonadism [35], but others found a low response to GnRH as we did [2]. LEPR is expressed by mouse gonadotropes and may be important in the regulation of GnRH binding [38]. This suggests that hypogonadism in LEPR deficiency may be due to a defect both at the hypothalamic and at the pituitary level. However, hypogonadism may change over
time, as a low response to GnRH was reported in a 16-year-old LEPR-deficient boy who had a normal response at the age of 19 years [6]. Another case with a low response to GnRH as a teenager later had spontaneous pubertal development and a natural pregnancy [39].

In previous studies, hyperinsulinemia or insulin resistance was observed in 10/10 and 5/9 LEPR-deficient subjects studied; 2 middle-aged subjects and 1 20-year-old had type 2 diabetes mellitus [3]. Patient 2 from the current study already has impaired glucose tolerance, suggesting that progression of hyperinsulinemia to type 2 diabetes mellitus may occur at a young age. Neither of the 2 patients had immune defects in contrast to 9 previously studied LEPR-deficient children [3, 4, 7].

In conclusion, we describe 2 girls with severe obesity caused by novel LEPR mutations. The longitudinal growth data show normal childhood growth; the older girl reached normal adult height despite hypogonadotropic hypogonadism and the lack of an obvious pubertal growth spurt. BA is remarkably advanced in the younger girl, and BMI is high in both girls, which might be related to leptin resistance. With the description of more patients with LEPR deficiency, it becomes obvious that the phenotype is variable, with early-onset obesity as a common feature and hypogonadotropic hypogonadism and immune deficiency for example as more variable features. More data on bone development in LEPR-deficient subjects should help further unravel the role of leptin in human bone.

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Disclosure Statement

J.M.W. is a consultant for OPKO, Versartis, Biopartners, Merck-Serono, Teva and Ammonett. The other authors have no conflicts of interest to declare.

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